

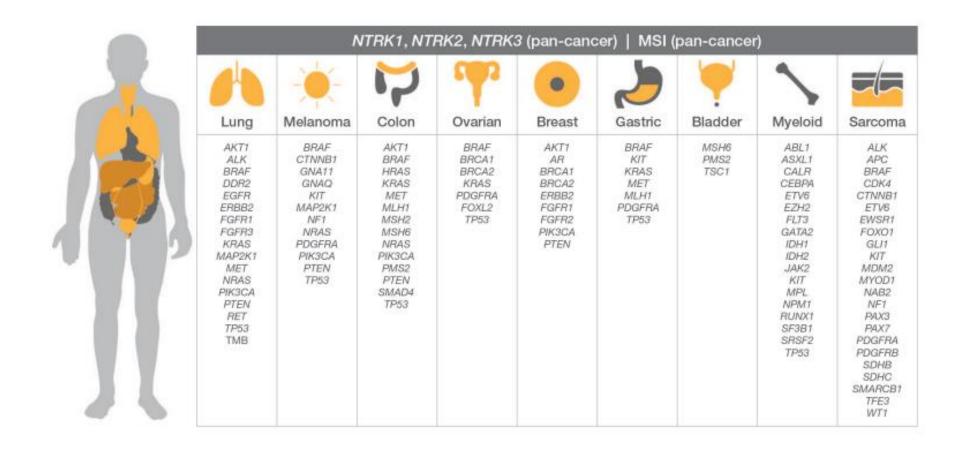


# TruSight<sup>™</sup> Oncology 500 Assay (TSO 500)

Target Specifications
523 genes (1.94 Mb) Encompasses TruSight Tumor 170, MSK-IMPACT™ and FoundationOne®
TruSight Oncology with unique molecular identifiers (UMIs)
5% LOD for DNA variants >96% analytical sensitivity; >99.99% analytical specificity
40 ng DNA from FFPE samples
Single nucleotide variants (SNVs), indels, tumor mutational burden (TMB), microsatellite instability (MSI)
Local Docker-Based Variant Calling Software
8 samples per NextSeq™ run
Variant file with all passing, non-synonymous variants
NextSeq , NextSeq 550Dx



# Some targets covered by TSO500:





# TSO500: Software TS Training overview

- TSO 500 Assay
  - Library Prep and Enrichment Workflow
  - Overview of library construction
- Sequencing Recommendations
- Analysis: TSO 500 Local App
  - Docker, Singularity container platforms
  - Field support model
  - Output folders and data metrics
  - Tumor Mutational Burden score
  - Microsatellite Instability score
  - TST 170 Local App for RNA Analysis
- Other options and future plans
- Partnership with PierianDx

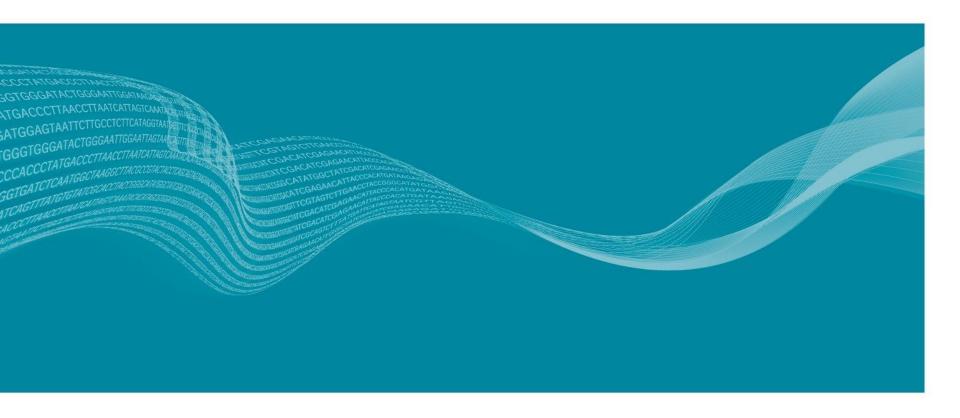








# **TSO 500 Library Preparation:** Quick review



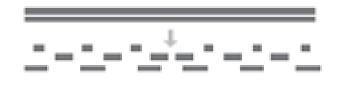


# **Library Prep Step 1: Fragment genomic DNA**



# Fragment gDNA

Hands-on: 10 minutes Total: 120 minutes Reagents: TEB



- Covaris focused-ultrasonicator shears genomic DNA
- Generate 90 250 bp dsDNA fragments with 3' and 5' overhangs

# Library Prep Step 2: End Repair and A-Tailing

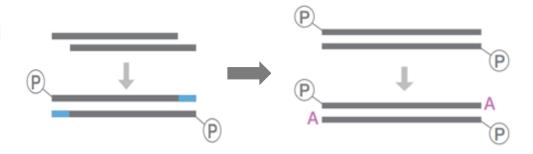


## End Repair and A-Tailing

Hands-on: 10 minutes

Total: 70 minutes

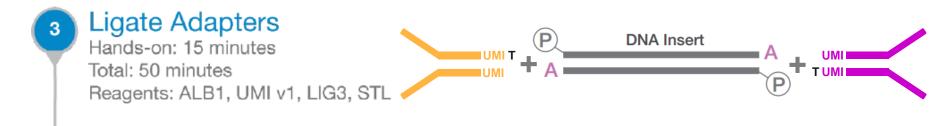
Reagents: ERA1-A, ERA1-B





# **Library Prep Step 3: Ligate Adapters to DNA fragments**

Unique Molecular Identifiers



# Library Prep Step 4: Ligation Clean Up

Purify DNA fragments with Sample Purification Beads

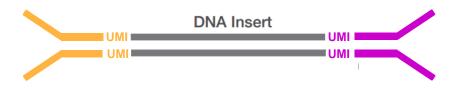


## Ligation Clean Up

Hands-on: 40 minutes

Total: 50 minutes

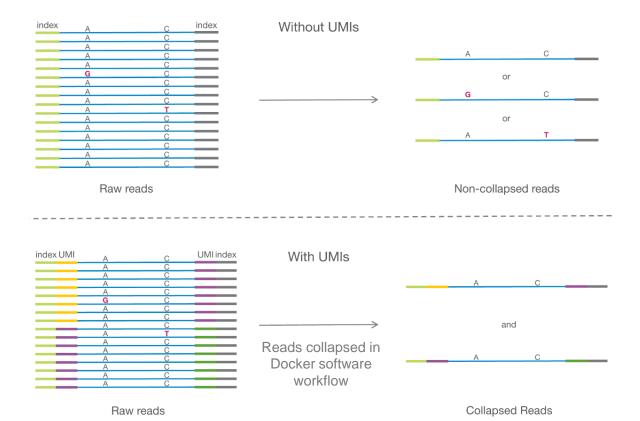
Reagents: SPB, RSB, 80% EtOH





# **Unique Molecular Identifiers (UMIs)**

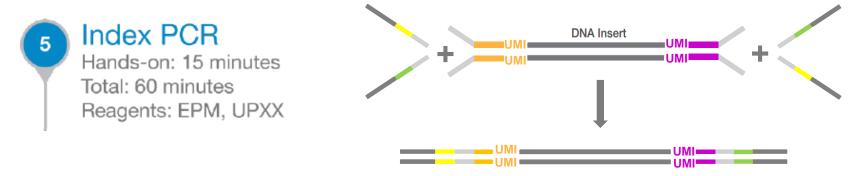
- Every individual fragment is tagged for identification
- Reads derived from the same fragment have the same sequence
- UMIs enable Error Correction: distinction between errors and <u>variants</u>
- Reduce error rate to ≤ 0.007%



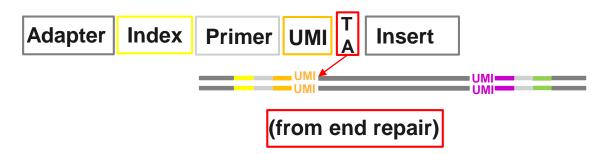


# **Library Prep Step 5: Index PCR**

Amplify DNA and add indexed adapters for sample multiplexing

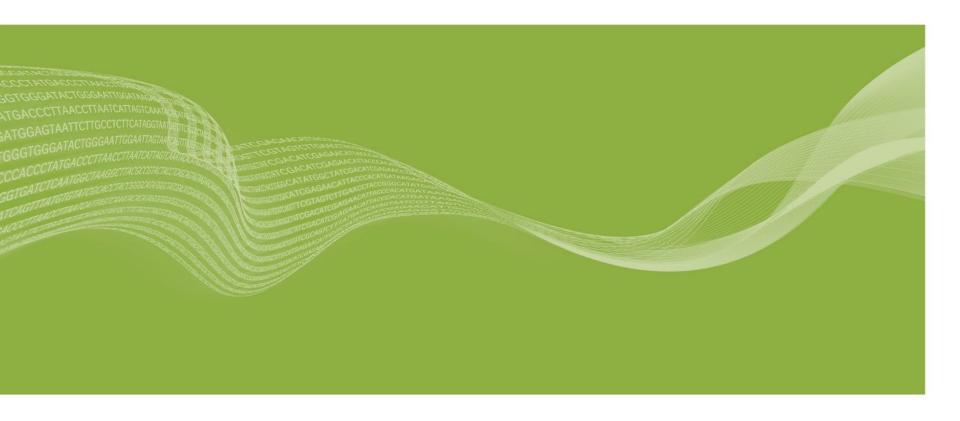


# **Library Prep Step 5: Final Library**





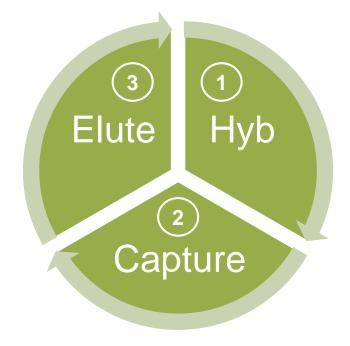
# **TSO 500 Enrichment**

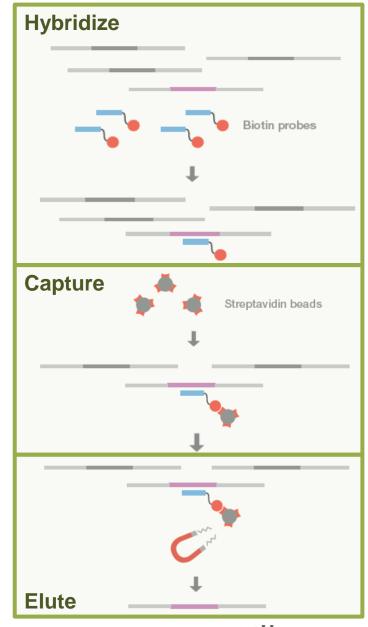




# **Enrichment**

 TSO 500 target probes capture and enrich library fragments that contain targeted regions







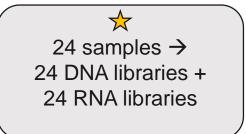
**TruSight Oncology 500 Kits** 

DNA-only or DNA+RNA reagents

- TSO 500 DNA Kit (20028213)
  - Library Prep and Enrichment Reagents
  - No NextSeq core reagents
  - 16 indexes, 48 samples



- TSO 500 DNA Kit, plus NextSeq kit (20028214)
  - Library Prep and Enrichment Reagents
  - 16 indexes, 48 samples
- TSO 500 DNA/RNA Kit (20028215)
  - Library Prep and Enrichment Reagents
  - No NextSeq core reagents
  - 16 indexes, <mark>24</mark> samples☆

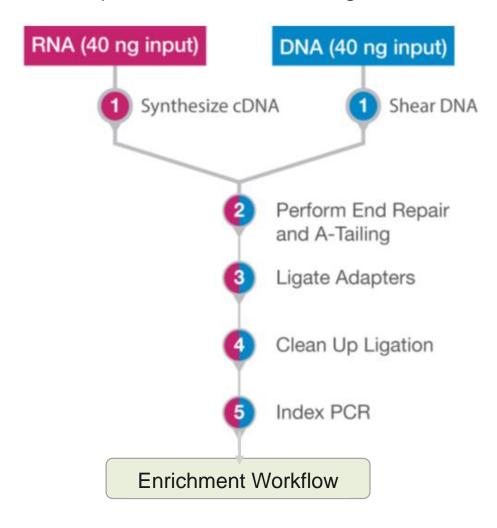


- TSO 500 DNA/RNA Kit, plus NextSeq kit (20028216)
  - Library Prep and Enrichment Reagents
  - 16 indexes, <mark>24</mark> samples★



### TSO 500 DNA + TST 170 RNA Workflow Overview

- Extract RNA and process in parallel with DNA
- Detect fusions and splice variants from 55 genes





# **TSO 500 Sequencing**





# **Recommended Sequencing Configuration**

- Compatible platforms:
  - NextSeq 500/550
  - NextSeq 550Dx (RUO Mode)
  - Other platforms have not been tested and are not supported
  - Possible NovaSeq support in future
- 2 x 101 Run in Standalone mode
  - BaseSpace uploads not supported
- Paired-End (required), Dual Index (required)
- Minimum of 3 libraries for sufficient diversity
  - UPXX index primer sets for low-plexity pooling provided in TSO 500 Reference Guide
- 8 <u>samples</u> maximum for optimal coverage
  - 8 DNA libraries + 8 RNA libraries
  - 80M PE reads per library

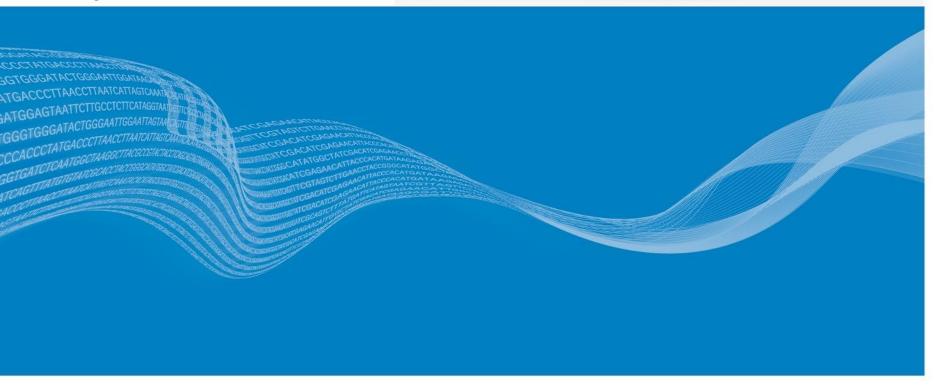






# Prepare library Sequence Analyze data

# **TruSight Oncology 500 Analysis**





# **TSO 500 Local App Agenda**

### TSO 500 Local App

- Overview
- Installation
- Launching analysis

### Output folders and data metrics

- Tumor Mutational Burden score
- Microsatellite Instability score
- TST 170 RNA Analysis



# **TSO 500 Local App**

### Overview



### A software package that reports:

- Single Nucleotide Variants (SNVs)
- Small Insertions and Deletions (indels)
- Tumor Mutational Burden score (TMB)
- Microsatellite Instability score (MSI)



### Analyzes BCL files from a NextSeq:TSO 500 run folder

- NextSeq 500/550 or NextSeq 550Dx (RUO Mode)
- Requires Sample Sheet (on support site)
- FASTQ analysis workflow available

### Output files:

- Variant Call Files (VCFs)
- Metrics Report
- Biomarker Reports with TMB score and MSI score



# TSO 500 Local App: Docker or Singularity



### What is Docker?

An open platform for easily developing and running software

- Docker engine is installed on a Linux server (CentOS)
- Applications are packaged in containers and run on Docker
- Uses a command line interface

### What is Singularity?

Another open platform option for TSO 500

### What is a Container?

A standardized unit of software

- A standalone, executable software package
- Includes everything needed to run the app
- Depends on elements of the OS

# App App Docker Host Operating System

Containerized Applications

Source: www.docker.com/resources/what-container



# **TSO 500 Local App Installation**



### How to access the TSO 500 local app?

- Install Docker or Singularity on CentOS 7.3
- Download TSO 500 app bundle with HTTP client (Curl, Wget)
  - Container Resources folder includes manifest, sample sheet template, and hg19 genome
  - Illumina Technical Support will provide link to customer by email
- Install TSO 500 per User Guide instructions

### User Requirements

- Basic user knowledge of:
  - Linux operating system
  - Docker software container platform
- Confirm OS meets computing requirements in User Guide
  - ILMN can recommend server if needed



# TSO 500 Local App: Launching the Analysis



- Instructions and command line entries provided in User Guide
- One analysis at a time recommended
  - Wait for any running TSO 500 container to complete before launching new analysis
- Start software with TruSight\_Oncology\_500.sh script on command line as described in User Guide

Resource:

**TruSight Oncology 500 Local App User Guide** 



# TSO 500 Local App: Required Inputs

Docker analysis from BCL files



Paths in command line must be absolute paths:

- Path to the TruSight Oncology 500 Resources folder
- Path to an empty output folder for analysis results
  - User must create this folder first
- Path to the sequencer run data folder -OR- a FASTQ folder
- Path to SampleSheet.csv file if not located in run folder
  - Example on TSO 500 Support page
  - Must be named "SampleSheet.csv"
  - Specific instructions provided in Local app User Guide

Resource:

**TruSight Oncology 500 Local App User Guide** 



# **TSO 500 SampleSheet.csv**



[Header]								
IEMFileVersion	4	,						
Investigator Name	User Name							
Experiment Name	Experiment							
Date	10/3/2018	i	Valid Co	- ala C	N- a at a war			
Workflow	GenerateFASTQ ద		Valid Sa	ampie c	Sheet exar	npies:		
Application	NextSeq FASTQ Only		•	Includ	ded in TS0	2 500 la	ncal ann	
Assay							July 1	
Description					urces fold			
Chemistry	Default		•	Provi	ded on TS	SO 500	support pa	age
[Reads]			<ul> <li>Nan</li> </ul>	ne "San	npleSheet	csv"		
101			• Add	to run	folder or p	rovide	path	
101								
[Settings]								
Adapter	AGATCGGAAGAGCACACGTCTGAACTCCAGTCA							
AdapterRead2	AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGT		Base 8 is f	rom er	nd repair.			
Read1UMILength	7	<b>☆</b>						
Read2UMILength	7		skipped for	allaly	515		J	
Read1StartFromCycle	9							
Read2StartFromCycle	9	$\Rightarrow$			Delete	unused	indexes	
						m temp		
[Data]					110	III temp	laic	
Sample_ID 🗙	Sample_Name	Sample_	Plate Sample_Well	Index_ID	index	I7_Index_	index2	I5_Index_ID
Test_Sample_UP01				UP01	TCCGGAGA	D702	AGGATAGG	D503
Test_Sample_UP02	Sample_Name:			UP02	CTGAAGCT	D707	TCAGAGCC	D504
Test_Sample_UP03	Blank or identical to			UP03	CGTAGCTC	D717	CATCCGAA	D509
Test_Sample_UP04				UP04	GAATTCGT	D706	TTATGAGT	D510
Test_Sample_UP05	Sample_ID			UP05	AGCGATAG	D712	ACGAATAA	D513
Test_Sample_UP06				UP06	GCGATTAA	D724	GATCTGCT	D515
Test_Sample_UP07				UP07	ATTCAGAA	D705	AGGCTATA	D501
Test_Sample_UP08				UP08	GAATAATC	D713	GCCTCTAT	D502

# TSO 500 Local App: Launch Analysis

Example launch from run folder with BCL files



### At command prompt enter:

```
{APP_PATH}/{Version}/TruSight_Oncology_500.sh --user=$UID --remove \
--analysisFolder /full/path/to/output/analysisFolder \
--resourcesFolder /{RESOURCE_FOLDER}/resources \
--runFolder /full/path/to/runFolder
```

### Variables:

- APP\_PATH: /scratch
- Version: 1.3.0
- /full/path/to/output/analysisFolder: /scratch/tso500\_180914\_NB501433\_0065\_AH3CVYBGX9
- RESOURCE FOLDER: /scratch/1.3.0/
- /full/path/to/runFolder: /scratch/180914\_NB501433\_0065\_AH3CVYBGX9

### Complete example:

/scratch/1.3.0/TruSight\_Oncology\_500.sh --user=\$UID --remove --analysisFolder /scratch/tso500\_180914\_NB501433\_0065\_AH3CVYBGX9 --resourcesFolder /scratch/1.3.0/resources --runFolder /scratch/180914\_NB501433\_0065\_AH3CVYBGX9

# TSO 500 Local App: Required Inputs

Launch analysis from BCL files



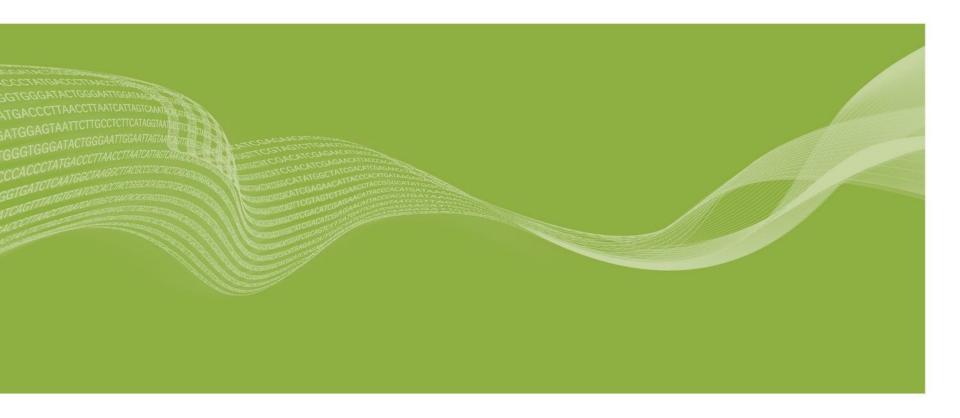
```
login as: oncofas
oncofas@10.31.34.51's password:
Last login: Tue Nov 27 23:14:57 2018 from ussd-olw-018537.illumina.com
[oncofas@EHV-docker2 ~]$ [

/scratch/1.3.0/TruSight_Oncology_500.sh --user=$UID --remove --analysisFolder
/scratch/tso500_180914_NB501433_0065_AH3CVYBGX9 --resourcesFolder
/scratch/1.3.0/resources --runFolder /scratch/180914_NB501433_0065_AH3CVYBGX9
```

 Analysis can also be started from FASTQs, assuming FASTQs have been generated with appropriate UMI settings (directions in user guide).

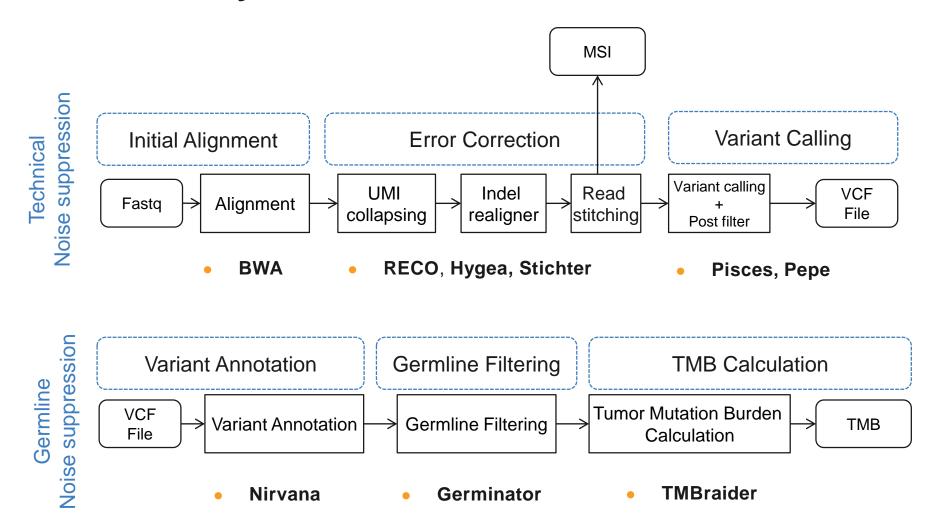


# TSO 500 Local App Output





# **TSO500** Analysis workflow





# TSO 500 Local App Analysis Output Folder



- Logs\_Intermediates
  - Annotation
  - CollapsedReads
  - Contamination
  - CvAnnotation
  - CvDnaAlignment
  - DnaAlignment
  - DnaQCMetrics
  - DnaRealignment
  - FastqGeneration
    - Sample1
      - Sample1\_S1\_L001\_R1\_001.fastq.gz
      - Sample1\_S1\_L001\_R2\_001.fastq.gz
      - Sample1\_S1\_L002\_R1\_001.fastq.gz
      - Sample1\_S1\_L002\_R2\_001.fastq.gz
      - Sample1 S1 L003 R1 001.fastq.qz
      - Sample1 S1 L003 R2 001.fastq.qz
      - Sample1\_S1\_L004\_R1\_001.fastq.gz
      - Sample1\_S1\_L004\_R2\_001.fastq.gz
  - FastqValidation
  - HypothesisCaller

- indelRealignment
- i MarkDuplicates
- MetricsAggregation
- Msi
- ResourceVerification
- RunQC
- SampleSheetValidation
- SmallVariantFilter
- StitchedReads
- StitchedReads
- i Tmb
- VariantCaller
- TruSight-Oncology-500-pipeline-<datetime>.log
- Results
  - </
  - MetricsReport.tsv



# **Results Folder: Metrics Report**

### **Metrics Report**



Combined key analysis metrics in a .tsv file

LSL and USL guidelines provided per sample metric





# MetricsReport.tsv

Con	

TruSight Oncology 500 - Metrics Repo	rt				
For Research Use Only. Not for use in	diagnostic procedures				
[Header]					
Report Date	1/4/2019				
Report Time	22:22:11				
Pipeline Version	1.3.1.3				
[Run Metrics]					
Metric (UOM)	LSL Guideline	USL Guideline	Value		
PCT_PF_READS (%)	80	NA	90.2		
PCT_Q30_R1 (%)	80	NA	90.4		
PCT_Q30_R2 (%)	80	NA	87.3		
[DNA Library QC Metrics]					
Metric (UOM)	LSL Guideline	USL Guideline	CCL-DNA	FFPE-Lung-DNA	HD753-DNA
CONTAMINATION_SCORE (NA)	0	3106	223	7535	30409
CONTAMINATION_P_VALUE (NA)	0	0.049	1	0	0.416
[DNA Library QC Metrics for Small Va	riant Calling and TMB]				
Metric (UOM)	LSL Guideline	USL Guideline	CCL-DNA	FFPE-Lung-DNA	HD753-DNA
MEDIAN_INSERT_SIZE (bp)	70	NA	104	108	118
MEDIAN_EXON_COVERAGE (Count)	150	NA	390	483	602
PCT_EXON_50X (%)	90	100	98.6	99.3	99
[DNA Library QC Metrics for MSI]					
Metric (UOM)	LSL Guideline	USL Guideline	CCL-DNA	FFPE-Lung-DNA	HD753-DNA
USABLE_MSI_SITES (Count)	40	NA	102	120	125

# **MetricsReport.tsv – Contamination Detection**



### **Contamination Detection**

SNPs in VCFs analyzed to detect foreign DNA contamination. Evaluate each sample using **both** Contamination Score and Contamination P value.

[DNA Library QC Metrics]			ı	No Contam	<b>CN Events</b>	CONTAMINAT
Metric (UOM)	LSL Guideline	USL Guide	eline	CCL-DNA	FFPE-Lung-DNA	HD753-DNA
CONTAMINATION_SCORE (NA)		0	3106	223	7535	30409
CONTAMINATION P VALUE (NA)		0	0.049	1	0	0.416

### **CONTAMINATION SCORE:**

- Evaluate common SNPS with expected allele frequencies of 0%, 50%, or 100%
- Frequencies shift when sample is contaminated
- Score is likelihood that observed allele frequency is due to mixing DNA from different samples

### CONTAMINATION\_P\_VALUE:

- Measures the uniformity of low frequency SNPs across the genome
- The probability that a truly contaminated sample has a <u>non-uniform SNP</u> distribution is very small (unless there have been copy number events)
- Measures certainty of copy number events



# **MetricsReport.tsv continued**



[DNA Expanded Metrics]					
Metric (UOM)	LSL Guideline	USL Guideline	CCL-DNA	FFPE-Lung-DNA	HD753-DNA
TOTAL_PF_READS (Count)	NA	NA	89277228	92653732	90620250
MEAN_FAMILY_SIZE (Count)	NA	NA	4.1	3.2	2.9
MEDIAN_TARGET_COVERAGE (Count)	NA	NA	372	425	580
PCT_CHIMERIC_READS (%)	NA	NA	0.1	0.8	0.1
PCT_EXON_100X (%)	NA	NA	96.8	98.3	98.3
PCT_READ_ENRICHMENT (%)	NA	NA	87.6	84.8	85.3
PCT_USABLE_UMI_READS (%)	NA	NA	99.7	99.8	99.8
MEAN_TARGET_COVERAGE (Count)	NA	NA	376	523.5	578
PCT_ALIGNED_READS (%)	NA	NA	95.3	95.6	95.5
PCT_CONTAMINATION_EST (%)	NA	NA	0	0.1	0.2
PCT_PF_UQ_READS (%)	NA	NA	100	100	100
PCT_TARGET_0.4X_MEAN (%)	NA	NA	91.1	82.8	93.1
PCT_TARGET_100X (%)	NA	NA	95.2	97.2	97.3
PCT_TARGET_250X (%)	NA	NA	77.1	75.1	92.4

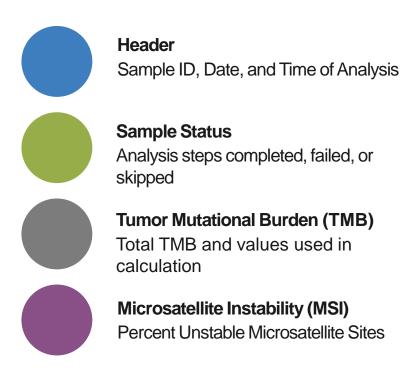


# Results Folder: Biomarker Report

### **Biomarker Report**



Final Report per sample that reports TMB and MSI metrics in a .txt file





# SampleID\_BiomarkerReport.txt



TruSight Oncology 500 - Biomarker Report			
For Research Use Only. Not for use in diagnostic proceed			
For QC Status and Information, reference the MetricsRe	in the Resu	ılts folder.	
[HEADER]			
Sample ID:	FFPE-Lung-DNA		
Report Date:	1/4/2019		
Report Time:	22:22:10		
Pipeline Version:	1.3.1.3		
[SAMPLE STATUS]			
Completed all Steps:	TRUE		
Failed Steps:	NA		
Steps not Executed:	NA		
[TMB]			
Total TMB:	4.708		
Nonsynonymous TMB:	1.569		
Coding Region Size in Megabases:	1.275		
Number of Passing Eligible Variants:	6		
Number of Passing Eligible Nonsynonymous Variants:	2		
[MSI]			
Usable MSI Sites:	108		
Total Microsatellite Sites Unstable:	5		
Percent Unstable Sites:	4.63		



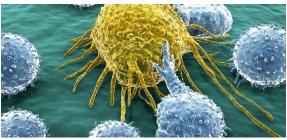
# **Tumor Mutational Burden (TMB)**

### Correlation to immunotherapy outcome

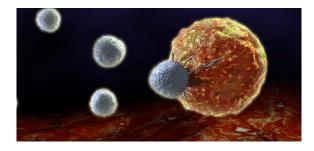
Not every tumor responds well to immunotherapy. High *Tumor Mutational Burden* has been linked to better responses



Tumor Cells with high TMB...<sup>1,2</sup>



...may have high neoantigen load...<sup>1,2</sup>



...which can lead to increased immunotherapy response.<sup>2-5</sup>

### **TUMOR MUTATIONAL BURDEN (TMB):**

- The number of nonsynonymous mutations carried by tumor cells
- TMB is emerging as a potential predictive biomarker to better segment patient populations for immunotherapies treatment <sup>6</sup>
  - 1. Schumacher TN, Schreiber RD. Science. 2015; 348(6230): 69-74.
  - 2. Kim JM, Chren DS. Ann.Oncol. 2016; 27(8):1492-1504.
  - 3. Liontos M et al. Ann Transl Med. 2016; 4(14): 264.
  - 4. Sharma P, Allison JP. Science. 2015; 348 (6230): 56-61.
  - 5: Giannakis M et al. Cell Rep. 2016; 15(4): 857-865.
  - 6. Chalmers ZR et al. Genome Med. 2017; 9(1):34.



# TSO 500 Output: TMB Score

Number of eligible somatic variants per Mb



BCL > FASTQ **Alignment** 

Read collapsing Realignment Stitching

**Variant** calling **Small Variant Filtering** 

**VCF** files

**TMB Calculations** 

TMB =

**N** = Total eligible variants\*

- Coding region
- Somatic variants only
- VAF ≥ 5%

- No hotspots
- No MNVs
- ≥ 50X coverage



M = Total coding region ≥ 50X coverage

he term describes mutations occurring at a chromosomal region, which is more susceptible to genetic damage/change than average sequences, or contains sequences that are more like by spontaneous mutations, such as CpG dinucleotides

**Biomarker** 

[TMB]

Total TMB: 298.989

237.618 **TMB** Nonsynonymous TMB:

Coding Region Size in Megabases: 1.271 M Number of Passing Eligible Variants:

Number of Passing Eligible Nonsynonymous Variants:

302 N

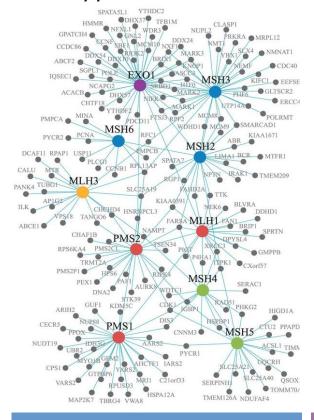
\*Listed in tmb.tsv file





# Microsatellite Instability (MSI)

FDA-approved biomarker for immunotherapy



### **Normal Microsatellite Allele:**

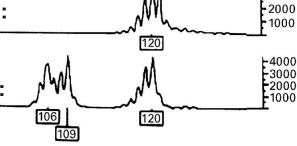
GGTA CA CA CA CA CA CA TTGC

### **Tumor MSI Alleles:**

GGTA CA CA CA CA CA TTGC



Tumor w/ MSI Allele:



### Mismatch Repair Pathways

- · Mismatch Detection
- · Messaging/Signaling/ Repair Activation
- Repair
- · Cell cycle arrest / Apoptosis activation
- · Defects drive Microsatellite Instability

### **Microsatellite**

- · Genomic region of repeated sequence
- Repeating motifs of ~1–9 bp
- Repeat ~5-50X
- All alleles shown in green are examples of microsatellites

### **Microsatellite Instability**

- Indicator of MMR pathway deficiency
- · Traditionally analyzed by PCR or IHC
- Can analyze more loci by NGS than PCR



3000

# TSO 500 Output: MSI Score







Read collapsing

Realignment

Stitching

StitchedReads.bam

MSI Status (Hubble)





MSI Score

≥ 20% MSI-High < 20% MS-Stable

**Total Assessed MSI sites** 

# Biomarker Report

[MSI]

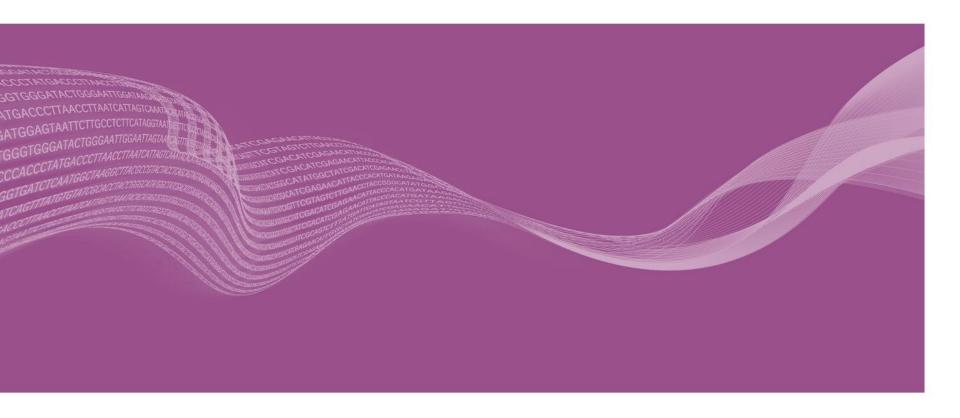
Usable MSI Sites: 102

Total Microsatellite Sites Unstable: 82

Percent Unstable Sites: 80.390

illumına<sup>®</sup>

# **TST 170 RNA Analysis**





# **TSO 170 RNA Analysis options**





# **TruSight Tumor 170 Local App**

- A Docker or Singularity based software package
- Requires separate SampleSheet.csv (RNA libraries only)
- Template available on TruSight Tumor 170 support page



### BaseSpace Sequence Hub TST 170 app

- Sequencing requirements for RNA analysis
  - Minimum of 16 M reads (8 M sequencing clusters) recommended
  - Paired-end samples with the same read lengths required



# **TSO 170 RNA Analysis output**



- Fusions (Manta): {SampleID}\_Fusions.csv
  - All candidate fusions identified by the RNA analysis pipeline
- Splice variants (STAR): {SampleID}\_SpliceVariants.vcf
  - All candidate splice variants identified by the RNA analysis pipeline
  - Annotated by Nirvana
- High Confidence Variants: {Sample\_ID}\_HighConfidenceVariants.csv
  - Fusions passing filter with at least one gene of the pair in the 55-gene target list
  - High confidence splice variants targeted by the panel (total of 3 genes) passing filter
- Published Fusions: {SampleID}\_PublishedFusions.csv
  - List of published fusions (Mitelman Database) targeted by TST 170 panel

Resources:

**TruSight Tumor 170 Local App User Guide** 



# **Questions**



