# Bioelectricity is a universal multifaced signaling cue in living organisms

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ABSTRACT The cellular electrical signals of living organisms were discovered more than a century ago and have been extensively investigated in the neuromuscular system. Neuronal depolarization and hyperpolarization are essential for our neuromuscular physiological and pathological functions. Bioelectricity is being recognized as an ancient, intrinsic, fundamental property of all living cells, and it is not limited to the neuromuscular system. Instead, emerging evidence supports a view of bioelectricity as an instructional signaling cue for fundamental cellular physiology, embryonic development, regeneration, and human diseases, including cancers. Here, we highlight the current understanding of bioelectricity and share our views on the challenges and perspectives.

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#### INTRODUCTION

Bioelectricity refers to the cellular electrical properties intrinsically caused by the uneven distributions of ions and charged molecules across cell membranes (Burr and Northrop, 1935). Usually, there is a high concentration of potassium ions and a low concentration of sodium ions in cytosol compared with the external cellular environment—the cellular resting membrane potential forms when these chemical concentrations and their electric charge gradients reach a balance. The ion channels, pumps, gap junctions, and solute carriers are the major contributors to this uneven distribution of ions and charged molecules, such as amino acids and proteins, essential for maintaining cell physiology. The types of these contributors can be variable, given one type of tissues or cells. Thus, different types of cells may possess a characteristic resting mem-

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Abbreviations used: ASAP1, Accelerated Sensor of Action Potentials 1; EV, Extracellular Vesicles; GEVI, Genetically Encoded Voltage Indicator; NADH/NAD+, Nicotinamide-adenine dinucleotide; TME, Tumor Microenvironment; TNT, Tunneling nanotubes.

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brane potential, which can be due to a unique profile of ion regulators or to its physiological history, as many of these ion regulators can open and close posttranslationally due to other events in the cell. From this perspective, the neuron is a specialized cell type with drastic electric changes. In this brief review, we focus on nonexcitable cells.

While bioelectricity is a phenomenon that is intrinsically rooted in individual cells, its influence is not limited to the cellular level (McCaig et al., 2009). Instead, it can manifest at various levels, including the organ, tissue, and even whole body (Figure. 1). This multilevel manifestation of bioelectricity plays a crucial role in various physiological processes, such as cell migration, proliferation, differentiation, and many diseases, including cancer metastasis. One important function of bioelectric signaling is to enable cellular collectives (tissue networks) to store and process information in ways that individual cells cannot. Much as neural bioelectricity in the brain underlies the emergence of a coherent organism that adaptively navigates three-dimensional space, non-neural bioelectricity allows groups of cells to traverse anatomical morphospace during embryogenesis and large-scale regeneration. Thus, bioelectricity could have diverse and far-reaching implications in biology.

## BIOELECTRICITY AND FUNDAMENTAL CELLULAR PHYSIOLOGY

The differential distributions of ions and charged molecules not only establish the cell membrane potential but also maintain cell volume. In animals, stable cell volume is maintained by pumping

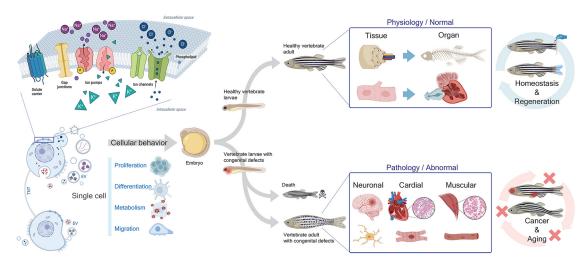


FIGURE 1: Bioelectricity is a multiple-faced signal essential from a single cell to higher levels of biological processes. Cellular bioelectric status is an overall signal with contributions from multiple ion channels, pumps, gap junctions, and solute carriers. The bioelectric sign plays essential roles in fundamental cell physiology, such as proliferation, differentiation, metabolism, and migration. Cellular bioelectricity can also be modified by EV (extracellular vesicles) and TNT (tunneling nanotube). Bioelectricity can be utilized during embryonic development as a signal for patterning and differentiation of tissues and organs. Once animals develop into adults, bioelectricity remains a critical signal for homeostasis, regeneration, and diseases such as cancers. Abnormal bioelectricity may lead to diseases such as birth defects, aging, and cancers. Some cartoons were adopted or modified from BioRendor.

Na<sup>+</sup> out and K<sup>+</sup> in through Na<sup>+</sup>/K<sup>+</sup> ATPase to counter the Donnan effect, unstable osmotic fluctuations caused by intracellular impermeant molecules from cell metabolism (Kay, 2017). Cellular shapes and sizes are critical for cell function and cell migration. Migrating cells are usually polarized and form leading-edge protrusions (e.g., lamellipodia, filopodia, and blebs). The local membrane potential and differential distribution of ion channels were found to contribute to the lamellipodial protrusion (Schwab, 2001). Indeed, the cell membrane potential also serves as a fundamental regulator of cell migration through calcium influx (voltage-gated ion channels), modulating cytoskeleton signaling such as ERK-Rho-kinase, and modulating cortical actin polymerization (Schwab et al., 2012). Various ion channels (e.g., K<sup>+</sup>, Na<sup>+</sup>, Ca<sup>2+</sup>, Cl<sup>-</sup>, TRP (Transient receptor potential) superfamily, and piezo channels) have been reported to be involved in cell migration (Schwab et al., 2012; Canales Coutino and Mayor, 2021).

Cellular bioelectric state and its related differential ion concentrations are also critical for cell metabolism. Intracellular ion concentration may directly affect key metabolic enzymes and essential regulator proteins. In addition, some solute carriers pair metabolites (e.g., glutamate) transport and specific ions (Zhang et al., 2019; Alam et al., 2023). Thus, bioelectric change inevitably influences metabolism. Furthermore, cellular membrane potential can interact with metabolic redox and energy carrier pairs (NADH/NAD+ and ADP/ATP) through membrane ATPase and membrane-bound dehydrogenases (Schofield et al., 2020).

Cell proliferation, another critical autonomous behavior, was also found to be tightly linked with cell membrane potential and ion channels (Blackiston et al., 2009). Each cell type may have a distinct bioelectric status. Generally, the cell membrane potentials of less proliferative cells are more electrically hyperpolarized, and proliferating cells are more depolarized (Blackiston et al., 2009; Abdul Kadir et al., 2018). In addition, cell membrane potentials and potassium ion concentration undergo a rhythmic change during cell cycles (Urrego et al., 2014). For example, MCF-7 mammary tumor cells were found to be depolarized toward  $G_0/G_1$ , interme-

diately hyperpolarized during the  $G_1/S$  transition, and then further hyperpolarized during the  $G_2/M$  stage (Wonderlin et al., 1995). Not only cell membrane potential but also mitochondrial membrane potential was reported to regulate cell cycle in yeasts (Gorospe et al., 2023). In addition, cell membrane potential was also found essential for bacterial cell division (Strahl and Hamoen, 2010), and bacterial biofilm formation and interbacteria communications can be mediated by bioelectric signals (Koshland, 1983; Prindle et al., 2015; Yang et al., 2020). Recently, we have observed that zebrafish embryonic cells show hyperpolarization before cytokinesis during cleavage stages (Silic et al., 2022). Alter cell membrane potential leads to cell proliferation changes, suggesting that bioelectricity may control the cell cycle, likely through the  $Ca^{2+}$  and  $K^+$  ion flux and cyclin/cyclin-dependent kinases (Whitaker, 1995; Abdul Kadir et al., 2018; Humeau et al., 2018).

Cell differentiation is generally coupled with cell proliferation and cell metabolism (Agathocleous and Harris, 2013). It is not surprising that bioelectricity also regulates cell differentiation (Sundelacruz et al., 2009). Mounting evidence revealed that the cell membrane potential is not only associated with cell differentiation but also plays an instructional role in multiple cell types, such as keratinocytes, neural cells, immune cells, and myocytes (Sundelacruz et al., 2008). For example, hyperpolarization may trigger tyrosine dephosphorylation of the Kir2.1 channel and cause an increase of myocyte transcription factors through Ca<sup>2+</sup> influx (Konig et al., 2004). Another fascinating example is that stem cells can be induced to differentiate into osteogenic or neurogenic tissues using electrical stimulation matching the endogenous membrane potentials (Zhang et al., 2023).

### **BIOELECTRICITY IN EMBRYONIC DEVELOPMENT**

Embryonic development is a robust system in which a fertilized egg proliferates, differentiates, and eventually forms an entire organism. Cell-autonomous behaviors such as proliferation and differentiation are essential. The intercell interactions (short and long ranges) are also critical for multicellular organisms to pattern and

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orient body axes, as well as pattern tissues such as bird feather systems (Jiang et al., 2021; Tseng et al., 2023) and create organs of the correct shapes and sizes (Takahashi et al., 2001; Lander, 2011). Morphogen proteins and transcription factors play crucial roles in embryogenesis (Wolpert, 1969; Pearson et al., 2005; Briscoe and Small, 2015; Negrete and Oates, 2021), and some of these are downstream of bioelectric cues (Vandenberg et al., 2011; Pai et al., 2015; Dahal et al., 2017; Belus et al., 2018). The role of bioelectricity is becoming evident in the developmental biology of multiple organisms such as Xenopus, zebrafish, mice, and fruit flies and has been extensively reviewed (Levin et al., 2017; Harris, 2021; Levin, 2021a; George and Bates, 2022; Silic and Zhang, 2023). The ectopic expression of ion channels, as well as dominant negative or hypermorphic mutations, leads to drastic morphological structure changes in animal models and human patients bearing channelopathies (Masotti et al., 2015; Adams et al., 2016; Sun et al., 2019). This finding suggests that bioelectricity may serve as organlevel or whole-body-level instructional signals. Bioelectric signals are critical for embryonic patterning that sets organ shapes and sizes, as reported in zebrafish fin-size - altered mutants caused by multiple K<sup>+</sup> channels, connexins, and solute carriers (lovine and Johnson, 2000; Iovine et al., 2005; Perathoner et al., 2014; Lanni et al., 2019; Silic et al., 2020; Daane et al., 2021; Stewart et al., 2021). In addition, changes in the bioelectric state of certain embryonic regions in a developmental stage can change the identity of that region to different organs. For example, ectopic eyes were induced via expression of a constitutively conductive EXP1 cation channel, glycine-gated chloride channel, or dominant negative Kir6.1 (Kcnj8) in Xenopus head or trunk regions (Pai et al., 2012), including gut tissues normally thought to be incompetent to acquire eye fate in vertebrates. A similar phenotype caused by multiple channels strongly supports the idea that bioelectricity, not individual ion channels, is the key to these embryonic phenotypes a conclusion also supported by the ability to swap vertebrate ion pumps for yeast pumps with no sequence or structural homology while keeping their functional effects in regeneration (Adams et al., 2007). Indeed, consistent with early studies using microprobes on Xenopus embryos, a recent investigation using the GEVI (genetically encoded voltage indicator), ASAP1, revealed characteristic voltage changes during zebrafish embryogenesis (Silic et al. 2022). Moreover, it was discovered that external electrical currents could regulate tissue size and shape in vitro (Shim et al., 2024). It is clear that bioelectricity plays a crucial instructional role in embryonic development. A few hypotheses, such as bioelectric code and integration with a morphogenic field, have been proposed as frameworks within which to understand how patterns of bioelectric state map onto anatomical outcomes (Tseng and Levin, 2013b; Levin and Martyniuk, 2018; Harris, 2021).

## BIOELECTRICITY IN TISSUE HOMEOSTASIS AND REGENERATION

Upon completion of embryonic development, tissue, and organlevel bioelectric signaling remains functional and is critical for tissue homeostasis, which is usually achieved through regeneration and tissue turnover. Turnover is a universal process of decay and regeneration of new components across biological systems (Reddien, 2024). Electrically coupled cells and collective electrical fields may serve as regenerative cues for other chemical and mechanical signals responsive to tissue integrity changes or injury (Levin, 2009; Tyler, 2017; McLaughlin and Levin, 2018). Consistent with this notion, electric currents have been recorded in the newt limb and Xenopus tadpole tail regeneration (Borgens et al., 1977b; Reid et al., 2009). Moreover, the regeneration can be modulated by external electric stimulation, V-ATPase, and voltage-gated Na<sup>+</sup> channel, Nav1.2, in tadpole tail regeneration (Borgens et al., 1977a; Adams et al., 2007; Tseng et al., 2010). In addition to newts and frogs, bioelectricity in regeneration was also reported in murids and zebrafish (Becker, 1972; Hechavarria et al., 2010; Monteiro et al., 2014; Leppik et al., 2015). Similarly, electric response and wound healing have been demonstrated in epithelial cells, and the regenerative bioelectricity has been found to be mediated by PI(3)K and redox signaling (Zhao et al., 2006; Reid and Zhao, 2014; Ferreira et al., 2016).

In addition to vertebrates, bioelectricity is evident in planarian regeneration, where it is responsible for the patterning of the anterior-posterior axis. Targeting the endogenous bioelectric circuit in regenerating planarian fragments can produce no-head and two-head animals (Beane et al., 2011; Beane et al., 2013; Durant et al., 2019). Strikingly, a temporary (as brief as 3 h) alteration of the bioelectric state gave rise to planaria, which have a permanently altered target morphology—their fragments continue to generate two heads in perpetuity with no further manipulation (Oviedo et al., 2010; Durant et al., 2017), motivating a model in which the bioelectric circuit holds anterior-posterior axis polarity as a kind of rewritable memory separate from the genetics (Pezzulo et al., 2021).

Altering the bioelectric state of the blastema by sodium ionophores was able to induce complete regeneration of tails and limbs in amphibia in normally nonregenerative conditions, acting as a brief trigger that kickstarts a complex program of changes in gene expression in anatomy (Tseng et al., 2010; Tseng and Levin, 2013a). The bioelectric role in regeneration is also being validated in other organisms. Fruit fly gut epithelial regeneration has also been reported to be bioelectric dependent on gap junction - mediated Ca<sup>2+</sup> currents triggered by neurons (Petsakou et al., 2023). Another exciting discovery is that Nav1.7 was identified as a critical chondrocyte regulator and therapeutic target for osteoarthritis, a joint degeneration disease. The Nav1.7 channel regulates the chondrocyte secretome through Ca<sup>2+</sup> signaling, and blocking this channel can facilitate the progression of osteoarthritis (Fu et al., 2024). As aging is naturally linked with tissue/organ homeostasis and regeneration, it is not a surprise that bioelectricity also plays a crucial role in aging (Silver and Nelson, 2018; Pio-Lopez and Levin, 2024).

## BIOELECTRICAL MALFUNCTIONS AND CANCER: A FAILURE OF MULTICELLULARITY

Cancer, though defined on the pathology level, is often thought of as a disease of genome decay, accompanied by genomic instability and a myriad of mutations from chromosome changes to single nucleotide mutations. So far, only a small number of genes have been defined as cancer driver genes (oncogenes or tumor suppressor genes), which are critical for cancer formation (Stratton et al., 2009). With the completion of many human cancer genomes, essentially all human genes are found mutated in some human cancer cell genomes (Ding et al., 2018; Martinez-Jimenez et al., 2020; Ostroverkhova et al., 2023). So, it is unsurprising that many ion channels were reported in cancers, especially given that more than 400 channels are in the human genome and are widely expressed in many cell types (Hutchings et al., 2019). Other models of cancer, favoring nongenetic mechanisms (Soto and Sonnenschein, 2011; Sonnenschein and Soto, 2016), likewise implicate impairments of

bioelectrical signaling as defects in the mechanisms that normally bind individual cells to a common purpose in normal tissue and organ morphostasis (Levin, 2021b).

A role for bioelectric signaling in cancer, via the role of sodium and potassium channels, is beginning to be dissected, and even some channel blockers were already in clinical trials (Ding et al., 2018; Prevarskaya et al., 2018; Martinez-Jimenez et al., 2020; Bell et al., 2023; Ostroverkhova et al., 2023). The detailed correlation evidence of ion channels in cancer has already been reviewed (Montgomery et al., 2020). As bioelectricity is an intrinsic signal and has a wide distribution of ion channels, it is undoubted that it has an essential cellular function in tumor initiation, progression, and metastasis from various human cancers (Yang and Brackenbury, 2013; Robinson et al., 2021; Sheth and Esfandiari, 2022). Glioma, a notorious brain tumor, is another of the most studied tumors for the role of bioelectricity. Glioma cells can interact with neurons through glioma synapses and gap junctions to change neuron bioelectric activities and cause disease symptoms. Recently, the in vivo glioma - neuron interactions were demonstrated in awake mice using a genetically encoded Ca<sup>2+</sup>reporter, GCaMP6f (Montgomery et al., 2020).

In animal models, bioelectric signals have been shown to induce melanoma in the absence of genetic defects or oncogene drivers (Morokuma et al., 2008; Lobikin et al., 2015), as well as to normalize tumors induced by human oncogenes such as TP53 and KRAS mutations (Chernet and Levin, 2013; Chernet et al., 2016). Recently, a neuron-like GABA-mediated bioelectrical activity between melanoma cells and keratinocytes was revealed in the zebrafish genetic melanoma model (Tagore et al., 2023). During cancer progression, cancer cells may depolarize cellular membrane potential to achieve independent cell growth and uncontrolled proliferation, and this process is tightly linked to ion channel alterations and Ca<sup>2+</sup> signals (Yang and Brackenbury, 2013). Depolarized glioma cell membrane potential was able to promote tumor proliferation, and hyperpolarization reduced the glioma progression (Venkatesh et al., 2019). Similarly, knockdown gap junction GJB2 could disrupt cancer cell nanotube projection and inhibit tumor invasiveness (Bahcheli et al., 2024). The cancer cell bioelectric changes may also influence cell differentiations and make some cancer stem-like cells (Biagiotti et al., 2006; Biella et al., 2007). The tumor microenvironment (TME) usually consists of an extracellular matrix and multiple cell types, including fibroblasts, immune cells, and endothelial cells. During cancer progression, TME profoundly impacts cancer cell bioelectricity and vice versa, along with interactions between bioelectric, biochemical, and mechanical signals (see details in next section) (Sheth and Esfandiari, 2022). Consistent with this notion, neuronal regulation in many types of cancer has recently been recognized since the bioelectric contributions from neurons can be relatively measured (Gillespie and Monje, 2020; Winkler et al., 2023).

Bioelectricity is also crucial for metastasis due to cancer cell bioelectric status and cancer microenvironment. (Payne et al., 2019). It is well known that ion channel distribution and related cell membrane potential are key regulators for cell migration (Schwab et al., 2012). Cancer cells may co-opt similar mechanisms for metastasis (Zhang et al., 2024). Many studies suggested that depolarized cancer cells may tend to be more metastatic (Ribeiro et al., 2020). On the tissue or organ level, bioelectricity may coordinate the metastasis from invasion to colonization through the TME and blood or lymphatic vessel environments. Collective migration and electric field were also frequently reported in cancers (Li et al., 2013; Garg et al., 2019; Zhu et al., 2020). Thus, ion channel blockers/agonists

and external electric fields may interrupt cancer development. Indeed, ion channels' function and potential as therapeutic targets are currently being investigated (Takayasu *et al.*, 2020; Abed *et al.*, 2023). In addition, electrotherapy and external electric devices have already been clinically applied to glioma based on the principle of bioelectricity (Rominiyi *et al.*, 2021).

#### CHALLENGES, NEW PROGRESS, AND PERSPECTIVES

One of the challenges is that the bioelectricity concept is not well accepted or even well known in many research areas. A prominent contributing factor is the misconception that bioelectricity is exclusively relevant in neuroscience, not nonexcitable tissues (Levin, 2021a). But this is gradually changing as more scientific discoveries are revealed and published. The bioelectricity subgroup in the Cell Bio 2023 is a great advance, at least to the cell biology research community (Pai et al., 2024). The special topic journal, like Bioelectricity, is an excellent move and a landmark for the bioelectricity field. The second barrier is the largely unrevealed connection with the current biomedical research regime centered on biochemistry and genetics. Genetics alone does not determine phenotypes entirely because the bioelectrical layer of control provides a kind of physiological "software" that provides extra plasticity to the genetically encoded hardware. For example, bioelectric status changes can induce persistent alternative anatomical structures after regeneration and even ones associated with different species than the genomic default (Sullivan et al., 2016; Levin, 2021a). Likewise, genetic state and phenotype diverge in animals bearing oncogenic mutation but no tumor due to bioelectric normalization.

Biochemical molecules are deemed as the mainstream signaling carriers (Lander, 2011). Bioelectricity is different from traditional biochemical signals in many ways. The bioelectric signals generally serve as overall body patterning instruction and coordinate and program overall body anatomy (Levin, 2021a). Essentially, the bioelectric control could be an epigenetic mechanism that guides morphogenesis usually through a robust and dynamic status (i.e., cellular voltage memory) (Levin et al., 2017; Levin and Martyniuk, 2018; Levin, 2021a). Furthermore, there is a lack of a 1:1 relationship between bioelectric outcome and gene products. This is because bioelectricity is an overall readout of multiple contributors (channels, gap junctions, etc.) and multiple downstream output routes to biochemical machinery on the cellular level (Levin, 2014, 2021a). Bioelectric patterns were found to impact important morphogen proteins such as BMP, SHH, and FGFs, which are critical for embryonic development, adult tissue, and organ regeneration. For example, BMP signaling was found to be regulated by KCNJ2-mediated bioelectricity in mice and fruit flies (George and Bates, 2022). Cell membrane potential stabilizes SMO and regulates hedgehog (HH) signaling in fruit fly wing discs (Emmons-Bell and Hariharan, 2021; Spannl et al., 2021). The Na<sup>+</sup> and K<sup>+</sup> gradients are essential for SHH secretion and SMO activation (Petrov et al., 2020). Likely due to the nature of secondary messenger, calcium signaling is often reported as a common downstream pathway. The calcium signal is similar to cellular voltage change during zebrafish embryos (Chen et al., 2017; Silic et al., 2022). In addition to calcineurin and PI(3)K pathways, Ca<sup>2+</sup> flux can modulate bioelectricity and tune the conductivity of ion channels that directly contribute to cell bioelectricity and excitability (Clapham, 2007). It is hard to distinguish the bioelectricity from the Ca<sup>2+</sup> signaling, but additional transduction mechanisms, such as serotonin movement (Fukumoto et al., 2005) and K-ras clustering (Zhou et al., 2015),

exist for converting bioelectric information into downstream transcriptional readouts.

In addition to biochemical interactions, bioelectricity is also tightly linked with mechanical transduction, which is involved in various physiological (e.g., touching, hearing, angiogenesis, development, and regeneration) and pathological processes (e.g., cardiac hypertrophy, muscular dystrophy, and polycystic kidney) (Martinac, 2014). Some ion channels, such as TRP, PIEZO, K2P, and ENaC (epithelial sodium channel), can directly sense cellular mechanical forces and convert mechanical changes into an electric signal or Ca<sup>2+</sup> flux (Martinac, 2014; Jin et al., 2020). Inversely, PIEZO channels were also found to be gated by voltage (Moroni et al., 2018). The extracellular matrix proteins can also directly influence ion channel conductivities on the cell membrane. For example, chondroitin sulfates were reported to modify voltage-dependent ion channels (Vigetti et al., 2008). Similarly, extracellular matrix proteins can also impact connexin functions (Imbeault et al., 2009). Thus, these mechanical, bioelectric, and biochemical signals can interplay and coordinately regulate complex biological processes such as stem cell differentiation and cancers (Engler et al., 2006; Discher et al., 2009; Karska et al., 2023).

Another challenge of bioelectricity research is the still limited available experimental approaches. The overall cellular bioelectric status is an integration from multiple inputs of ion channels, pumps, gap junctions, and solute carriers. Thus, the bioelectric status is robust and dynamic and is usually a component of physiological feedback, requiring the development of bioelectric simulator platforms (Pietak and Levin, 2016), and more broadly, new conceptual frameworks borrowed from multiscale neuroscience (Pezzulo and Levin, 2015). Genetic mutation of one of these contributors is generally insufficient to cause a phenotype unless the mutation is dominant negative or hypermorphic (Silic and Zhang, 2023). This phenomenon was noticed as bioelectricity does not reduce to molecular genetics (Levin, 2014). Thus, identifying a critical endogenous contributor is hard since forward genetic screening is less effective. As bioelectricity is highly dynamic, measuring in vivo cellular bioelectric status is also challenging using old-fashioned chemical dyes or patch clamp techniques. Fortunately, newly developed tools such as optical probes, GEVIs, optogenetics, and chemogenetics can readily be adapted to bioelectricity research using well-established model organisms such as zebrafish (Foust et al., 2023; Silic and Zhang, 2023). However, nonoptical reporter technologies will be needed to detect bioelectric states in deep tissues. Another exciting area of progress involves electroceuticals (Balasubramanian et al., 2024), including pharmacological and optogenetic modalities that target tissue native bioelectric networks instead of a specific pathway (Adams et al., 2013; Chernet et al., 2016; Pio-Lopez and Levin, 2023).

In addition to challenges, some new frontiers of cell biology related to bioelectricity have emerged. Extracellular vesicle (EV) is one of them. EVs are cell-derived membrane-enclosed particles found in essentially all cell types. Currently, EVs are known as a universal way of intracellular communication as they can deliver soluble mediators, such as proteins (cytokines, morphogens, and microRNAs, to the direct contact of neighboring cells. EVs can be classified into exosomes or ectosomes based on their biogenesis, and they have variable sizes (Zappulli et al., 2016; Salomon et al., 2022; van de Wakker et al., 2023). The EVs may directly distribute ion channels such as sodium channels, glutamate receptor subunits, solute transporters, connexin, and aquaporins (Faure et al., 2006; Soares et al., 2015; Pathare et al., 2017; Barros Lamus et al., 2021; Nouri et al., 2021). Thus, they may modulate cellular bioelectricity. On the other hand, bioelectricity could also regulate EV formation and release. P2X7R, an ATP-gated ion channel, was found in controlling EV release (Golia et al., 2023). Like EV, the cell membrane tunneling nanotubes (TNTs) are another type of intercellular communication for diverse contents from ions and proteins to organelles. However, the TNT working range (physical distance between cells) is longer than EVs. TNTs have an actin structure and portions of the plasma membrane between two cells and facilitate cytoplasm and organelle exchange (Cervantes and Zurzolo, 2021; Zurzolo, 2021; Wang et al., 2024). TNTs are also related to cellular bioelectricity since connexin and Ca<sup>2+</sup> flux were found to be generated and propagated within TNTs (Smith et al., 2011; Vargas et al., 2019). Moreover, cells connected by TNT can be electrically coupled through connexins (Wang et al., 2010). This TNT-mediated electrical coupling could modulate intracellular downstream signaling molecules such as PI3K (Abounit and Zurzolo, 2012). Thus, both EVs and TNTs expand the horizon of bioelectricity.

In conclusion, bioelectricity is a universal, multifaceted signaling modality for regulating autonomous collective cell behavior, embryonic development, regeneration, and diseases such as cancers. Although there are some challenges, significant progress has been made in recent years. With newly developed tools, we expect bioelectricity to find its way into cell, developmental, and evolutionary biology, as well as many other biomedical research areas.

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### **REFERENCES**

Abdul Kadir L, Stacey M, Barrett-Jolley R (2018). Emerging roles of the membrane potential: Action beyond the action potential. Front Physiol 9,

Abed T, Ganser K, Eckert F, Stransky N, Huber SM (2023). Ion channels as molecular targets of glioblastoma electrotherapy. Front Cell Neurosci

Abounit S, Zurzolo C (2012). Wiring through tunneling nanotubes - from electrical signals to organelle transfer. J Cell Sci 125, 1089-1098.

Adams DS, Masi A, Levin M (2007). H<sup>+</sup> pump-dependent changes in membrane voltage are an early mechanism necessary and sufficient to induce Xenopus tail regeneration. Development 134, 1323-1335.

Adams DS, Tseng AS, Levin M (2013). Light-activation of the Archaerhodopsin H(+)-pump reverses age-dependent loss of vertebrate regeneration: sparking system-level controls in vivo. Biol Open 2,

Adams DS, Uzel SG, Akagi J, Wlodkowic D, Andreeva V, Yelick PC, Devitt-Lee A, Pare JF, Levin M (2016). Bioelectric signaling via potassium channels: a mechanism for craniofacial dysmorphogenesis in KCNJ2associated Andersen-Tawil syndrome. J Physiol 594, 3245-3270.

Agathocleous M, Harris WA (2013). Metabolism in physiological cell proliferation and differentiation. Trends Cell Biol 23, 484-492.

Alam S, Doherty E, Ortega-Prieto P, Arizanova J, Fets L (2023). Membrane transporters in cell physiology, cancer metabolism and drug response. Dis Model Mech 16, dmm050404.

Bahcheli AT, Min HK, Bayati M, Zhao H, Fortuna A, Dong W, Dzneladze I, Chan J, Chen X, Guevara-Hoyer K, et al. (2024). Pan-cancer ion transport signature reveals functional regulators of glioblastoma aggression. EMBO J 43, 196-224.

Balasubramanian S, Weston DA, Levin M, Davidian DC (2024). Electroceuticals: Emerging applications beyond the nervous system and excitable tissues. Trends Pharmacol Sci 45, 391-394.

- Beane WS, Morokuma J, Adams DS, Levin M (2011). A chemical genetics approach reveals H,K-ATPase-mediated membrane voltage is required for planarian head regeneration. Chem Biol 18, 77–89.
- Beane WS, Morokuma J, Lemire JM, Levin M (2013). Bioelectric signaling regulates head and organ size during planarian regeneration. Development 140, 313–322.
- Becker RO (1972). Stimulation of partial limb regeneration in rats. Nature 235, 109–111.
- Bell DC, Leanza L, Gentile S, Sauter DR (2023). News and views on ion channels in cancer: Is cancer a channel opathy? Front Pharmacol 14, 1258933.
- Belus MT, Rogers MA, Elzubeir A, Josey M, Rose S, Andreeva V, Yelick PC, Bates EA (2018). Kir2.1 is important for efficient BMP signaling in mammalian face development. Dev Biol 444 Suppl 1, S297–S307.
- Biagiotti T, D'Amico M, Marzi I, Gennaro P, Arcangeli A, Wanke E, Olivotto M (2006). Cell renewing in neuroblastoma: electrophysiological and immunocytochemical characterization of stem cells and derivatives. Stem Cells 24, 443–453.
- Biella G, Di Febo F, Goffredo D, Moiana A, Taglietti V, Conti L, Cattaneo E, Toselli M (2007). Differentiating embryonic stem-derived neural stem cells show a maturation-dependent pattern of voltage-gated sodium current expression and graded action potentials. Neuroscience 149, 38–52.
- Blackiston DJ, McLaughlin KA, Levin M (2009). Bioelectric controls of cell proliferation: ion channels, membrane voltage and the cell cycle. Cell Cycle 8, 3527–3536.
- Borgens RB, Vanable JW, Jr., Jaffe LF (1977a). Bioelectricity and regeneration. I. Initiation of frog limb regeneration by minute currents. J Exp Zool 200, 403–416
- Borgens RB, Vanable JW, Jr., Jaffe LF (1977b). Bioelectricity and regeneration: Large currents leave the stumps of regenerating newt limbs. Proc Natl Acad Sci U S A 74, 4528–4532.
- Briscoe J, Small S (2015). Morphogen rules: design principles of gradient-mediated embryo patterning. Development 142, 3996–4009.
- Burr HS, Northrop FS (1935). The electro-dynamic theory of life. Q Rev Biol 10, 322–333.
- Canales BC, Mayor R (2021). Mechanosensitive ion channels in cell migration. Cells Dev 166, 203683.
- Cervantes DC, Zurzolo C (2021). Peering into tunneling nanotubes-the path forward. EMBO J 40, e105789.
- Chen J, Xia L, Bruchas MR, Solnica-Krezel L (2017). Imaging early embryonic calcium activity with GCaMP6s transgenic zebrafish. Dev Biol 430, 385–
- Chernet BT, Adams DS, Lobikin M, Levin M (2016). Use of genetically encoded, light-gated ion translocators to control tumorigenesis. Oncotarget 7, 19575–19588.
- Chernet BT, Levin M (2013). Transmembrane voltage potential is an essential cellular parameter for the detection and control of tumor development in a Xenopus model. Dis Model Mech 6, 595–607.
- Clapham DE (2007). Calcium signaling. Cell 131, 1047-1058.
- Daane JM, Blum N, Lanni J, Boldt H, Iovine MK, Higdon CW, Johnson SL, Lovejoy NR, Harris MP (2021). Modulation of bioelectric cues in the evolution of flying fishes. Curr Biol 31, 5052–5061.e8.
- Dahal GR, Pradhan SJ, Bates EA (2017). Inwardly rectifying potassium channels influence *Drosophila* wing morphogenesis by regulating Dpp release. Development 144, 2771–2783.
- Ding L, Bailey MH, Porta-Pardo E, Thorsson V, Colaprico A, Bertrand D, Gibbs DL, Weerasinghe A, Huang KL, Tokheim C, et al. (2018). Perspective on oncogenic processes at the end of the beginning of cancer genomics. Cell 173, 305–320.
- Discher DE, Mooney DJ, Zandstra PW (2009). Growth factors, matrices, and forces combine and control stem cells. Science 324, 1673–1677.
- Durant F, Bischof J, Fields C, Morokuma J, LaPalme J, Hoi A, Levin M (2019). The role of early bioelectric signals in the regeneration of planarian anterior/posterior polarity. Biophys J 116, 948–961.
- Durant F, Morokuma J, Fields C, Williams K, Adams DS, Levin M (2017). Long-term, stochastic editing of regenerative anatomy via targeting endogenous bioelectric gradients. Biophys J 112, 2231–2243.
- Emmons-Bell M, Hariharan IK (2021). Membrane potential regulates hedgehog signaling in the *Drosophila* wing imaginal disc. EMBO Rep 22, e51861.
- Engler AJ, Sen S, Sweeney HL, Discher DE (2006). Matrix elasticity directs stem cell lineage specification. Cell 126, 677–689.
- Faure J, Lachenal G, Court M, Hirrlinger J, Chatellard-Causse C, Blot B, Grange J, Schoehn G, Goldberg Y, Boyer V, et al. (2006). Exosomes are released by cultured cortical neurones. Mol Cell Neurosci 31, 642– 648.

- Ferreira F, Luxardi G, Reid B, Zhao M (2016). Early bioelectric activities mediate redox-modulated regeneration. Development 143, 4582–4594.
- Foust AJ, Quicke PE, Sun Y, Bakal C, Djamgoz M (2023). Voltage-sensitive optical probes for measuring cell membrane potentials: An update and applications to "nonexcitable" cells. Bioelectricity 5, 250–265.
- Fu W, Vasylyev D, Bi Y, Zhang M, Sun G, Khleborodova A, Huang G, Zhao L, Zhou R, Li Y, et al. (2024). Na(v)1.7 as a chondrocyte regulator and therapeutic target for osteoarthritis. Nature 625, 557–565.
- Fukumoto T, Kema I, Levin M (2005). Serotonin signaling is a very early step in patterning of the left-right axis in chick and frog embryos. Curr Biol 15, 794–803.
- Garg AA, Jones TH, Moss SM, Mishra S, Kaul K, Ahirwar DK, Ferree J, Kumar P, Subramaniam D, Ganju RK, et al. (2019). Electromagnetic fields alter the motility of metastatic breast cancer cells. Commun Biol 2, 303.
- George LF, Bates EA (2022). Mechanisms underlying influence of bioelectricity in development. Front Cell Dev Biol 10, 772230.
- Gillespie S, Monje M (2020). The neural regulation of cancer. Annu Rev Canc Biol 4, 371–390.
- Golia MT, Gabrielli M, Verderio C (2023). P2X(7) receptor and extracellular vesicle release. Int J Mol Sci 24, 9805.
- Gorospe CM, Carvalho G, Curbelo AH, Marchhart L, Mendes IC, Niedzwiecka K, Wanrooij PH (2023). Mitochondrial membrane potential acts as a retrograde signal to regulate cell cycle progression. Life Sci Alliance 6, e202302091.
- Harris MP (2021). Bioelectric signaling as a unique regulator of development and regeneration. Development 148, 180794.
- Hechavarria D, Dewilde AH, Braunhut SJ, Levin M, Kaplan DL (2010). BioDome regenerative sleeve for biochemical and biophysical stimulation of tissue regeneration. Med Eng Phys 32, 1065–1073.
- Humeau J, Bravo-San Pedro JM, Vitale I, Nunez L, Villalobos C, Kroemer G, Senovilla L (2018). Calcium signaling and cell cycle: Progression or death. Cell Calcium 70, 3–15.
- Hutchings CJ, Colussi P, Clark TG (2019). Ion channels as therapeutic antibody targets. MAbs 11, 265–296.
- Imbeault S, Gauvin LG, Toeg HD, Pettit A, Sorbara CD, Migahed L, DesRoches R, Menzies AS, Nishii K, Paul DL, et al. (2009). The extracellular matrix controls gap junction protein expression and function in postnatal hippocampal neural progenitor cells. BMC Neurosci 10, 13.
- Iovine MK, Higgins EP, Hindes A, Coblitz B, Johnson SL (2005). Mutations in connexin43 (GJA1) perturb bone growth in zebrafish fins. Dev Biol 278, 208–219.
- lovine MK, Johnson SL (2000). Genetic analysis of isometric growth control mechanisms in the zebrafish caudal Fin. Genetics 155, 1321–1329.
- Jiang TX, Li A, Lin CM, Chiu C, Cho JH, Reid B, Zhao M, Chow RH, Widelitz RB, Chuong CM (2021). Global feather orientations changed by electric current. iScience 24, 102671.
- Jin P, Jan LY, Jan YN (2020). Mechanosensitive ion channels: Structural features relevant to mechanotransduction mechanisms. Annu Rev Neurosci 43, 207–229.
- Karska J, Kowalski S, Saczko J, Moisescu MG, Kulbacka J (2023). Mechanosensitive ion channels and their role in cancer cells. Membranes (Basel) 13, 167.
- Kay AR (2017). How cells can control their size by pumping ions. Front Cell Dev Biol 5, 41.
- Konig S, Hinard V, Arnaudeau S, Holzer N, Potter G, Bader CR, Bernheim L (2004). Membrane hyperpolarization triggers myogenin and myocyte enhancer factor-2 expression during human myoblast differentiation. J Biol Chem 279, 28187–28196.
- Koshland DE (1983). The bacterium as a model neuron. Trends Neurosci 6, 133–137.
- Lamus ER, Carotti V, Vries CR, Witsel F, Arntz OJ, Loo FA, Carvajal CA, Bindels RJ, Hoenderop JG, Rigalli JP (2021). Extracellular vesicles regulate purinergic signaling and epithelial sodium channel expression in renal collecting duct cells. FASEB J 35, e21506.
- Lander AD (2011). Pattern, growth, and control. Cell 144, 955–969.
- Lanni JS, Peal D, Ekstrom L, Chen H, Stanclift C, Bowen ME, Mercado A, Gamba G, Kahle KT, Harris MP (2019). Integrated K+ channel and K+Clcotransporter functions are required for the coordination of size and proportion during development. Dev Biol 456, 164–178.
- Leppik LP, Froemel D, Slavici A, Ovadia ZN, Hudak L, Henrich D, Marzi I, Barker JH (2015). Effects of electrical stimulation on rat limb regeneration, a new look at an old model. Sci Rep 5, 8353.
- Levin M (2009). Bioelectric mechanisms in regeneration: Unique aspects and future perspectives. Semin Cell Dev Biol 20, 543–556.

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- Levin M (2014). Molecular bioelectricity: How endogenous voltage potentials control cell behavior and instruct pattern regulation in vivo. Mol Biol Cell 25, 3835–3850.
- Levin M (2021a). Bioelectric signaling: Reprogrammable circuits underlying embryogenesis, regeneration, and cancer. Cell 184, 1971–1989.
- Levin M (2021b). Bioelectrical approaches to cancer as a problem of the scaling of the cellular self. Prog Biophys Mol Biol 165, 102–113.
- Levin M, Martyniuk CJ (2018). The bioelectric code: An ancient computational medium for dynamic control of growth and form. Biosystems 164, 76–93.
- Levin M, Pezzulo G, Finkelstein JM (2017). Endogenous bioelectric signaling networks: Exploiting voltage gradients for control of growth and form. Annu Rev Biomed Eng 19, 353–387.
- Li L, He Y, Zhao M, Jiang J (2013). Collective cell migration: Implications for wound healing and cancer invasion. Burns Trauma 1, 21–26.
- Lobikin M, Lobo D, Blackiston DJ, Martyniuk CJ, Tkachenko E, Levin M (2015). Serotonergic regulation of melanocyte conversion: A bioelectrically regulated network for stochastic all-or-none hyperpigmentation. Sci Signal 8, ra99.
- Martinac B (2014). The ion channels to cytoskeleton connection as potential mechanism of mechanosensitivity. Biochim Biophys Acta 1838, 682–691.
- Martinez-Jimenez F, Muinos F, Sentis I, Deu-Pons J, Reyes-Salazar I, Arnedo-Pac C, Mularoni L, Pich O, Bonet J, Kranas H, et al. (2020). A compendium of mutational cancer driver genes. Nat Rev Cancer 20, 555–572.
- Masotti A, Uva P, Davis-Keppen L, Basel-Vanagaite L, Cohen L, Pisaneschi E, Celluzzi A, Bencivenga P, Fang M, Tian M, et al. (2015). Keppen-Lubinsky syndrome is caused by mutations in the inwardly rectifying K(+) channel encoded by KCNJ6. Am J Hum Genet 96, 295–300.
- McCaig CD, Song B, Rajnicek AM (2009). Electrical dimensions in cell science. J Cell Sci 122, 4267–4276.
- McLaughlin KA, Levin M (2018). Bioelectric signaling in regeneration: Mechanisms of ionic controls of growth and form. Dev Biol 433, 177–189.
- Monteiro J, Aires R, Becker JD, Jacinto A, Certal AC, Rodríguez-León J (2014). V-ATPase proton pumping activity is required for adult zebrafish appendage regeneration. PLoS One 9, e92594.
- Montgomery MK, Kim SH, Dovas A, Zhao HT, Goldberg AR, Xu W, Yagielski AJ, Cambareri MK, Patel KB, Mela A, et al. (2020). Glioma-induced alterations in neuronal activity and neurovascular coupling during disease progression. Cell Rep 31, 107500.
- Morokuma J, Blackiston D, Adams DS, Seebohm G, Trimmer B, Levin M (2008). Modulation of potassium channel function confers a hyperproliferative invasive phenotype on embryonic stem cells. Proc Natl Acad Sci U S A 105, 16608–16613.
- Moroni M, Servin-Vences MR, Fleischer R, Sanchez-Carranza O, Lewin GR (2018). Voltage gating of mechanosensitive PIEZO channels. Nat Commun 9, 1096.
- Negrete J, Jr., Oates AC. (2021). Towards a physical understanding of developmental patterning. Nat Rev Genet 22, 518–531.
- Nouri MZ, Yu L, Liu LP, Chacko KM, Denslow ND, LaDisa JF, Jr., Alli AA (2021). Increased endothelial sodium channel activity by extracellular vesicles in human aortic endothelial cells: putative role of MLP1 and bioactive lipids. Am J Physiol Cell Physiol 321, C535–C548.
- Ostroverkhova D, Przytycka TM, Panchenko AR (2023). Cancer driver mutations: Predictions and reality. Trends Mol Med 29, 554–566.
- Oviedo NJ, Morokuma J, Walentek P, Kema IP, Gu MB, Ahn JM, Hwang JS, Gojobori T, Levin M (2010). Long-range neural and gap junction protein-mediated cues control polarity during planarian regeneration. Dev Biol 339, 188–199.
- Pai VP, Aw S, Shomrat T, Lemire JM, Levin M (2012). Transmembrane voltage potential controls embryonic eye patterning in Xenopus laevis. Development 139, 313–323.
- Pai VP, Lemire JM, Pare JF, Lin G, Chen Y, Levin M (2015). Endogenous gradients of resting potential instructively pattern embryonic neural tissue via notch signaling and regulation of proliferation. J Neurosci 35, 4366–4385.
- Pai VP, Zhang G, Levin M (2024). "Bioelectricity in development, regeneration, and cancers" cell bio 2023: A joint meeting of the American society of cell biology and European Molecular Biology Organization December 2–6, 2023, in Boston, MA, USA. Bioelectricity 6, 65–68.
- Pathare G, Tutakhel OA, van der Wel MC, Shelton LM, Deinum J, Lenders JW, Hoenderop JG, Bindels RJM (2017). Hydrochlorothiazide treatment increases the abundance of the NaCl cotransporter in urinary extracellular vesicles of essential hypertensive patients. Am J Physiol Renal Physiol 312, F1063–F1072.

- Payne SL, Levin M, Oudin MJ (2019). Bioelectric control of metastasis in solid tumors. Bioelectricity 1, 114–130.
- Pearson JC, Lemons D, McGinnis W (2005). Modulating Hox gene functions during animal body patterning. Nat Rev Genet 6, 893–904.
- Perathoner S, Daane JM, Henrion U, Seebohm G, Higdon CW, Johnson SL, Nüsslein-Volhard C, Harris MP (2014). Bioelectric signaling regulates size in zebrafish fins. PLoS Genet 10, e1004080.
- Petrov K, Wierbowski BM, Liu J, Salic A (2020). Distinct cation gradients power cholesterol transport at different key points in the Hedgehog signaling pathway. Dev Cell 55, 314–327.e7.
- Petsakou A, Liu Y, Liu Y, Comjean A, Hu Y, Perrimon N (2023). Cholinergic neurons trigger epithelial Ca<sup>2+</sup> currents to heal the gut. Nature 623, 122– 131
- Pezzulo G, LaPalme J, Durant F, Levin M (2021). Bistability of somatic pattern memories: Stochastic outcomes in bioelectric circuits underlying regeneration. Philos Trans R Soc Lond B Biol Sci 376, 20190765.
- Pezzulo G, Levin M (2015). Re-membering the body: applications of computational neuroscience to the top-down control of regeneration of limbs and other complex organs. Integr Biol (Camb) 7, 1487–1517.
- Pietak A, Levin M (2016). Exploring instructive physiological signaling with the bioelectric tissue simulation engine (BETSE). Front Bioeng Biotechnol 4, 55.
- Pio-Lopez L, Levin M (2023). Morphoceuticals: Perspectives for discovery of drugs targeting anatomical control mechanisms in regenerative medicine, cancer and aging. Drug Discov Today 28, 103585.
- Pio-Lopez L, Levin M (2024). Aging as a loss of morphostatic information: A developmental bioelectricity perspective. Ageing Res Rev 97, 102310.
- Prevarskaya N, Skryma R, Shuba Y (2018). Ion channels in cancer: Are cancer hallmarks oncochannelopathies? Physiol Rev 98, 559–621.
- Prindle A, Liu J, Asally M, Ly S, Garcia-Ojalvo J, Suel GM (2015). Ion channels enable electrical communication in bacterial communities. Nature 527, 59–63.
- Reddien PW (2024). The purpose and ubiquity of turnover. Cell 187, 2657–2681.
- Reid B, Song B, Zhao M (2009). Electric currents in Xenopus tadpole tail regeneration. Dev Biol 335, 198–207.
- Reid B, Zhao M (2014). The electrical response to injury: Molecular mechanisms and wound healing. Adv Wound Care (New Rochelle) 3, 184–201.
- Ribeiro M, Elghajiji A, Fraser SP, Burke ZD, Tosh D, Djamgoz MBA, Rocha PRF (2020). Human breast cancer cells demonstrate electrical excitability. Front Neurosci 14, 404.
- Robinson AJ, Jain A, Sherman HG, Hague RJM, Rahman R, Sanjuan-Alberte P, Rawson FJ (2021). Toward hijacking bioelectricity in cancer to develop new bioelectronic medicine. Adv Ther 4, 2000248.
- Rominiyi O, Vanderlinden A, Clenton SJ, Bridgewater C, Al-Tamimi Y, Collis SJ (2021). Tumour treating fields therapy for glioblastoma: current advances and future directions. Br J Cancer 124, 697–709.
- Salomon C, Das S, Erdbrügger U, Kalluri R, Lim SK, Olefsky JM, Rice GE, Sahoo S, Tao WA, Vader P, et al. (2022). Extracellular vesicles and their emerging roles as cellular messengers in endocrinology: An endocrine society scientific statement. Endocr Rev 43, 441–468.
- Schofield Z, Meloni GN, Tran P, Zerfass C, Sena G, Hayashi Y, Grant M, Contera SA, Minteer SD, Kim M, et al. (2020). Bioelectrical understanding and engineering of cell biology. J R Soc Interface 17, 20200013.
- Schwab A (2001). Function and spatial distribution of ion channels and transporters in cell migration. Am J of Physiol 280, F739–747.
- Schwab A, Fabian A, Hanley PJ, Stock C (2012). Role of ion channels and transporters in cell migration. Physiol Rev 92, 1865–1913.
- Sheth M, Esfandiari L (2022). Bioelectric dysregulation in cancer initiation, promotion, and progression. Front Oncol 12, 846917.
- Shim G, Breinyn IB, Martinez-Calvo A, Rao S, Cohen DJ (2024). Bioelectric stimulation controls tissue shape and size. Nat Commun 15, 2938.
- Silic MR, Dong Z, Chen Y, Kimbrough A, Zhang G (2022). Zebrafish embryos display characteristic bioelectric signals during early development. Cells 11. 3586.
- Silic MR, Wu Q, Kim BH, Golling G, Chen KH, Freitas R, Chubykin AA, Mittal SK, Zhang G (2020). Potassium channel-associated bioelectricity of the dermomyotome determines fin patterning in zebrafish. Genetics 215, 1067–1084.
- Silic MR, Zhang G (2023). Bioelectricity in developmental patterning and size control: Evidence and genetically encoded tools in the zebrafish model. Cells 12, 1148.
- Silver BB, Nelson CM (2018). The bioelectric code: Reprogramming cancer and aging from the interface of mechanical and chemical microenvironments. Front Cell Dev Biol 6, 21.

- Smith IF, Shuai J, Parker I (2011). Active generation and propagation of Ca<sup>2+</sup> signals within tunneling membrane nanotubes. Biophys J 100, L37–L39.
- Soares AR, Martins-Marques T, Ribeiro-Rodrigues T, Ferreira JV, Catarino S, Pinho MJ, Zuzarte M, Anjo SI, Manadas B, Sluijter JPG, et al. (2015). Gap junctional protein Cx43 is involved in the communication between extracellular vesicles and mammalian cells (vol 5, 13243, 2015). Sci Rep 5, 13243.
- Sonnenschein C, Soto AM (2016). Carcinogenesis explained within the context of a theory of organisms. Prog Biophys Mol Biol 122, 70–76.
- Soto AM, Sonnenschein C (2011). The tissue organization field theory of cancer: a testable replacement for the somatic mutation theory. Bioessays 33, 332–340.
- Spannl S, Buhl T, Nellas I, Zeidan SA, Venkatesan Iyer K, Khaliullina H, Schultz C, Nadler A, Dye NA, Eaton S (2021). Glycolysis regulates Hedgehog signaling via the plasma membrane potential. EMBO J 40, e107925
- Stewart S, Le Bleu HK, Yette GA, Henner AL, Robbins AE, Braunstein JA, Stankunas K (2021). longfin causes cis-ectopic expression of the kcnh2a ether-a-go-go K+ channel to autonomously prolong fin outgrowth. Development 148, 199384.
- Strahl H, Hamoen LW (2010). Membrane potential is important for bacterial cell division. Proc Natl Acad Sci U S A 107, 12281–12286.
- Stratton MR, Campbell PJ, Futreal PA (2009). The cancer genome. Nature 458, 719–724.
- Sullivan KG, Emmons-Bell M, Levin M (2016). Physiological inputs regulate species-specific anatomy during embryogenesis and regeneration. Commun Integr Biol 9, e1192733.
- Sun AX, Yuan Q, Fukuda M, Yu W, Yan H, Lim GGY, Nai MH, D'Agostino GA, Tran HD, Itahana Y, et al. (2019). Potassium channel dysfunction in human neuronal models of Angelman syndrome. Science 366, 1486–1492.
- Sundelacruz S, Levin M, Kaplan DL (2008). Membrane potential controls adipogenic and osteogenic differentiation of mesenchymal stem cells. PLoS One 3, e3737.
- Sundelacruz S, Levin M, Kaplan DL (2009). Role of membrane potential in the regulation of cell proliferation and differentiation. Stem Cell Rev Rep 5, 231–246
- Tagore M, Hergenreder E, Perlee SC, Cruz NM, Menocal L, Suresh S, Chan E, Baron M, Melendez S, Dave A, et al. (2023). GABA regulates electrical activity and tumor initiation in melanoma. Cancer Discov 13, 2270–2291.
- Takahashi Y, Osumi N, Patel NH (2001). Body patterning. Proc Natl Acad Sci U S A 98, 12338–12339.
- Takayasu T, Kurisu K, Esquenazi Y, Ballester LY (2020). Ion channels and their role in the pathophysiology of gliomas. Mol Cancer Ther 19, 1959–1969.
- Tseng A, Levin M (2013a). Cracking the bioelectric code: Probing endogenous ionic controls of pattern formation. Commun Integ Biol 6, 1–8.
- Tseng A, Levin M (2013b). Cracking the bioelectric code: Probing endogenous ionic controls of pattern formation. Commun Integr Biol 6, e22595.
- Tseng AS, Beane WS, Lemire JM, Masi A, Levin M (2010). Induction of vertebrate regeneration by a transient sodium current. J Neurosci 30, 13192– 13200
- Tseng CC, Woolley TE, Jiang TX, Wu P, Maini PK, Widelitz RB, Chuong CM (2023). Gap junctions in Turing-type periodic feather pattern formation. bioRxiv.
- Tyler SEB (2017). Nature's electric potential: A systematic review of the role of bioelectricity in wound healing and regenerative processes in animals, humans, and plants. Front Physiol 8, 627.
- Urrego D, Tomczak AP, Zahed F, Stuhmer W, Pardo LA (2014). Potassium channels in cell cycle and cell proliferation. Philos Trans R Soc Lond B Biol Sci 369, 20130094.
- van de Wakker SI, Meijers FM, Sluijter JPG, Vader P (2023). Extracellular vesicle heterogeneity and its impact for regenerative medicine applications. Pharmacol Rev 75, 1043–1061.

- Vandenberg LN, Morrie RD, Adams DS (2011). V-ATPase-dependent ectodermal voltage and pH regionalization are required for craniofacial morphogenesis. Dev Dyn 240, 1889–1904.
- Vargas JY, Loria F, Wu YJ, Cordova G, Nonaka T, Bellow S, Syan S, Hasegawa M, van Woerden GM, Trollet C, Zurzolo C (2019). The Wnt/Ca(2+) pathway is involved in interneuronal communication mediated by tunneling nanotubes. EMBO J 38, e101230.
- Venkatesh HS, Morishita W, Geraghty AC, Silverbush D, Gillespie SM, Arzt M, Tam LT, Espenel C, Ponnuswami A, Ni L, et al. (2019). Electrical and synaptic integration of glioma into neural circuits. Nature 573, 539–545.
- Vigetti D, Andrini O, Clerici M, Negrini D, Passi A, Moriondo A (2008). Chondroitin sulfates act as extracellular gating modifiers on voltage-dependent ion channels. Cell Physiol Biochem 22, 137–146.
- Wang X, Veruki ML, Bukoreshtliev NV, Hartveit E, Gerdes HH (2010). Animal cells connected by nanotubes can be electrically coupled through interposed gap-junction channels. Proc Natl Acad Sci U S A 107, 17194–17199.
- Wang Y, Han X, Deng L, Wang X (2024). Tunneling nanotube-transmitted mechanical signal and its cellular response. Biochem Biophys Res Commun 693, 149368.
- Whitaker M (1995). Regulation of the cell division cycle by inositol trisphosphate and the calcium signaling pathway. Adv Second Messenger Phosphoprotein Res 30, 299–310.
- Winkler F, Venkatesh HS, Amit M, Batchelor T, Demir IE, Deneen B, Gutmann DH, Hervey-Jumper S, Kuner T, Mabbott D, et al. (2023). Cancer neuroscience: State of the field, emerging directions. Cell 186, 1689–1707.
- Wolpert L (1969). Positional information and the spatial pattern of cellular differentiation. J Theor Biol 25, 1–47.
- Wonderlin WF, Woodfork KA, Strobl JS (1995). Changes in membrane potential during the progression of MCF-7 human mammary tumor cells through the cell cycle. J Cell Physiol 165, 177–185.
- Yang CY, Bialecka-Fórnal M, Weatherwax C, Larkin JW, Prindle A, Liu J, Garcia-Ojalvo J, Süel GM (2020). Encoding membrane-potential-based memory within a microbial community. Cell Syst 10, 417–423.
- Yang M, Brackenbury WJ (2013). Membrane potential and cancer progression. Front Physiol 4, 185.
- Zappulli V, Friis KP, Fitzpatrick Z, Maguire CA, Breakefield XO (2016). Extracellular vesicles and intercellular communication within the nervous system. J Clin Invest 126, 1198–1207.
- Zhang FY, Yan XY, Wu MY, Chen YM, Zhao H, Zhang CG, Dang PR, Wei L, Zhu FY, Chen Y, et al. (2023). Modulating lineage specification in stem cell differentiation via bioelectrical stimulation intensity matching. Adv Mater Interfaces 11, 2300609.
- Zhang L, Gu H, Li X, Wang Y, Yao S, Chen X, Zheng L, Yang X, Du Q, An J, et al. (2024). Pathophysiological role of ion channels and transporters in hepatocellular carcinoma. Cancer Gene Ther 31, 1611–1618.
- Zhang Y, Zhang Y, Sun K, Meng Z, Chen L (2019). The SLC transporter in nutrient and metabolic sensing, regulation, and drug development. J Mol Cell Biol 11, 1–13.
- Zhao M, Song B, Pu J, Wada T, Reid B, Tai GP, Wang F, Guo AH, Walczysko P, Gu Y, et al. (2006). Electrical signals control wound healing through phosphatidylinositol-3-OH kinase-gamma and PTEN. Nature 442, 457–460.
- Zhou Y, Wong CO, Cho KJ, van der Hoeven D, Liang H, Thakur DP, Luo J, Babic M, Zinsmaier KE, Zhu MX, et al. (2015). SIGNAL TRANSDUCTION. Membrane potential modulates plasma membrane phospholipid dynamics and K-Ras signaling. Science 349, 873–876.
- Zhu K, Hum NR, Reid B, Sun Q, Loots GG, Zhao M (2020). Electric fields at breast cancer and cancer cell collective galvanotaxis. Sci Rep 10, 8712.
- Zurzolo C (2021). Tunneling nanotubes: Reshaping connectivity. Curr Opin Cell Biol 71, 139–147.

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