

MG7: Configurable and scalable 16S metagenomics data analysis – new methods optimized for massive cloud computing

Alexey Alekhin¹ Evdokim Kovach¹ Marina Manrique¹ Pablo Pareja¹ Eduardo Pareja¹ Raquel Tobes¹ and Eduardo Pareja-Tobes¹,*

 1 Oh no sequences! Research Group, Era 7 Bioinformatics, Granada, Spain

Correspondence*:

Corresponding Author

Oh no sequences! Research Group, Era7 Bioinformatics, Plaza Campo Verde 3, Granada, 18001, Spain, eparejatobes@ohnosequences.com

2 ABSTRACT

- 3 No abstract yet. Will be here.
- 4 Keywords: Metagenomics, 16S, Bacterial diversity profile, Bio4j, Graph databases, Cloud computing, NGS, Genomic big data

1 INTRODUCTION

- 5 Metagenomics data analysis is growing at exponential rate during the last years. The increasing throughput
- 6 of massively parallel sequencing technologies, the derived decreasing cost, and the high impact of
- 7 metagenomics studies, especially in human health (diagnostics, treatments, drug response, prevention), are
- 8 crucial reasons responsible for this growth of Metagenomics. There is a growing interest in sequencing
- 9 all kind of microbiomes (gut, mouth, skin, urinary tract, airway, milk, bladder), in different conditions of
- 10 health and disease, or after different treatments. Metagenomics is also impacting environmental sciences,
- 11 crop sciences, agrifood sector and biotechnology in general. This new possibilities for exploring the
- 12 diversity of micro-organisms in the most diverse environments is opening many new research areas but,
- 13 due to this wide interest, it is expected that the amount of data will be overwhelming in the short time
- 14 (Stephens et al., 2015).
- Genome researchers have raised the alarm over big data in the past nature news add ref but even a more
- 16 serious challenge might be faced with the metagenomics boom/ upswing. If we compare metagenomics
- 17 data with other genomics data used in clinical genotyping we find a differential feature: the key role of
- 18 time. Thus, for example, in some longitudinal studies, serial sampling of the same patient along several
- 19 weeks (or years) is being used for the follow up of some intestinal pathologies, for studying the evolution
- 20 of gut microbiome after antibiotic treatment, or for colon cancer early detection (Zeller et al., 2014).
- 21 This need of sampling across time adds more complexity to metagenomics data storage and demands
- 22 adapted algorithms to detect state variations across time as well as idiosyncratic commonalities of the
- 23 microbiome of each individual (Franzosa et al., 2015). In addition to the intra-individual sampling-time
- 24 dependence, metagenomic clinical test results vary depending on the specific region of extraction of
- 25 the clinical specimen. This local variability adds complexity to the analysis since different localizations
- 26 (different tissues, different anatomical regions, healthy or tumour tissues) are required to have a sufficiently

complete landscape of the human microbiome. Moreover, reanalysis of old samples using new tools and
better reference databases might be also demanded from time to time.

During the last years other sciences as astronomy or particle physics are facing the big data challenge 29 but, at least, these science have standards for data processing (Stephens et al., 2015). Global standards 30 for converting raw sequence data into processed data are not yet well defined in metagenomics and 31 there are shortcomings derived from the fact that many bioinformatics methodologies currently used for 32 metagenomics data analysis were designed for a scenario very different that the current one. These are 33 some of the aspects that have suffered crucial changes and advances with a direct impact in metagenomics 34 data analysis. i. The first aspect is related to the sequences to be analyzed: the reads are larger, the 35 36 sequencing depth and the number of samples of each project are considerably bigger. The first metagenomics 37 studies were very local projects, while nowadays the most fruitful studies are done at a global level (international, continental, national). This kind of global studies has yielded the discovery of clinical 38 39 biomarkers for diseases of the importance of cancer, obesity or inflammatory bowel diseases and has allowed exploring the biodiversity in many earth environments ii. The second aspect derives from the 40 41 impressive genomics explosion, its effect being felt in this case in the reference sequences. The immense 42 amount of sequences available in public repositories demands new approaches in curation, update and storage for metagenomics reference databases: current models will or already have problems to face 43 the future avalanche of metagenomic sequences. iii. The third aspect to consider for metagenomics data 45 analysis is related to the appearance of new models for massive computation and storage and to the new programming methodologies (Scala, ...) and new cloud models and resources. The immense new 46 possibilities that these advances offer must have a direct impact in the metagenomics data analysis. iv. 48 And finally the new social manner to do science, and especially genomic science is the fourth aspect 49 to consider. Metagenomics evolves in a social and global scenario following a science democratization trend in which many small research groups from distant countries share a common big metagenomics 50 51 project. This global cooperation demands systems allowing following exactly the same pipelines using equivalent cloud resources to modularly execute the analysis in an asynchronous way of working between 52 different groups. This definitively new scenario demands new methods and tools to handle the current and 53 future volume of metagenomic data with the sufficient speed of analysis. Considering all these aspects we 54 have designed a new open source methodology for analyzing metagenomics data that exploits the new 55 possibilities that cloud computing offers to get a system robust, programmatically configurable, modular, distributed, flexible, scalable and traceable in which the biological databases of reference sequences can be 57 easily updated and/or frequently substituted by new ones or by databases specifically designed for focused 58 projects. 59

2 MATERIALS AND METHODS

50 2.1 Amazon Web Services

61 **2.2 Scala**

Scala is a hybrid object-functional programming language which runs on Java Virtual Machine. It has support for type-level programming, type-dependent types (through type members) and singleton types, which permits a restricted form of dependent types where types can depend essentially on values determined at compile time (through their corresponding singleton types). Conversely, through implicits

one can retrieve the value corresponding to a singleton type.

- The other key feature for us is Java interoperability, which let us build on the vast number of existing Java libraries; we take advantage of this when using Bio4j as an API for the NCBI taxonomy.
- 69 MG7 itself and all the libraries used are written in Scala 2.11.

70 2.3 Statika

- 71 Statika is a Scala library developed by so and so which serves as a way of defining and composing
- 72 machine behaviors statically. The main component are bundles. Each bundle declares a sequence of
- 73 computations (its behavior) which will be executed in an **environment**. A bundle can *depend* on other
- 54 bundles, and when being executed by an environment, its DAG of dependencies is linearized and run in
- 75 sequence. In our use, bundles correspond to what an EC2 instance should do and an environment to an
- 76 image (AMI: Amazon Machine Image) which prepares the basic configuration, downloads the Scala code
- 77 and runs it.

78 2.4 Datasets

- 79 Datasets is a Scala library developed by so and so to declare datasets and their locations. Data is
- 80 represented as type-indexed fields: Keys are modeled as singleton types, and values correspond to what
- 81 could be called a denotation of the key: a value of type Location tagged with the key type. Then a
- 82 **Dataset** is essentially a collection of data, which are guaranteed statically to be different through type-level
- 83 predicates, making use of the value type correspondence which can be established through singleton types
- 84 and implicits. A dataset location is then just a list of locations formed by locations of each data member of
- 85 that dataset.
- Data keys can further have a reference to a **data type**, which, as the name hints at, can help in providing
- 87 information about the type of data we are working with. For example, when declaring Illumina reads as a
- 88 data, a data type containing information about the read length, insert size or end type (single or paired) is
- 89 used.
- A **location** can be, for example, an S3 object or a local file; by leaving the location type used to denote
- 91 particular data free we can work with different "physical" representations, while keeping track of to which
- 92 logical data they are a representation of. Thus, a process can generate locally a .fastq file representing
- 93 the merged reads, while another can put it in S3 with the fact that they all correspond to the "same" merged
- 94 reads is always present, as the data that those "physical" representations denote.

95 **2.5 Loquat**

- Loquat is a library developed by **so and so** designed for the execution of embarrassingly parallel tasks
- 97 using S3, SQS and EC2.
- 98 A **loquat** executes a process with explicit input and output datasets (declared using the *Datasets* library
- 99 described above). Workers (EC2 instances) read from an SQS queue the S3 locations for both input and
- output data; then they download the input to local files, and pass these file locations to the process to be
- 101 executed. The output is then put in the corresponding S3 locations.
- A manager instance is used to monitor workers, provide initial data to be put in the SQS queue and
- 103 optionally release resources depending on a set of configurable conditions.
- Both worker and manager instances are Statika bundles. In the case of the worker, it can declare any
- 105 dependencies needed to perform its task: other tools, libraries, or data.

Frontiers 3

- All configuration such as the number of workers or the instance types is declared statically, the specification of a loquat being ultimately a Scala object. There are deploy and resource management methods, making it easy to use an existing loquat either as a library or from (for example) a Scala REPL.
- The input and output (and their locations) being defined statically has several critical advantages. First,
- 110 composing different loquats is easy and safe; just use the output types and locations of the first one as input
- 111 for the second one. Second, data and their types help in not mixing different resources when implementing
- 112 a process, while serving as a safe and convenient mechanism for writing generic processing tasks. For
- 113 example, merging paired-end Illumina reads generically is easy as the data type includes the relevant
- 114 information (insert size, read length, etc) to pass to a tool such as FLASH.

115 2.6 Type-safe DSLs for BLAST and FLASH

- We developed our own type-safe DSLs (Domain Specific Language) for FLASH and BLAST expressions
- 117 and their execution.
- 118 2.6.1 BLAST DSL
- In the case of BLAST we use a model for expressions where we can guarantee for each BLAST command
- 120 expression at compile time
- all required arguments are provided
- only valid options are provided
- correct types for each option value
- valid output record specification
- Generic type-safe parsers returning an heterogeneous record of BLAST output fields are also available,
- 126 together with output data defined using *Datasets* which have a reference to the exact BLAST command
- 127 options which yielded that output. This let us provide generic parsers for BLAST output which are
- 128 guaranteed to be correct, for example.
- 129 2.6.2 FLASH DSL
- 130 In the same spirit as for BLAST,
- 131 **2.7 Bio4j**
- Bio4j is a data platform integrating data from different resources such as UniProt or GO in a graph data
- paradigm. We use the module containing the NCBI Taxonomy, and the use their Java API from Scala in the
- 134 assignment phase.

3 RESULTS

35 3.1 Overview

- To tackle the challenges posed by metagenomics big data analysis outlined in the Introduction,
- AWS resources in Scala (??) A new approach to data analysis specification, management and
- 138 specification based on working with it in exactly the same way as for a software project, together with the
- 139 extensive use of compile-time structures and checks. Parallelization and distributed analysis based on
- 140 AWS, with on-demand infrastructure as the basic paradigm fully automated processes, data and cloud
- 141 resources management. Static reproducible specification of dependencies and behavior of the different

- 142 components using Statika and Datasets Definition of complex pipelines using Loquat a composable
- 143 system for scaling/parallelizing stateless computations especially designed for Amazon Web Services
- 144 (AWS) Modeling of the taxonomy tree using the new paradigm of graph databases (Bio4j). It facilitates
- 145 the taxonomic assignment tasks and the calculation of the taxa abundance values considering the hierarchic
- 146 structure of taxonomy tree (cumulative values). per-read assignment (??)

147 3.2 16S Reference Database Construction

- 148 Our 16S Reference Database is a curated subset of sequences from NCBI nucleotide database nt. The 149 sequences included were selected by similarity with the bacterial and archaeal reference sequences downloaded from the RDP database (Cole et al., 2013). RDP unaligned sequences were used to 150 capture new 16S sequences from **nt** using BLAST similarity search strategies and then, performing 151 152 additional curation steps to remove sequences with poor taxonomic assignments to taxonomic nodes close to the root of the taxonomy tree. All the nucleotide sequences included in nt database has a 153 taxonomic assignment provided by the Genbank sequence submitter. NCBI provides a table (available 154 155 at ftp://ftp.ncbi.nlm.nih.gov/pub/taxonomy/) to do the mapping of any Genbank Identifier (GI) to its Taxonomy Identifier (TaxID). Thus, we are based on a crowdsourced submitter-maintained taxonomic 156 annotation system for reference sequences. It supposes a sustainable system able to face the expected 157 158 number of reference sequences that will populate the public global nucleotide databases in the near future. Another advantageous point is that we are based on NCBI taxonomy, the *de facto* standard taxonomic 159 160 classification for biomolecular data (Cochrane and Galperin, 2010). NCBI taxonomy is, undoubtedly, the most used taxonomy all over the world and the most similar to the official taxonomies of each specific field. 161 This is a crucial point because all the type-culture and tissue databanks follow this official taxonomical 162 163 classification and, in addition, all the knowledge accumulated during last decades is referred to this 164 taxonomy. In addition NCBI provides a direct connection between taxonomical formal names and the 165 physical specimens that serve as exemplars for the species (Federhen, 2014).
- 166 Certainly, if metagenomics results are easily integrated with the theoretical and experimental knowledge of each specific area, the impact of metagenomics will be higher that if metagenomics progresses as a 167 disconnected research branch. Considering that metagenomics data interoperability, which is especially 168 critical in clinical environments, requires a stable taxonomy to be used as reference, we decided to rely on 169 the most widely used taxonomy: the NCBI taxonomy. In addition, the biggest global sequence database 170 GenBank follows this taxonomy to register the origin of all their submitted sequences. Our 16S database 171 building strategy allows the substitution of the 16S database by any other subset of **nt**, even by the complete 172 **nt** database if it would be needed, for example, for analyzing shotgun metagenomics data. This possibility 173 of changing the reference database provides flexibility to the system enabling it for easy updating and 174 project-driven personalization. 175
- 176 3.3 Bio4j and Graph Databases
- 177 3.4 MG7 Pipeline Description
- 178 3.5 Taxonomic Assignment Algorithms
- 179 3.5.1 Lowest Common Ancestor based Taxonomic Assignment
- 180 For each read:
- 1. Select only one BLASTN alignment (HSP) per reference sequence (the HSP with lowest e value) 2.
- 182 Filter all the HSPs with bitscore below a defined BLASTN bitscore threshold s_0 3. Find the best bitscore

Frontiers 5

- value S in the set of BLASTN HSPs corresponding to hits of that read 4. Filter all the alignments with
- bitscore below p * S (where p is a fixed by the user coefficient to define the bitscore required, e.g. if p=0.9 184
- and S=700 the required bitscore threshold would be 630) 5. Select all the taxonomic nodes to which map 185
- the reference sequences involved in the selected HSPs: If all the selected taxonomic nodes forms a line 186
- in the taxonomy tree (are located in a not branched lineage to the tree root) we should choose the most 187
- specific taxID as the final assignment for that read If not, we should search for the (sensu stricto) Lowest 188
- Common Ancestor (LCA) of all the selected taxonomic nodes (See Figure X) 189
- In this approach the value used for evaluating the similarity is the bitscore that is a value that increases 190
- when similarity is higher and depends a lot on the length of the HSP 191
- 3.5.2 Best BLAST hit taxonomic assignment
- We have maintained the simpler method of Best BLAST Hit (BBH) taxonomic assignment because, in 193
- some cases, it can provide information about the sequences that can be more useful than the obtained using 194
- LCA algorithm. Using LCA algorithm when some reference sequences with BLAST alignments over the 195
- required thresholds map to a not sufficiently specific taxID, the read can be assigned to an unspecific taxon
- near to the root. If the BBH reference sequence maps to a more specific taxa this method, in that case, gives 197
- us useful information. 198

Using MG7 with some example data-sets 3.6 199

We selected the datasets described in [Kennedy-2014] (??) 200

3.7 MG7 availability 201

MG7 is open source, available at https://github.com/ohnosequences/mg7 under an AGPLv3 license. 202

DISCUSSION

4.1 What MG7 brings

- We could summarize the most innovative ideas and developments in MG7: 204
- 1. Treat data analysis as a software project. This makes for radical improvements in *reproducibility*, *reuse*, 205 versioning, safety, automation and expressiveness 206
- 2. input and output data, their locations and type are expressible and checked at compile-time using our 207
- Scala library datasets 3. management of dependencies and machine configurations using our Scala 208
- library Statika 209
- 3. automation of AWS cloud resources and processes, including distribution and parallelization through 210 the use of *Loquat* 211
- 4. taxonomic data and related operations are treated natively as what they are: graphs, through the use of 212 Bio4i 213
- 214 5. MG7 provides a sustainable model for taxonomic assignment, appropriate to face the challenging amount of data that high throughput sequencing technologies generate 215

4.2 A new approach to data analysis 216

- General approach. An analysis is defined as a software project. It can evolve in the same way. We can run 217
- the analysis in a test phase, review configuration and changes, etc. Key advantages of this approach are 218

- **Reproducibility** the same analysis can be run again with exactly the same configuration in a trivial way.
- **Versioning** The analysis is a software project so it goes through the same stages, there can be different versions, stable releases, etc.
- Reuse we can build standard configurations on top of this and reuse them for subsequent data analysis.
- **Decoupling** We can start working on the analysis specification, without any need for data in a much easier way.
- Expresiveness and safety choose only from valid Illumina read types, build default FLASH command based on that, ...
- 228 4.3 Inputs, outputs, data: compile-time, expressive, composable
- 229 4.4 Tools, data, dependencies and machine configurations
- 230 4.5 Parallel cloud execution ??
- 231 4.6 Taxonomy and Bio4j
- 232 4.7 Future-proof
- 233 4.8 MG7 Future developments
- Other possible uses of the general schema: statika, loquat, ...
- 235 4.8.1 Comparison of groups of samples
- 236 4.8.2 Interactive visualizations using the output files of MG7 (Biographika project)

5 ACKNOWLEDGEMENTS

237 INTERCROSSING (Grant 289974)

REFERENCES

- 238 Cochrane, G. R. and Galperin, M. Y. (2010). The 2010 nucleic acids research database issue and online
- database collection: a community of data resources. Nucleic acids research 38, D1–D4
- 240 Cole, J. R., Wang, Q., Fish, J. A., Chai, B., McGarrell, D. M., Sun, Y., et al. (2013). Ribosomal database
- project: data and tools for high throughput rrna analysis. *Nucleic acids research*, gkt1244
- 242 Federhen, S. (2014). Type material in the ncbi taxonomy database. *Nucleic acids research*, gku1127
- 243 Franzosa, E. A., Huang, K., Meadow, J. F., Gevers, D., Lemon, K. P., Bohannan, B. J., et al. (2015).
- 244 Identifying personal microbiomes using metagenomic codes. *Proceedings of the National Academy of*
- 245 *Sciences*, 201423854
- 246 Stephens, Z. D., Lee, S. Y., Faghri, F., Campbell, R. H., Zhai, C., Efron, M. J., et al. (2015). Big data:
- 247 Astronomical or genomical? *PLoS Biol* 13, e1002195
- 248 Zeller, G., Tap, J., Voigt, A. Y., Sunagawa, S., Kultima, J. R., Costea, P. I., et al. (2014). Potential of fecal
- 249 microbiota for early-stage detection of colorectal cancer. *Molecular systems biology* 10, 766

Frontiers 7