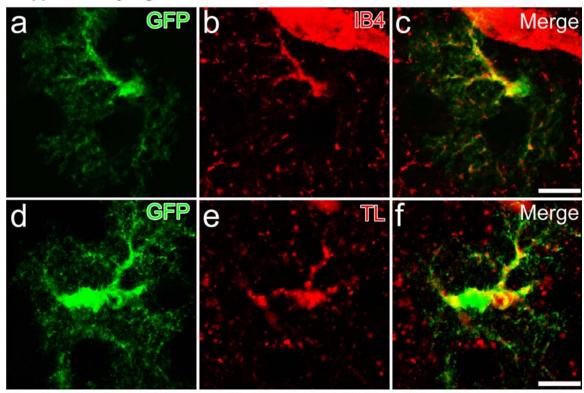


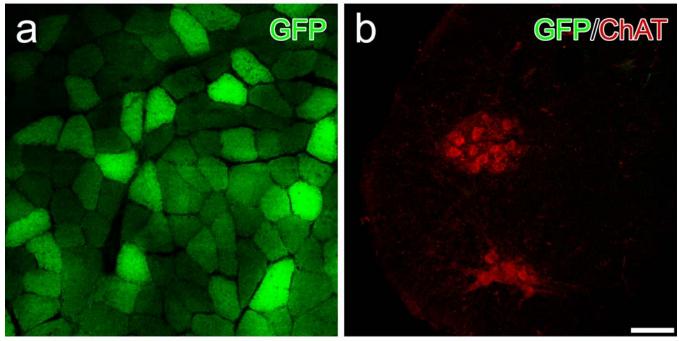
Supplementary Figure 1 Intravenous injection of AAV9 leads to widespread and long-term neonatal spinal cord transduction in lumbar motor neurons. Z-series confocal microscopy showing GFP-expression in 21-day-old mice that received 4x10¹¹ particles of scAAV9-CB-GFP intravenous injections on postnatal day-1. Z-stack images of GFP (a), ChAT (b), GFAP (c) and merged (d) demonstrating persistent GFP-expression in motor neurons and astrocytes (d) for at least three-weeks following scAAV9-CB-GFP injection. Orthoganol views (e-g) further confirm co-localization between GFP and ChAT labeled cells (e-f) and a GFP and GFAP labeled cell (g). Scale bar, 20µm (d).



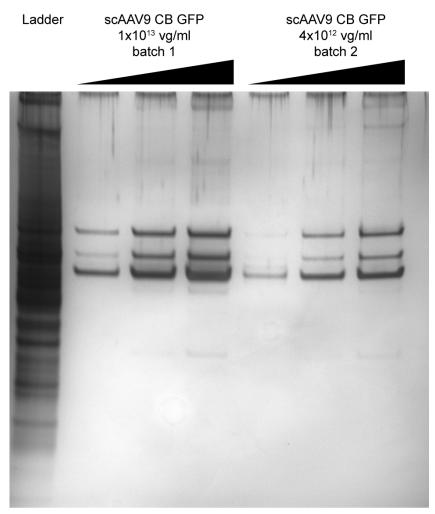
Supplementary Figure 2 Intravenous injection of scAAV9 CB GFP resulted in GFP expression within microglia 21 days after postnatal day-1 administration. Immunofluorescent detection of GFP (\mathbf{a} , \mathbf{d}) and isolectin B₄ (\mathbf{b}) or tomato lectin (\mathbf{e}) mediated labeling demonstrates AAV9 transduction of microglia (\mathbf{c} , \mathbf{f}) in the neonate. Scale bars, 10µm (\mathbf{c} and \mathbf{f})

Supplementary Figure 3 b a e h k

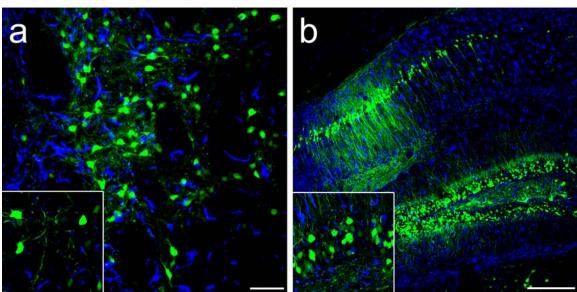
Supplementary Figure 3 Co-localization of GFP positive cells with GAD67. Immunohistochemical detection of GFP (a,d,g,j) and GAD67 (b,e,h,k) expression within select regions of mouse brain 21-days following postnatal day-1 injection of 4x10¹¹ particles of scAAV9-CB-GFP. Merged images (c,f,i,l) and insets show limited co-localization of GFP and GAD67 signals in the cingulate gyrus (a-c), the dentate gyrus (d-f) and the hippocampus (g-i), but numerous GFP/GAD67 Purkinje cells within the cerebellum(I). Scale bars, 100µm (c), 50µm (a-b,d-l)



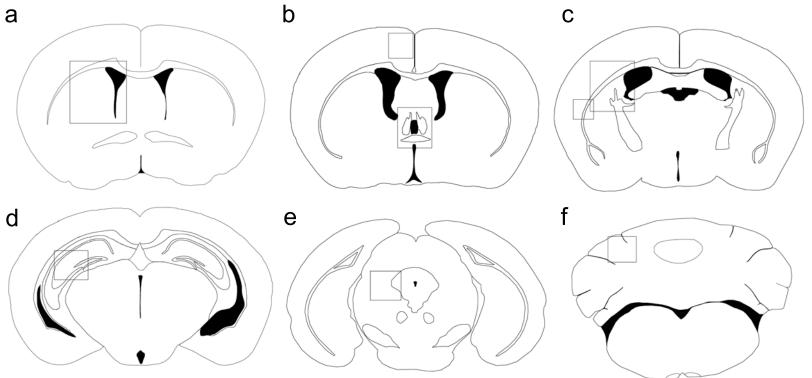
Supplementary Figure 4 Intramuscular injection of scAAV9 CB GFP resulted in robust GFP expression in the injected muscle (a) but a lack of expression within innervating lower motor neurons (b). Tissues were harvested two-weeks post intramuscular injection of $5x10^{10}$ particles of scAAV9 CB GFP. Scale bar, $100\mu m$ (b)



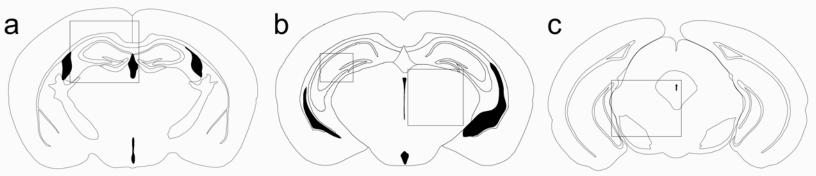
Supplementary Figure 5 Gel Electrophoresis and silver staining of various AAV9-CB-GFP vector preparations demonstrates high purity of research grade virus utilized in studies. Shown are 2 vector batches at varying concentrations demonstrating the predominant 3 viral proteins (VP); VP1, 2, 3 as the significant components of the preparation. 1µl, 5µl, and 10µl were loaded of each respective batch of virus.



Supplementary Figure 6 Direct injection of scAAV9-CB-GFP into the adult brain demonstrates predominant neuronal transduction. Injection of virus into the striatum (**a**) and hippocampus (**b**) resulted in the familiar neuronal transduction pattern as expected (*n*=3). Co-labeling for GFP (green) and GFAP (blue) demonstrate a lack of astrocyte transduction in the injected structures with significant neuronal cell transduction. Insets of striatum (**a**) and dentate gyrus (**b**) show high magnification merged images of GFP (green) and GFAP (blue) labeling futher confirming a lack of astrocyte transduction. Scale bars, 50µm (**a**), 200µm (**b**)



Supplementary Figure 7 Diagrams of coronal sections throughout the mouse brain corresponding to the approximate locations shown in **figure 3(a-h)** for postnatal day-1 injected neonatal mouse brains. The box in **(a)** corresponds to the location of **(Fig. 3a)**. The smaller box in **(b)** corresponds to **(Fig. 3b)** and the larger box to **(Fig. 3c)**. The larger box in **(c)** corresponds to **(Fig. 3d)** while the smaller box in **(c)** represents **(Fig. 3e)**. Finally, **(d-f)** correspond to **(Fig. 3 f-h)** respectively.



Supplementary Figure 8 Diagrams of coronal sections throughout the mouse brain corresponding to the approximate locations shown in (**Fig. 5m-p**). The box in (**a**) corresponds to the location shown in (**Fig. 5m**). The smaller box in (**b**) corresponds to (**Fig. 5n**) and the larger box to (**Fig. 5o**). Finally, the approximate location of (**Fig. 5p**) is shown in (**c**).