

568Project

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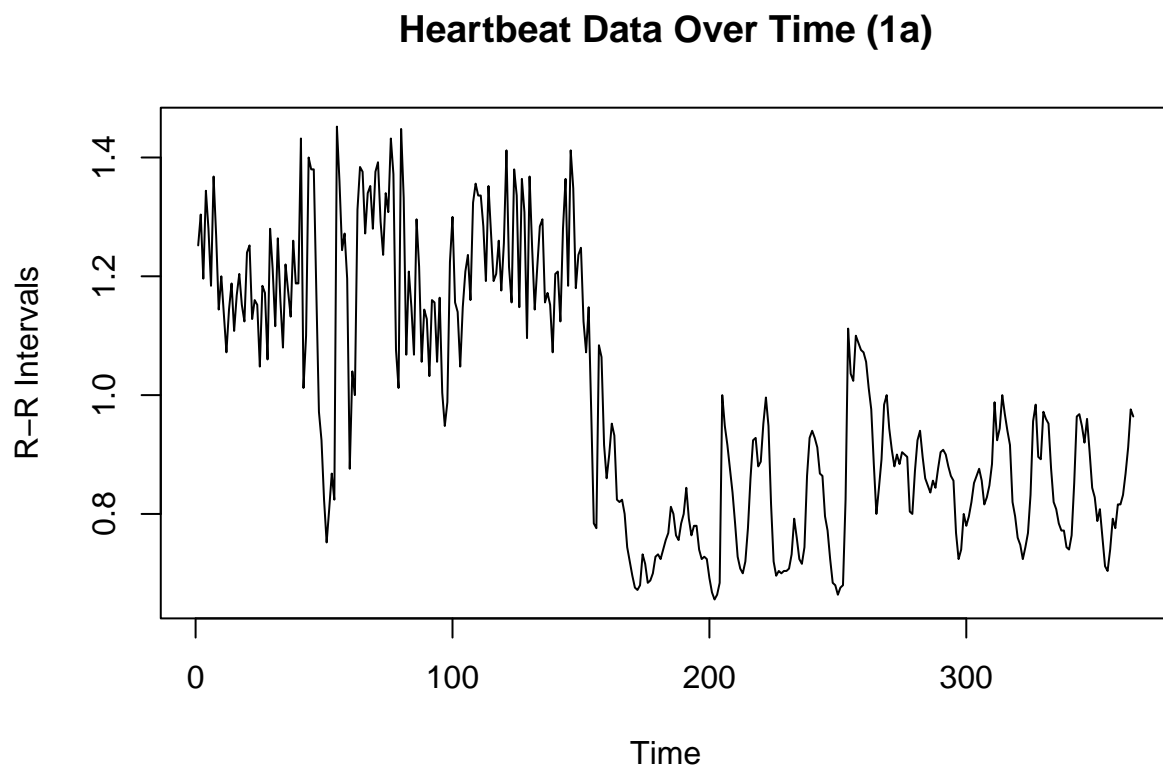
```
## subject 1
data1a <- read.table('data/subj1a.txt', header = FALSE)
data1b <- read.table('data/subj1b.txt',header = FALSE)

## subject 2
data2a <- read.table('data/subj2a.txt', header = FALSE)
data2b <- read.table('data/subj2b.txt',header = FALSE)

## subject 3
data3a <- read.table('data/subj3a.txt', header = FALSE)
data3b <- read.table('data/subj3b.txt',header = FALSE)

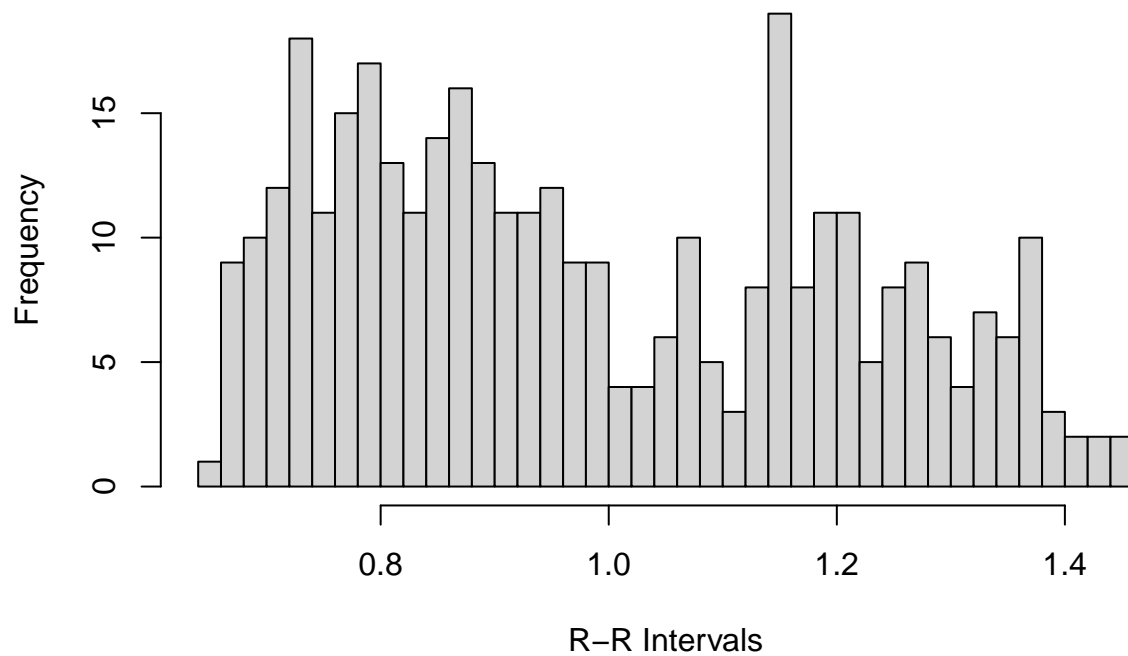
## subject 4
data4a <- read.table('data/subj4a.txt', header = FALSE)
data4b <- read.table('data/subj4b.txt',header = FALSE)

plot(data1a$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (1a)
```



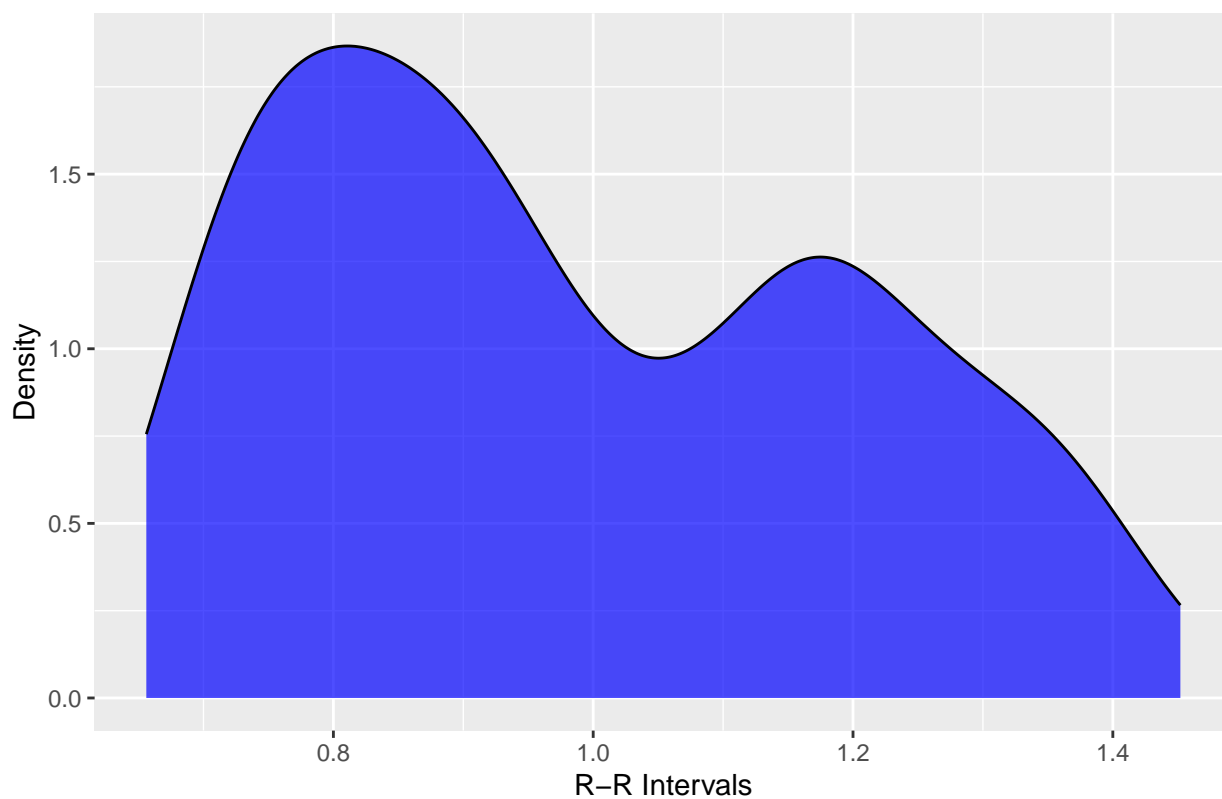
```
hist(data1a$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (1a)")
library(ggplot2)
```

Distribution of R-R Intervals (1a)



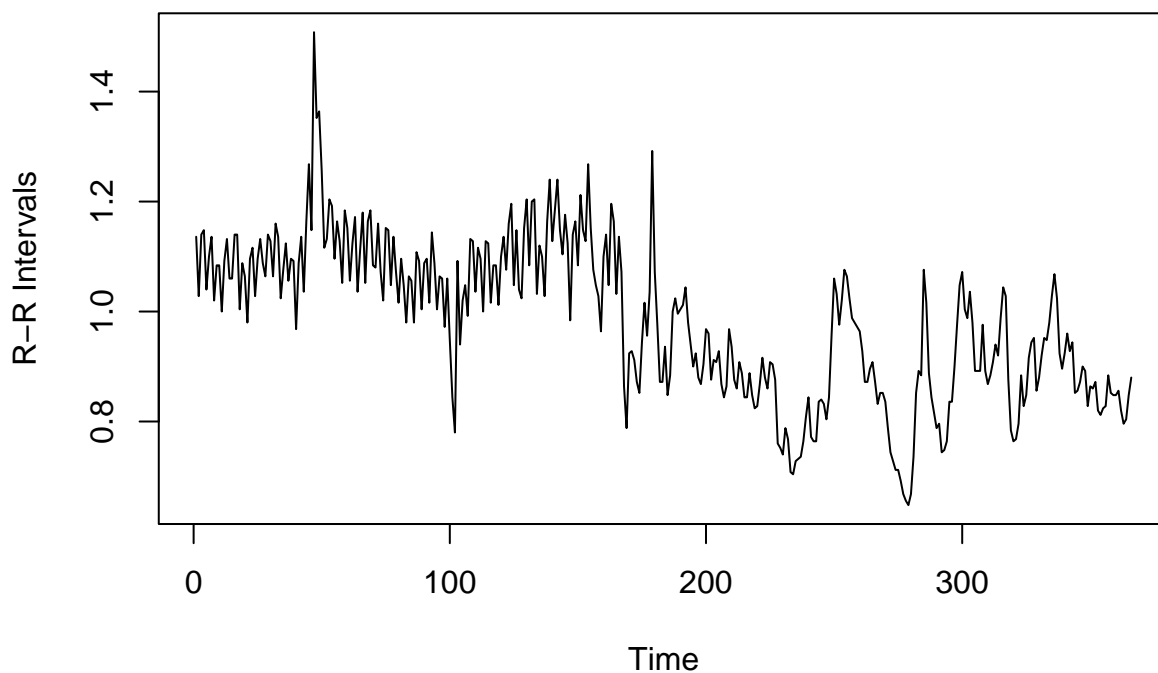
```
ggplot(data1a, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (1a)")
```

Density Plot of R-R Intervals (1a)



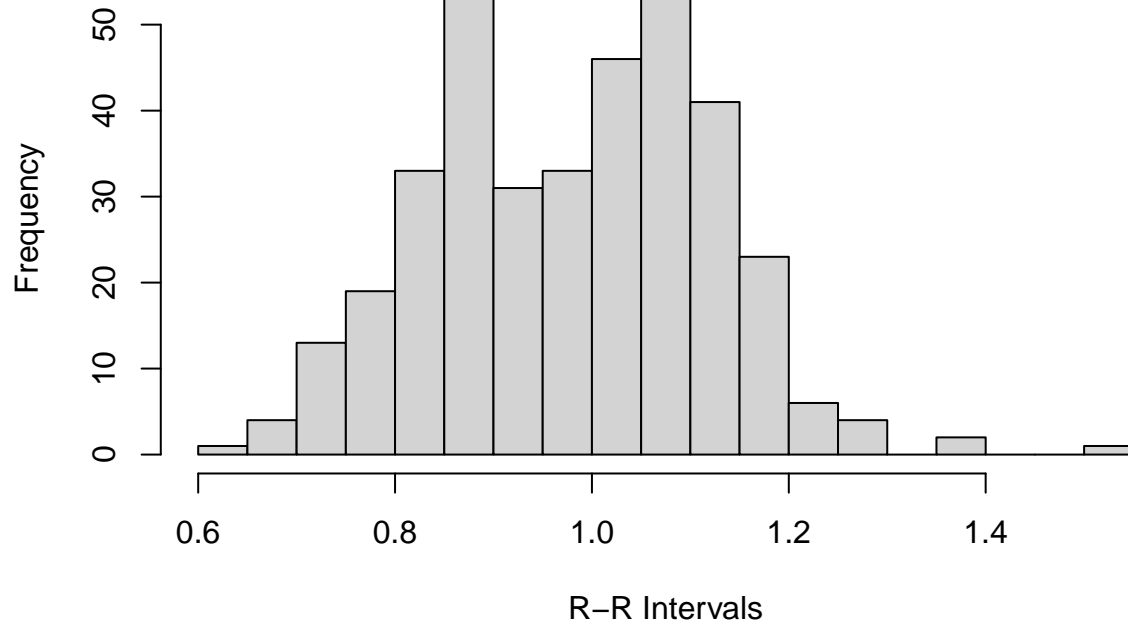
```
plot(data1b$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (1b)
```

Heartbeat Data Over Time (1b)



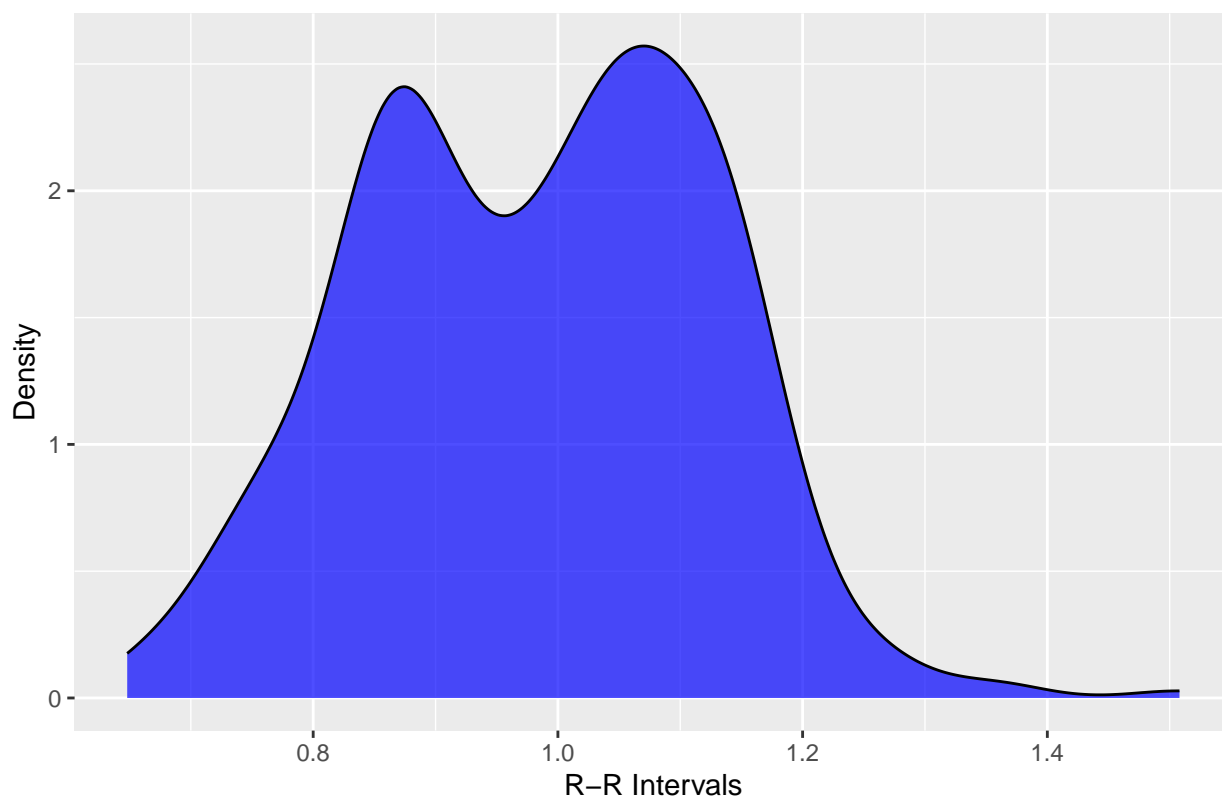
```
hist(data1b$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (1b)")
```

Distribution of R-R Intervals (1b)



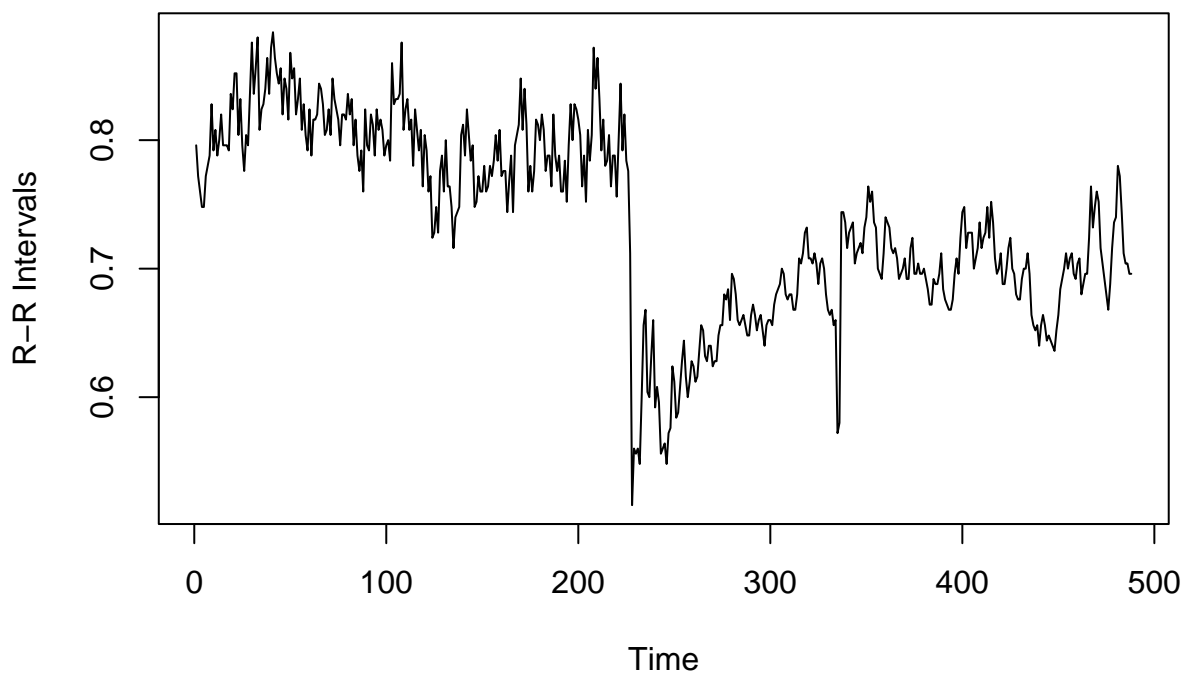
```
library(ggplot2)
ggplot(data1b, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (1b)")
```

Density Plot of R-R Intervals (1b)



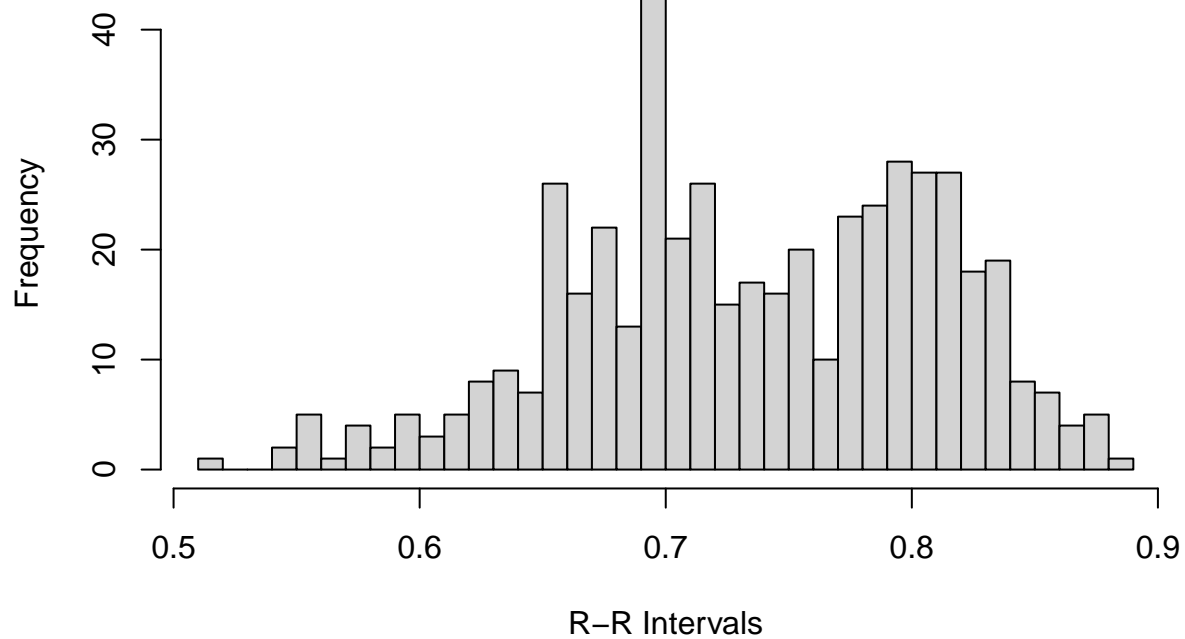
```
plot(data2a$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (2a)
```

Heartbeat Data Over Time (2a)



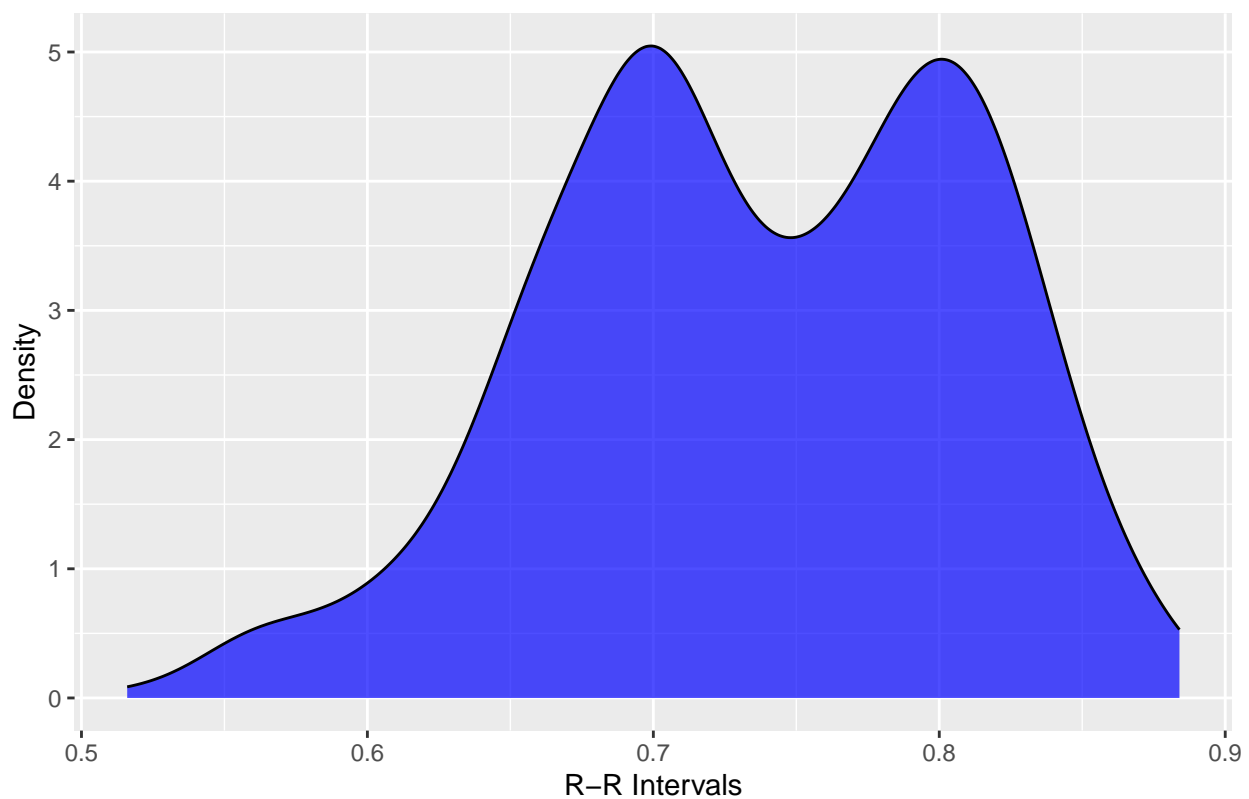
```
hist(data2a$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (2a)")
```

Distribution of R-R Intervals (2a)



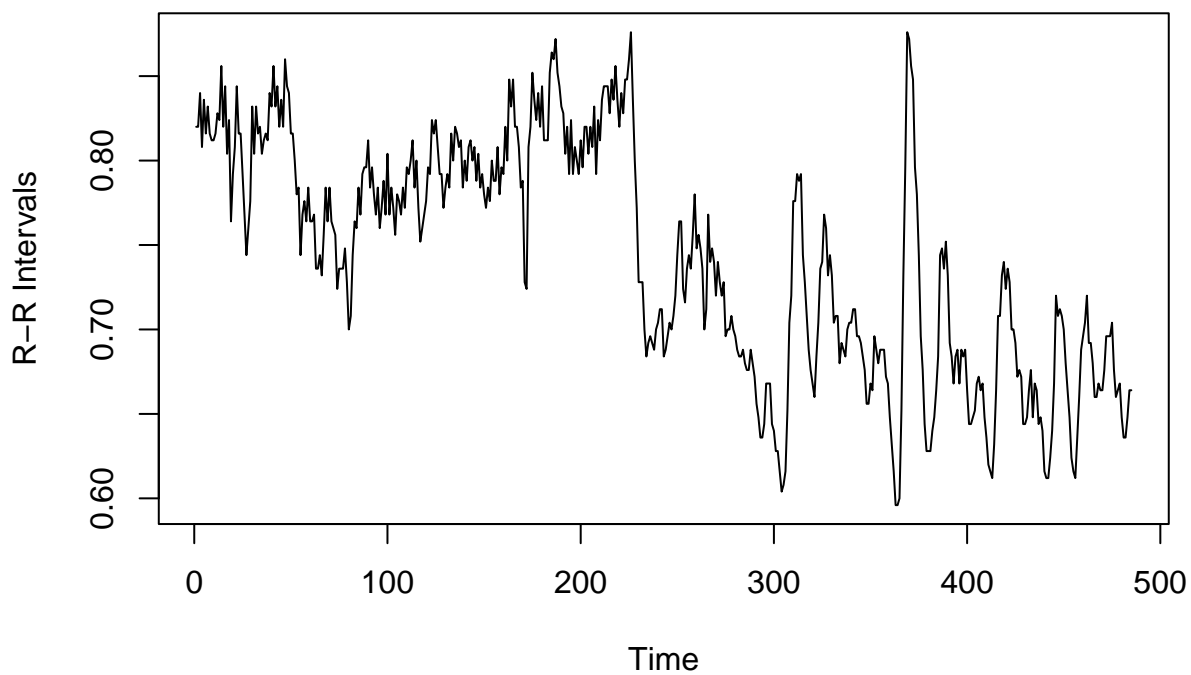
```
library(ggplot2)
ggplot(data2a, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (2a)")
```

Density Plot of R-R Intervals (2a)



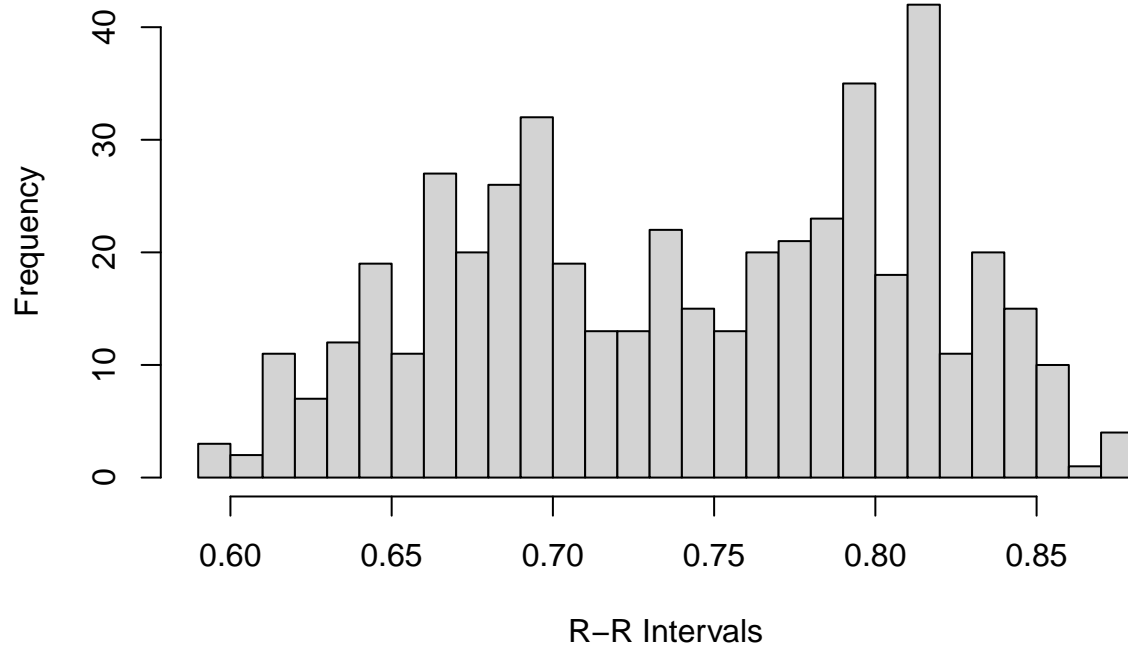
```
plot(data2b$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (2b)
```

Heartbeat Data Over Time (2b)



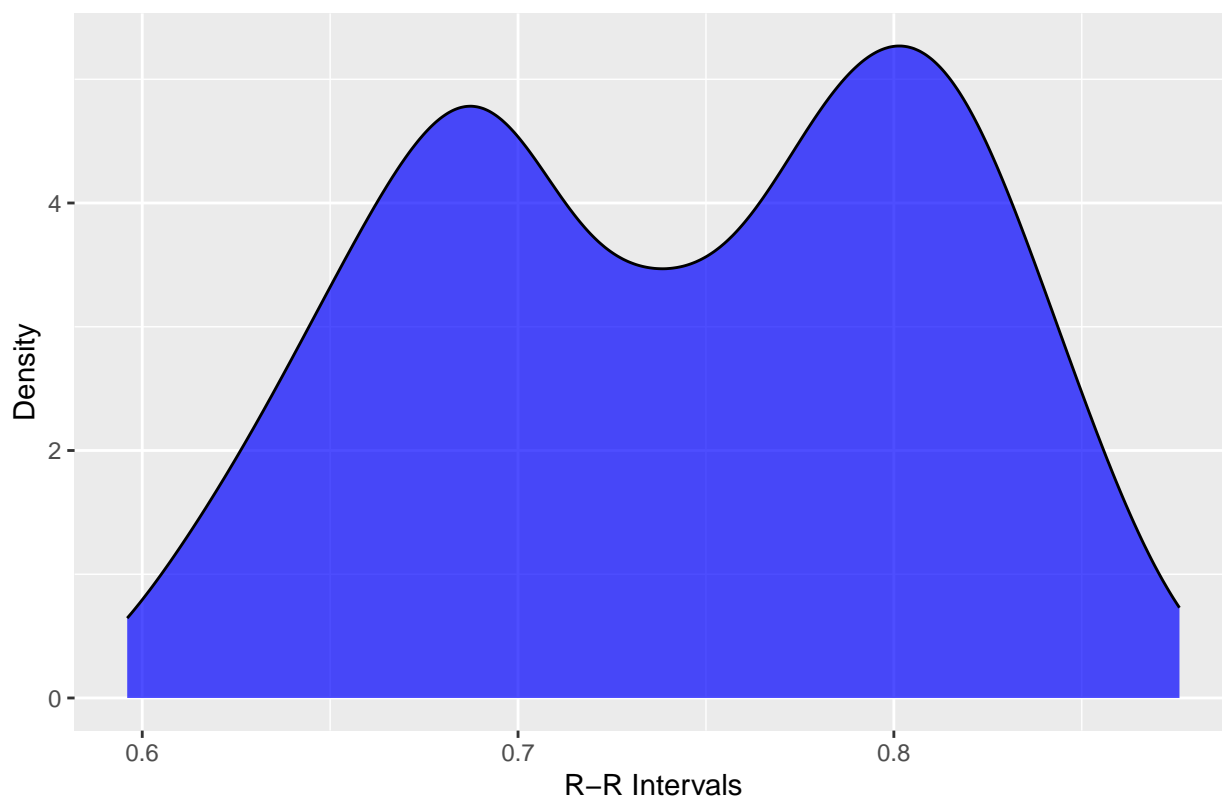
```
hist(data2b$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (2b)")
```

Distribution of R-R Intervals (2b)



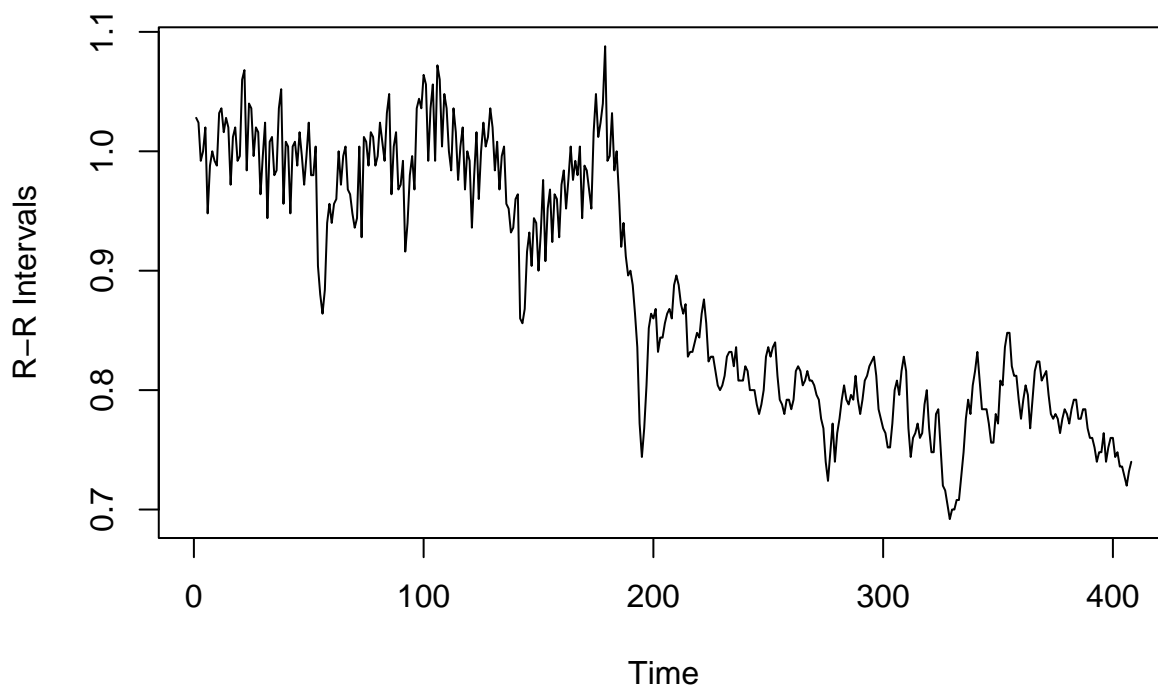
```
library(ggplot2)
ggplot(data2b, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (2b)")
```


Density Plot of R-R Intervals (2b)



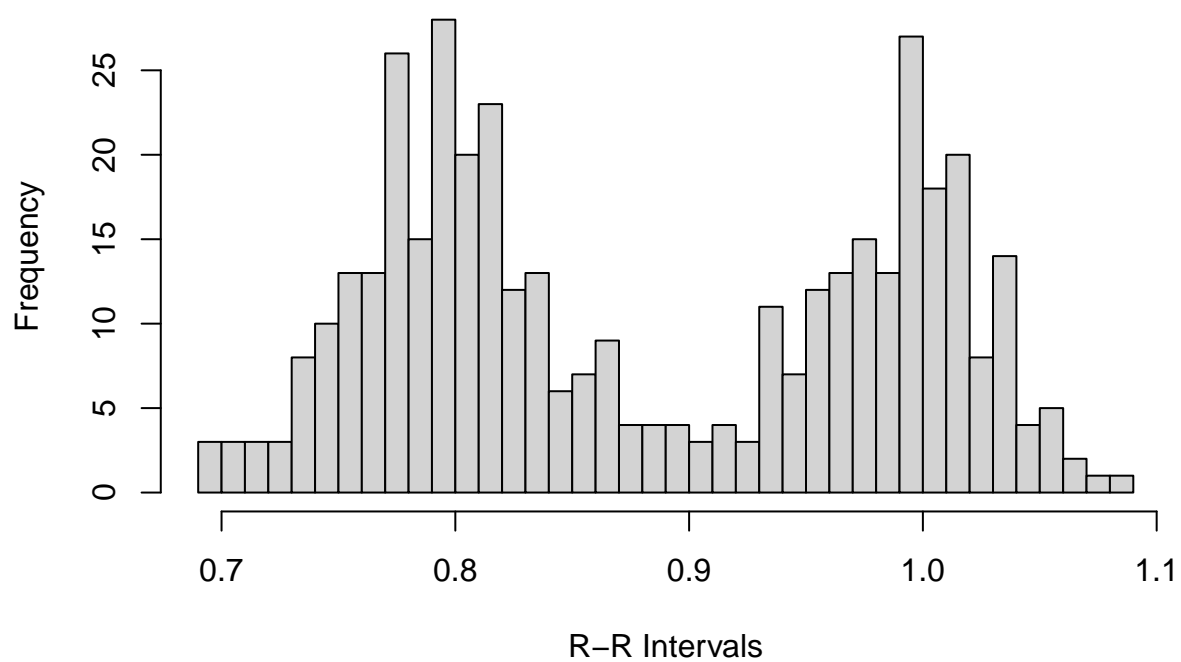
```
plot(data3a$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (3a)
```

Heartbeat Data Over Time (3a)



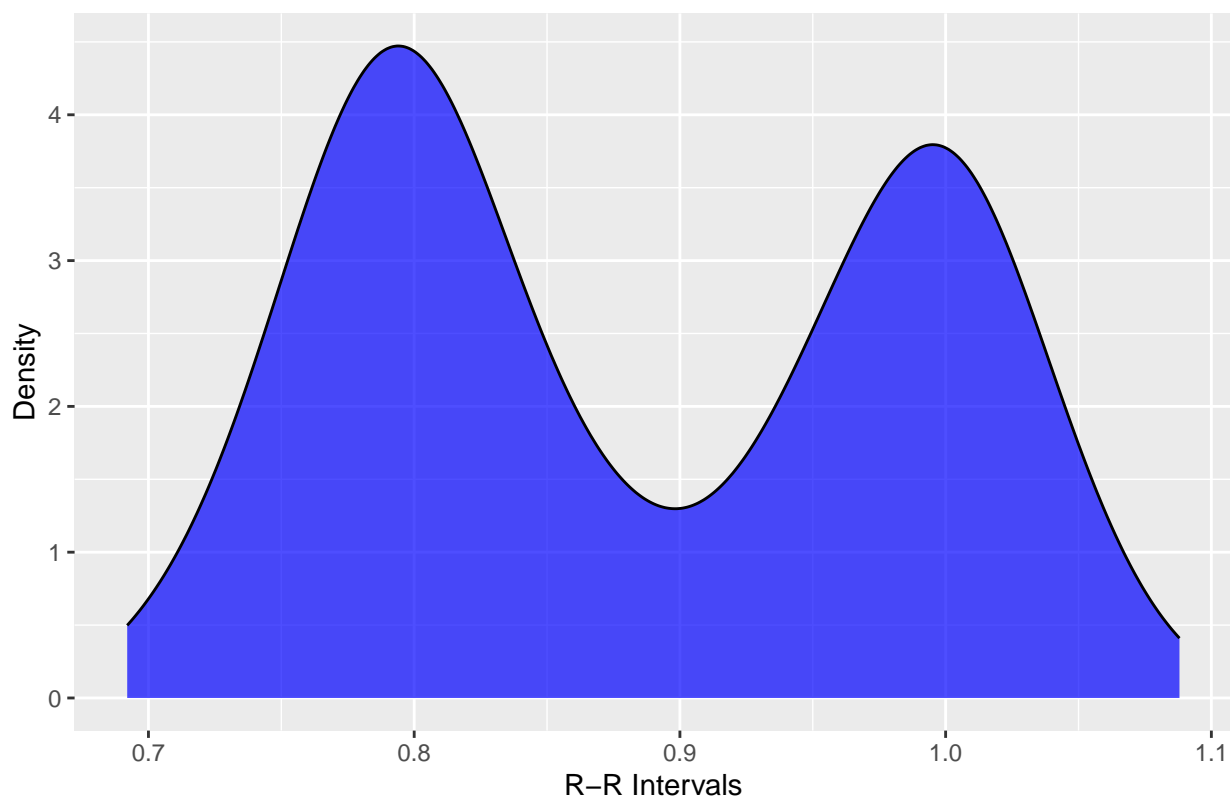
```
hist(data3a$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (3a)")
```

Distribution of R-R Intervals (3a)



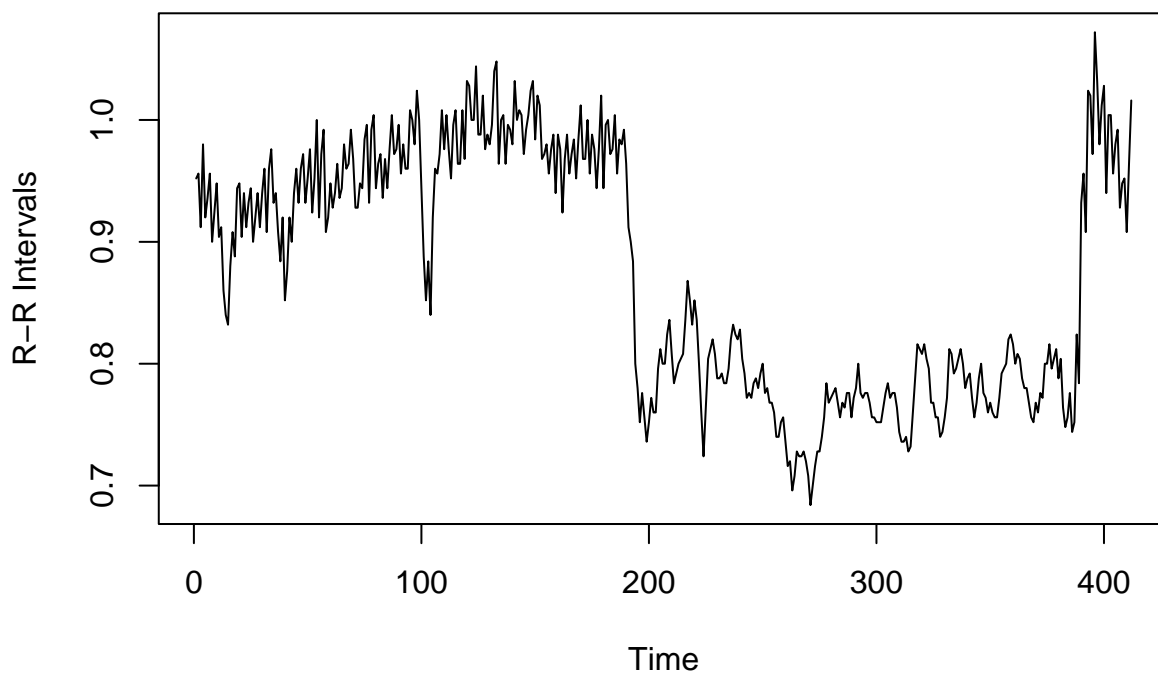
```
library(ggplot2)
ggplot(data3a, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (3a)")
```

Density Plot of R-R Intervals (3a)



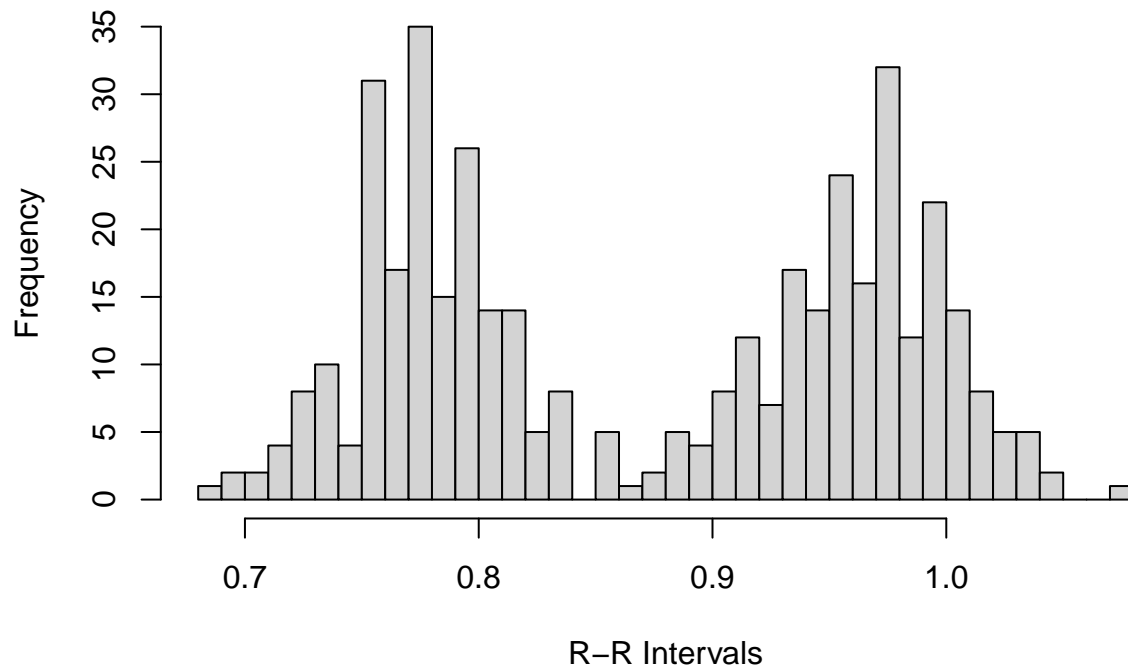
```
plot(data3b$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (3b)
```

Heartbeat Data Over Time (3b)



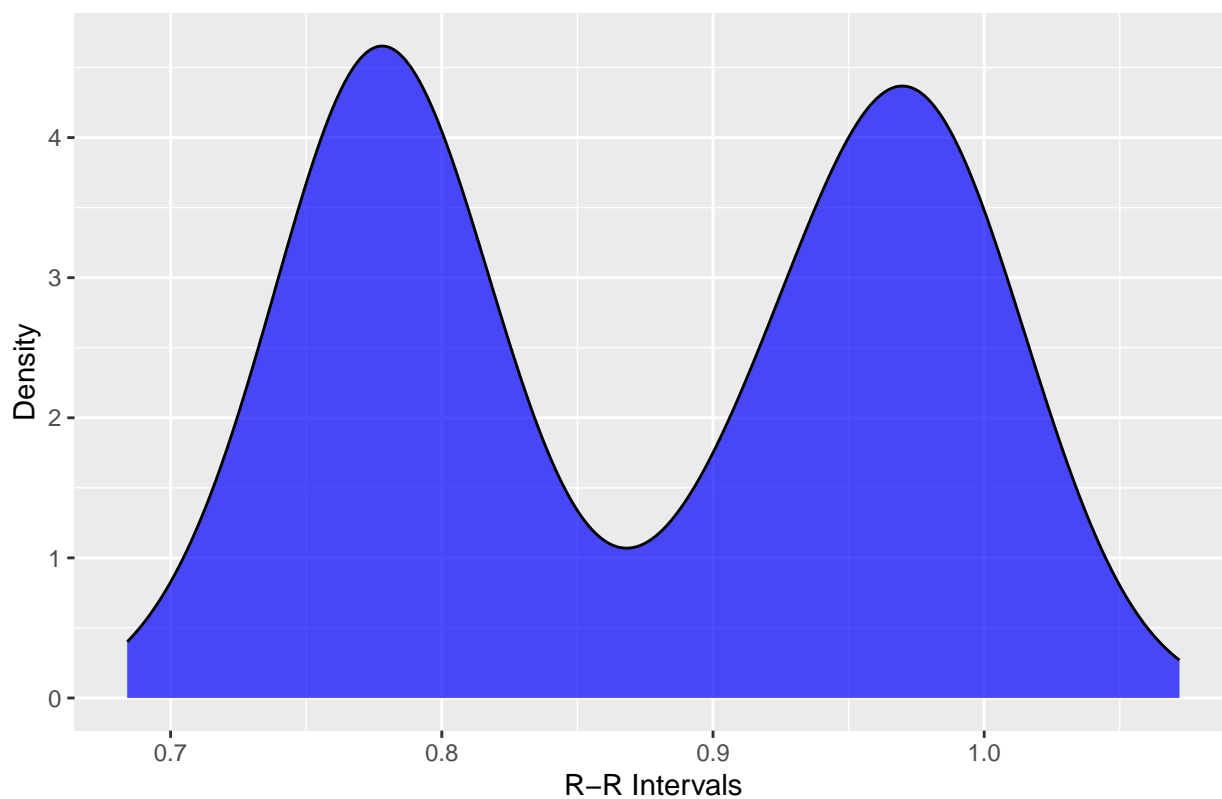
```
hist(data3b$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (3b)")
```

Distribution of R-R Intervals (3b)



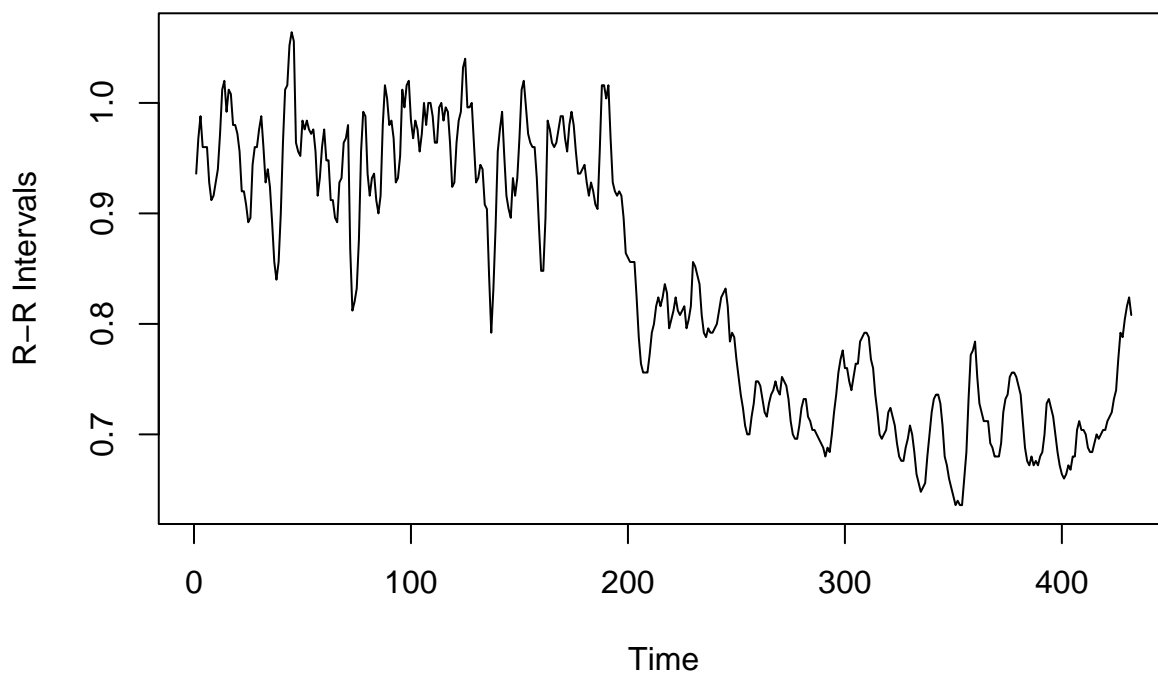
```
library(ggplot2)
ggplot(data3b, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (3b)")
```

Density Plot of R-R Intervals (3b)



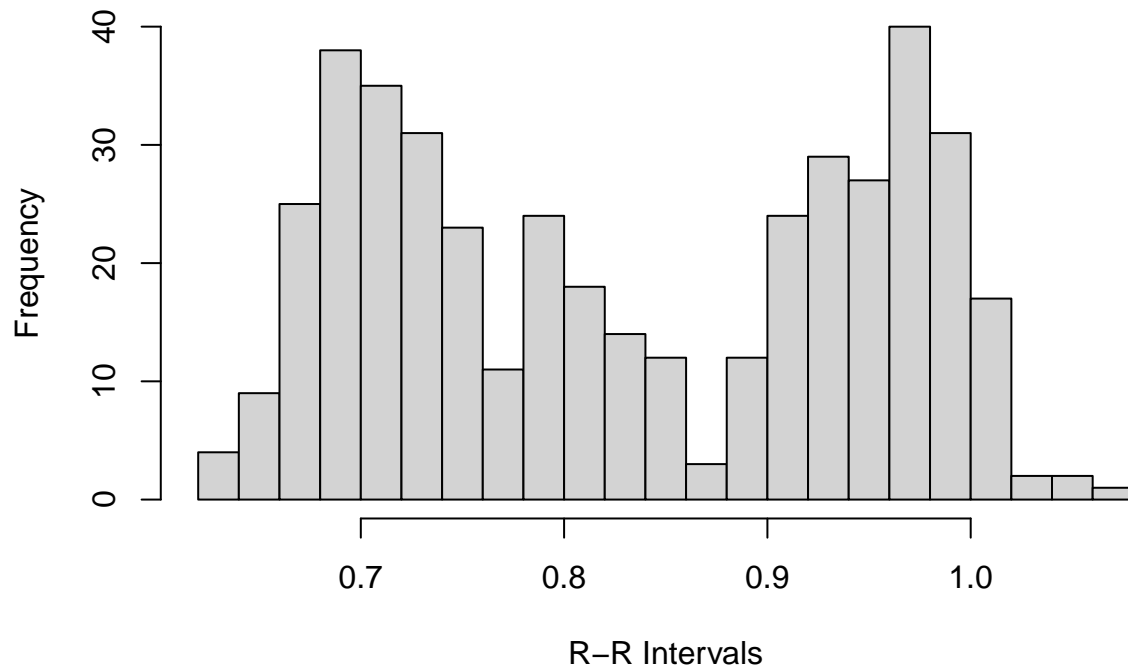
```
plot(data4a$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (4a)
```

Heartbeat Data Over Time (4a)



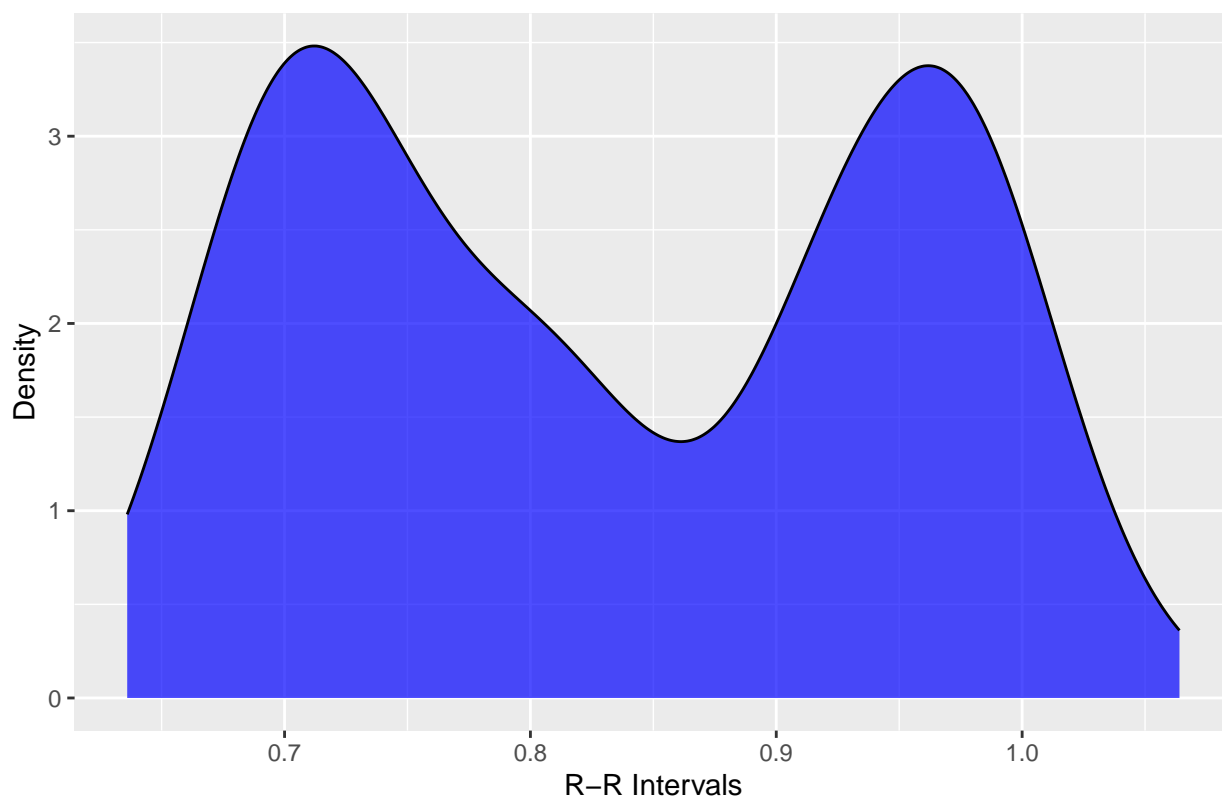
```
hist(data4a$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (4a)")
```

Distribution of R-R Intervals (4a)



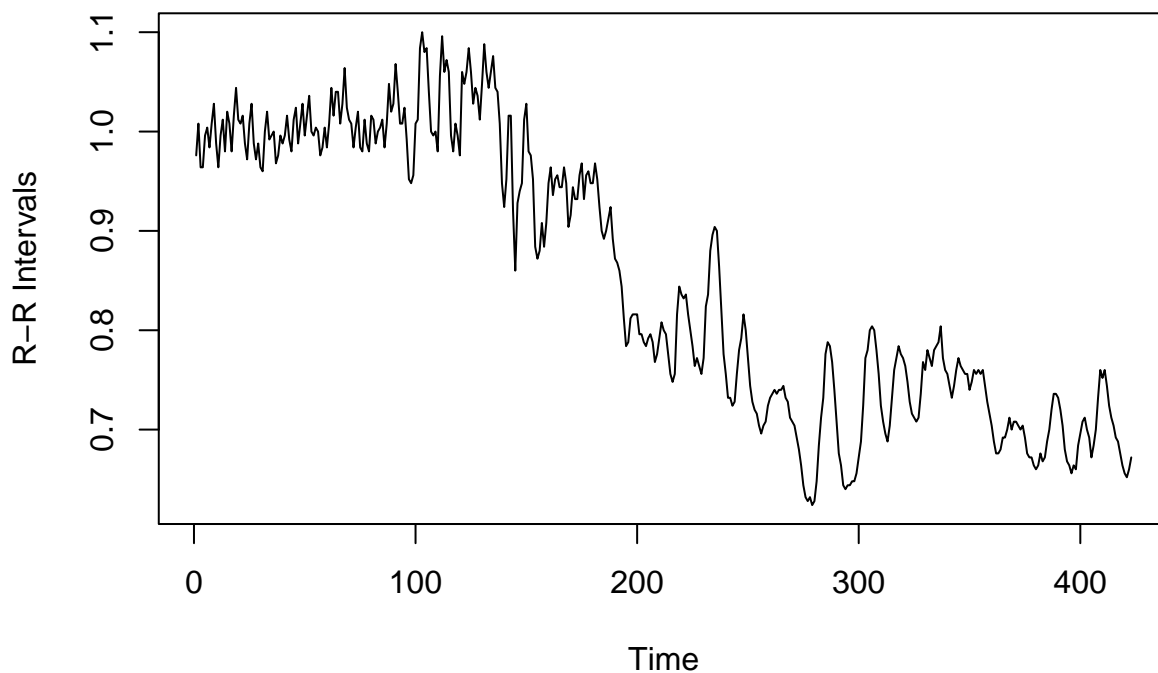
```
library(ggplot2)
ggplot(data4a, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (4a)")
```

Density Plot of R-R Intervals (4a)



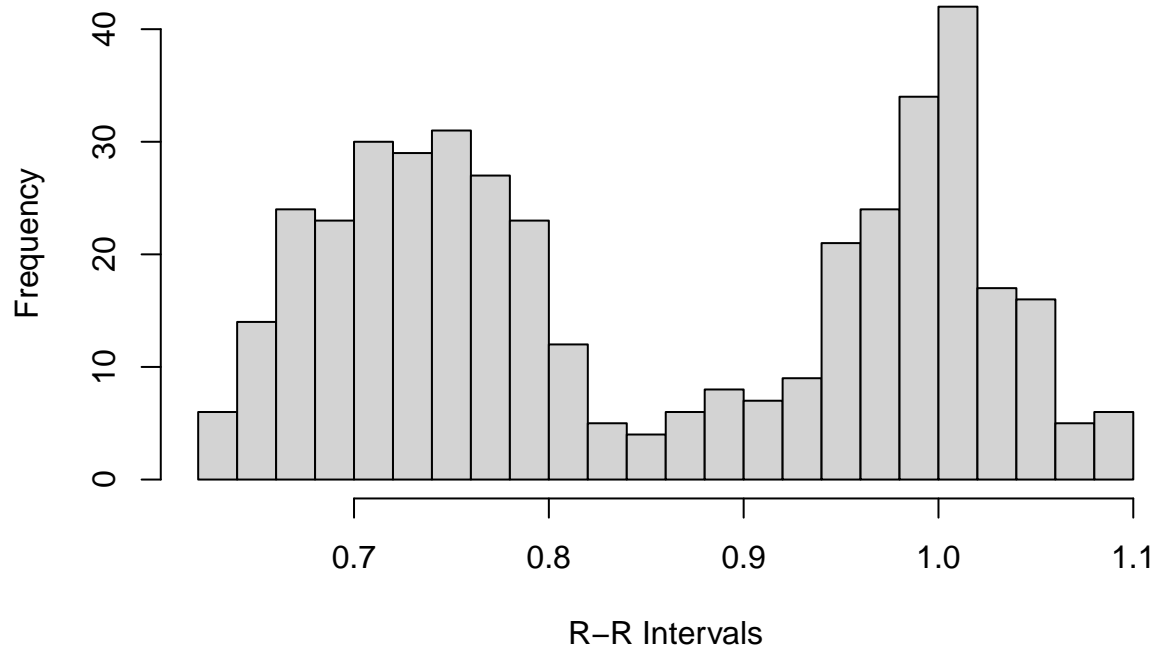
```
plot(data4b$V2, type = "l", xlab = "Time", ylab = "R-R Intervals", main = "Heartbeat Data Over Time (4b)
```

Heartbeat Data Over Time (4b)



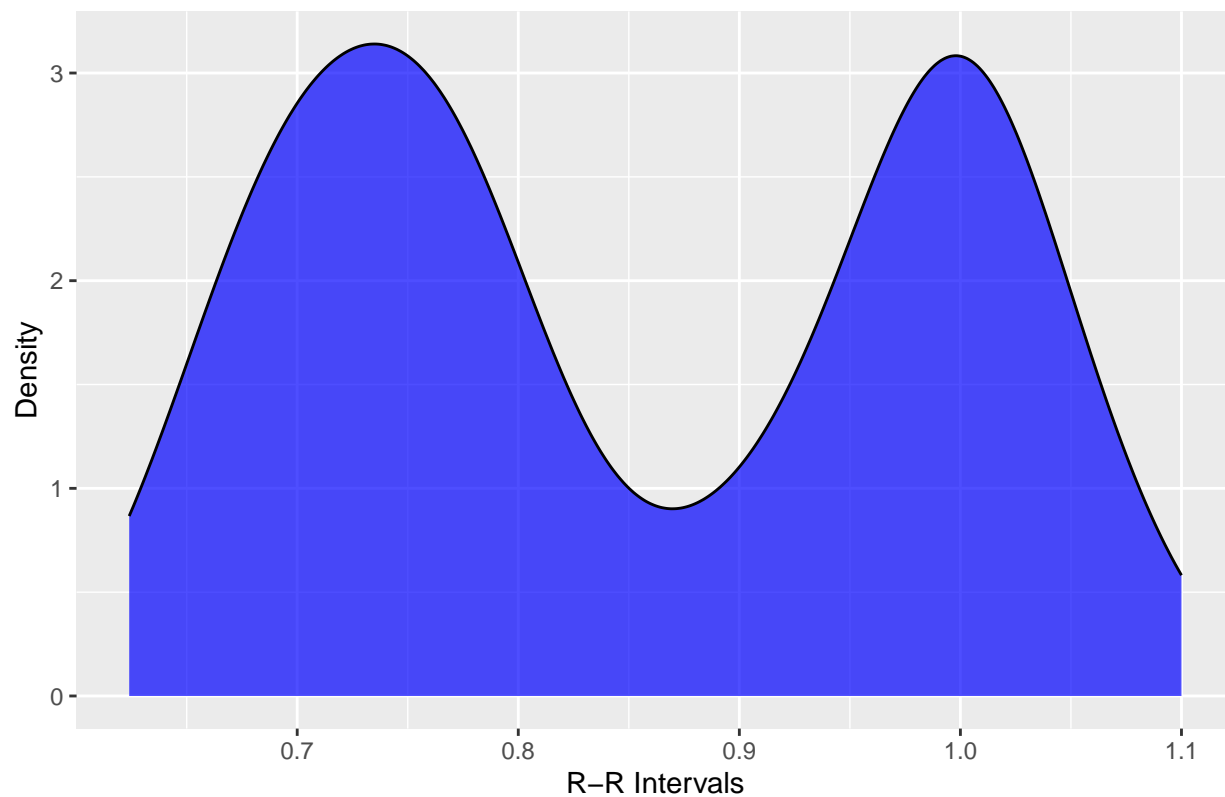
```
hist(data4b$V2, breaks = 30, xlab = "R-R Intervals", main = "Distribution of R-R Intervals (4b)")
```

Distribution of R-R Intervals (4b)



```
library(ggplot2)
ggplot(data4b, aes(x = V2)) + geom_density(fill = "blue", alpha = 0.7) +
  labs(x = "R-R Intervals", y = "Density", title = "Density Plot of R-R Intervals (4b)")
```


Density Plot of R-R Intervals (4b)



```
library(stats)
rr1a <- data1a$V2

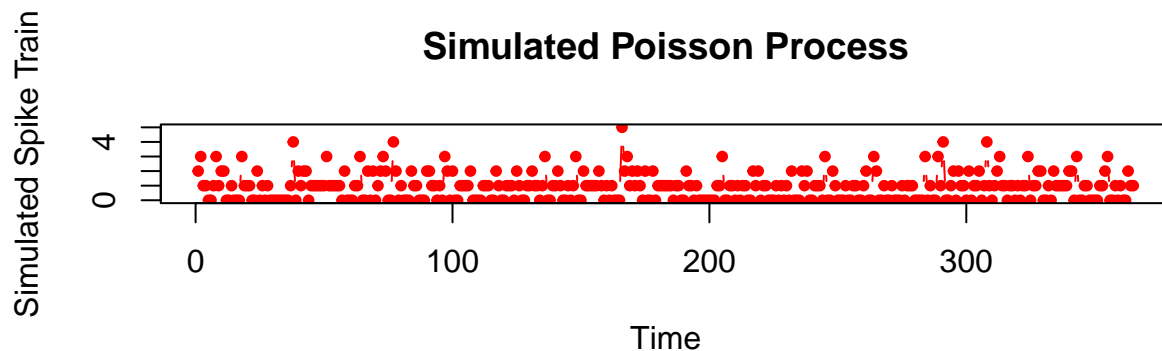
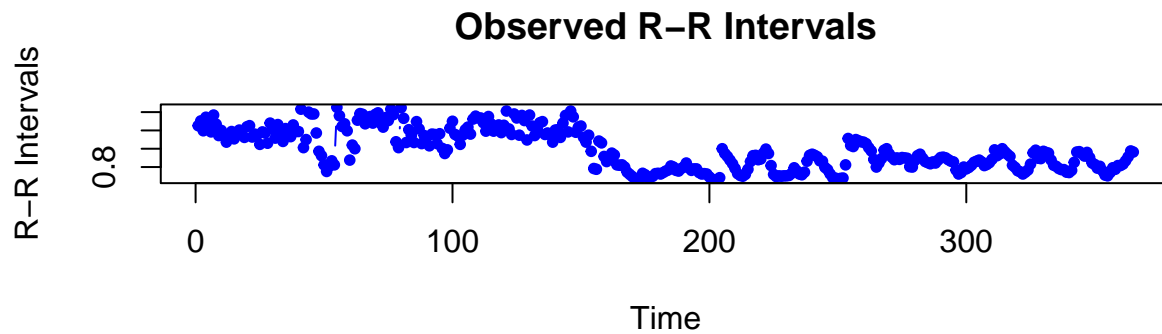
lambda_est <- mean(rr1a)

cat("Estimated rate parameter (lambda):", lambda_est, "\n")

## Estimated rate parameter (lambda): 0.9876822
simulated_spike_train <- rpois(length(rr1a), lambda = lambda_est)

par(mfrow = c(2, 1))
plot(rr1a, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'R-R Intervals',
     main = 'Observed R-R Intervals')

plot(simulated_spike_train, type = 'b', col = 'red', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train',
     main = 'Simulated Poisson Process')
```



```
par(mfrow = c(1, 1))

# Perform KS test
ks_result <- ks.test(rr1a, "pexp", rate = 1/lambda_est)

# Display the results of the KS test
cat("KS Test Results:\n")
```

KS Test Results:

```
print(ks_result)
```

```
##
## Asymptotic one-sample Kolmogorov-Smirnov test
##
## data: rr1a
## D = 0.48672, p-value < 2.2e-16
## alternative hypothesis: two-sided
```

DISTRIBUTIONS NOT IDENTICAL. POISSON PROCESS IS NOT A GOOD FIT.

```
# Load the required library
library(stats)

# Extract the R-R intervals from the second column and rename the variable
rr1b <- data1b$V2

# Calculate the mean R-R interval as an estimate for the rate parameter (lambda)
lambda_est_b <- mean(rr1b)

# Display the estimated rate parameter
```

```

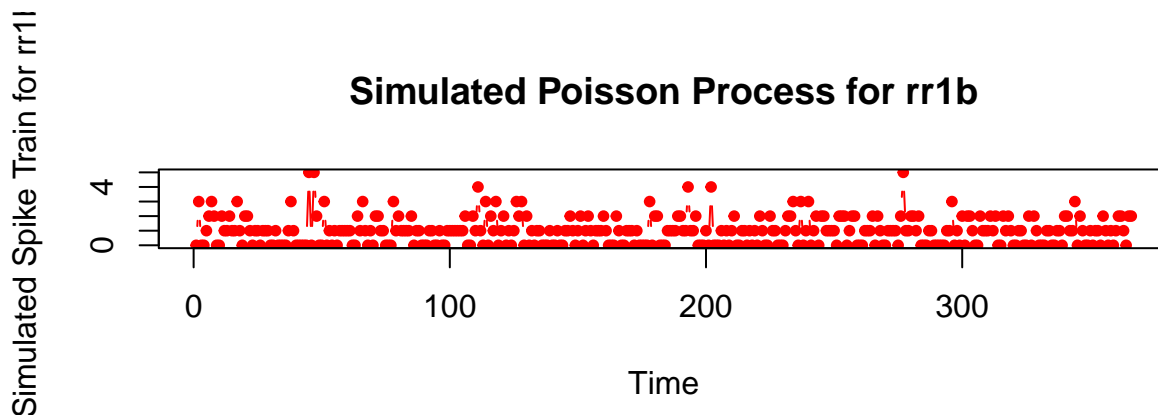
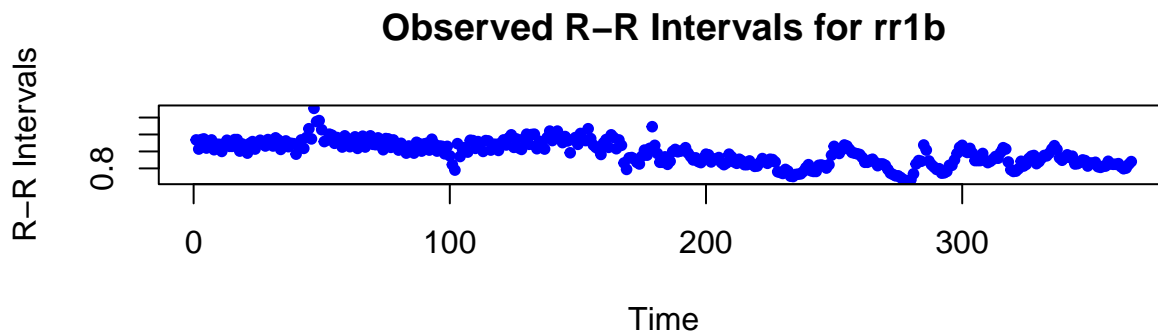
cat("Estimated rate parameter (lambda) for rr1b:", lambda_est_b, "\n")

## Estimated rate parameter (lambda) for rr1b: 0.9828197
# Simulate a Poisson process based on the estimated lambda
simulated_spike_train_b <- rpois(length(rr1b), lambda = lambda_est_b)

# Plot the observed R-R intervals and the simulated spike train for rr1b
par(mfrow = c(2, 1))
plot(rr1b, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'R-R Intervals',
     main = 'Observed R-R Intervals for rr1b')

plot(simulated_spike_train_b, type = 'b', col = 'red', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train for rr1b',
     main = 'Simulated Poisson Process for rr1b')

```



```

# Reset the plotting layout
par(mfrow = c(1, 1))

# Perform KS test for rr1b
ks_result_b <- ks.test(rr1b, "pexp", rate = 1/lambda_est_b)

# Display the results of the KS test for rr1b
cat("KS Test Results for rr1b:\n")

```

```
## KS Test Results for rr1b:
```

```
print(ks_result_b)
```

```
##
```

```
## Asymptotic one-sample Kolmogorov-Smirnov test
##
## data: rr1b
## D = 0.49864, p-value < 2.2e-16
## alternative hypothesis: two-sided

# Extract the R-R intervals from the second column and rename the variable
rr2a <- data2a$V2

# Calculate the mean R-R interval as an estimate for the rate parameter (lambda)
lambda_est_2a <- mean(rr2a)

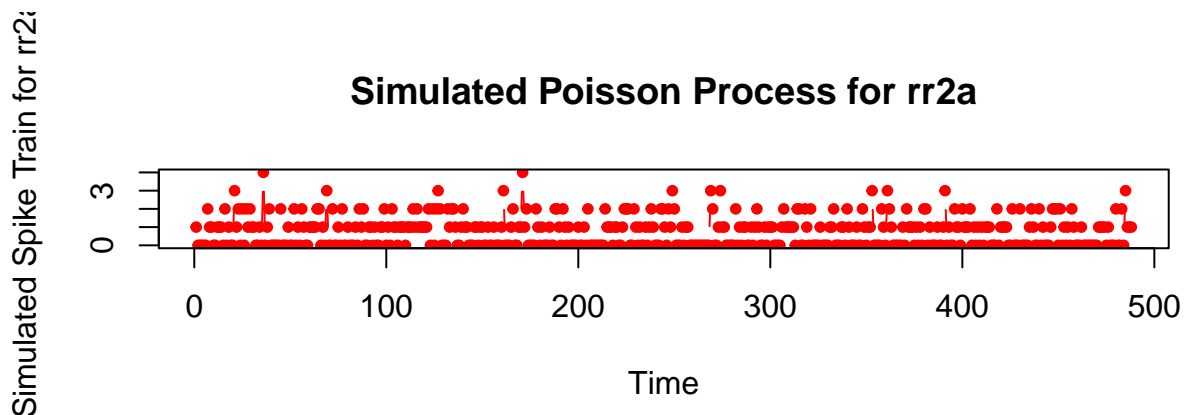
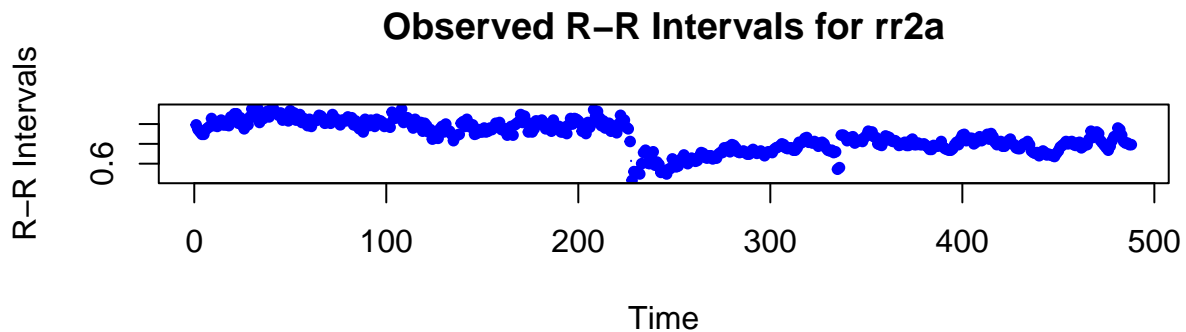
# Display the estimated rate parameter
cat("Estimated rate parameter (lambda) for rr2a:", lambda_est_2a, "\n")

## Estimated rate parameter (lambda) for rr2a: 0.7369344

# Simulate a Poisson process based on the estimated lambda
simulated_spike_train_2a <- rpois(length(rr2a), lambda = lambda_est_2a)

# Plot the observed R-R intervals and the simulated spike train for rr2a
par(mfrow = c(2, 1))
plot(rr2a, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'R-R Intervals',
     main = 'Observed R-R Intervals for rr2a')

plot(simulated_spike_train_2a, type = 'b', col = 'red', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train for rr2a',
     main = 'Simulated Poisson Process for rr2a')
```



```

# Reset the plotting layout
par(mfrow = c(1, 1))

# Perform KS test for rr2a
ks_result_2a <- ks.test(rr2a, "pexp", rate = 1/lambda_est_2a)

# Display the results of the KS test for rr2a
cat("KS Test Results for rr2a:\n")

## KS Test Results for rr2a:
print(ks_result_2a)

##
## Asymptotic one-sample Kolmogorov-Smirnov test
##
## data: rr2a
## D = 0.5236, p-value < 2.2e-16
## alternative hypothesis: two-sided

# Load data for subject 2b
data2b <- read.table('data/subj2b.txt', header = FALSE)

# Extract the R-R intervals from the second column and rename the variable
rr2b <- data2b$V2

# Calculate the mean R-R interval as an estimate for the rate parameter (lambda)
lambda_est_2b <- mean(rr2b)

# Display the estimated rate parameter
cat("Estimated rate parameter (lambda) for rr2b:", lambda_est_2b, "\n")

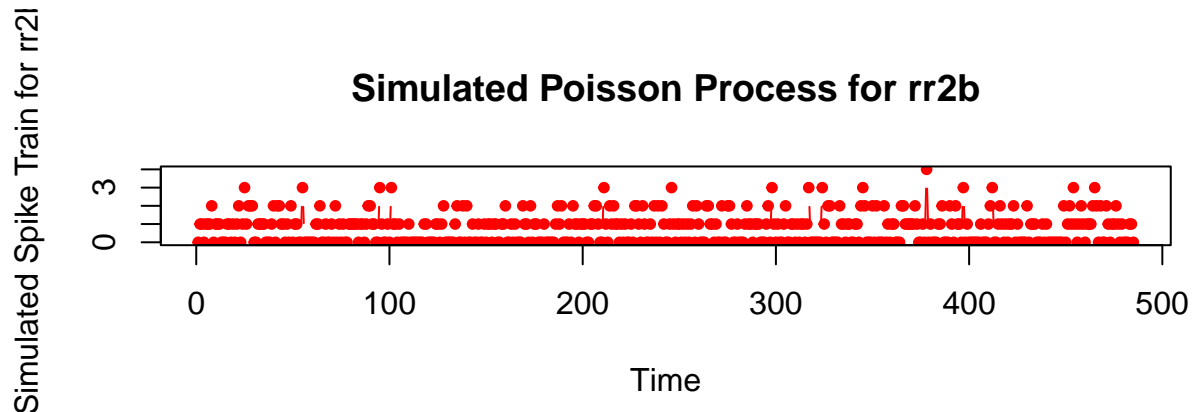
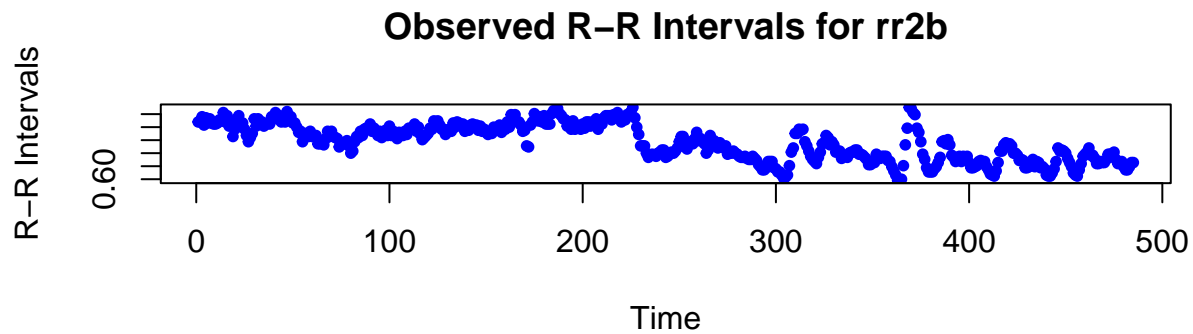
## Estimated rate parameter (lambda) for rr2b: 0.7423423

# Simulate a Poisson process based on the estimated lambda
simulated_spike_train_2b <- rpois(length(rr2b), lambda = lambda_est_2b)

# Plot the observed R-R intervals and the simulated spike train for rr2b
par(mfrow = c(2, 1))
plot(rr2b, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'R-R Intervals',
     main = 'Observed R-R Intervals for rr2b')

plot(simulated_spike_train_2b, type = 'b', col = 'red', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train for rr2b',
     main = 'Simulated Poisson Process for rr2b')

```



```
# Reset the plotting layout
par(mfrow = c(1, 1))

# Perform KS test for rr2b
ks_result_2b <- ks.test(rr2b, "pexp", rate = 1/lambda_est_2b)

# Display the results of the KS test for rr2b
cat("KS Test Results for rr2b:\n")

## KS Test Results for rr2b:

print(ks_result_2b)

##
## Asymptotic one-sample Kolmogorov-Smirnov test
##
## data: rr2b
## D = 0.55196, p-value < 2.2e-16
## alternative hypothesis: two-sided

# Load the required library
library(stats)

# Load data for subject 1a
data1a <- read.table('data/subj1a.txt', header = FALSE)

# Extract the R-R intervals from the second column and rename the variable
rr1a <- data1a$V2

# Create a time variable (assuming time increments of 1, adjust as needed)
time <- 1:length(rr1a)
```

```

# Fit a linear model to estimate the intensity function
model <- lm(rr1a ~ time)

# Extract the coefficients
beta0 <- coef(model)[1]
beta1 <- coef(model)[2]

# Define the estimated intensity function
lambda_est <- exp(beta0 + beta1 * time)

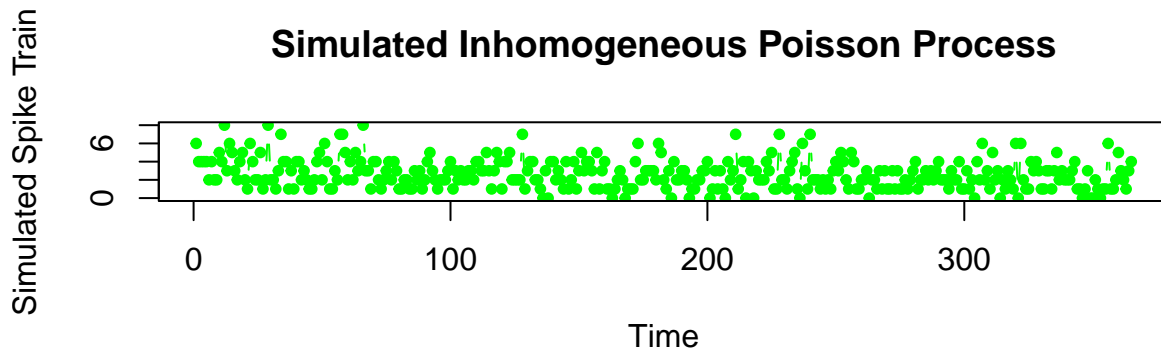
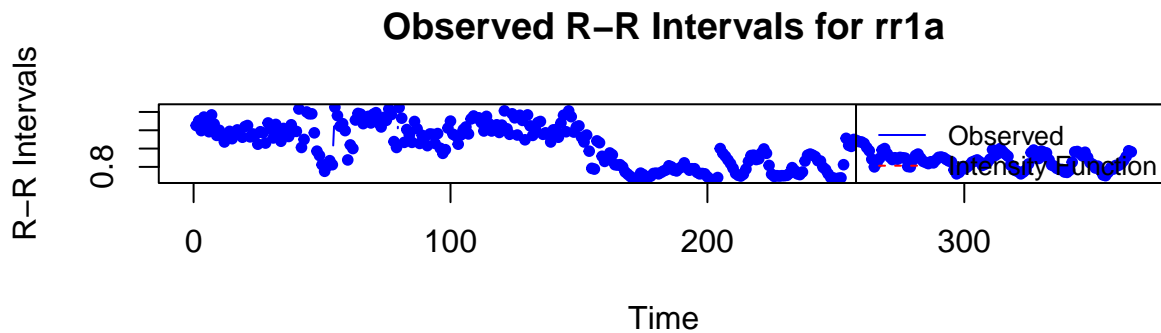
# Simulate an inhomogeneous Poisson process based on the estimated intensity
simulated_spike_train <- rpois(length(rr1a), lambda = lambda_est)

# Plot the observed R-R intervals and the estimated intensity function
par(mfrow = c(2, 1))
plot(rr1a, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'R-R Intervals',
     main = 'Observed R-R Intervals for rr1a')

lines(time, lambda_est, col = 'red', lty = 2)
legend("topright", legend = c("Observed", "Intensity Function"),
     col = c("blue", "red"), lty = c(1, 2), cex = 0.8)

# Plot the simulated inhomogeneous Poisson process
plot(simulated_spike_train, type = 'b', col = 'green', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train',
     main = 'Simulated Inhomogeneous Poisson Process')

```



```

# Reset the plotting layout
par(mfrow = c(1, 1))

# Perform KS test for inhomogeneous Poisson process
ks_result_inh <- ks.test(rr1a, lambda_est)

## Warning in ks.test.default(rr1a, lambda_est): p-value will be approximate in
## the presence of ties

# Display the results of the KS test for inhomogeneous Poisson process
cat("KS Test Results for Inhomogeneous Poisson Process:\n")

## KS Test Results for Inhomogeneous Poisson Process:
print(ks_result_inh)

##
## Asymptotic two-sample Kolmogorov-Smirnov test
##
## data: rr1a and lambda_est
## D = 1, p-value < 2.2e-16
## alternative hypothesis: two-sided

library(stats)

# Transform R-R intervals to log(R-R)
log_rr1a <- log(rr1a)

# Remove non-positive values
log_rr1a <- log_rr1a[log_rr1a > 0]

# Check if there are still non-positive values
if (any(log_rr1a <= 0)) {
  stop("After log transformation, there are still non-positive values in log(R-R) intervals.")
}

# Estimate rate parameter for the transformed data
lambda_est_log <- mean(log_rr1a)

cat("Estimated rate parameter (lambda) for log(R-R):", lambda_est_log, "\n")

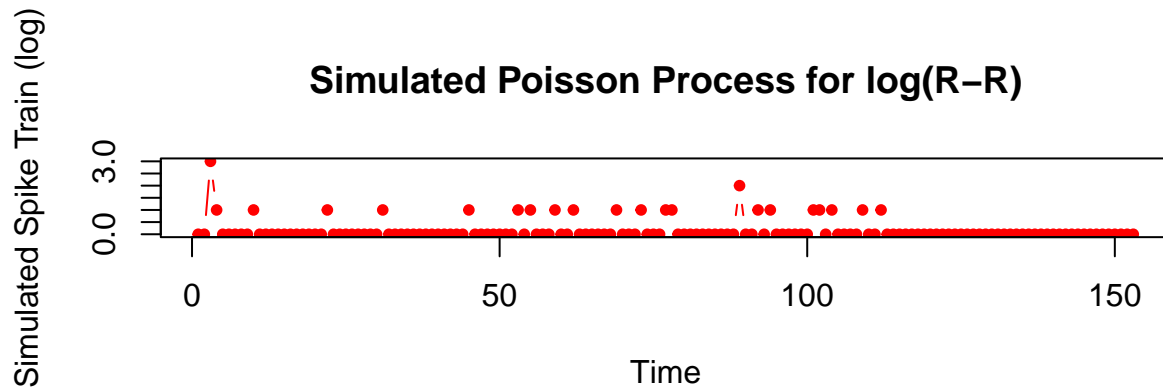
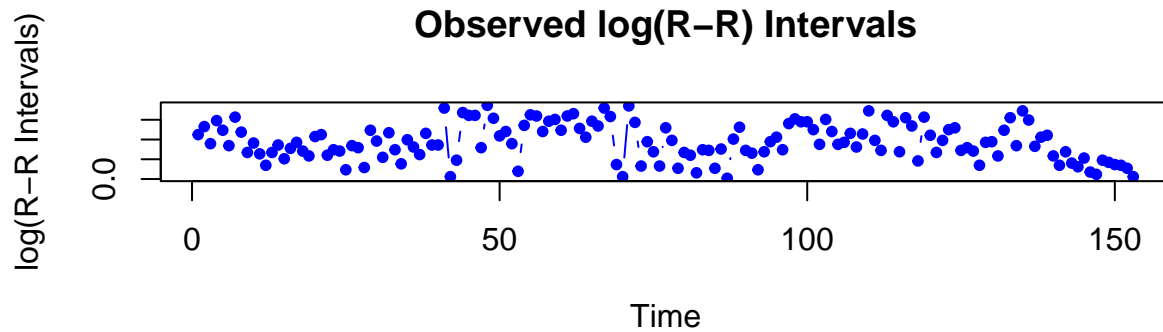
## Estimated rate parameter (lambda) for log(R-R): 0.1874978

# Simulate Poisson process for log(R-R)
simulated_spike_train_log <- rpois(length(log_rr1a), lambda = lambda_est_log)

# Plot the observed log(R-R) intervals and the simulated spike train
par(mfrow = c(2, 1))
plot(log_rr1a, type = 'b', col = 'blue', pch = 20,
     xlab = 'Time', ylab = 'log(R-R Intervals)',
     main = 'Observed log(R-R) Intervals')

plot(simulated_spike_train_log, type = 'b', col = 'red', pch = 20,
     xlab = 'Time', ylab = 'Simulated Spike Train (log)',
     main = 'Simulated Poisson Process for log(R-R)')

```

```
# Reset the plotting layout
par(mfrow = c(1, 1))
```

```
# Perform KS test for log(R-R)
ks_result_log <- ks.test(log_rr1a, "pexp", rate = 1/lambda_est_log)
```

```
## Warning in ks.test.default(log_rr1a, "pexp", rate = 1/lambda_est_log): ties
## should not be present for the Kolmogorov-Smirnov test
```

```
# Display the results of the KS test for log(R-R)
cat("KS Test Results for log(R-R):\n")
```

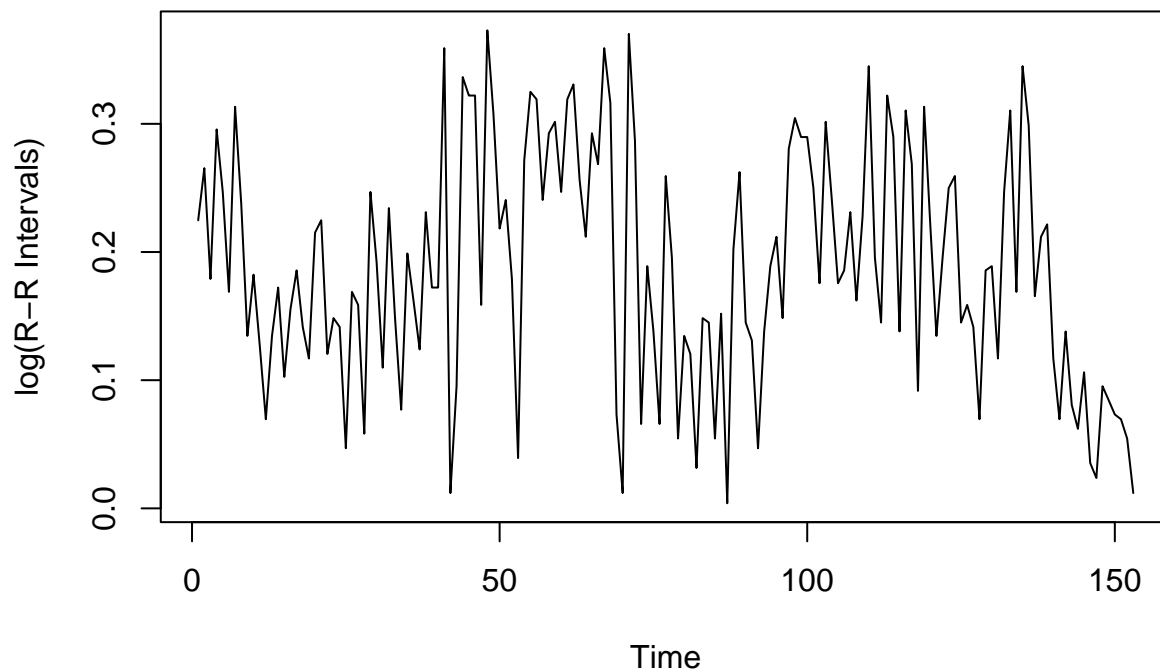
```
## KS Test Results for log(R-R):
```

```
print(ks_result_log)
```

```
##
## Asymptotic one-sample Kolmogorov-Smirnov test
##
## data: log_rr1a
## D = 0.25475, p-value = 4.746e-09
## alternative hypothesis: two-sided
```

```
plot(log_rr1a, type = "l", xlab = "Time", ylab = "log(R-R Intervals)", main = "Heartbeat Data Over Time")
```

Heartbeat Data Over Time (log1a)



```
library(forecast)
```

```
## Registered S3 method overwritten by 'quantmod':
```

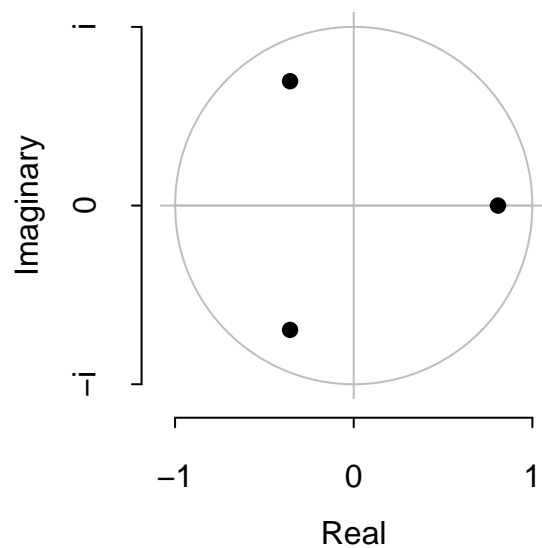
```
##   method      from
```

```
## as.zoo.data.frame zoo
```

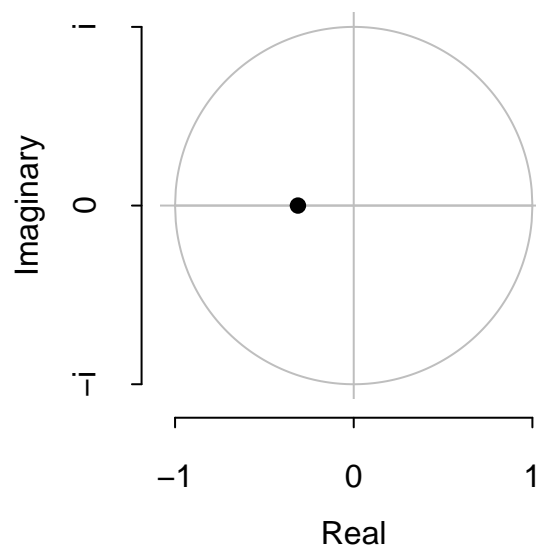
```
res <- auto.arima(log_rr1a)
```

```
plot(res)
```

Inverse AR roots



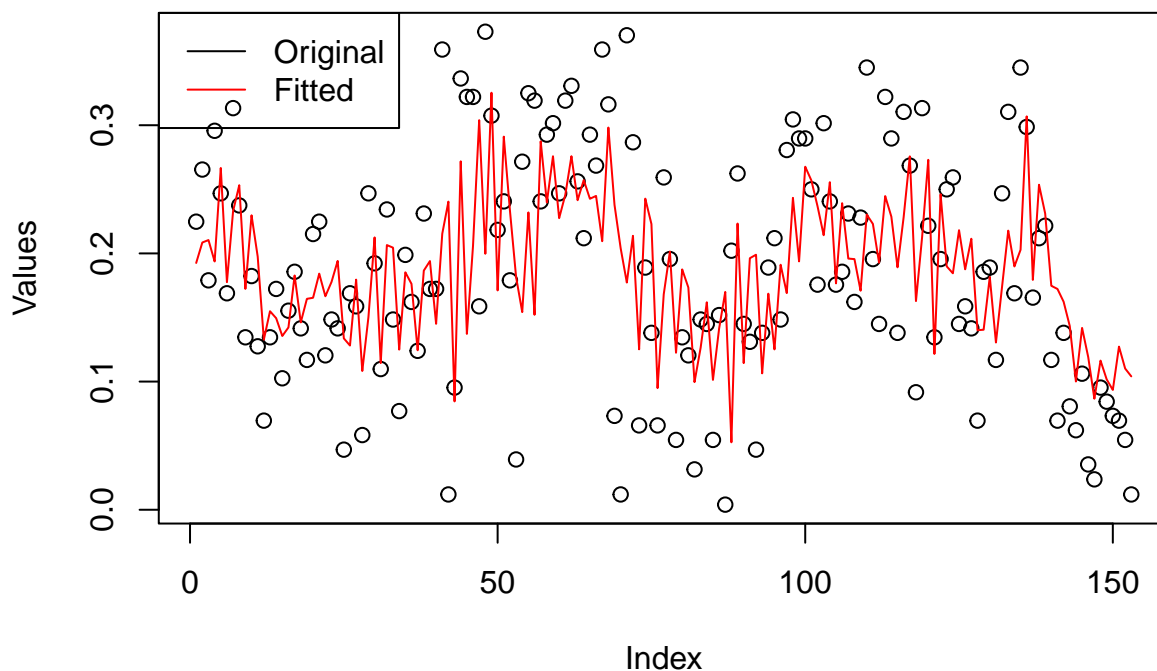
Inverse MA roots



```
# Plot the original time series data
plot(log_rr1a, main = "Original Time Series Data", ylab = "Values")

# Add the fitted values to the plot
lines(fitted(res), col = "red")
legend("topleft", legend = c("Original", "Fitted"), col = c("black", "red"), lty = 1)
```

Original Time Series Data



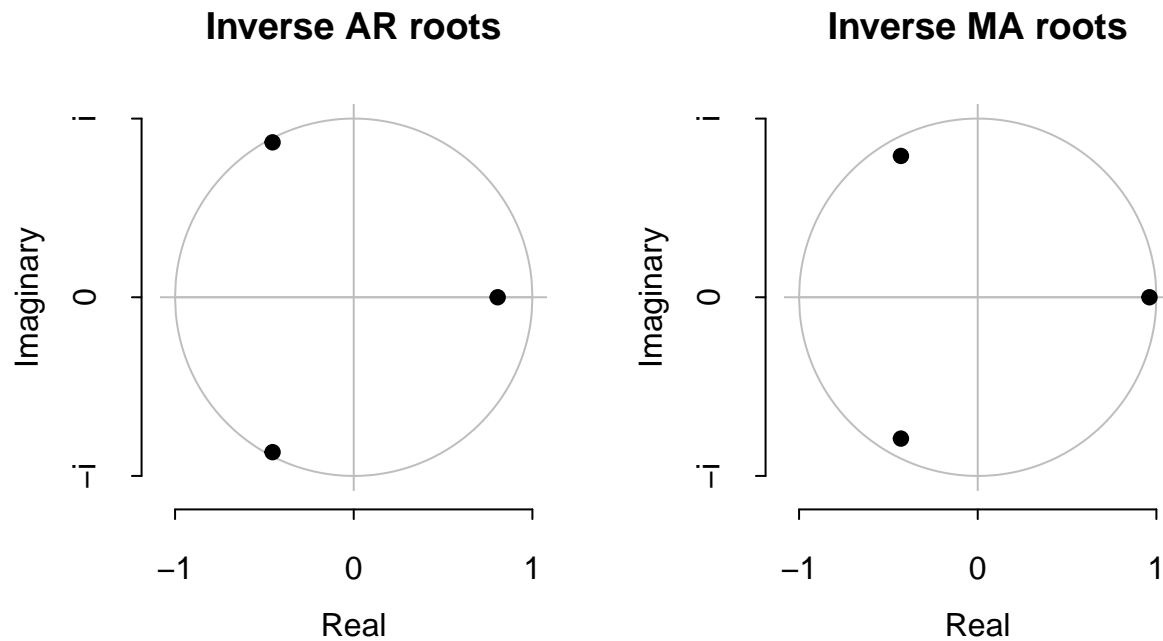
```
summary(res)
```

```
## Series: log_rr1a
## ARIMA(3,0,1) with non-zero mean
##
## Coefficients:
##      ar1      ar2      ar3      ma1      mean
##      0.0949 -0.0360  0.4936  0.3123  0.1846
## s.e.  0.1354   0.0846  0.0719  0.1556  0.0171
##
## sigma^2 = 0.005612: log likelihood = 181.4
## AIC=-350.8   AICc=-350.23   BIC=-332.62
##
## Training set error measures:
##              ME      RMSE      MAE      MPE      MAPE      MASE
## Training set -0.0003254295 0.0736823 0.05801808 -74.22299 95.18879 0.7177017
##              ACF1
## Training set -0.007719581
```

```
# Transform R-R intervals to log(R-R)
log_rr1b <- log(rr1b)

resb <- auto.arima(log_rr1b)
```

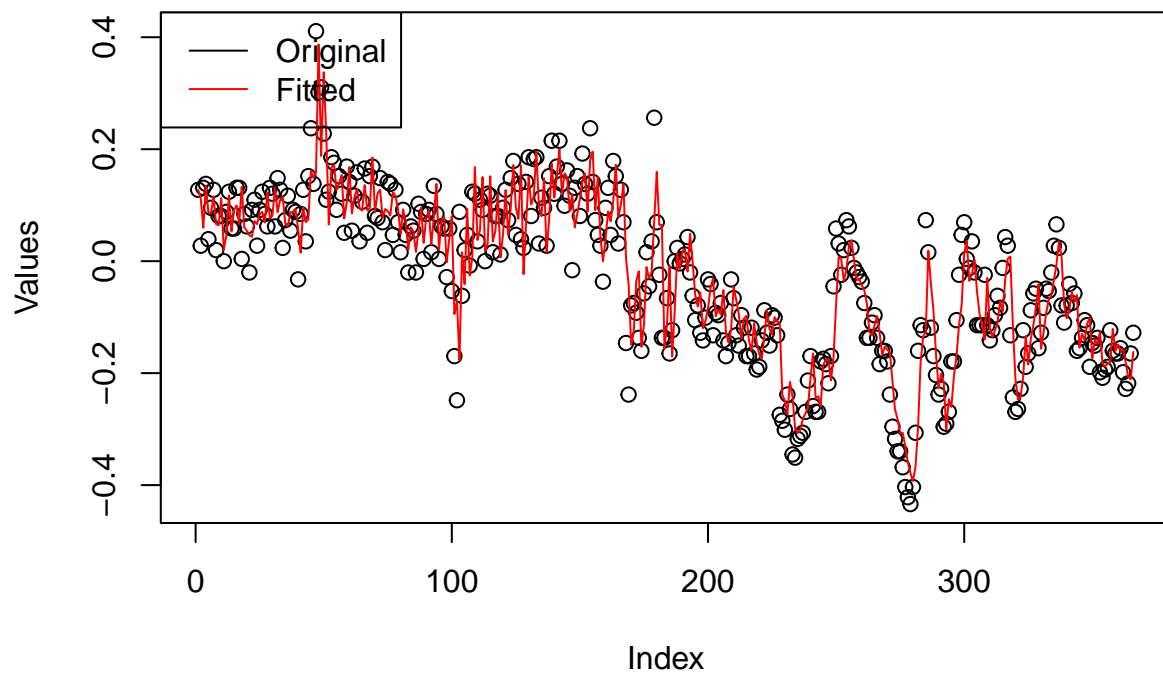
```
plot(resb)
```



```
# Plot the original time series data
plot(log_rr1b, main = "Original Time Series Data", ylab = "Values")

# Add the fitted values to the plot
lines(fitted(resb), col = "red")
legend("topleft", legend = c("Original", "Fitted"), col = c("black", "red"), lty = 1)
```

Original Time Series Data



```
summary(resb)
```

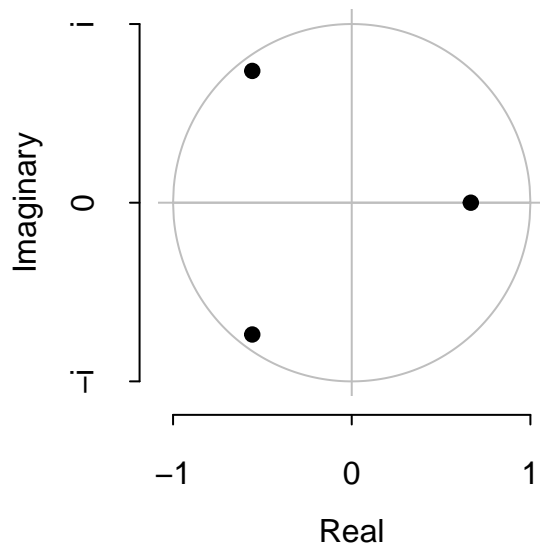
```
## Series: log_rr1b
## ARIMA(3,1,3)
##
## Coefficients:
##          ar1      ar2      ar3      ma1      ma2      ma3
##      -0.1030  -0.2262  0.7720  -0.1015  -0.0158  -0.7791
## s.e.   0.0503   0.0452  0.0447   0.0495   0.0534   0.0484
##
## sigma^2 = 0.003821:  log likelihood = 500.2
## AIC=-986.4   AICc=-986.08   BIC=-959.1
##
## Training set error measures:
##              ME      RMSE      MAE MPE MAPE      MASE      ACF1
## Training set -0.00347232 0.06122252 0.04618269 NaN  Inf 0.808039 0.0182391
```

```
library(forecast)
```

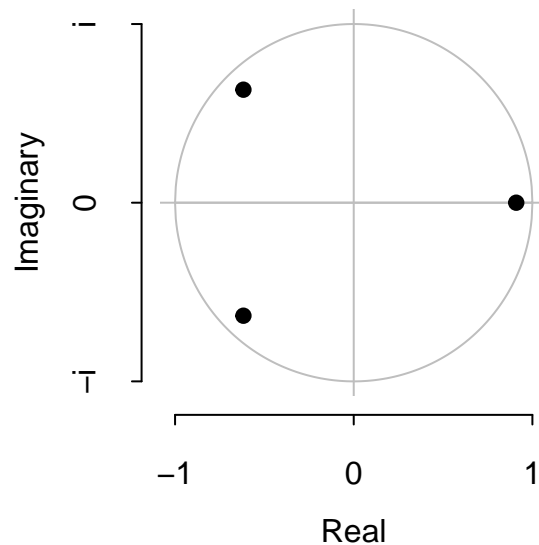
```
res1a <- auto.arima(rr1a)
```

```
plot(res1a)
```

Inverse AR roots



Inverse MA roots



```
# Plot the original time series data
```

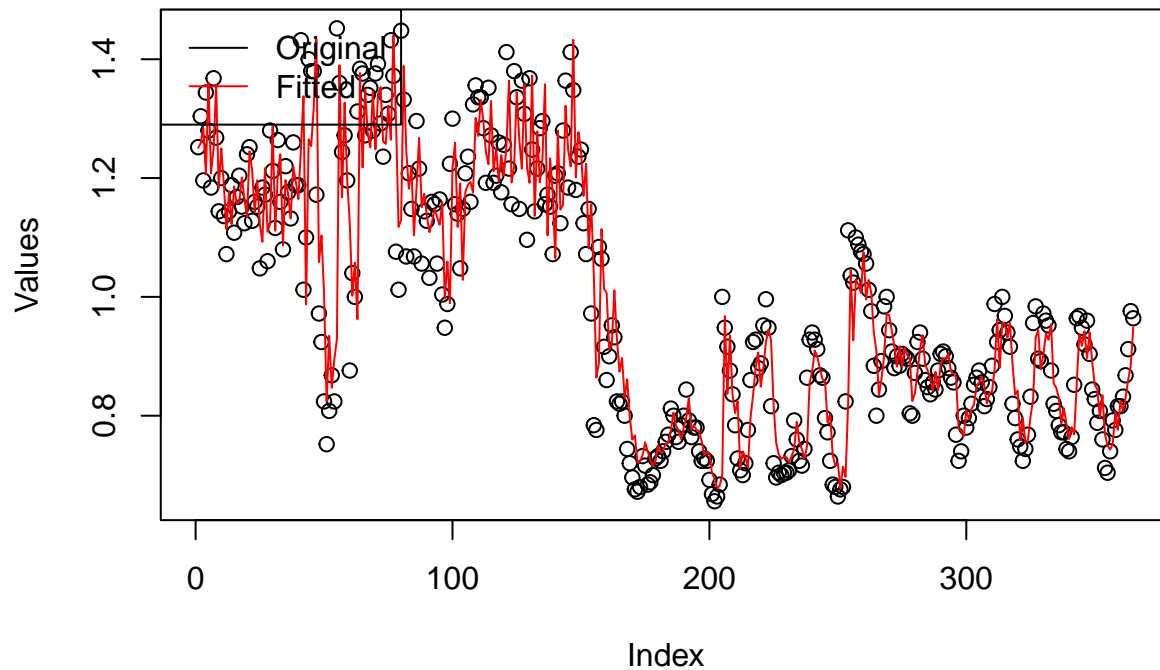
```
plot(rr1a, main = "Original Time Series Data", ylab = "Values")
```

```
# Add the fitted values to the plot
```

```
lines(fitted(res1a), col = "red")
```

```
legend("topleft", legend = c("Original", "Fitted"), col = c("black", "red"), lty = 1)
```

Original Time Series Data

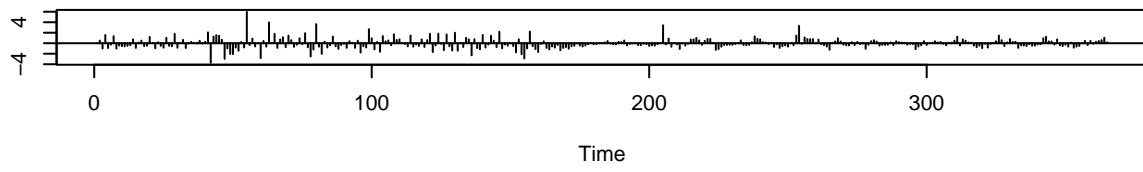


```
summary(res1a)
```

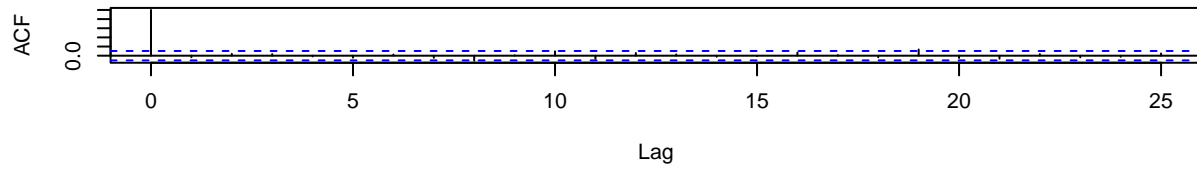
```
## Series: rr1a
## ARIMA(3,1,3)
##
## Coefficients:
##      ar1      ar2      ar3      ma1      ma2      ma3
##    -0.4470 -0.1119  0.5694  0.3254 -0.3419 -0.7106
## s.e.   0.1188   0.0758  0.0692  0.1201   0.0534   0.0820
##
## sigma^2 = 0.007718: log likelihood = 371.19
## AIC=-728.37  AICc=-728.06  BIC=-701.09
##
## Training set error measures:
##              ME      RMSE      MAE      MPE      MAPE      MASE
## Training set -0.003635937 0.08700585 0.06114694 -0.9751233 6.029105 0.9197308
##              ACF1
## Training set -0.03399185
```

```
tsdiag(res1a)
```

Standardized Residuals



ACF of Residuals



p values for Ljung-Box statistic

