Using R for Step Detection Original 2012-05-25

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Abstract

The codes were run in Mac OSX and Windows XP without problems. We cannot guarantee its performance in other platforms/ systems. Please do report any bugs to the author but unfortunately we cannot guarantee a timely solution.

In the Folder:

- 1. Manual
- 2. GLS-STEP.R
- 3. MyCode.R
- 4. MyCode2.R
- $5.\ msProcess_1.0.7.tar.gz$

Installing the package msProcess

As the package msProcess is no longer actively maintained by its author as of this writing, the archived version of the package is provided with this code. The package is found in the file $msProcess_1.0.7.tar.gz$. This will be a one time process to install the package (or whenever you update R).

- 1. Install the latest version of R from http://www.r-project.org/
- 2. You are provided msProcess_1.0.7.tar.gz along with this code.
- 3. Note down the path where you have saved msProcess_1.0.7.tar.gz
- 4. In R type the following command: msProcess.install()
- 5. Next in R type the following : install.packages("path/filename",repos=NULL,type="source") where path is the path you noted down earlier and filename is $msProcess_1.0.7.tar.gz$
- 6. The above steps should have installed msProcess in R.

1 Step Detection

- 1. Assume all relevant codes, datasets are in the same directory.
- 2. Change the working directory to where you have stored the code GLS-STEP.R using setwd().
- 3. Source the code in by

```
source("GLS-STEP.R")
```

You can check if the codes are sourced in by ls() 4. Input the sample data set by RNA<-read.table("RNAtrace1.txt",sep="\t",header=TRUE)</pre> 5. You can see the first few lines of the data by head(RNA) 6. Set window sizes by w < -c(seq(10,90,by=10),seq(100,1000,by=25))7. Set y<-RNA[,1] 8. Set times<-RNA[,2] 9. Plot your trace and check plot(y,type="1") 10. Run the command check.packages() to install/ load required R packages. 11. Getting the η statistics: chpt0<-get.zstat(y,times,w)</pre> 12. Doing the step Fit: chpt1<-ChPt(chpt0)</pre> 13. Plot Step: plot.step(chpt0,chpt1,type="bic") You can choose type = "aic" for an AIC fit. 14. To get the results: RNA.RESULT1<-get.results(chpt1,type="bic")</pre>

You can choose type = "aic" for an AIC fit. Check results by

```
RNA.RESULT1
```

15. If you want to plot to a pdf file:

```
plot.step(chpt0,chpt1,type="bic",pdfname="MyRNAPlot.pdf")
```

16. The command

As the names imply these are - Step size estimates, the variance-covariance matrix of the step estimates, the t-statistic for the steps for hypothesis testing, the fitted step, the residuals from the fit, the variance of the noise, the p-value of the steps, the aic value, the bic value, The AR-order of the noise and the AR coefficients.

17. One can look at these value as follows. So to see the AR order of the noise one types

```
MyModel$AR-order
and to view the BIC value
myModel$bic

18. To look at the ACF and PACF of residuals
acf(myModel$res$)
and
pacf(myModel$res$)
```

2 Step Detection - For Large Traces

It can be very time consuming. We suggest that you run the code in the background in the UNIX shell. Save the following code in say MyCode.R

```
source("GLS-STEP.R")
check.packages()
RNA<-read.table("RNAtrace1.txt",sep="\t",header=TRUE)
w<-c(seq(10,90,by=10),seq(100,1000,by=25))
Y<-RNA[,1]
times<-RNA[,2]
chpt0<-get.zstat(y,times,w)
chpt1<-ChPt(chpt0)
plot.step(chpt0,chpt1,pdfname="RNAFIG1.pdf")
RNA.RESULT1<-get.results(chpt1)
write.table(RNA.RESULT1,file="RNA-RESULT1.txt",sep="\t",header=TRUE)
myModel<-get.model(chpt1,type="bic")
save(myModel,file="All-results-1.Rdata")
#load("All-results-1.Rdata")</pre>
```

- 1. Open X11
- 2. Change the working directory to where the data, codes are located
- 3. Type in

```
nohup R CMD BATCH MyCode.R myLOG.txt &
```

4. For Windows try the following. Make sure you have the corrects paths.

```
"PATH\R.exe" CMD BATCH --vanilla --slave "PATH\MyCode.R"
```

5. Once the analysis is done check RNA-RESULT1.txt for the results and RNAFIG1.pdf for the plot.

3 To Run Multiple Traces

To run more than one trace at a time - create a folder say MyTraces and place all your traces in this folder. Save the following code in say MyCode2.R

```
source("GLS-STEP.R")
 check.packages()
 setwd("MyTraces FOLDER") # Set the My traces Folder Directory path
 file.names<-system("ls",intern=TRUE)</pre>
                                         # This should get the names of the trace files in the folder.
 for(i in 1:length(file.names)){
  RNA<-read.table(file.names[i],sep="\t",header=TRUE)</pre>
  w < -c(seq(10,90,by=10),seq(100,1000,by=25))
  Y<-RNA[,1]
  times<-RNA[,2]
  chpt0<-get.zstat(y,times,w)</pre>
  chpt1<-ChPt(chpt0)</pre>
  plot.step(chpt0,chpt1,pdfname=paste("RNAFIG-",i,".pdf",sep=""))
  RNA.RESULT1<-get.results(chpt1)</pre>
  write.table(RNA.RESULT1,file=paste("RNA-RESULT-",i,".txt",sep=""),sep="\t",header=TRUE)
  myModel<-get.model(chpt1,type="bic")</pre>
  save(myModel,file=paste("myModel-",i,".Rdata",sep=""))
  # This is saves as a R file .Rdata, you can load it into R by load("myModel-1.Rdata")
}
```

- 1. Open X11
- 2. Change the working directory to where the data, codes are located
- 3. Type in

```
nohup R CMD BATCH MyCode2.R myLOG.txt &
```

4. For Windows try the following. Make sure you have the corrects paths.

```
"PATH\R.exe" CMD BATCH --vanilla --slave "PATH\MyCode2.R"
```

5. Once the analysis is done check RNA-RESULT1.txt for the results and RNAFIG1.pdf for the plot and other files.