# 2 Zooplankton abundance and biomass size spectra in the East Antarctic sea-

# ice zone during the winter-spring transition

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#### Abstract

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Sea ice is an influential feature in Southern Ocean- Antarctic marine environments creating a 21 22 2-phase vertical ecosystem. The lack of information on how this system influences community structure during the winter-spring transition however is largely lacking. 23 Zooplankton form the link that bridges these environments, with the meiofaunal and algal 24 25 communities within sea ice directly influencing the epipelagic zooplankton community at the ice-water interface. A combination of methods including sea ice coring, umbrella net 26 sampling and Laser Optical Plankton Counter were used to describe the vertical structure of 27 zooplankton and meiofaunal communities. The distribution of meiofauna and chlorophyll a 28 both played important roles in structuring the zooplankton community within this dynamic 29 region. Many dominant taxa, including Calanus propinguus and Oithona similis, directly 30 responded to the high availability of algae present within the bottom strata of sea ice. The 31 sea-ice associated species *Stephos longipes* represented a strong link between this 2-phase 32 ecosystem. Observations of the vertical distribution of biomass obtained from the LOPC 33 34 suggests that the responses of these species to the sea ice directly influences the vertical structure of zooplankton during the winter-spring transition. 35

- 36 Key words: Antarctica, sea ice, cryo-pelagic coupling, Calanus propinquus, Oithona similis,
- 37 Stephos longipes, optical plankton counter

#### 1. Introduction

Zooplankton play an integral role in Southern Ocean ecosystem structure and function, yet limited work has been undertaken to describe and understand patterns in their distribution in the Antarctic sea-ice zone, especially during the winter-spring transition when sea ice is at its maximum extent. This period is critical for the establishment and recruitment of many dominant zooplankton species, with the convergence of abiotic and biotic cues triggering the cessation of diapause in some species and initiating reproduction in preparation for the spring phytoplankton bloom (Hunt and Hosie, 2006a).

Sea ice has a dynamic role in Southern Ocean physical processes. Its optical and insulative properties, combined with its freeze and melt cycles, strongly affect physical characteristics of the water column below, regulating light availability, temperature, salinity and stratification (Massom and Stammerjohn, 2010). Vast regions of the Southern Ocean are seasonally ice-covered, with winter extent reaching 19 x 10<sup>6</sup> km<sup>2</sup> and decreasing to approximately 4 x 10<sup>6</sup> km<sup>2</sup> during summer. There is a high degree of regional variation in both sea-ice extent and duration due to prevailing differences in wind, oceanic circulation and bathymetry (Stammerjohn et al., 2012; Massom et al., 2013). Sea ice harbours a network of micro-environments for ice-associated (=sympagic) organisms, including bacteria, algae and metazoans, which form communities in different depth horizons of the ice (Kramer et al., 2011).

The number of studies on the diversity of ice-associated metazoans ( $>20~\mu m$ , hereafter referred to as meiofauna) is limited, particularly during the winter months. This impedes our ability to understand how these communities intersect with the pelagic environment; e.g. there is evidence to suggest that grazing on ice-associated detritus, algae and meiofauna forms an important trophic pathway for some pelagic zooplankton groups (Werner, 2005; Kramer et al., 2011). Therefore intra-regional and inter-annual variation in sea-ice physical

properties have the potential to alter both sea-ice meiofaunal and under-ice epipelagic zooplankton communities.

The timing of sea-ice advance and retreat, and spatial variation in thickness and age, are controlling factors of annual patterns of primary and secondary production (Thomas and Dieckmann, 2010). The biomass of the standing stock of epipelagic zooplankton varies in response to these patterns in sea-ice growth and decay. Zooplankton presence and activity in the vicinity of the sea ice-water interface have been shown to increase during the onset of spring, when environmental signals trigger the end of diapause events and the beginning of breeding (Schnack-Schiel and Hagen, 1994). The seasonal succession of zooplankton is an important regulator of ecosystem processes such as grazing and biogeochemical flux, however, zooplankton, in turn, are controlled by the physical environment and primary producers. This 'bottom-up' control is characteristic of the Southern Ocean (Hunt and Hosie, 2006a; b). Bottom ice-algal communities occurring at the sea ice – water interface are influential in shaping the community of epipelagic zooplankton communities, presenting a resource that is up to 50 times more concentrated than that found within the upper water column (Arrigo and Thomas, 2004), with the out-flux of biogenic particles from ageing and decaying sea ice most likely adding to resource availability (Lizotte, 2001; Kramer et al., 2011). Pack ice often drifts up to 20 km day<sup>-1</sup>, whilst the water column beneath can reach speeds in excess of 17 km day<sup>-1</sup> with varied directionality due to mesoscale currents, eddies and wind stress (Williams et al., 2011). Consequently, the sea ice and water column move independently of one another, yet the movements and behaviour of zooplankton within and between these dynamic environments directly couples this 2-phase ecosystem.

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The aims of this study were to identify drivers of, and describe patterns in, the distribution of sea-ice meiofauna and under-ice zooplankton, at both the community and individual taxon levels, off the Budd/Sabrina coast of East Antarctica. Sea-ice cores and net tows were used to describe community composition during the winter-spring transition in 2007 and 2012. To assess how the biomass of plankton was distributed vertically in the surface waters under the sea ice, we deployed a Laser Optical Plankton Counter (LOPC), in conjunction with our net sampling. The LOPC has rarely been deployed in Southern Ocean waters yet it provides a relatively rapid means of comparing plankton biomass and community size structure between sites, independent of the species present.

# 2. Methods

2.1 Field

Sea ice was sampled between 114°E and 130°E (Fig. 1), from mid-September to early October 2007, and again from late September to the end of October 2012, as part of the Sea Ice Physics and Ecosystems eXperiment (SIPEX) and SIPEX-2 voyages. During both years sea ice was encountered at ~61°S, extending as first-year pack until fast ice occurred at ~65.5°S. The maximum extent of sea ice for this area is characteristically small compared with flanking regions to the east and west, with the sea-ice zone reaching only 300 km off the coastline as it is constricted by high wind and water velocities (Williams et al., 2011; Massom et al., 2013).

Twelve sites were sampled in 2007 and eight in 2012 (Fig. 1). Of the 12 sites in 2007, 11 were sampled for epipelagic zooplankton and 12 for sea-ice meiofauna. In 2012, seven sites were sampled for epipelagic zooplankton whilst only five were accessible for sea-ice coring due to thick, rafted sea ice in excess of 2 m. All ice cores were taken from within a 4

 $m^2$  area at each site using a SIPRE ice corer with a 0.02 m internal diameter. A single sea- ice core was taken from each site during 2007 for quantifying meiofauna abundance and distribution, while two to six ice cores were collected during 2012. All cores were sectioned (at 0.1 m intervals) and melted in filtered seawater at 4 °C. Afterwards the contents of each segment were concentrated using a 20  $\mu$ m sieve to collect meiofauna above this size. During 2012, sites 2 and 3 were pre-drilled with a Jiffy drill and only cores for the bottom 0.02 and 0.03 m, respectively, were processed.

Epipelagic zooplankton were sampled using an umbrella net (mouth opening: 0.078 m², mesh size: 100 μm) deployed through holes widened after sea ice coring took place (Loots et al. 2009). Due to heavy ice conditions encountered during 2012 some sites were sampled from the trawl deck of *RV Aurora Australis*. Zooplankton were collected by vertical hauls through the top 50 m of the water column during 2007 and 60 m during 2012. All zooplankton collected were preserved in 4% borax-buffered formaldehyde.

A Laser Optical Plankton Counter (LOPC; Herman, 2004) was deployed vertically at sites 3, 4, 6, 7 and 8 during 2012, accompanying umbrella net sampling at each site. The LOPC was lowered from the surface to 60 m at a rate of 1 m s<sup>-1</sup>. At sites 7 and 8 the LOPC was deployed manually through a hole drilled in the sea ice, while at sites 3, 4 and 6 it was lowered from the stern of the vessel. The LOPC was housed within a steel frame, and consisted of a 7 x 7 cm sampling tunnel, with a 35 mm x 1 mm laser that was reflected back onto a 35 element (1 mm spacing) photo-diode array by a prism. When a particle passed through the emitted or reflected beam, the size of the shadow was detected and recorded as the corresponding equivalent spherical diameter (ESD) of the particle. The data were recorded on a data-logger (ODIM Brooke Ocean, Dartmouth, Canada) housed within the frame, and a conductivity—temperature—depth (CTD) sensor (Model SBE- 37, Sea-Bird Electronics, Inc., Washington, U.S.A) measured the depth profile.

Environmental variables were recorded during both voyages. The thickness of the sea ice and the snow cover were measured at the site of each ice core. Chlorophyll *a* concentration and salinity and temperature of the sea ice cores were profiled as per Meiners et al. (2011). Brine volumes were calculated from the temperature and salinity profiles using the equations of Cox and Weeks (1983).

A ship-based CTD-rosette was used to measure temperature, salinity, fluorescence and PAR of the top 200 m of the water column. Chlorophyll *a* concentrations of the upper water column were determined by High Performance Liquid Chromatography (HPLC) at depth intervals of approximately 15 m, according to methods described by Wright and van den Enden (2000). Linear regressions were established between chlorophyll *a* measurements and CTD fluorescence measurements at the same depths. The regression equations were then used to convert the remaining fluorescence data in order to obtain high-resolution profiles of the water-column. Integrated measurements were calculated for the top 150 m.

# 2.2 Laboratory

Preserved zooplankton and meiofauna were identified to the lowest taxonomic level possible. Replicate umbrella net samples collected in 2012 were used to estimate biomass (dry weight) by filtering zooplankton onto pre-combusted (450 °C for 12 h) 50 µm pore-size glass-fibre filters (Macherey-Nagel) and dried at 60 °C until constant mass was reached. Additional biomass estimates for both years, expressed as mg C m<sup>-3</sup> were calculated by multiplying the abundance of each zooplankton species by its carbon content (determined by Jia et al., this volume).

The LOPC particle sizes were classified into size bins from 306 µm–3080 µm. The size bins were calculated based on the geometric mean of the squares of two consecutive whole

numbers from 17 to 55 (Moore and Suthers, 2006). Size bins from the first zero-count and larger were removed from the analysis. Biomass of zooplankton measured using the LOPC was estimated using the method of (Suthers et al., 2004). ESD values ( $\mu$ m) were converted to biomass assuming the volume of a sphere and the density of water ( $\rho$  =10<sup>9</sup> mg m<sup>-3</sup>) using:

Biomass 
$$(mg \ m^{-3}) = \frac{4}{3}\pi \left(\frac{ESD}{2}\right)^3 \rho$$

Normalised biomass (NB; m<sup>-3</sup>) was calculated by dividing the biomass of each bin by the mass range of that bin (Platt, 1985; Platt and Denman, 1978). The normalised biomass is independent of any specified body size interval, allowing for comparison across systems and studies (Krupica et al., 2012). The normalised biomass  $\beta(w)$  is the total biomass b(w) in the size class characterised by weight (w) divided by the width of the size class  $\Delta w$ :

$$\beta(w) = b(w)/\Delta w$$

The linear slope and intercept is given by the least-squares linear regression of the  $log_{10}$  NB against the  $log_{10}$  body mass (mg). Due to the dominance of crustaceans identified in umbrella net samples, total biomass derived from LOPC was then converted to a dry weight using the conversion factor of 0.171 outlined by Young et al. (1996).

# 2.3 Statistical analysis

Analyses were performed using the statistical packages PRIMER 6 (version 6.1.13) and R (version 3.1.1). To explore the relationships between epipelagic zooplankton assemblages and sampling sites, a canonical analysis of principal coordinates (CAP) was used to find the axes that best discriminated between the samples from the two years, while a vector overlay showed which species were responsible for differences in the years. SIMPER (similarity

percentages) analysis was conducted to determine which species contributed to greater than 10% of total abundance in each year.

Environmental data at each site were summarised, and tested for statistical differences in the means between years using Welch's t-test; this test was chosen due to heterogeneity in the variances. There was a high degree of variability in brine and chlorophyll *a* content over the depth of the sea ice; therefore, brine volume and chlorophyll a concentration in the bottom 0.01 m of the sea ice were analysed separately to the remainder of the core (hereafter referred to as 'bottom ice chlorophyll' and 'bottom ice brine volume', respectively).

R-mode analysis (PRIMER) was undertaken on 17 epipelagic taxa that were common to both sampling periods based upon cluster analysis performed using unweighted pair-group arithmetic averages (UPGMA). Non-metric Dimensional Scaling (NMDS) was then used to provide zooplankton associations defined by the cluster analysis. Indicator Value (IndVal) analysis was performed to identify 'indicator' species within each sampling year. Each IndVal was calculated as:

$$IndVal_{ii} = A_{ii} * B_{ii} * 100$$

Where  $A_{ij} = Nindividuals_{ij}/Nindividuals_i$  and  $B_{ij} = Nsites_{ij}/Nsites_j$ 

 $A_{ij}$  is a measure of specificity, where Nindividuals<sub>ij</sub> is the mean number of individuals of species i across all sites and Nindividuals<sub>i</sub> is the sum of the mean of species i across all sites.  $B_{ij}$  is a measure of fidelity where Nsites<sub>ij</sub> is the number of sites where species i is present, with Nsites<sub>j</sub> the total number of sites sampled in that year (Dufrêne and Legendre, 1997). A taxon with an IndVal of 25% or greater was considered to be indicative of a community, showing that it was present in at least 50% of the samples for 50% of the time.

A BIOENV (biological-environmental) analysis using Spearman's rank correlation and Euclidean distance was used to describe the environmental variables which were associated with the patterns in zooplankton community structure identified. Due to missing variables, site 7 form 2007 and sites 5, 7 and 8 from 2012 were omitted from the BIOENV analysis.

To understand how environmental variables influenced individual taxa, generalised linear models (GLM) were applied to the numerically abundant genera or species common to both years. A generalised additive model (GAM) with all predictor variables was initially used to assess the relationship of each predictor variable to each taxon's abundance to assess the need to incorporate polynomial terms in subsequent GLM. GLM used a Poisson distribution and when tests indicated that abundance data were overdispersed a negative binomial model was used instead. Backwards selection was used to select the most parsimonious model based upon Akaike Criterion Information (AIC), with the model presenting the lowest AIC deemed the most appropriate. The goodness-of-fit of the simplified model was then tested for using a Pearson's chi-squared test based upon the residual deviance and degrees of freedom of the model.

#### 3. Results

3.1 Environmental variation

Environmental variables that were measured during both years are summarised in Table 1. The water column features that differed significantly between years were temperature, depth and chlorophyll *a* concentration. Water temperatures were slightly warmer, surface layer chlorophyll *a* concentrations were higher and mean depth of sampling sites deeper in 2012 than in 2007. Both snow depth and ice thickness were also greater in 2012, although these

features were more variable between sites, ranging between 0.008 m and 0.1 m for snow cover and from 0.075 m to 2.5 m for ice thickness.

3.2 Epipelagic zooplankton community composition and species assemblages

Twenty-one taxa were identified in 2007 and 38 in 2012 from epipelagic umbrella net samples. Copepods, including nauplii, contributed the most to total abundance (63-96%) at most sites during 2007, with the exception of sites 1 and 3, where they contributed 13 and 41%, respectively (Fig. 2a). Tintinnids, dominated by members of the genus *Cymatocylis*, accounted for 57% of total abundance at site 1 and 84% at site 3; they comprised from 1 to 35% of abundance at the other sites. The foraminifera *Neogloboquadrina pachyderma* and radiolarians comprised 1-8% of overall abundance, with all other groups, including euphausiids, ostracods and pteropods, accounting for less than 1%. In 2012, copepods were the most abundant group at all sites, comprising 68-97% of total abundance (Fig. 2b). Those samples also contained protists (0-21%) and pteropods (0-7%). All other taxonomic groups, including ostracods, polychaetes, chaetognaths, siphonophores, euphausiids and appendicularians contributed less than 3%. Total abundance did not differ significantly between years (*p*-value = 0.18), based on a non-parametric Welch's test.

Seventeen taxa co-occurred in net samples from both SIPEX voyages (Table 2). These included the numerically abundant cyclopoid *Oitona similis*, the harpacticoid *Microsetella norvegica*, the small calanoid *Microcalanus pygmaeus* and copepod nauplii. The large calanoid copepods, *Calanus propinquus* and *Calanus simillimus*, were present in relatively low abundances. Total zooplankton abundance showed little variation between years, however, there was high spatial variability, with large ranges for each site in both years. Six distinctive assemblages were identified at 40% similarity, although *Stephos longipes*,

Ctenocalanus sp., Thysanoessa macrura and Neocalanus tonsus each formed solitary clusters (clusters C-F). Maupasia sp. and Calanus simillimus formed a single cluster, B, and the remaining 11 taxa grouped to form the largest cluster, A (Fig. 3a). Whilst group A consisted of the most individuals, several sub-clusters were evident at 70% similarity.

3.3 Environmental drivers of epipelagic community composition

Results from the CAP comparing water column communities between years are shown in Fig. 3b. A clear distinction between years is evident based upon species composition and abundance. BIOENV indicated that the environmental variables that best described the community composition of epipelagic zooplankton were longitude, snow thickness and chlorophyll *a* concentration in the water column, with a correlation of 0.384.

GAM indicated that all predictor variables had a linear relationship with species abundance and therefore no polynomial terms were used in fitting each GLM. The taxa assessed were *Calanus propinquus, Microsetella norvegica, Oithona similis,*Neogloboquadrina pachyderma and tintinnids (Table 3). Due to the lack of sufficient data from both years, *Microcalanus pygmaeus, Stephos longipes* and *Limacina helicina* could not be assessed. In each case, the most parsimonious model, as selected by AIC, provided a better fit than the saturated model.

Calanus propinquus was mainly influenced by total sea-ice chlorophyll a concentration, with its abundance increasing with increasing chlorophyll a content.

Microsetella norvegica was influenced by bottom-ice chlorophyll a, with an increase in abundance corresponding to increased chlorophyll a concentration. Oithona similis was influenced by the depth of the site, snow thickness, sea ice chlorophyll a concentration and latitude, whereby its abundance increased with a decrease in snow depth, and increased

latitude and sea ice chlorophyll *a* content. *Neogloboquadrina pachyderma* responded to snow depth, chlorophyll *a* concentrations within the sea ice and latitude, with abundances increasing with deeper snow, less sea-ice chlorophyll *a* and a decrease in latitude. Tintinnids were most influenced by bottom-ice chlorophyll concentration, latitude and longitude. As bottom-ice chlorophyll concentration increased so did the abundance of tintinnids, with abundances also being higher close to the coast on the western boundary of the sampling region (corresponding to an increase in longitude and a decrease in latitude).

3.4 Sea-ice meiofauna: temporal variation and vertical structure

Ten taxa were identified in sea-ice cores collected during 2007 and five in 2012. Of these taxa, N. pachyderma, Paralabidocera antarctica, Drescheriella glacialis, Stephos longipes and copepod nauplii were common to both years. Integrated abundances of each species across the length of their respective ice core tended to be higher in 2007 than in 2012 (Fig 4). During 2007, N. pachyderma was the most abundant species, accounting for 42 to 100% of total integrated abundance. A variable number of nauplii was found (0-58%), and all other taxa contributed less than 12% together. In 2012, N. pachyderma contributed from 22 to 98% to total abundance. Copepod nauplii and D. glacialis were present in approximately equal proportions, contributing 1-38 and 3-30%, respectively. Stephos longipes reached its highest proportion at site 6 (10%), and contributed less than 1% at all other sites, while P. antarctica represented less than 2% at all sites. Total meiofauna abundance did not differ significantly between years (p = 0.231), based on a non-parametric Welch's test.

Figure 5 a-c depicts representative cores from 2007 and figure 5 d and e from 2012. During 2007 meiofaunal communities just below the snow-ice interface were observed at sites 1, 3, 8 and 13 (Fig. 5 and b). The surface communities tended to be dominated by *N*.

pachyderma and nauplii, when present. Neogloboquadrina pachyderma dominated the bottom 50% of sea ice cores at all sites except 3 and 8. When present, *D. glacialis*, *S. longipes* and copepod nauplii also followed this trend but in lower abundances (Fig. 5 c). In 2012, surface communities were dominated by *N. pachyderma*, with low numbers of *P. antarctica*, *D. glacialis* and *S. longipes* (Fig. 5d). Sites 2, 3 and 6 possessed very similar meiofaunal communites, with approximately equal proportions of *N. pachyderma*, *D. glacialis* and *P. anatarctica* in the bottom 0.02 m of ice (Fig. 5e). Nauplii were also recorded at site 2 whilst *S. longipes* were recorded at site 3.

Vertical profiles through the sea ice in 2007 showed L-shaped profiles with lowest chlorophyll *a* concentrations at the surface of the ice, remaining low throughout the length of each core and then increasing within the bottom 20% of each core (Fig. 6a). Sea ice sampled during 2012 lacked a similar trend (Fig. 6b). Sites 2 and 3 had chlorophyll maxima mid-way down the length of the core, whilst site 4 showed a reverse trend from that recorded during 2007, with the highest chlorophyll concentration at the surface of the core and decreasing to the ice-water interface (Fig. 6b). Ice brine volumes showed generally C-shaped profiles, with the lowest volumes occurring in the interior of the ice and higher volumes near the surface and at the ice-water interface (Figs. 6c, d).

Carbon biomass did not differ significantly between years within the water column or the sea ice (p = 0.055 and p = 0.349, respectively; Table 4), although carbon content was much higher in the sea ice than the water column in both years. Biomass derived from the LOPC deployments in 2012, was lowest at sites 3 and 6 (24 and 26 mg m<sup>-3</sup>, respectively), peaked at site 4 (231 mg m<sup>-3</sup>) and remained relatively constant at sites 7 and 8 (70 and 48 mg m<sup>-3</sup>

3.5 Carbon content and vertical biomass distribution of epipelagic communities

respectively; Fig. 7). The normalised biomass size spectrum (NBSS) at site 7 indicated a shallow slope (-0.59) while site 4 had a steeper slope (-1.06). Sites 3, 6 and 8 had slopes of -0.63, -0.70 and -0.73, respectively. Sites 3, 6 and 8 had distinct nonlinearities (doming) at sizes greater than 0.58 mm (Fig. 7), indicating an increased biomass at these larger sizes.

A breakpoint of 2 mm length was used to separate the zooplankton identified during 2012 into size classes, separating protists, molluses and small copepods such as *O. similis*, *C. citer* and *S. longipes* from larger, biomass dominating species including the copepod *C. propinquus* and euphausiid, *T. macrura*. Assuming an elliptical shape and a 3:1 length:width ratio of the zooplankton individuals, the ESD size classes of greater and less than 1.154 mm were used to define this break allowing for an LOPC derived vertical structure of biomass at each site. Integrated biomass of the upper 60 m of the water column indicated a clear disjunction in the contribution of each of these size classes to overall biomass. The larger size class contributed 38% or less to total biomass derived from LOPC deployments. The LOPC data were used to examine vertical distributions of biomass over the sampling depth at five sites. Most of the biomass was distributed in the top 20 m at sites 3, 4 and 6 (Fig. 8). At sites 7 and 8, however, the biomass was more evenly distributed, with the highest proportion found at depth.

# 4. Discussion

4.1 Drivers of the epipelagic zooplankton

The composition of zooplankton collected during 2007 and 2012 was typical of the Pacific/Indian sectors of the Southern Ocean (Hunt and Hosie, 2006a; Ojima et al., 2013). The dominant taxa recorded from both years were indicative of Antarctic waters south of the Southern Antarctic Circumpolar Current Front (SACCF) (Atkinson, 1991; Hunt and Hosie, 2006a), with copepods and protists forming the dominant taxonomic groups. Considerable

variability in these two groups was observed, with tintinnids contributing greater proportions to total zooplankton abundance than copepods at the first three sites sampled in 2007. However, while 21 and 38 taxa were identified during 2007 and 2012, respectively, the 17 taxa common to both years generally accounted for greater than 95% of total zooplankton abundance. Therefore, the separation seen in epipelagic zooplankton assemblages in the two years was likely due to changes in ratios of these common taxa rather than substantial differences in the species collected; *e.g.* cluster B developed from the ordination analysis was predominantly comprised of those species that contributed to greater than 10% of total abundance for each year.

Over larger regions of the Southern Ocean distinct associations of species assemblages with major water masses have been observed in large-scale synoptic surveys of the east Antarctic SIZ, such as BROKE and BROKE-West (Hosie et al., 2000; Swadling et al., 2010). The hydrography of this region of East Antarctica has been extensively studied over the past 20 years, with oceanographic features including the SACCF and the Southern Boundary of the Antarctic Circumpolar Current (SBACC) well understood (Williams et al., 2011). There were relatively small degrees of variation in both temperature and salinity of the upper water column, both between sites and between years; however, the mixed layers were considerably deeper in 2007 (100 – 400 m; Williams et al., 2011) than in 2012 (90 – 160 m; Schallenberg et al., this volume). Water column characteristics, with temperatures below -1 °C and salinities of 34.2 confirmed that epipelagic zooplankton communities were sampled south of the SBACC (Hunt and Hosie, 2006a; Williams et al., 2011).

Meso-scale changes in water column chlorophyll *a* (a proxy for phytoplankton biomass and available resources for grazers), snow depth and longitude were shown to be the variables most likely driving the communities of zooplankton observed. Snow depth was

generally ten times higher in 2012 than in 2007. Given its high albedo and high attenuation coefficient it is likely that snow depth was responsible for the changes observed in water column chlorophyll *a* and thus was indicated as a driving feature of epipelagic zooplankton communities. As snow significantly reduces the amount of photosynthetically active radiation (PAR) that reaches the sea-ice interface, algal communities present in the bottom layers of the ice are likely to be negatively impacted, reducing potential algal biomass despite low-light adaptations of these algal communities (Smith et al., 2000). As the surface roughness and structural heterogeneity of sea ice influences snow accumulation (Sturm and Massom, 2010), sea ice may have played an indirect role in under-ice zooplankton community structure. Counter-intuitively, under-ice water chlorophyll *a* concentrations were higher during 2012 when snow depth was significantly greater. However this could be an indication of dark-adaptation, e.g. a higher cell-specific chlorophyll *a* content in phytoplankton. Alternatively, it could reflect the slightly later sampling time in 2012, with phytoplankton populations more developed.

Longitude also played an important role in zooplankton community composition, chiefly for those taxa that grouped independently when the cluster analysis was applied. *Maupausia* sp. and *C. simillimus* were found only at the longitudinal extremes of the study region, while *S. longipes* and *T. macrura* possessed no pattern with longitude. 122°E formed a breakpoint, with *Ctenocalanus* sp. found to the east of this point and *N. tonsus* to the west. Whilst *N. tonsus* and *C. simillimus* are considered to be sub-Antarctic species, both occurring north of the SB, the presence of *C. simillimus* in waters below this boundary is not uncommon (Hunt and Hosie, 2006a). *Calanus simillimus* presence has been attributed to southern eddy-transport from the SBACC; however, there was a lack of warm water "hotspots" that could explain its presence in 2007 and 2012, indicating that seeding of individuals may have occurred from eddy activity earlier in both years.

Although BIOENV analysis revealed physical characteristics of the water column to be important in structuring communities, individual taxon-based analysis using GLM highlighted the influence of the sea ice as an important driver of populations. The results of GLM indicated that two different aspects of sea ice properties influenced the dominant taxa of the epipelagic communities below: (i) depth-integrated ice algal chlorophyll a (i.e. in the entire ice cores), and bottom ice chlorophyll a (bottom 0.01 m in direct contact with the under-ice realm). The abundances of C. propinguus and O. similis increased with an increase in integrated sea-ice chlorophyll a content, whilst N. pachyderma decreased. Microsetella norvegica and tintinnids, however, were only influenced by bottom ice chlorophyll. The feeding of C. propinguus is not fully understood, with suggestions of ice algal grazing at the ice-ocean interface as well as feeding in the water column ingesting pelagic phytoplankton, protists and microzooplankton (Bathmann et al., 1993; Pasternak and Schnack-Schiel, 2001). Jia et al. (this volume) found, via carbon and nitrogen isotope analysis, that C. propinguus fed predominantly upon phytoplankton within the water column. Whilst our results of the GLM indicate that C. propinguus was most influenced by the algae present within the sea ice, it is likely, given these two pieces of information, that *C. propinguus* formed close associations with the sea ice, feeding sporadically on ice algae at the sea-ice interface. Atkinson et al (2001) described this species as an indiscriminate feeder favouring large, non-spinous diatoms, and therefore algal composition of both the sea ice and water column may have influenced these results. Ice algal bottom communities were dominated by pennate diatoms, in particular Fragilariopsis spp., during both 2007 and 2012 (Meiners et al., 2011; Ugalde et al., this volume).

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The diet of *O. similis* is perhaps better understood than that of *C. propinquus*, and it is known to rely upon microplankton (both autotrophic and heterotrophic) and detrital material including faecal pellets, readily switching between any available resources. Consequently the

presence of a concentrated food source, which is often comprised of small diatoms and the presence of large numbers of other species close to the sea-ice interface allow *O. similis* to exploit its flexible diet and copraphageous nature (Pond and Ward, 2011). However, a lack of stable isotope analysis for this species makes it unclear which of these potential factors influenced the trend uncovered by GLM in the present study.

The negative relationship between integrated chlorophyll *a* in the sea ice and *N*. *pachyderma* is interesting, and might point to strong grazing pressure. Conversely, the positive relationship between abundance and snow depth suggests that *N. pachyderma* accumulate in areas of low chlorophyll concentration where light attenuation by snow limits algal growth. It is possible that, given their omnivorous diet, a shift is seen with the onset of spring whereby individuals switch from grazing on phytoplankton to other protists.

Reproduction of *N. pachyderma* begins in spring, so this switch may occur in preparation for coinciding with the mass release of entrained individuals during ice melt (Spindler and Dieckmann, 1986).

Bottom ice chlorophyll *a* concentration was an important driver of *M. norvegica* and tintinnid distribution. *Microsetella norvegica* is known to inhabit both the epipelagic and the lower portion of sea ice; therefore it relies heavily upon sea-ice algal communities for its diet (Dahms et al., 1990). The distribution and feeding ecology of tintinnids is poorly understood. The tintinnid bloom recorded during 2007 points to their possible importance in the sea-ice zone off East Antarctica. The tintinnids found within this bloom were predominantly of the genus *Cymatocylis*, which are cosmopolitan filter feeders consuming phytoplankton of approximately 30 µm (Dolan et al., 2012; Alder and Boltovskoy, 1991). The high abundances of tinitinnids associated with high bottom ice chlorophyll *a* suggests their ability to capitalise on bottom-ice algal communities, with relatively stable water column environmental conditions ideal for proliferation.

# 4.2 Distribution of sea-ice meiofauna

Our study suggests that a strong degree of coupling exists between the sea ice and epipelagic environments. The environmental stability and supply of concentrated food resources provided by sea ice during winter and early spring is a driving force in shaping sea-ice associated zooplankton communities. Moreover, the presence, abundance and vertical distribution of sea-ice meiofauna contributes to the coupling of this 2-phase system. The sea-ice meiofaunal communities found during 2007 and 2012 are similar to previous studies off East Antarctica during maximum sea-ice extent (Swadling et al., 1997; Loots et al. 2009). Brine volume, an indicator of habitable space, was the dominant factor influencing meiofauna distribution, with high brine volumes correlated with high abundances and diversity. Whilst meiofauna abundances tend to increase with ice algal concentration (Kramer et al. 2011), our study indicated that proximity to the ice-water interface was a key factor, confirming a previous observation that sea-ice meiofauna maxima generally occur within this sea-ice strata (Schnack-Schiel et al., 2001).

The vertical distribution of the three copepods identified in sea-ice cores reflects their respective life histories. The low abundances of adult *P. antarctica* is a common observation in sea ice (Swadling et al., 2004). Adults generally do not directly inhabit sea ice, although their nauplii and young copepodid stages use it as the main overwintering habitat (Loots et al. 2009). Nauplii that were found at the ice-water interface were likely entrained during initial sea-ice formation, e.g. in autumn (Swadling et al., 2004). *Stephos longipes* have been shown to migrate actively between the water column and sea ice habitats, where naupliar stages are able to move towards the interior of the sea ice when sufficient 'space' is available (Kurbjeweit et al., 1993). In the present study *S. longipes* was only found within the water column when it was also present within the sea ice above, demonstrating their ability to migrate freely between the ice and the water, and therefore forming a substantial link

between the two habitats. In contrast, *D. glacialis* appears to be completely reliant upon the sea ice as no individuals were identified within the water column from either year. Whilst previous studies have demonstrated an active migration of this species between the sea ice, pelagic realm and benthos this generally occurred in autumn, prior to sea-ice formation when the nauplii migrate to the sea-ice interface and are likely scavenged during sea-ice formation (Loots et al., 2009). The large numbers of *S. longipes* and *D. glacialis* nauplii in both the lower and surface layers of sea ice suggests active breeding by these ice-associated species. Given their small size, naupliar stages of these two species are able to inhabit sea-ice layers with small brine volumes and correspondingly small brine channel diameters. The distribution of *S. longipes* across distinct sea-ice layers and in the under-ice waters indicates their ability to migrate to and within sea ice, therefore representing an, so far understudied, link in cryopelagic coupling,

# 4.3 Vertical distribution of biomass

Despite variations in total zooplankton abundance and species composition, no statistical difference in carbon biomass was recorded between 2007 and 2012. Dry weight and LOPC derived biomass estimates were only conducted for 2012, and biomass calculated from the LOPC were considerably higher than from the other methods applied. Given that LOPC derived biomass is comparable to a 'wet-weight', higher LOPC estimates of biomass is not unexpected, however when LOPC biomass is converted to a dry weight, biomass estimates were considerably larger than the other methods applied. Trends in total biomass were consistent between each of the three methods used for sites during 2012. Previous attempts to use LOPC to quantify biomass of the water column *in situ* generally always yielded higher values than those by traditional means and is attributed to the detection by the LOPC of non-

zooplankton particles, e.g. detritus and algal material (Herman and Harvey, 2006). The separation of particles based upon size class in the present study, i.e. smaller and larger than 2 mm, reduces this problem for the larger size class at least, given that detrital and algal material would rarely fall within that category. The vertical stratification of biomass based upon this size class separation therefore provides a relatively robust insight into the distribution of the dominant taxa in the upper water column. A dominant change in the distribution of biomass in both size classes was seen at sites 7 and 8 during 2012. At these sites *C. propinquus* was the most numerically abundant zooplankter within the greater than 2 mm size fraction. Its presence in abundances greater than 1 ind. m<sup>-3</sup> coincided with the trend seen in LOPC data at sites 3, 4 and 6 during 2012. High biomasses of particles within close proximity to the sea-ice interface could not be directly resolved due to the interference of ice particles with LOPC measurements.

In conclusion, our study suggests an effect of coupled sea ice physical-biological properties on under-ice zooplankton abundance and species composition. Our study supports the observations that copepod species *Oithona similis* and *Calanus propinquus* use the pelagic habitat as well as the sea-ice interface, whilst *Stephos longipes* use both, as well as the interior of the ice. We suggest that these species contribute significantly to exchange of material and carbon flow across the sea ice – ocean interface. To our knowledge, this study presents the first LOPC data on under-ice plankton in the high latitude Southern Ocean. LOPC data, albeit providing larger estimates of biomass, demonstrated site specific trends that were consistent with classical methods indicating that LOPC may provide a useful method to study under-ice zooplankton in a cost- and time-effective manner. Out study confirms previous findings that Antarctic sea ice supports dense communities of ice-associated meiofauna.

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Table 1. Environmental variables recorded for the water column and sea ice for 2007 and 2012 (showing means and ranges). F-values were calculated using Welch's tests to test for significant differences between years. \*denotes p-value  $\leq$ 0.05 and \*\* denotes p-value  $\leq$ 0.005 for calculated F values.

	2007	2012	F value
Water column			
Number of samples	11	19	
Latitude °S	64.94 (64.23 – 65.58)	65.01 (64.4 – 65.26)	0.179
Longitude °E	121.93 (116.19 – 128.10)	119.52 (116.16 – 121.67)	2.913
Depth, m	2000(750 - 3000)	3000(2000 - 3500)	8.339*
Temperature, °C	-1.85 (-1.83 – -1.86)	-1.86 (-1.85 – -1.87)	11.69**
Salinity	34.24 (34.10 - 34.41)	34.28 (34.21 - 34.33)	0.166
Chlorophyll a, mg m <sup>-3</sup>	0.05(0.01-0.10)	0.15(0.10-0.25)	19.087**
Sea ice			
Number of samples	12	5	
Latitude °S	64.97 (64.32 – 65.58)	64.91 (63.88 – 65.40)	0.047
Longitude °E	121.65 (116.82 – 128.10)	120.33 (119.91 - 120.92)	1.082
Snow depth, m	0.0035 (0 - 0.008)	0.0055 (0.008 - 0.1)	33.79**
Ice thickness, m	0.0078 (0.0058 - 0.132)	0.768 (0.075 - 2.5)	6.10*
Freeboard, m	0.0024 (-0.004 - 0.008)	0(-0.009 - 0.005)	0.273
Chlorophyll <i>a</i> in bottom 0.01 m, mg m <sup>-3</sup>	0.144 (0.0040 - 0.647)	0.089 (0.00036 - 0.318)	0.483
Integrated chlorophyll a, mg m <sup>-2</sup>	2.14(0.08 - 8.43)	3.02(0.41-1.06)	0.013
Brine volume - bottom 0.01 m, %	20.04 (13.24 – 34.82)	18.26 (8.64 – 26.92)	0.19
Brine volume - remainder, %	7.60(5.37 - 11.79)	11.40(6.69 - 14.57)	4.047

Table 2. List of the common zooplankton taxa. Abundances are means for all sites within each sampling year (ind.  $m^{-3}$ ). **Bold** indicates a statistically significant Indicator Value of greater than 25% for that year. \*distinguishes those species that contributed  $\geq 10\%$  to abundance for that year (SIMPER analysis).

	2007	2012
Calanus propinquus	0.95	1.22
Calanus simillimus	0.05	0.53
Ctenocalanus sp.	0.05	0.05
Microcalanus pygmaeus	0.44	6.30*
Microsetella norvegica	1.09	3.47*
Neocalanus tonsus	0.72	0.04
Oithona similis	94.5*	58.5*
Stephos longipes	0.12	1.29
Nauplii	38.4*	34.4*
Neogloboquadrina pachyderma	1.99*	0.82
Radiolarians	2.55*	2.89
Tintinnids	85.0*	0.91
Gastropod veliger	0.09	2.16
Limacina helicina	0.26	2.59
Maupasia sp.	0.02	0.36
Ostracods	0.07	0.95
Thysanoessa macrura	0.05	0.19

Table 3. The most parsimonious GLM based upon AIC selection criteria. Predictor variables used are indicated and when appropriate, significant parameters ( $p \le 0.05$ ) are indicated. The fit of the model as determined by Chi-squared analysis is also provided.

Species	Model predictors	Significant Parameter/s	coeff	± SE	Z-value	AIC	Fit of model
C. propinquus <sup>1</sup>	Chla <sub>water</sub> , snow depth, Chla <sub>ice</sub> , Chla <sub>bottom 0.01 m</sub> , longitude, year	$Chla_{ice}$	1.53	0.76	2.01	67.81	0.101
M. norvegica <sup>1</sup>	Chla <sub>water</sub> , Chla <sub>bottom 0.01 m</sub> , depth	Chlabottom 0.01 m	0.98	0.29	3.39	76.02	0.039
		depth	-0.93	0.341	-2.72		
O. similis <sup>2</sup>	snow depth, Chla <sub>ice</sub> , Chla <sub>bottom 0.01 m</sub> , latitude	intercept	76.69	22.48	3.42	215.1	0.143
		snow depth	-0.013	0.01	-2.97		
		$Chla_{ice}$	0.39	0.15	2.67		
		latitude	-1.12	0.35	-3.24		
N. pachyderma <sup>1</sup>	longitude, latitude, snow depth, $Chla_{ice}$	intercept	106.6	33.86	3.14	56.92	0.913
		snow depth	0.03	0.01	2.09		
		latitude	-1.82	0.54	-3.33		
		$Chla_{ice}$	-37.12	15.91	-2.33		
Tintinnids <sup>2</sup>	Chl $a_{\text{water}}$ , salinity, Chl $a_{\text{bottom 0.01 m}}$ , latitude, longitude	Chlabottom 0.01m	0.625	0.16	4.00	108.5	0.092
	-	latitude	-1.432	0.59	-2.41		
		longitude	0.54	0.09	5.60		

<sup>664</sup> GLM based upon a Poisson distribution

<sup>&</sup>lt;sup>2</sup>GLM based upon a negative binomial distribution due to overdispersion of the abundance data

Table 4. Zooplankton and sea-ice meiofauna. Abundance and biomass in the water column and sea ice during winter 2007 and 2012 (showing means and ranges). F-values were calculated using Welch's tests.

	2007	2012	F value
Water column			
Total net zooplankton abundance, ind. m <sup>-3</sup>	226.7 (33.9 – 725.8)	120.0(30.1 - 315.1)	2.462
Total number of species per site, S	9(6-15)	12(6-20)	-
Dry weight (DW), mg m <sup>-3</sup>	-	0.11(0.00-20)	-
Carbon, mg m <sup>-3</sup>	0.83(0.041 - 2.77)	1.54 (0.23 - 4.44)	4.07
LOPC predicted biomass mg m <sup>-3</sup>	-	80.89 (24.93 – 231.06)	-
LOPC converted biomass (DW) mg m <sup>-3</sup>	-	13.65 (4.10 – 39.50)	
Sea ice			
Total meiofauna abundance, ind. m <sup>-3</sup>	27483 (0 - 117472)	61189 (24036 – 140890)	1.797
Total number of species per site, S	3(1-5)	4(3-4)	2.233
Carbon, mg m <sup>-3</sup>	61.07 (0 - 266.3)	114.8 (14.33 - 279.7)	1.031

# Figure Captions

- Figure 1. Map of the sites sampled during 2007 (solid) and 2012 (open) voyages, showing
- features mentioned in the text.
- Figure 2. Abundance (ind. m<sup>-3</sup>) of main epipelagic zooplankton groups identified from the
- upper water column during (a) 2007 and (b) 2012.
- Figure 3. Multivariate analyses of epipelagic species and sites for 2007 and 2012. (a) Species
- assemblages identified by cluster analysis indicating those clusters identified at 40%
- similarity and sub-clusters at 70% and (b) CAP analysis of sites, with dominant species as
- 679 vectors.
- Figure 4. Sea-ice meiofauna integrated abundance (ind. m<sup>-2</sup>) during (a) 2007 and (b) 2012.
- Figure 5. Vertical distribution and abundance of meiofauna in sea ice cores from the sea ice
- surface from representative sea ice cores during 2007 (a-c) and 2012 (d, e).
- Figure 6. Vertical distribution of brine volume and chlorophyll a (mg m<sup>-3</sup>) within sea ice
- during 2007 (a, b) and 2012 (c, d).
- Figure 7. Normalised Biomass Size Spectrum (NBSS) for each of the 5 sites (A-E) derived
- from the LOPC deployments in 2012. Slope intercept, biomass and counts are shown on each
- figure.
- Figure 8. Vertical structure of epipelagic biomass (mg m<sup>-3</sup>) for zooplankton less than (solid)
- and greater than (open) 2 mm length as derived from the vertical deployments of the LOPC in
- 690 2012.