300967

MENTAL RETARDATION, X-LINKED, SYNDROMIC 34; MRXS34 Alternative titles: symbols

MENTAL RETARDATION, X-LINKED, SYNDROMIC, MIRCSOF-LANGOUET

TYPE; MRXSML

ORPHA: 466791; DO: 0060817;

Xq13.1 Mental retardation, X-linked, syndromic 34 300967 X-linked 3 NONO 300084

TEXT

A number sign (#) is used with this entry because of evidence that X-linked syndromic mental retardation-34 (MRXS34) is caused by mutation in the NONO gene (300084) on chromosome Xq13.

Description

X-linked syndromic mental retardation-34 is an X-linked recessive neurodevelopmental disordercharacterized by delayed psychomotor development, intellectual disability with poor speech, dysmorphic facial features, and mild structural brain abnormalities, including thickening of the corpus callosum (summary by Mircsof et al., 2015).

Clinical Features

Mircsof et al. (2015) reported 3 unrelated males, aged 15 to 20 years, with syndromic form of intellectual disability. The patients had delayed psychomotor development with poor nasal speech, and a shy, gentle, and cheerful demeanor. They had a slender build, scoliosis or kyphosis, pes planus, and ankylosis of the metacarpophalangeal joint of P1. Facial features included macrocephaly, long face with upslanting palpebral fissures, malar hypoplasia, thin and high nasal root, small open mouth, and narrow high-arched palate with dental crowding. Additional variable features included strabismus, myopia, and hypotonia with poor sucking in infancy, One patient had seizures and 2 had delayed puberty. Brain imaging showed thickening or dysgenesis of the corpus callosum in 2 patients; 1 patient had a Chiari type I malformation and another had a hypoplastic cerebellum.

Reinstein et al. (2016) reported a 17-year-old Ashkenazi Jewish boy of Libyan origin who had developmental delay, macrocephaly, dysmorphic facies, and left ventricular noncompaction (LVNC). At age 1 month, he presented with macrocephaly, axial hypotonia, and head lag. At 8 months, chest x-ray revealed an enlarged cardiac silhouette, and echocardiography showed LVNC. In addition, he exhibited low muscle tone, and all developmental milestones were delayed. At 4 years of age, he was diagnosed with autism and mild intellectual disability, and at age 12, he developed aggressive behavior. Examination at age 16 showed a shy and anxious patient, with verbal dyspraxia, intention tremor, and mild ataxia. He had mild joint laxity, macrocephaly, and dysmorphic features that included long face, prominent nose, wide mouth, thick lips, and short philtrum. Neurologic examination demonstrated general hypotonia, increased patellar and Achilles reflexes without clonus, kyphoscoliosis, and bilateral hallux valgus. Brain imaging showed a thick calvarium, with a short and hypoplastic corpus callosum, but no cerebellar abnormalities. His parents and 2 sibs were healthy.

Scott et al. (2017) studied 3 unrelated Hispanic boys who all exhibited developmental and

intellectual delays, relative macrocephaly, dysmorphic features, and cardiac defects, including LVNC, atrial and ventricular septal defects, patent ductus arteriosus, and patent foramen ovale. MRI of the brain showed corpus callosum defects in 2 of the patients.

Inheritance

The transmission pattern of MRXS34 in one of the families reported by Mircsof et al. (2015) was consistent with autosomal recessive inheritance.

Molecular Genetics

Mircsof et al. (2015) reported 3 different hemizygous mutations in the NONO gene (300084.0001-300084.0003) in 3 unrelated males with MRXS34. One patient inherited the mutation from an unaffected mother; another patient had a de novo mutation and also carried a hemizygous deletion of chromosome 15q13, which may have contributed to the phenotype. The mutations were found by whole-exome sequencing; Western blot analysis of patient cells showed loss of NONO protein. Patient fibroblasts showed increased expression of 2 other splicing genes PSPC1 (612408) and SFPQ (605199), as well as a reduced amplitude of circadian oscillations.

In a 17-year-old Ashkenazi Jewish boy of Libyan origin with developmental delay, macrocephaly, dysmorphism, and LVNC, Reinstein et al. (2016) performed exome sequencing and identified hemizygosity for a de novo splice site mutation in the NONO gene (300084.0004).

Noting that the cardiac phenotype was not reported in the patients described by Mircsof et al. (2015), Reinstein et al. (2016) suggested that LVNC might be part of MRXS34. By exome sequencing in 2 unrelated Hispanic boys with global developmental delay, relative macrocephaly, dysmorphic features, and cardiac anomalies including LVNC, Scott et al. (2017) identified mutations in the NONO gene, the previously identified 1-bp duplication (300084.0002) and the R365X substitution (300084.0003), respectively. In a similarly affected Hispanic male infant, the authors identified a maternally inherited Xq13.1 deletion that included the first 3 coding exons of the NONO gene. The authors noted that all 6 reported patients with mutations in the NONO gene exhibited relative macrocephaly and intellectual disability and/or global developmental delay, and 5 of the 6 had abnormalities of the corpus callosum; however, other features showed greater variability. Given that 2 of the patients with LVNC carried the identical mutation as previously reported patients without cardiac defects, Scott et al. (2017) suggested that loss of NONO function might predispose males to the development of congenital heart defects and LVNC, with the penetrance of these cardiac-related problems being influenced by genetic, epigenetic, environmental, or stochastic differences that affect the expression or function of other cardiac genes or compensatory mechanisms.

Animal Model

Mircsof et al. (2015) found that mice with disruption of the Nono gene had a small cerebellum, impaired spatial memory performance, and increased anxiety compared to control mice. Mutant mouse brains and neurons showed decreased levels of Gabra2 (137140) associated with altered postsynaptic clustering of gephyrin (GPHN; 603930). Expression of Gabra2 rescued the defect in gephyrin clustering. The findings suggested that Nono can regulate Gabra2 levels and thus has a role in regulating inhibitory synaptic biology in the brain, which can affect behavior.

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3.

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OMIM Entry - # 300967 - MENTAL RETARDATION, X-LINKED, SYNDROMIC 34, MRXS34; https://www.omim.org/entry/300967

NON-POU DOMAIN-CONTAINING OCTAMER-BINDING PROTEIN; NONO

NUCLEAR RNA-BINDING PROTEIN, 54-KD; NRB54 p54NRB p54(NRB)

Other entities represented in this entry: NONO/TFE3 FUSION GENE, INCLUDED HGNC Approved Gene Symbol: NONO

Cytogenetic location: Xq13.1 Genomic coordinates (GRCh38): X:71,283,191-71,301,167 (from NCBI)

Xq13.1 Mental retardation, X-linked, syndromic 34 300967.

Description

The NONO gene encodes a protein that belongs to the highly conserved Drosophila behavior/human splicing (DBHS) protein family. This family includes 3 members in mammals: NONO, PSPC1 (612408), and SFPQ (605199). DBHS proteins are nuclear proteins involved in various aspects of RNA metabolism (summary by Mircsof et al., 2015).

Cloning and Expression

Dong et al. (1993) purified and cloned the cDNA of HeLa cell p54nrb, a nuclear protein with 2 RNA recognition motifs and extensive homology to human splicing factor PSF (SFPQ; 605199) and Drosophila NONA/BJ6. By mass spectrometry of proteins isolated from purified HeLa cell nucleoli, Andersen et al. (2002) identified p54(NRB). By SDS-PAGE, p54(NRB) had an apparent molecular mass of about 55.4 kD. Fox et al. (2002) found that p54NRB colocalized with PSP1 (PSPC1; 612408) and PSP2 (RBM14;612409) in nucleoplasmic structures called paraspeckles, which were adjacent to but distinct from SC35 (SFRS2; 600813)-containing splicing speckles.

Mircsof et al. (2015) found expression of the Nono gene in several regions of mouse brain, including cortex and hippocampus. Immunostaining was strong in neurons and granule cells, and absent in astrocytes.

Gene Structure

Peters et al. (1997) determined that the NRB54 gene contains 12 exons ranging in size from 40 to 1,227 bp. The start codon is in exon 3 and the stop codon in exon 12.

Mapping

Brown et al. (1997) examined the expression of 33 X-linked genes in 8 mouse/human somatic cell hybrids that contained either the human active (3 hybrids) or inactive (5 hybrids) X chromosome. They found that the p54nrb gene was expressed only in those hybrids with the active human X. Using a megabase scale from pter to qter devised by Nelson et al. (1995), they noted that the approximate physical position of the gene was 70 Mb from pter. Brown et al. (1997) placed it in almost exactly the same position as the CCG1 gene (TAF2A;

313650), which had been mapped to Xq13-q27, and approximately 2 Mb proximal to PHKA1 (311870), which had been mapped to Xq13. Thus, Xq13 is the likely location. Peters et al. (1997) stated that the AFX1 gene (300033) and the NRB54 gene map to a YAC contig of Xq13.1.

Cytogenetics

NONO/TFE3 Fusion Gene

In papillary renal cell carcinoma tissue (RCCX1; 300854), Clark et al. (1997) identified an X-chromosome inversion inv(X)(p11.2;q12) that resulted in the fusion of the NONO gene to the TFE3 gene (314310).

Gene Function

Fox et al. (2002) found that PSP1, PSP2, and p54NRB relocalized from paraspeckles to the perinucleolar cap region upon transcriptional blockade.

Brown et al. (2005) identified 2 PER1 (602260)-associated factors, NONO and WDR5 (609012), that modulate PER activity. The reduction of NONO expression by RNA interference (RNAi) attenuated circadian rhythms in mammalian cells, and fruit flies carrying a hypomorphic allele were nearly arrhythmic. WDR5, a subunit of histone methyltransferase complexes, augmented PER-mediated transcriptional repression, and its reduction by RNAi diminished circadian histone methylation at the promoter of a clock gene.

Using a high-throughput screen, Amelio et al. (2007) found that endogenous NONO interacted with the CREB (123810) coactivators TORC1 (MECT1; 607536), TORC2 (CRTC2; 608972), and TORC3 (608986) in human embryonic kidney cells. RNAi experiments showed that NONO was necessary for cAMP-dependent activation of CREB target genes in vivo. TORC2 and NONO interacted on cAMP-responsive promoters, and NONO acted as a bridge between the CREB/TORC complex and RNA polymerase II (see 180660).

In a screen to identify factors that interacted with Sox9 (608160) to promote chondrocyte differentiation and Col2a1 (120140) expression in the mouse chondrogenic cell line ATDC5, Hata et al. (2008) identified p54nrb. p54nrb physically interacted with Sox9 and enhanced Sox9-dependent transactivation of the Col2a1 promoter. In ATDC5 cells, p54nrb colocalized with Sox9 in nuclear paraspeckle bodies, and knockdown of p54nrb suppressed Sox9-dependent Col2a1 expression and promoter activity. Overexpression of a mutant p54nrb protein lacking the RNA recognition motifs markedly altered the appearance of paraspeckle bodies and inhibited maturation of Col2a1 mRNA in ATDC5 cells. Mutant p54nrb inhibited chondrocyte differentiation of mesenchymal cells and mouse metatarsal explants, and transgenic mice expressing mutant p54nrb in the chondrocyte lineage exhibited dwarfism associated with impaired chondrogenesis.

Liu et al. (2014) identified a long noncoding RNA, lncUSMYCN (MYCNUT; 615968), that was coamplified with MYCN (164840) in a subset of primary neuroblastomas and neuroblastoma cell lines. Knockdown of lncUSMYCN via small interfering RNA reduced MYCN mRNA expression in a human neuroblastoma cell line, and ectopic expression of lncUSMYCN upregulated exogenous MYCN mRNA and induced cell proliferation. Unlike other lncRNAs, lncUSMYCN did not directly modulate MYCN promoter activity, but it bound directly to NONO, which then increased MYCN expression. High lncUSMYCN or NONO expression in neuroblastoma tissue independently predicted poor patient prognosis. Knockdown of lncUSMYCN reduced tumor growth in neuroblastoma-bearing mice. Liu et al. (2014) concluded that lncUSMYCN and NONO play important roles in regulating MYCN expression and neuroblastoma oncogenesis.

Chaoui et al. (2015) found that p54NRB interacted with SOX10 (602229) to enhance

expression of SOX10 target genes. However, p54NRB did not activate SOX10 target gene transcription alone. Overexpression of p54NRB caused redistribution of SOX10 to nuclear bodies.

Molecular Genetics

Mircsof et al. (2015) reported 3 different hemizygous mutations in the NONO gene (300084.0001-300084.0003) in 3 unrelated males with X-linked syndromic mental retardation-34 (MRXS34; 300967). One patient inherited the mutation from an unaffected mother; another patient had a de novo mutation and also carried a hemizygous deletion of chromosome 15q13, which may have contributed to the phenotype. The mutations were found by whole-exome sequencing; Western blot analysis of patient cells showed loss of NONO protein. Patient fibroblasts showed increased expression of 2 other splicing genes, PSPC1 (612408) and SFPQ (605199), as well as a reduced amplitude of circadian oscillations.

In a 17-year-old Ashkenazi Jewish boy of Libyan origin who had developmental delay, macrocephaly, dysmorphism, and left ventricular noncompaction (LVNC), Reinstein et al. (2016) performed exome sequencing and identified hemizygosity for a de novo splice site mutation in the NONO gene (300084.0004).

In 2 unrelated Hispanic boys with global developmental delay, relative macrocephaly, dysmorphic features, and cardiac anomalies including LVNC, Scott et al. (2017) identified mutations in the NONO gene, the previously identified 1-bp duplication (300084.0002) and the R365X substitution (300084.0003), respectively. In a similarly affected Hispanic male infant, the authors identified a maternally inherited Xq13.1 deletion encompassing all of the NONO noncoding exons, 3 coding exons, and at least 19 kb of sequence upstream of the translational start site. Given that 2 of the patients with LVNC carried the identical mutation as previously reported patients without cardiac defects, Scott et al. (2017) suggested that loss of NONO function might predispose males to the development of congenital heart defects and LVNC, with penetrance influenced by additional factors.

Animal Model

Mircsof et al. (2015) found that mice with disruption of the Nono gene had a small cerebellum, impaired spatial memory performance, and increased anxiety compared to control mice. Mutant mouse brains and neurons showed decreased levels of Gabra2 (137140) associated with altered postsynaptic clustering of gephyrin (GPHN; 603930). Expression of Gabra2 rescued the defect in gephyrin clustering. The findings suggested that Nono can regulate Gabra2 levels and thus has a role in regulating inhibitory synaptic biology in the brain, which can affect behavior.

ALLELIC VARIANTS 4 Selected Examples):

.0001 MENTAL RETARDATION, X-LINKED, SYNDROMIC 34

NONO, 1131G-A rs869025343

In a 17-year-old boy (MCCID1) with X-linked syndromic mental retardation-34 (MRXS34; 300967), Mircsof et al. (2015) identified a de novo hemizygous c.1131G-A transition (c.1131G-A,NM_001145408.1) in the last base of exon 10 of the NONO gene, resulting in a splice site defect. The mutation, which was found by whole-exome sequencing and confirmed by Sanger sequencing, was filtered against the dbSNP (build 130), 1000 Genomes Project, and Exome Variant Server databases, as well as an in-house database. Western blot analysis of patient cells showed little or no NONO protein. The patient also carried a de novo heterozygous deletion of chromosome 15q13.3, which may have contributed to the

.0002 MENTAL RETARDATION, X-LINKED, SYNDROMIC 34

NONO, 1-BP DUP, NT1394 rs869025344

In a 15-year-old boy (MCCID2) with X-linked syndromic mental retardation-34 (MRXS34; 300967), Mircsof et al. (2015) identified a hemizygous 1-bp duplication (c.1394dup, NM_001145408.1) in the last coding exon of the NONO gene, resulting in a frameshift and premature termination (Asn466LysfsTer13). The mutation, which was found by whole exome sequencing and confirmed by Sanger sequencing, was filtered against the dbSNP (build 130), 1000 Genomes Project, and Exome Variant server databases, as well as an in-house database. The mutation was inherited from his unaffected mother. Western blot analysis of patient cells showed little or no NONO protein.

In a 5-year-old Hispanic boy with global developmental delay, relative macrocephaly, and cardiac anomalies including left ventricular noncompaction, Scott et al. (2017) identified the c.1394dupC mutation in the NONO gene. The mutation was not found in his unaffected parents or in the Exome Variant Server, 1000 Genomes Project (phase 3), or ExAC (v.0.3) databases.

.0003 MENTAL RETARDATION, X-LINKED, SYNDROMIC 34

NONO, ARG365TER rs869025345

In a 20-year-old man with X-linked syndromic mental retardation-34 (MRXS34; 300967), Mircsof et al. (2015) reported a hemizygous c.1093C-T transition (c.1093C-T, NM_001145408.1) in the NONO gene, resulting in an arg365-to-ter (R365X) substitution in the coiled-coil domain. The patient was part of the Deciphering Developmental Disorders (DDD) study.

In a 10-year-old Hispanic boy with global developmental delay, relative macrocephaly, and cardiac anomalies including left ventricular noncompaction, Scott et al. (2017) identified the R365X mutation in exon 10 of the NONO gene. The mutation was not found in his unaffected parents or in the Exome Variant Server, 1000 Genomes Project (phase 3), or ExAC (v.0.3) databases. Quantitative RT-PCR analysis of patient lymphoblastoid cell lines revealed an 84% reduction in mRNA compared to that of his parents, consistent with nonsense-medicated decay, and Western blot of the same cell lines did not detect any NONO protein.

.0004 MENTAL RETARDATION, X-LINKED, SYNDROMIC 34

NONO, IVS11DS, G-T, +1 rs1114167441

In a 17-year-old Ashkenazi Jewish boy of Libyan origin who had developmental delay, dysmorphism, and left ventricular noncompaction (MRXS34; 300967), Reinstein et al. (2016) identified a de novo c.1171G-T transversion (c.1171G-T, NM_001145408.1) in intron 11 of the NONO gene, predicted to abolish the splice donor site.

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300967

MENTAL RETARDATION, X-LINKED, SYNDROMIC 34; MRXS34

ORPHA: 466791; DO: 0060817;

INHERITANCE

- X-linked

GROWTH

Other

- Slender build

HEAD & NECK

Head

- Macrocephaly, relative or absolute

Face

- Long face
- Frontal bossing
- Malar hypoplasia

Eves

- Upslanting palpebral fissures
- Strabismus
- Myopia

Nose

- Thin nasal root
- High nasal root
- Deviated nasal septum (in some patients)
- Prominent nose

Mouth

- Open mouth
- Small mouth
- High narrow palate
- Wide mouth (in some patients)

Teeth

- Crowded teeth
- Widely spaced teeth (in some patients)

CARDIOVASCULAR

Heart

- Left ventricular noncompaction
- Atrial septal defect
- Ventricular septal defect
- Patent ductus arteriosus
- Patent foramen ovale
- Right ventricular hypertrophy

ABDOMEN

Gastrointestinal

- Poor sucking in infancy
- Gastroesophageal reflux (in some patients)

GENITOURINARY

Internal Genitalia (Male)

- Cryptorchidism (in some patients)

SKELETAL

Skull

- Macrocephaly, relative or absolute
- Thickened calvarium

Spine

- Scoliosis
- Kyphosis

Hands

- Ankylosis of the metacarpophalangeal joint of P1

Feet

- Pes planus
- Hallux valgus

MUSCLE, SOFT TISSUES

- Hypotonia, neonatal

NEUROLOGIC

Central Nervous System

- Delayed psychomotor development
- Intellectual disability
- Poor language
- No language
- Seizures (in 1 patient)
- Thickened corpus callosum
- Dysgenesis of the corpus callosum
- Cerebellar hypoplasia (in some patients)

Behavioral Psychiatric Manifestations

- Shy, gentle, cheerful demeanor (in some patients)
- Anxious demeanor (in some patients)
- Aggressive behavior (in some patients)

VOICE

- Nasal speech

ENDOCRINE FEATURES

- Delayed puberty (in some patients)

MISCELLANEOUS

- Three unrelated males have been reported (last curated February 2016)

MOLECULAR BASIS

- Caused by mutation in the non-POU domain-containing octamer-binding protein gene (NONO, 300084.0001)

Contributors: Marla J. F. O'Neill - updated: 06/15/2017

Creation Date: Cassandra L. Kniffin: 2/18/2016

NOTE: OMIM is intended for use primarily by physicians and other professionals concerned with genetic disorders, by genetics

researchers, and by advanced students in science and medicine. While the OMIM database is open to the public, users seeking

information about a personal medical or genetic condition are urged to consult with a qualified physician for diagnosis and for

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answers to personal questions.

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