

# Lesson 4.

## Iterated filtering: principles and practice

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## 1 Introduction

### Introduction

- This tutorial covers likelihood estimation via the method of iterated filtering.
- It presupposes familiarity with building partially observed Markov process (POMP) objects in the R package **pomp** (King *et al.*, 2016).
- This tutorial follows on from the [topic of particle filtering](#) (also known as sequential Monte Carlo) via **pfilter** in **pomp**.

### Objectives

1. To review the available options for inference on POMP models, to put iterated filtering in context.
2. To understand how iterated filtering algorithms carry out repeated particle filtering operations, with randomly perturbed parameter values, in order to maximize the likelihood.
3. To gain experience carrying out statistical investigations using iterated filtering in a relatively simple situation: fitting an SIR model to data from a measles outbreak.

## 2 Classification of statistical methods for POMP models

### Classification of statistical methods for POMP models

- Many, many statistical methods have been proposed for inference on POMP models (He *et al.*, 2010; King *et al.*, 2016).
- The volume of research indicates both the importance and the difficulty of the problem.
- Let's start by considering three criteria to categorize inference methods:
  - the plug-and-play property
  - full-information or feature-based
  - frequentist or Bayesian

### The plug-and-play property

#### Plug-and-play (also called simulation-based) methods

- Inference methodology that calls `rprocess` but not `dprocess` is said to be *plug-and-play*. All popular modern Monte Carlo methods fall into this category.
- “Simulation-based” is equivalent to “plug-and-play”.
- Historically, simulation-based meant simulating forward from initial conditions to the end of the time series.
- However, particle filtering methods instead consider each observation interval sequentially. They carry out multiple, carefully selected, simulations over each interval.
- Plug-and-play methods can call `dmeasure`. A method that uses only `rprocess` and `rmeasure` is called “doubly plug-and-play”.
- Two *non-plug-and-play* methods—expectation-maximization (EM) and Markov chain Monte Carlo (MCMC)—have theoretical convergence problems for nonlinear POMP models. The failures of these two workhorses of statistical computation have prompted development of alternative methodologies.

### Full information vs. feature-based methods

#### Full-information and feature-based methods

- *Full-information* methods are defined to be those based on the likelihood function for the full data (i.e., likelihood-based frequentist inference and Bayesian inference).
- *Feature-based* methods either consider a summary statistic (a function of the data) or work with an alternative to the likelihood.
- Asymptotically, full-information methods are statistically efficient and feature-based methods are not.
- In some cases, loss of statistical efficiency might be an acceptable tradeoff for advantages in computational efficiency.
- However:
  - Good low-dimensional summary statistics can be hard to find.

- When using statistically inefficient methods, it can be hard to know how much information you are losing.
- Intuition and scientific reasoning can be inadequate tools to derive informative low-dimensional summary statistics (Shrestha *et al.*, 2011; Ionides, 2011).

## Bayesian vs. frequentist approaches

### Bayesian and frequentist methods

- Recently, plug-and-play Bayesian methods have been discovered:
  - particle Markov chain Monte Carlo (PMCMC) (Andrieu *et al.*, 2010).
  - approximate Bayesian computation (ABC) (Toni *et al.*, 2009).
- Prior belief specification is both the strength and weakness of Bayesian methodology:
- The likelihood surface for nonlinear POMP models often contains nonlinear ridges and variations in curvature.
- These situations bring into question the appropriateness of independent priors derived from expert opinion on marginal distributions of parameters.
- They also are problematic for specification of “flat” or “uninformative” prior beliefs.
- Expert opinion can be treated as data for non-Bayesian analysis. However, our primary task is to identify the information in the data under investigation, so it can be helpful to use methods that do not force us to make our conclusions dependent on quantification of prior beliefs.

## Summary

### POMP inference methodologies

	Frequentist	Bayesian
Plug-and-play		
Full-information	iterated filtering	particle MCMC
Feature-based	simulated moments synthetic likelihood (SL) nonlinear forecasting	ABC SL-based MCMC
Not plug-and-play		
Full-information	EM algorithm Kalman filter	MCMC
Feature-based	Yule-Walker <sup>1</sup> extended Kalman filter??	extended Kalman filter <sup>2</sup>

## 3 Iterated filtering in theory

### Full-information, plug-and-play, frequentist methods

- Iterated filtering methods (Ionides *et al.*, 2006, 2015) are the only currently available, full-information, plug-and-play, frequentist methods for POMP models.
- Iterated filtering methods have been shown to solve likelihood-based inference problems for epidemiological situations which are computationally intractable for available Bayesian methodology (Ionides *et al.*, 2015).

### An iterated filtering algorithm (IF2)

We focus on the IF2 algorithm of Ionides *et al.* (2015). In this algorithm:

- Each iteration consists of a particle filter, carried out with the parameter vector, for each particle, doing a random walk.
- At the end of the time series, the collection of parameter vectors is recycled as starting parameters for the next iteration.
- The random-walk variance decreases at each iteration.

In theory, this procedure converges toward the region of parameter space maximizing the maximum likelihood. In practice, we can test this claim on examples.

### IF2 algorithm pseudocode

#### Input:

- simulators for  $f_{X_0}(x_0; \theta)$  and  $f_{X_n|X_{n-1}}(x_n|x_{n-1}; \theta)$ ;
- evaluator for  $f_{Y_n|X_n}(y_n|x_n; \theta)$ ;
- data,  $y_{1:N}^*$

#### Algorithmic parameters:

- number of iterations,  $M$ ;
- number of particles,  $J$ ;
- initial parameter swarm,  $\{\Theta_j^0, j = 1, \dots, J\}$ ;
- perturbation density,  $h_n(\theta|\varphi; \sigma)$ ;
- perturbation scale,  $\sigma_{1:M}$

#### Output:

- final parameter swarm,  $\{\Theta_j^M, j = 1, \dots, J\}$

### IF2 algorithm pseudocode II

#### Procedure:

1. For  $m$  in  $1:M$
2.      $\Theta_{0,j}^{F,m} \sim h_0(\theta|\Theta_j^{m-1}; \sigma_m)$  for  $j$  in  $1:J$
3.      $X_{0,j}^{F,m} \sim f_{X_0}(x_0; \Theta_{0,j}^{F,m})$  for  $j$  in  $1:J$
4.     For  $n$  in  $1:N$
5.          $\Theta_{n,j}^{P,m} \sim h_n(\theta|\Theta_{n-1,j}^{F,m}, \sigma_m)$  for  $j$  in  $1:J$
6.          $X_{n,j}^{P,m} \sim f_{X_n|X_{n-1}}(x_n|X_{n-1,j}^{F,m}; \Theta_{n,j}^{P,m})$  for  $j$  in  $1:J$
7.          $w_{n,j}^m = f_{Y_n|X_n}(y_n^*|X_{n,j}^{P,m}; \Theta_{n,j}^{P,m})$  for  $j$  in  $1:J$
8.         Draw  $k_{1:J}$  with  $P[k_j = i] = w_{n,i}^m / \sum_{u=1}^J w_{n,u}^m$
9.          $\Theta_{n,j}^{F,m} = \Theta_{n,k_j}^{P,m}$  and  $X_{n,j}^{F,m} = X_{n,k_j}^{P,m}$  for  $j$  in  $1:J$
10.     End For
11.     Set  $\Theta_j^m = \Theta_{N,j}^{F,m}$  for  $j$  in  $1:J$
12. End For

## IF2 algorithm pseudocode III

### Remarks:

- The  $N$  loop (lines 4 through 10) is a basic particle filter applied to a model with stochastic perturbations to the parameters.
- The  $M$  loop repeats this particle filter with decreasing perturbations.
- The superscript  $F$  in  $\Theta_{n,j}^{F,m}$  and  $X_{n,j}^{F,m}$  denote solutions to the *filtering problem*, with the particles  $j = 1, \dots, J$  providing a Monte Carlo representation of the conditional distribution at time  $n$  given data  $y_{1:n}^*$  for filtering iteration  $m$ .
- The superscript  $P$  in  $\Theta_{n,j}^{P,m}$  and  $X_{n,j}^{P,m}$  denote solutions to the *prediction problem*, with the particles  $j = 1, \dots, J$  providing a Monte Carlo representation of the conditional distribution at time  $n$  given data  $y_{1:n-1}^*$  for filtering iteration  $m$ .
- The *weight*  $w_{n,j}^m$  gives the likelihood of the data at time  $n$  for particle  $j$  in filtering iteration  $m$ .

### Analogy with evolution by natural selection

- The parameters characterize the *genotype*.
- The swarm of particles is a *population*.
- The likelihood, a measure of the compatibility between the parameters and the data, is the analogue of *fitness*.
- Each successive observation is a new *generation*.
- Since particles reproduce in each generation in proportion to their likelihood, the particle filter acts like *natural selection*.
- The artificial perturbations augment the “genetic” variance and therefore correspond to *mutation*.
- IF2 increases the *fitness* of the population of particles.
- However, because our scientific interest focuses on the model without the artificial perturbations, we decrease the intensity of the latter with successive iterations.

## 4 Iterated filtering in practice

### An example problem

#### Applying IF2 to the Consett measles outbreak

To apply IF2 to a relatively simple epidemiological example, we consider fitting a stochastic SIR model to data from a measles outbreak in the small English town of Consett.

Reports consist of the number of cases for each week of the year.

The population of the town was approximately 38,000 and, since the outbreak is confined to less than one year, we will ignore births and deaths.

The data are available on the course website:

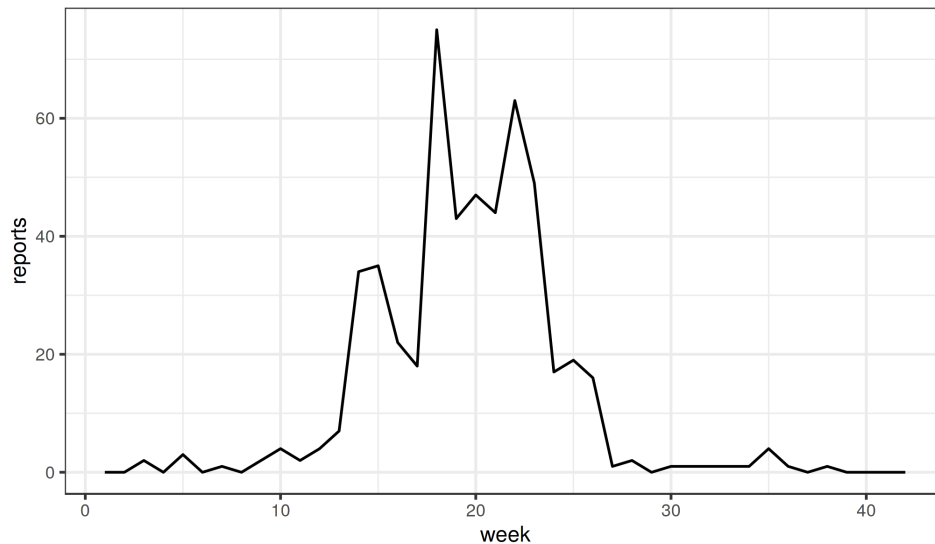
```
library(tidyverse)

courseurl <- "https://kingaa.github.io/sbied/"
datafile <- "mif/Measles_Consett_1948.csv"
```

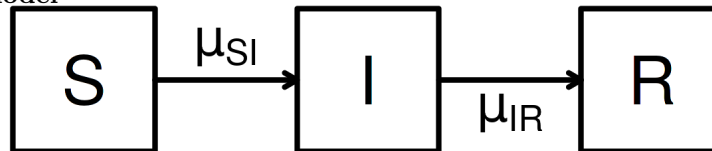
```
read_csv(paste0(courseurl,datafile)) %>%
  select(week,reports=cases) %>%
  filter(week<=42) -> dat
```

## Applying IF2 to the Consett measles outbreak II

```
dat %>%
  ggplot(aes(x=week,y=reports))+
  geom_line()
```



## Stochastic SIR model



- Our model is a variation on a basic SIR Markov chain
- State:  $X(t) = (S(t), I(t), R(t))$ ; numbers of hosts in susceptible, infectious, and recovered classes.
- Assume: a single infection in week 0, i.e., that  $I(0) = 1$ .
- Markov transmission model:
  - Each individual in  $S$  transitions to  $I$  at rate  $\mu_{SI} = \beta I(t)/N$ .
  - Each individual in  $I$  transitions at rate  $\mu_{IR}$  to  $R$ .
  - $1/\mu_{IR}$  is the mean infectious period.
- All rates will have units  $\text{wk}^{-1}$ .
- This model has limitations and weaknesses, but writing down and fitting a model is a starting point for data analysis, not an end point.
- In particular, having fit one model, one should certainly examine alternative models.

- For example, one could include a latency period for infections, or one could modify the model to give a better description of the diagnosis, bed-confinement, and convalescence processes.
- Notice that we do not need to track  $R$  since this variable has consequences neither for the dynamics of the state process nor for the data.

## Implementation in pomp

As before, we code the model using C snippets

```
library(pomp)

sir_step <- Csnippet("
double dN_SI = rbinom(S,1-exp(-Beta*I/N*dt));
double dN_IR = rbinom(I,1-exp(-mu_IR*dt));
S -= dN_SI;
I += dN_SI - dN_IR;
H += dN_IR;
")
```

The `rinit` component:

```
sir_init <- Csnippet("
S = nearbyint(eta*N);
I = 1;
H = 0;
")
```

The measurement model:

```
dmeas <- Csnippet("
lik = dbinom(reports,H,rho,give_log);
")

rmeas <- Csnippet("
reports = rbinom(H,rho);
")
```

```
dat %>%
  pomp(
    times="week",t0=0,
    rprocess=euler(sir_step,delta.t=1/7),
    rinit=sir_init,
    rmeasure=rmeas,
    dmeasure=dmeas,
    accumvars="H",
    partrans=parameter_trans(
      log=c("Beta"),
      logit=c("rho","eta")
    ),
    statenames=c("S","I","H"),
    paramnames=c("Beta","mu_IR","eta","rho","N")
  ) -> measSIR
```

## Testing the codes

To develop and debug code, it is useful to have testing codes that run quickly and fail if the codes are not working correctly.

As such a test, here we run some simulations and a particle filter.

We'll use the following parameters, derived from our earlier explorations:

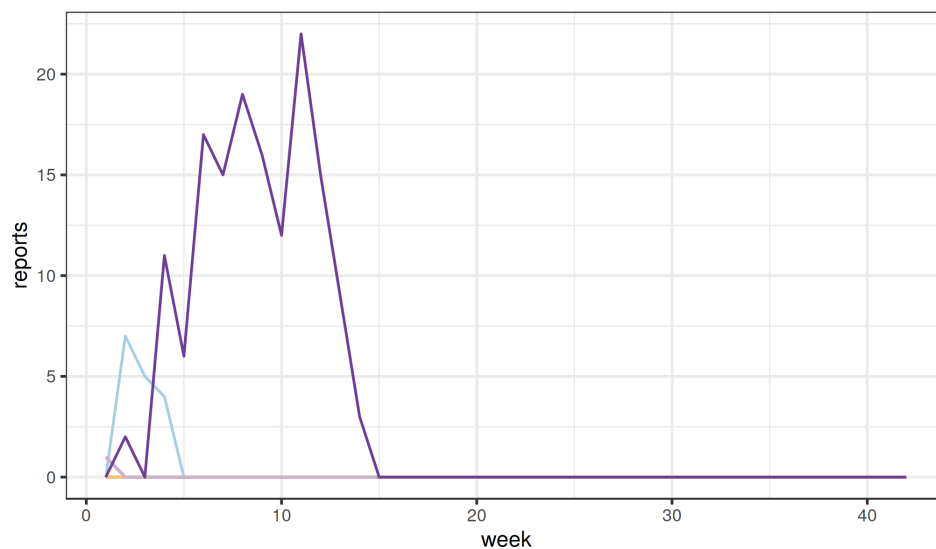
```
params <- c(Beta=20,mu_IR=2,rho=0.5,eta=0.1,N=38000)
```

## Testing the codes: simulation

Now to run and plot some simulations:

```
measSIR %>%  
  simulate(params=params,nsim=10,format="data.frame") -> y
```

```
y %>%  
  ggplot(aes(x=week,y=reports,group=.id,color=factor(.id)))+  
  geom_line()+  
  scale_color_brewer(type="qual",palette=3)+  
  guides(color=FALSE)
```



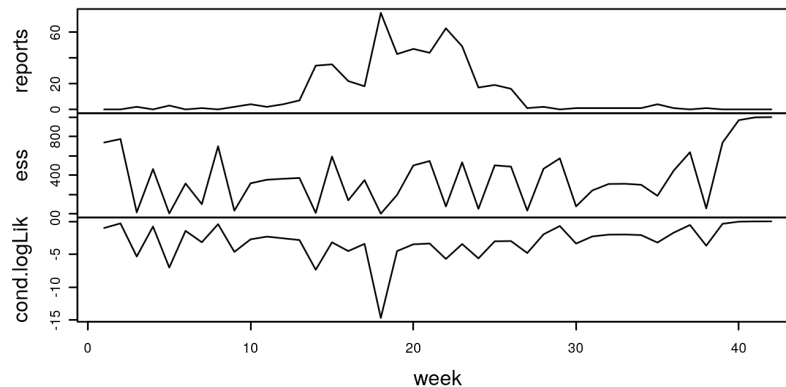
## Testing the codes: filtering

Before engaging in iterated filtering, it is a good idea to check that the basic particle filter is working since we can't iterate something unless we can run it once! The simulations above check the `rprocess` and `rmeasure` codes; the particle filter depends on the `rprocess` and `dmeasure` codes and so is a check of the latter.

```
measSIR %>%  
  pfilter(Np=1000,params=params) -> pf
```

```
plot(pf)
```





The above plot shows the data (**reports**), along with the *effective sample size* (ESS) of the particle filter (**ess**) and the log likelihood of each observation conditional on the preceding ones (**cond.logLik**). The ESS is the equivalent number of independent particles. In this case, the ESS is very small at several points.

## Setting up the estimation problem

### Setting up the estimation problem

Let's assume that the population size,  $N$ , is known accurately. We'll fix that parameter.

Let's also imagine that we have access to the results of household and clinical studies that have concluded that infected patients shed the virus for 3–4 da. We'll use these results to constrain the infectious period in our model to 3.5 da, i.e.,  $\mu_{IR} = 2 \text{ wk}^{-1}$ . Later, we'll relax this assumption.

```
fixed_params <- c(N=38000, mu_IR=2)
```

We proceed to estimate  $\beta$ ,  $\eta$ , and  $\rho$ .

### Parallel computing

It will be helpful to parallelize most of the computations. Most machines nowadays have multiple cores and using this computational capacity is as simple as:

- (i) letting R know you plan to use multiple processors;
- (ii) using the parallel for loop provided by the **foreach** package; and
- (iii) paying proper attention to the use of parallel random number generators (RNG).

For example:

```
library(foreach)
library(doParallel)
registerDoParallel()
```

The first two lines above load the **foreach** and **doParallel** packages, the latter being a “backend” for the **foreach** package. The next line tells **foreach** that we will use the **doParallel** backend. By default, R will guess how many cores are available and will run about half this number of concurrent R processes.

## Parallel random number generators (RNG)

To initialize a parallel RNG, we use the **doRNG** package. The following ensures that the parallel computations will be both mutually independent and reproducible.

```
library(doRNG)
registerDoRNG(625904618)
```

## Running a particle filter

We proceed to carry out replicated particle filters at an initial guess of  $\beta = 20$ ,  $\eta = 0.1$ , and  $\rho = 0.5$ .

```
foreach(i=1:10,.combine=c) %dopar% {
  library(pomp)
  measSIR %>% pfilter(params=params,Np=10000)
} -> pf

pf %>% logLik() %>% logmeanexp(se=TRUE) -> L_pf
L_pf
```

	se
-126.5611272	0.4642203

In 1.24 seconds, using 10 cores, we obtain an unbiased likelihood estimate of -126.6 with a Monte Carlo standard error of 0.46.

## Building up a picture of the likelihood surface

- Given a model and a set of data, the likelihood surface is well defined, though it may be difficult to visualize.
- We can develop a progressively more complete picture of this surface by storing likelihood estimates whenever we compute them.
- It is a very good idea to set up a database within which to store the likelihood of every point for which we have an estimated likelihood.
- This will become larger and more complete as our parameter-space search goes on and will be a basis for a variety of explorations.

At this point, we've computed the likelihood at a single point. Let's store this point, together with the estimated likelihood and our estimate of the standard error on that likelihood, in a CSV file:

```
pf[[1]] %>% coef() %>% bind_rows() %>%
  bind_cols(loglik=L_pf[1],loglik.se=L_pf[2]) %>%
  write_csv("measles_params.csv")
```

## A local search of the likelihood surface

### A local search of the likelihood surface

Let's carry out a local search using `mif2` around this point in parameter space.

- We need to choose the `rw.sd` and `cooling.fraction.50` algorithmic parameters.

- Since  $\beta$  and  $\mu_{IR}$  will be estimated on the log scale, and we expect that multiplicative perturbations of these parameters will have roughly similar effects on the likelihood, we'll use a perturbation size of 0.02, which we imagine will have a small but non-negligible effect.
- For simplicity, we'll use the same perturbation size on  $\rho$ .
- We fix `cooling.fraction.50=0.5`, so that after 50 `mif2` iterations, the perturbations are reduced to half their original magnitudes.

```
foreach(i=1:20,.combine=c) %dopar% {
  library(pomp)
  library(tidyverse)
  measSIR %>%
    mif2(
      params=params,
      Np=2000, Nmif=50,
      cooling.fraction.50=0.5,
      rw.sd=rw.sd(Beta=0.02, rho=0.02, eta=ivp(0.02))
    )
} -> mifs_local
```

## Windows issues

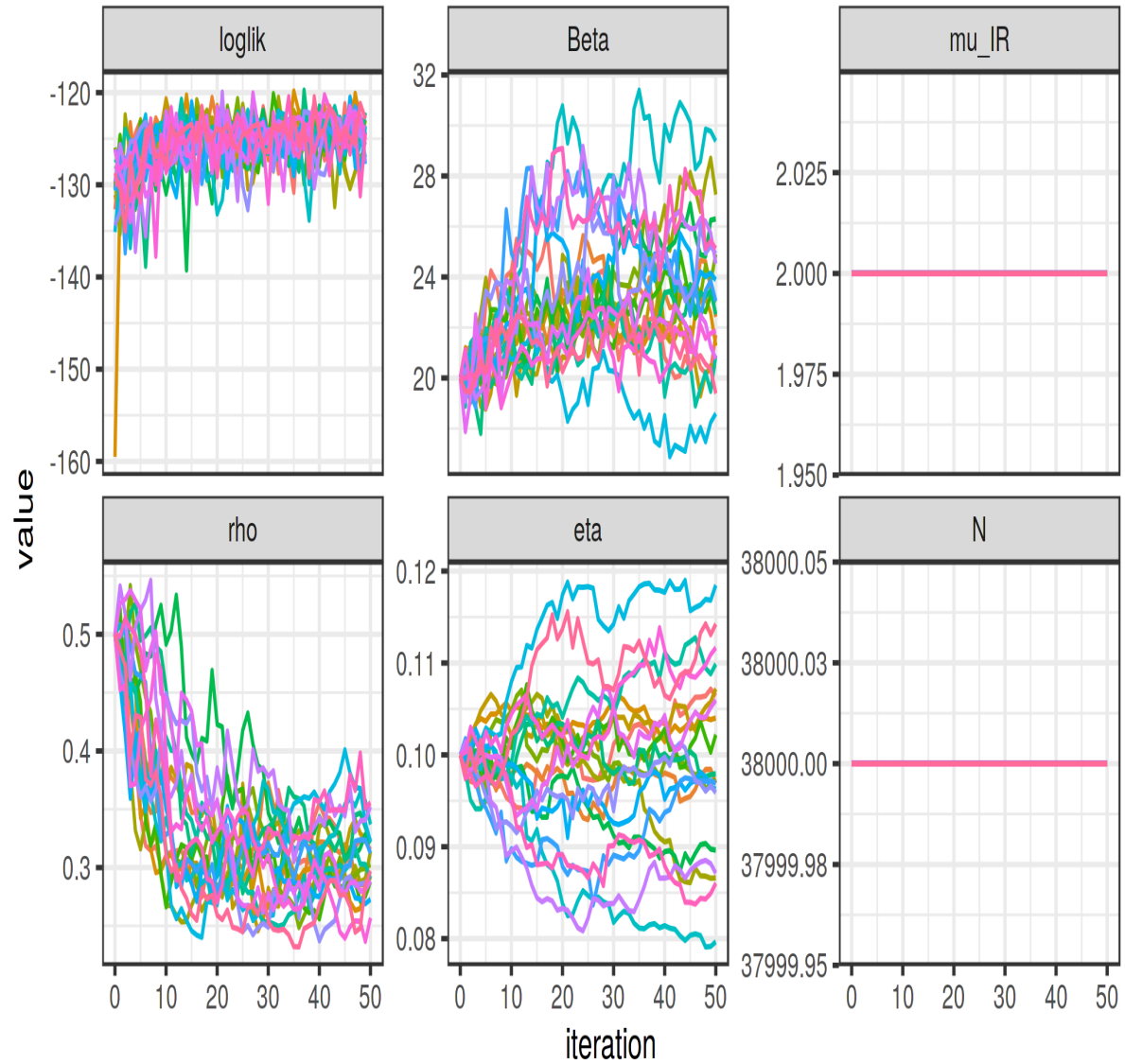
**NB:** Some Windows users have reported trouble with the above code. This appears to be due to certain Windows security features. It has been possible to circumvent this problem by adding `cdir="."` and `cfile=<filename>` as arguments in the call that created `measSIR`. Thus, for example,

```
dat %>%
  pomp(
    times="week", t0=0,
    rprocess=euler(sir_step, delta.t=1/7),
    rinit=sir_init, rmeasure=rmeas, dmeasure=dmeas,
    partrans=parameter_trans(log="Beta", logit=c("rho", "eta")),
    accumvars="H", statenames=c("S", "I", "H"),
    paramnames=c("Beta", "mu_IR", "eta", "rho", "N"),
    cdir=".", cfile="measSIR"
  ) -> measSIR
```

## Iterated filtering diagnostics

We obtain some diagnostic plots with the `plot` command applied to `mifs_local`. Here is a way to get a prettier version:

```
mifs_local %>%
  traces() %>%
  melt() %>%
  ggplot(aes(x=iteration, y=value, group=L1, color=factor(L1)))+
  geom_line()+
  guides(color=FALSE)+
  facet_wrap(~variable, scales="free_y")
```



- We see that the likelihood eventually increases as the iterations proceed, though there is considerable variability due to
  - (a) the poorness of our starting guess and
  - (b) the stochastic nature of this Monte Carlo algorithm.
- We see movement in the parameters, though considerable variability remains.

### Estimating the likelihood

Although the filtering carried out by `mif2` in the final filtering iteration generates an approximation to the likelihood at the resulting point estimate, this is not good enough for reliable inference.

- Partly, this is because parameter perturbations are applied in the last filtering iteration, so that the likelihood reported by `mif2` is not identical to that of the model of interest.

- Partly, this is because `mif2` is usually carried out with fewer particles than are needed for a good likelihood evaluation.

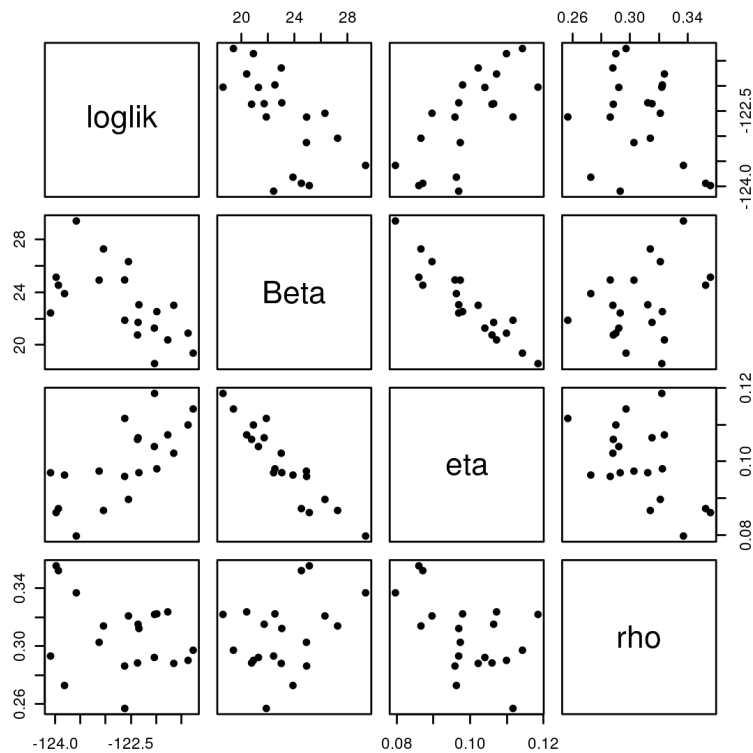
Therefore, we evaluate the likelihood, together with a standard error, using replicated particle filters at each point estimate.

```
foreach(mf=mifs_local,.combine=rbind) %dopar% {
  library(pomp)
  library(tidyverse)
  evals <- replicate(10, logLik(pfilter(mf,Np=20000)))
  ll <- logmeanexp(evals,se=TRUE)
  mf %>% coef() %>% bind_rows() %>%
    bind_cols(loglik=ll[1],loglik.se=ll[2])
} -> results
```

On 12 processors, this local investigation took 16 sec for the maximization and 24 sec for the likelihood evaluation.

These repeated stochastic maximizations can also show us the geometry of the likelihood surface in a neighborhood of this point estimate:

```
pairs(~loglik+Beta+eta+rho,data=results,pch=16)
```



### Building up a picture of the likelihood surface

This plot shows a hint of a ridge in the likelihood surface (cf. the  $\beta$ - $\eta$  panel). However, the sampling is as yet too sparse to give a clear picture.

We add these newly explored points to our database,

```
read_csv("measles_params.csv") %>%
  bind_rows(results) %>%
  arrange(-loglik) %>%
  write_csv("measles_params.csv")
```

and move on to a more thorough exploration of the likelihood surface.

## 5 Searching for the MLE

### A global search

#### A global search of the likelihood surface

- When carrying out parameter estimation for dynamic systems, we need to specify beginning values for both the dynamic system (in the state space) and the parameters (in the parameter space).
- To avoid confusion, we use the term “initial values” to refer to the state of the system at  $t_0$  and “starting values” to refer to the point in parameter space at which a search is initialized.
- Practical parameter estimation involves trying many starting values for the parameters.
- One way to approach this is to choose a large box in parameter space that contains all remotely sensible parameter vectors.
- If an estimation method gives stable conclusions with starting values drawn randomly from this box, this gives some confidence that an adequate global search has been carried out.

For our measles model, a box containing reasonable parameter values might be  $\beta \in (5, 80)$ ,  $\rho \in (0.2, 0.9)$ ,  $\eta \in (0, 0.4)$ .

We are now ready to carry out likelihood maximizations from diverse starting points.

```
set.seed(2062379496)

runif_design(
  lower=c(Beta=5, rho=0.2, eta=0),
  upper=c(Beta=80, rho=0.9, eta=0.4),
  nseq=300
) -> guesses

mf1 <- mifs_local[[1]]

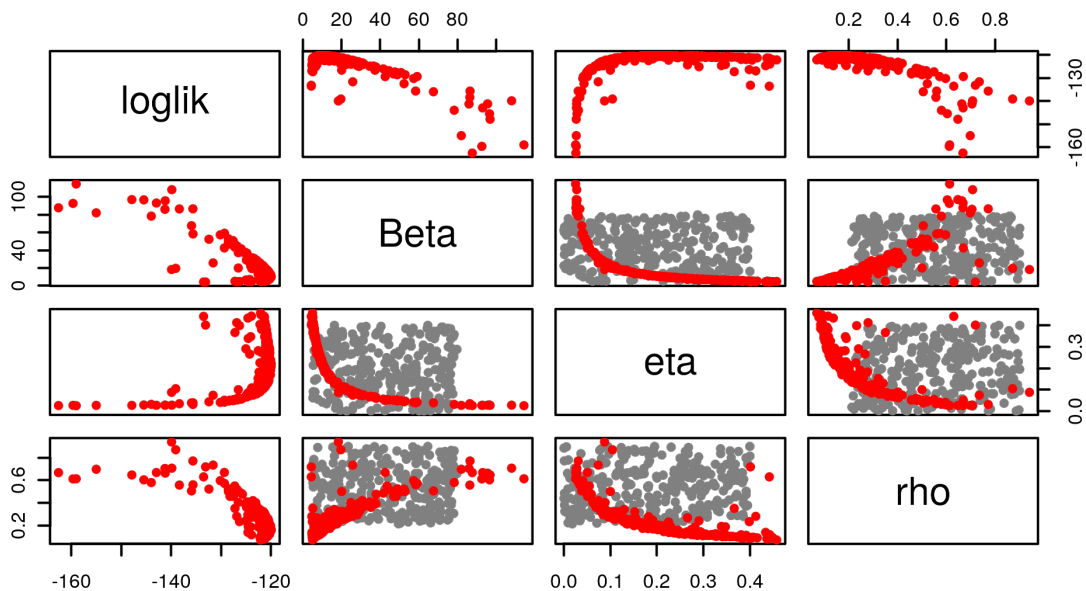
registerDoRNG(1270401374)
foreach(guess=iter(guesses, "row"), .combine=rbind) %dopar% {
  library(pomp)
  library(tidyverse)
  mf1 %>%
    mif2(params=c(unlist(guess), fixed_params)) %>%
    mif2(Nmif=100) -> mf
  replicate(
    10,
    mf %>% pfilter(Np=100000) %>% logLik()
  ) %>%
    logmeanexp(se=TRUE) -> ll
  mf %>% coef() %>% bind_rows() %>%
    bind_cols(loglik=ll[1], loglik.se=ll[2])
} -> results
```

- The above codes run one search from each of 300 starting values.
- Each search consists of an initial run of 50 IF2 iterations, followed by another 100 iterations.
- These codes exhibit a general **pomp** behavior:
  - Re-running a command on an object (i.e., `mif2` on `mf1`) created by the same command preserves the algorithmic arguments.
  - In particular, running `mif2` on the result of a `mif2` computation re-runs IF2 from the endpoint of the first run.
  - In the second computation, by default, all algorithmic parameters are preserved; here we overrode the default choice of `Nmif`.
- Following the `mif2` computations, the particle filter is used to evaluate the likelihood, as before.
- In contrast to the local-search codes above, here we return only the endpoint of the search, together with the likelihood estimate and its standard error in a named vector.
- The best result of this search had a likelihood of -120 with a standard error of 0.03.
- This took 4.3 minutes altogether using 250 processors.

Again, we attempt to visualize the global geometry of the likelihood surface using a scatterplot matrix. In particular, here we plot both the starting values (grey) and the IF2 estimates (red).

```
read_csv("measles_params.csv") %>%
  filter(loglik>max(loglik)-50) %>%
  bind_rows(guesses) %>%
  mutate(type=if_else(is.na(loglik),"guess","result")) %>%
  arrange(type) -> all

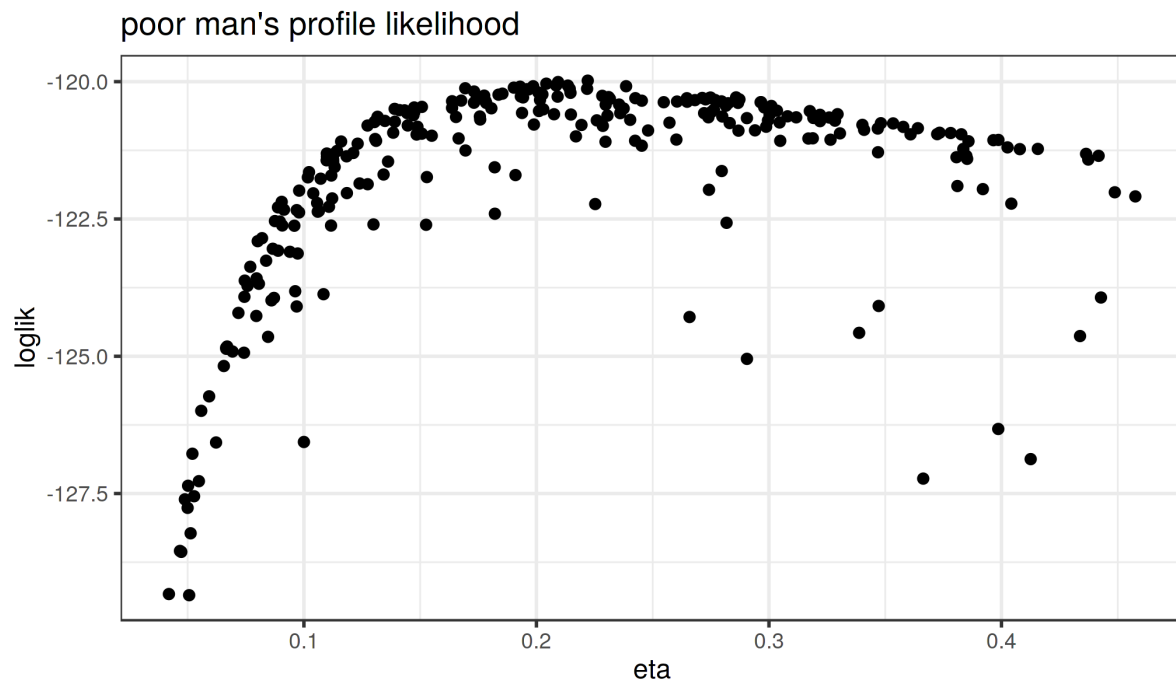
pairs(~loglik+Beta+eta+rho, data=all,
      col=ifelse(all$type=="guess",grey(0.5),"red"),pch=16)
```



- We see that optimization attempts from diverse remote starting points converge on a particular region in parameter space.
- The estimates have comparable likelihoods, despite their considerable variability.
- This gives us some confidence in our maximization procedure.

The projections of the estimates give us “poor man’s profiles”:

```
all %>%
  filter(type=="result") %>%
  filter(loglik>max(loglik)-10) %>%
  ggplot(aes(x=eta,y=loglik))+
  geom_point()+
  labs(
    x=expression("eta"),
    title="poor man's profile likelihood"
  )
```



## Profile likelihood

### Profile likelihood over $\eta$

- The curvature displayed in the upper envelope of the above plot suggests that there is indeed information in the data with respect to the susceptible fraction,  $\eta$ .
- To solidify this evidence, let’s compute a profile likelihood over this parameter.
- Recall that this means determining, for each value of  $\eta$ , the best likelihood that the model can achieve.
- To do this, we’ll first bound the uncertainty by putting a box around the highest-likelihood estimates we’ve found so far.



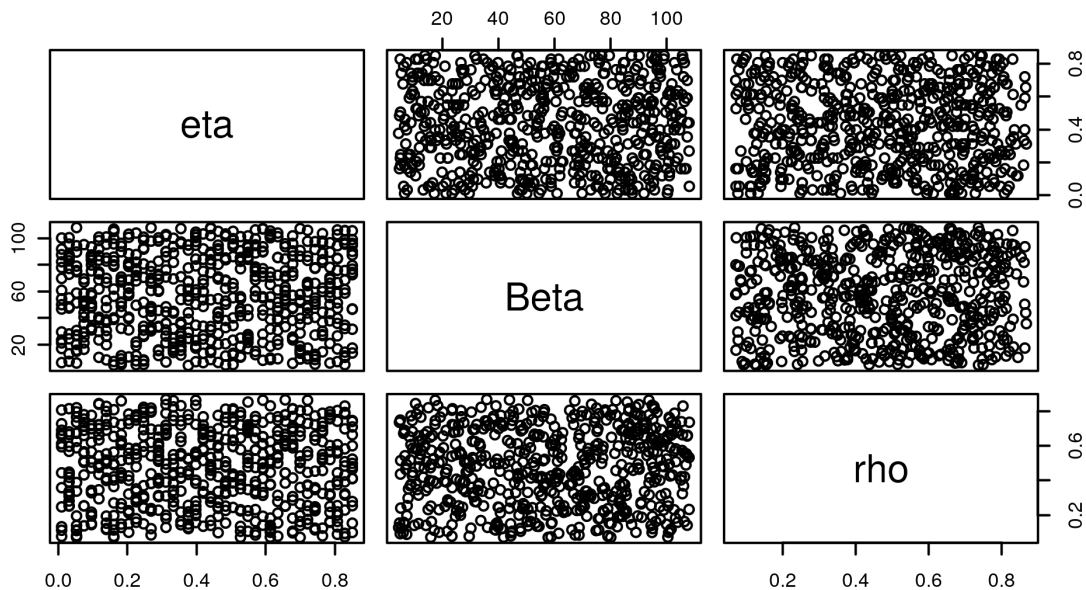
- Within this box, we'll choose some random starting points, for each of several values of  $\eta$ .

```
read_csv("measles_params.csv") %>%
  filter(loglik>max(loglik)-20,loglik.se<2) %>%
  sapply(range) -> box
box
```

	Beta	mu_IR	rho	eta	N	loglik
[1,]	4.261594	2	0.07025239	0.0277604	38000	-139.8695
[2,]	108.122779	2	0.87040501	0.4575748	38000	-119.9813

```
loglik.se
[1,] 0.01083245
[2,] 0.50246062
```

```
set.seed(1196696958)
profile_design(
  eta=seq(0.01,0.85,length=40),
  lower=box[1,c("Beta","rho")],
  upper=box[2,c("Beta","rho")],
  nprof=15, type="runif"
) -> guesses
plot(guesses)
```



- Now, we'll start one independent sequence of iterated filtering operations from each of these points.
- We'll be careful to keep  $\eta$  fixed.
- This is accomplished by not giving this parameter a random perturbation in the `mif2` call.

```

foreach(guess=iter(guesses,"row"), .combine=rbind) %dopar% {
  library(pomp)
  library(tidyverse)
  mf1 %>%
    mif2(params=c(unlist(guess),fixed_params),
          rw.sd=rw.sd(Beta=0.02,rho=0.02)) %>%
    mif2(Nmif=100,cooling.fraction.50=0.3) -> mf
  replicate(
    10,
    mf %>% pfilter(Np=100000) %>% logLik() %>%
    logmeanexp(se=TRUE) -> ll
  mf %>% coef() %>% bind_rows() %>%
    bind_cols(loglik=ll[1],loglik.se=ll[2])
} -> results

```

## Visualizing profile likelihood

As always, we save the results in our global database and plot the results.

```

read_csv("measles_params.csv") %>%
  bind_rows(results) %>%
  filter(is.finite(loglik)) %>%
  arrange(-loglik) %>%
  write_csv("measles_params.csv")

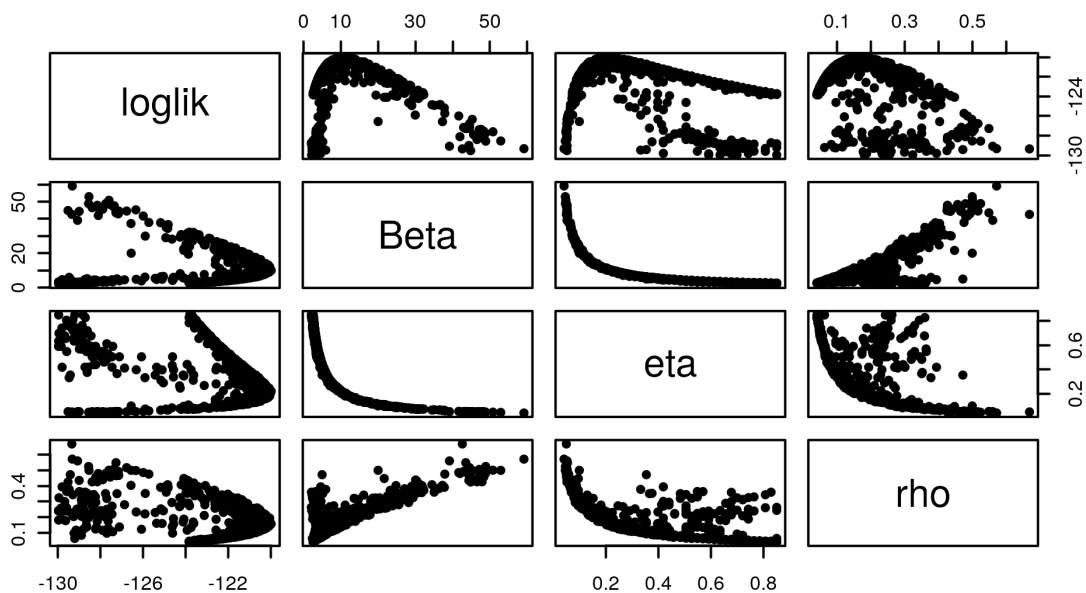
```

```

read_csv("measles_params.csv") %>%
  filter(loglik>max(loglik)-10) -> all

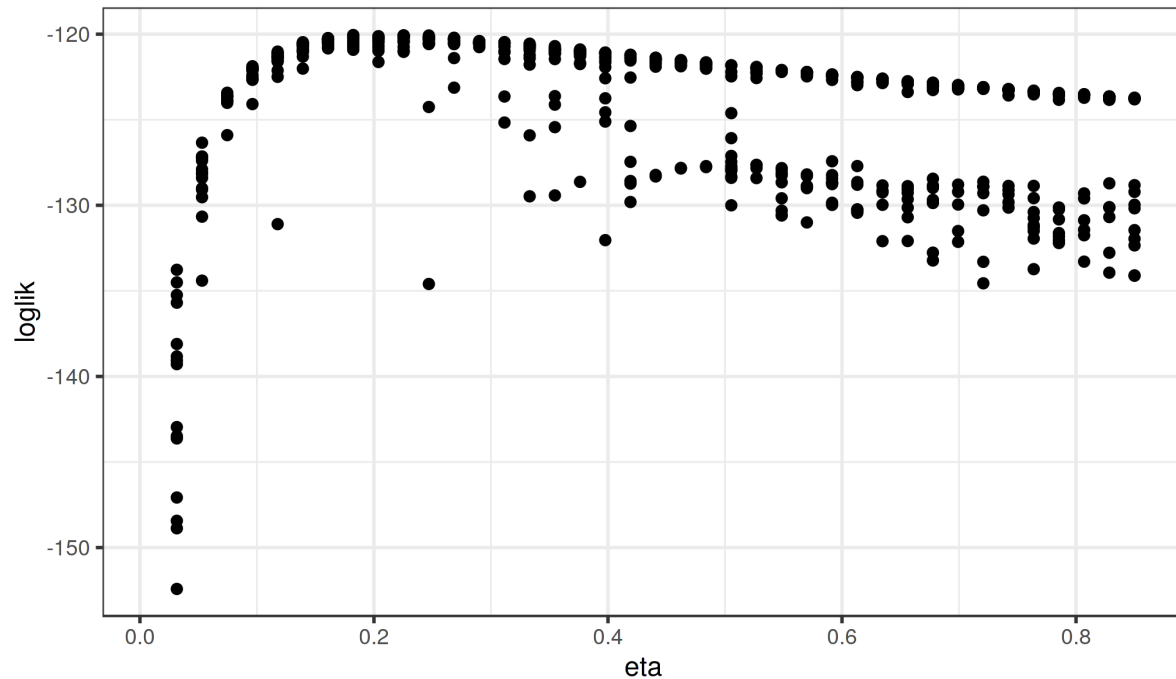
pairs(~loglik+Beta+eta+rho,data=all,pch=16)

```



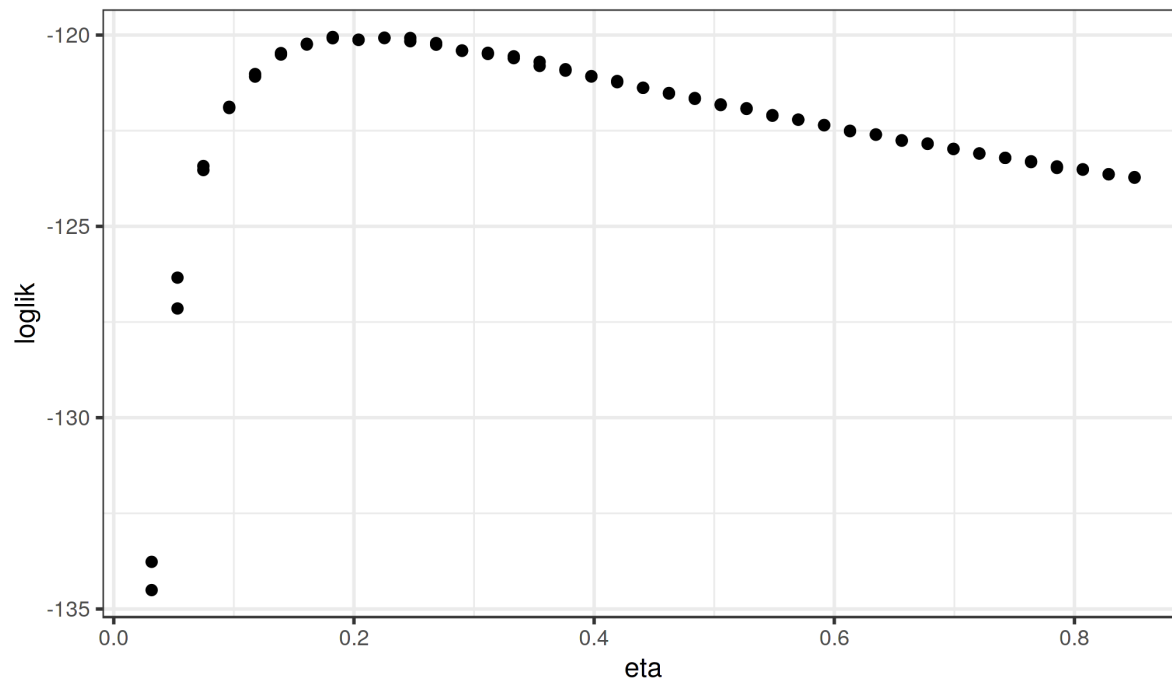
Plotting just the results of the profile calculation reveals that, while some of the IF2 runs either become “stuck” on local minima or run out of opportunity to reach the heights of the likelihood surface, many of the runs converge on high likelihoods.

```
results %>%  
  ggplot(aes(x=eta,y=loglik))+  
  geom_point()
```



A closer look shows what at first appears to be quite a flat surface over much of the explored range of  $\eta$ . Note that this appearance is due to the vertical scale, which is driven by the very low likelihoods associated with the smallest values of  $\eta$ .

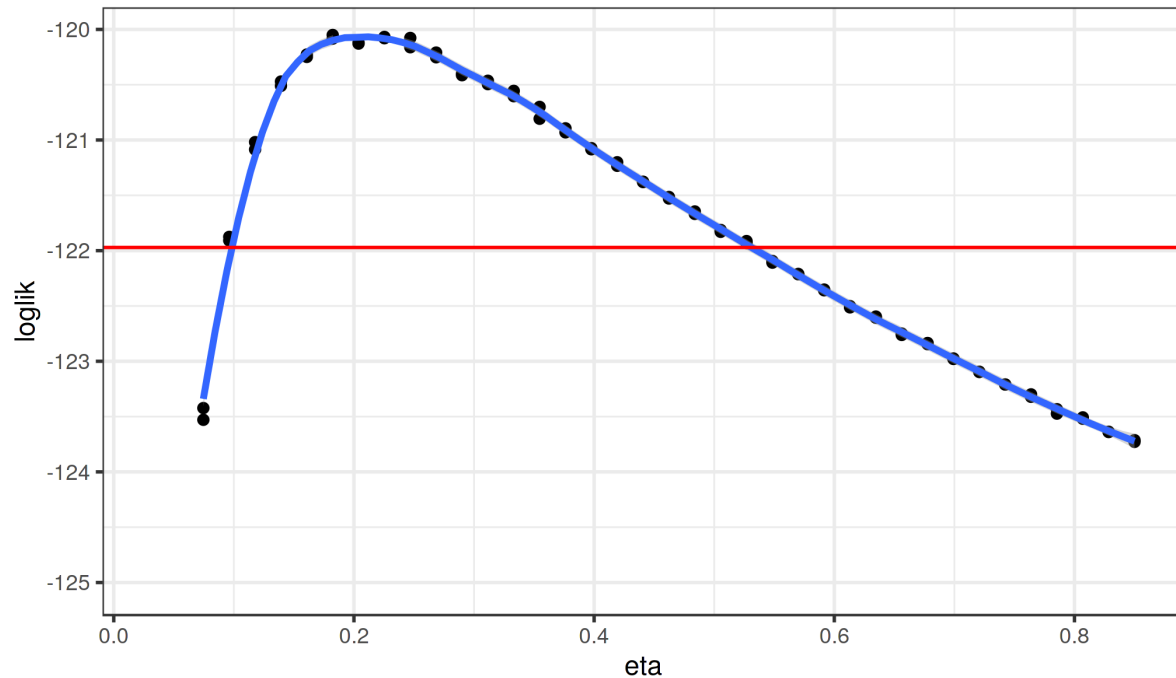
```
results %>%  
  filter(is.finite(loglik)) %>%  
  group_by(round(eta,5)) %>%  
  filter(rank(-loglik)<3) %>%  
  ungroup() %>%  
  filter(loglik>max(loglik)-20) %>%  
  ggplot(aes(x=eta,y=loglik))+  
  geom_point()
```



Focusing on just the top of the surface shows that, in fact, one is able to estimate  $\eta$  using these data. In the following plot, the cutoff for the 95% confidence interval (CI) is shown.

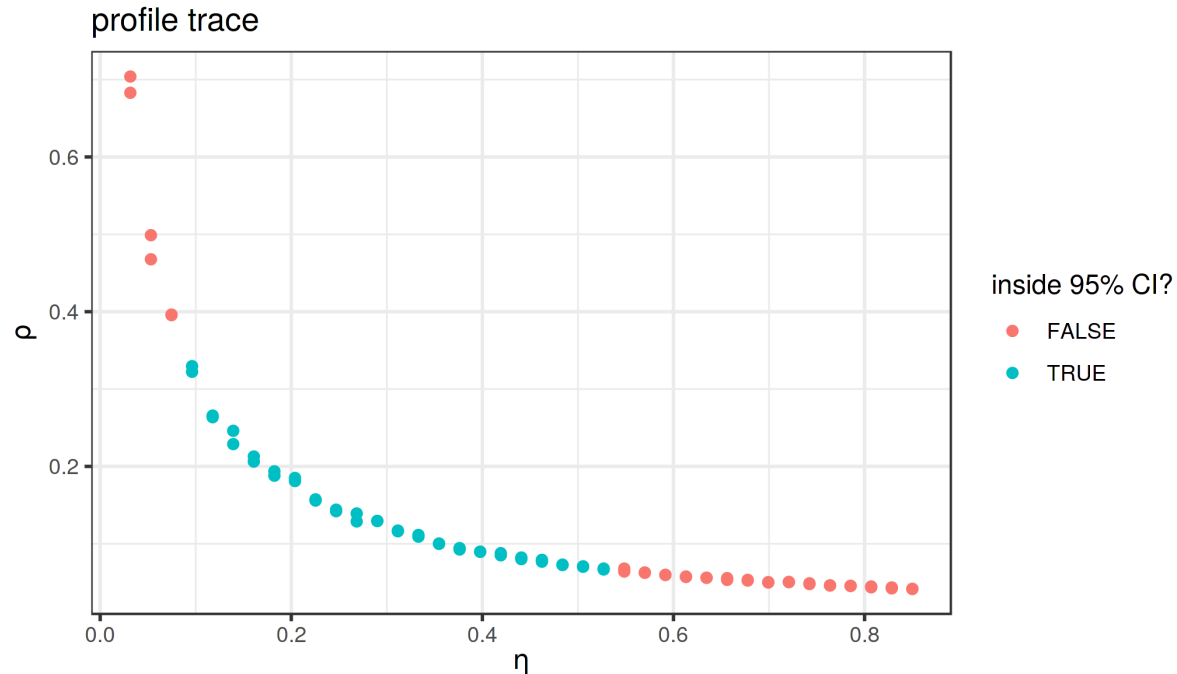
```
maxloglik <- max(results$loglik, na.rm=TRUE)
ci.cutoff <- maxloglik - 0.5 * qchisq(df=1, p=0.95)

results %>%
  filter(is.finite(loglik)) %>%
  group_by(round(eta, 5)) %>%
  filter(rank(-loglik) < 3) %>%
  ungroup() %>%
  ggplot(aes(x=eta, y=loglik)) +
  geom_point() +
  geom_smooth(method="loess", span=0.25) +
  geom_hline(color="red", yintercept=ci.cutoff) +
  lims(y=maxloglik - c(5, 0))
```



- As one varies  $\eta$  across the profile, the model compensates by adjusting the other parameters.
- It can be very instructive to understand how the model does this.
- For example, how does the reporting efficiency,  $\rho$ , change as  $\eta$  is varied?
- We can plot  $\rho$  vs  $\eta$  across the profile.
- This is called a *profile trace*.

```
results %>%
  filter(is.finite(loglik)) %>%
  group_by(round(eta,5)) %>%
  filter(rank(-loglik)<3) %>%
  ungroup() %>%
  mutate(in_ci=loglik>max(loglik)-1.92) %>%
  ggplot(aes(x=eta,y=rho,color=in_ci))+
  geom_point()+
  labs(
    color="inside 95% CI?",
    x=expression(eta),
    y=expression(rho),
    title="profile trace"
  )
```



### Profile over $\rho$

While the above profile trace is suggestive that the 95% CI for  $\rho$  must be between roughly 7% and 30%, to confirm this, we should construct a proper profile likelihood over  $\rho$ . We do so now.

This time, we will initialize the IF2 computations at points we have already established have high likelihoods.

```
read_csv("measles_params.csv") %>%
  group_by(cut=round(rho,2)) %>%
  filter(rank(-loglik)<=10) %>%
  ungroup() %>%
  select(-cut,-loglik,-loglik.se) -> guesses

foreach(guess=iter(guesses,"row"), .combine=rbind) %dopar% {
  library(pomp)
  library(tidyverse)
  mf1 %>%
    mif2(params=guess,
          rw.sd=rw.sd(Beta=0.02,eta=ivp(0.02))) %>%
    mif2(Nmif=100,cooling.fraction.50=0.3) %>%
    mif2() -> mf
  replicate(
    10,
    mf %>% pfilter(Np=100000) %>% logLik() %>%
    logmeanexp(se=TRUE) -> ll
    mf %>% coef() %>% bind_rows() %>%
    bind_cols(loglik=ll[1],loglik.se=ll[2])
  } -> results
```

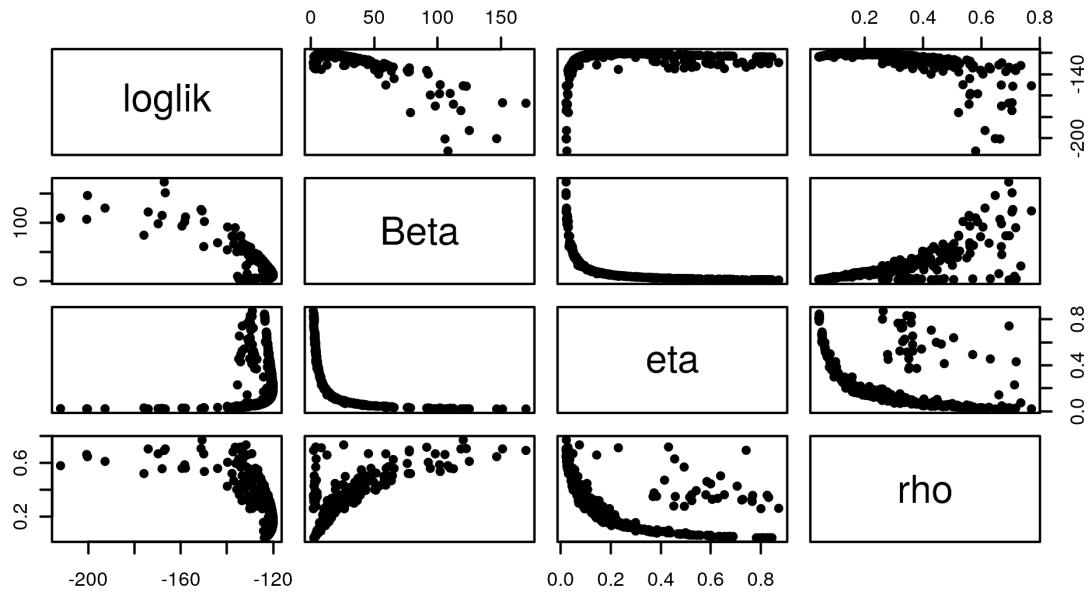
### Profile over $\rho$ : results

```

results %>%
  filter(is.finite(loglik)) -> results

pairs(~loglik+Beta+eta+rho,data=results,pch=16)

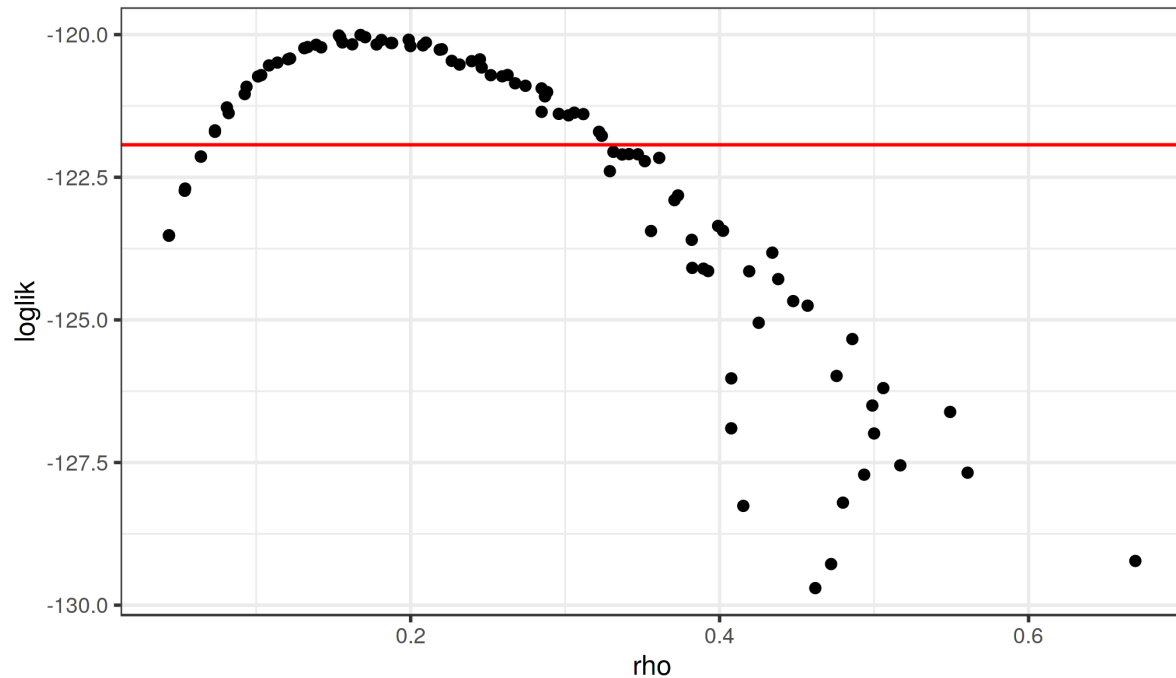
```



```

results %>%
  filter(loglik>max(loglik)-10,loglik.se<1) %>%
  group_by(round(rho,2)) %>%
  filter(rank(-loglik)<3) %>%
  ungroup() %>%
  ggplot(aes(x=rho,y=loglik))+
  geom_point()+
  geom_hline(
    color="red",
    yintercept=max(results$loglik)-0.5*qchisq(df=1,p=0.95)
  )

```



```
results %>%
  filter(loglik>max(loglik)-0.5*qchisq(df=1,p=0.95)) %>%
  summarize(min=min(rho),max=max(rho)) -> rho_ci
```

The data appear to be consistent with reporting efficiencies in the 7–32% range (95% CI).

## 6 The investigation continues....

### Making predictions

#### Parameter estimates as model predictions

- The estimated parameters are one kind of model prediction.
- When we can estimate parameters using other data, we can test these predictions.
- In the case of a highly contagious, immunizing childhood infection such as measles, we can obtain an estimate of the reporting efficiency,  $\rho$  by simply regressing cumulative cases on cumulative births (Anderson and May, 1991) over many years.
- When we do this for Consett, we see that the reporting efficiency is roughly 60%.
- Since such a value makes the outbreak data quite unlikely, the prediction does not appear to be borne out.
- We can conclude that one or more of our model assumptions is inconsistent with the data.
- Let's revisit our assumption that the infectious period is known to be 0.5 wk.
- Indeed, it would not be surprising were we to find that the *effective* infectious period, at the population scale, were somewhat shorter than the *clinical* infectious period.



- For example, confinement of patients should reduce contact rates, and might therefore curtail the effective infectious period.
- To investigate this, we'll relax our assumption about the value of  $\mu_{IR}$ .

## Searching in another direction

### Another global search

We will estimate the model under the assumption that  $\rho = 0.6$ , but without making assumptions about the duration of the infectious period. As before, we'll construct a random design of starting parameters.

```
set.seed(55266255)
runif_design(
  lower=c(Beta=5,mu_IR=0.2,eta=0),
  upper=c(Beta=80,mu_IR=5,eta=0.4),
  nseq=1000
) %>%
  mutate(
    rho=0.6,
    N=38000
  ) -> guesses
```

- For each of these starting points, we'll run a series of IF2 computations.
- Since we have gained some experience applying mif2 to this model and these data, we have some expectation about how much computation is required.
- In the following, we'll use a lot more computational power than we have so far.

For each of the starting points, we'll first perform 100 IF2 iterations:

```
library(pomp)
library(tidyverse)
measSIR %>%
  mif2(params=gues, Np=2000, Nmif=100,
        cooling.fraction.50=0.5,
        partrans=parameter_trans(
          log=c("Beta", "mu_IR"),
          logit="eta"), paramnames=c("Beta", "mu_IR", "eta"),
        rw.sd=rw.sd(Beta=0.02,mu_IR=0.02,eta=ivp(0.02))) -> mf
```

We use random perturbations of the same magnitude as before, taking care to transform the parameters we are estimating.

Next, we repeat the calculations several times. Each additional set of IF2 iterations restarts the previous calculation at its endpoint.

```
mf %>%
  mif2(Nmif=100) %>%
  mif2(Nmif=100) %>%
  mif2(Nmif=100) -> mf
```

In each of the previous calculations, we have used a relatively high cooling fraction. We'll now reduce this gradually.

```
mf %>%
  mif2(Nmif=100,cooling.fraction.50=0.3) %>%
  mif2(Nmif=100,cooling.fraction.50=0.3) %>%
  mif2(Nmif=100,cooling.fraction.50=0.3) %>%
  mif2(Nmif=100,cooling.fraction.50=0.1) %>%
  mif2(Nmif=100,cooling.fraction.50=0.1) %>%
  mif2(Nmif=100,cooling.fraction.50=0.1) -> mf
```

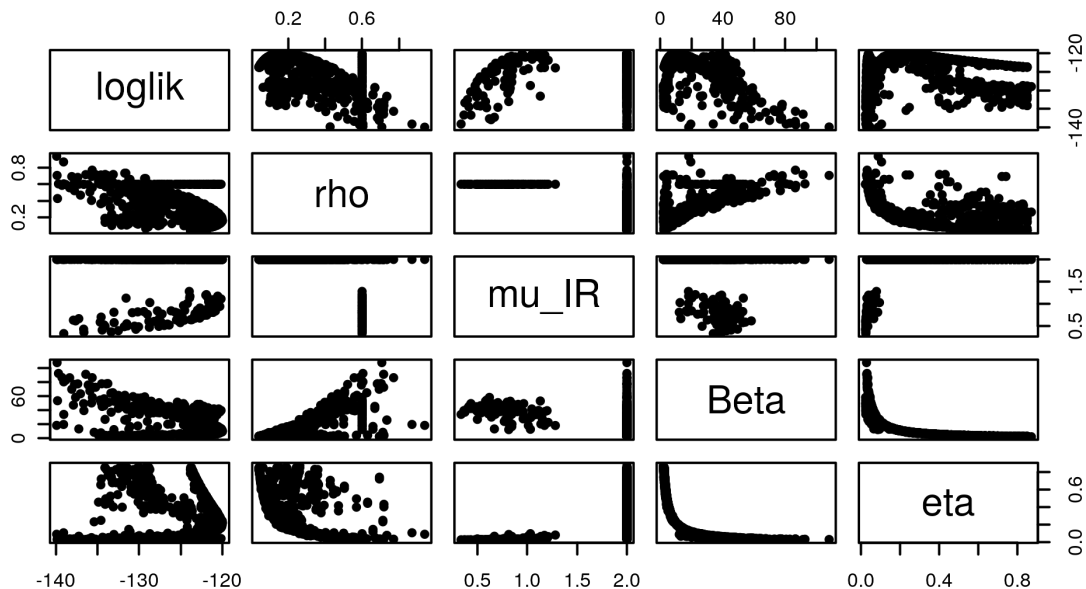
We wrap the above in a **foreach** loop as before and take care to evaluate the likelihood at each end-point using **pfilter**.

See the [R code for this lesson](#) to see exactly how this is done.

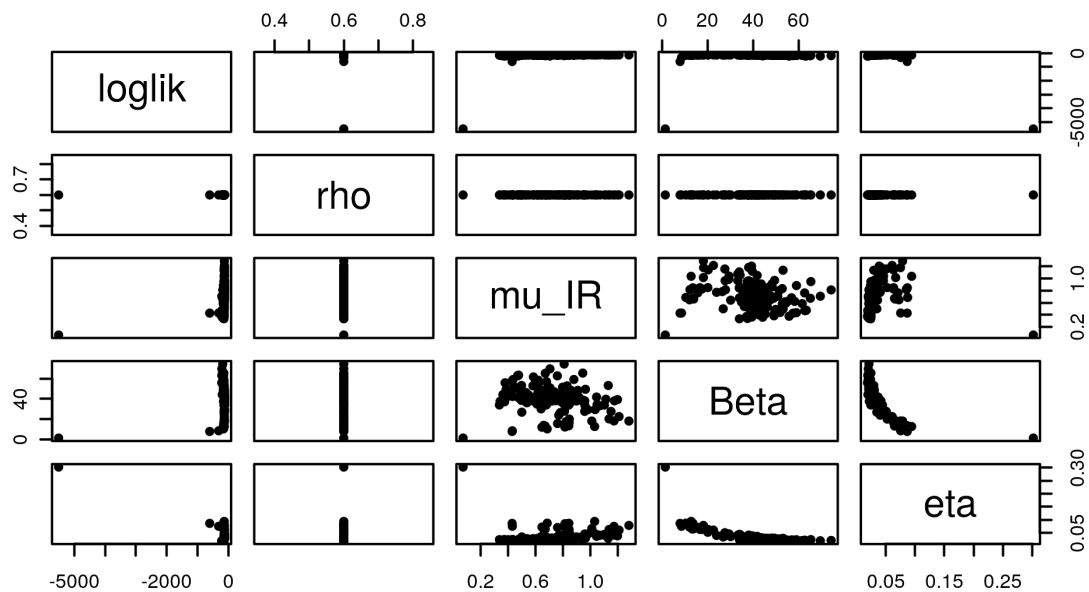
The computations above required 19.4 minutes on 250 processors.

```
read_csv("measles_params.csv") %>%
  filter(loglik>max(loglik)-20) -> all

pairs(~loglik+rho+mu_IR+Beta+eta,data=all,pch=16)
```



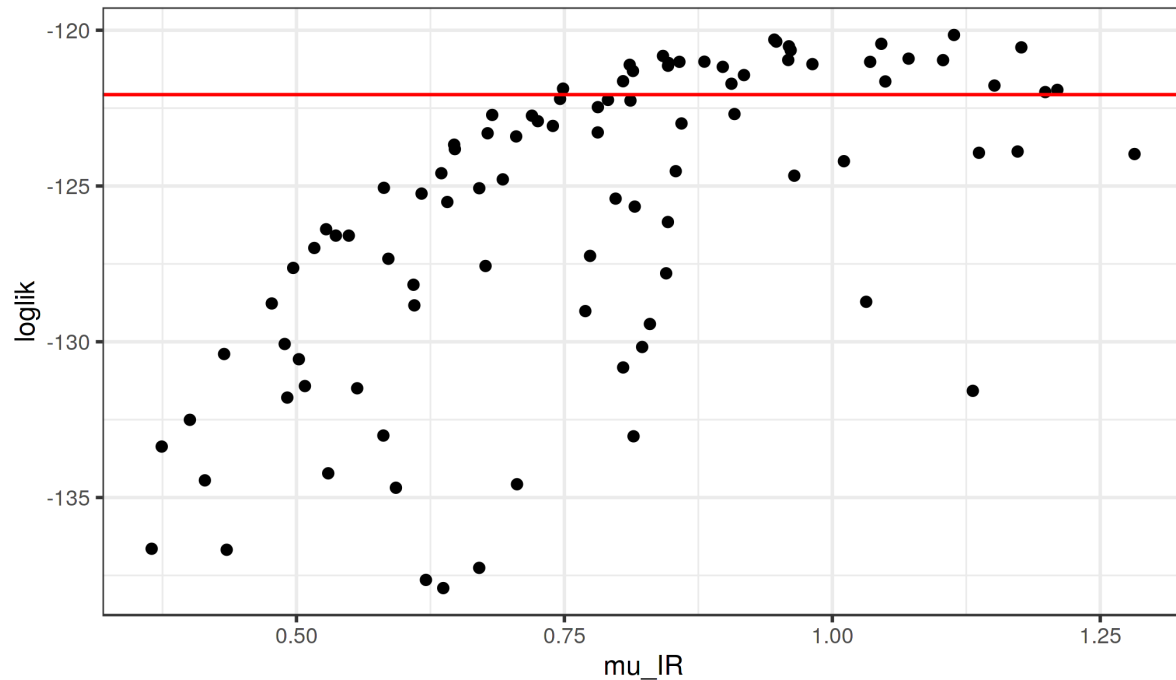
```
pairs(~loglik+rho+mu_IR+Beta+eta,data=results,pch=16)
```



```

results %>%
  filter(loglik>max(loglik)-20,loglik.se<1) %>%
  ggplot(aes(x=mu_IR,y=loglik))+
  geom_point()+
  geom_hline(
    color="red",
    yintercept=max(results$loglik)-0.5*qchisq(df=1,p=0.95)
  )

```



### Profile over infectious period

To make inferences about  $\mu_{IR}$ , we can again compute a profile likelihood. As before, we bound the region we will search:

```
read_csv("measles_params.csv") %>%
  filter(
    loglik > max(loglik) - 20,
    loglik.se < 2,
    abs(rho - 0.6) < 0.01
  ) %>%
  sample_n(1000) -> box
```

```
set.seed(610408798)
profile_design(
  mu_IR = seq(0.5, 2, by = 0.1),
  lower = box[1, c("Beta", "eta")],
  upper = box[2, c("Beta", "eta")],
  nprof = 100, type = "runif"
) %>%
  mutate(
    N = 38000,
    rho = 0.6
  ) -> guesses
```

```
foreach(guess = iter(guesses, "row"), .combine = rbind) %dopar% {
  library(pomp)
  library(tidyverse)
  measSIR %>%
    mif2(params = guess, Np = 2000, Nmif = 100,
```

```

partrans=parameter_trans(log="Beta",logit="eta"),
paramnames=c("Beta","eta"), cooling.fraction.50=0.5,
rw.sd=rw.sd(Beta=0.02,eta=ivp(0.02))
) %>% mif2(Nmif=100) %>%
mif2(Nmif=100,cooling.fraction.50=0.3) %>%
mif2(Nmif=100,cooling.fraction.50=0.1) -> mf
replicate(10,mf %>% pfilter(Np=100000) %>% logLik()) %>%
logmeanexp(se=TRUE) -> ll
mf %>% coef() %>% bind_rows() %>%
bind_cols(loglik=ll[1],loglik.se=ll[2])
} -> results

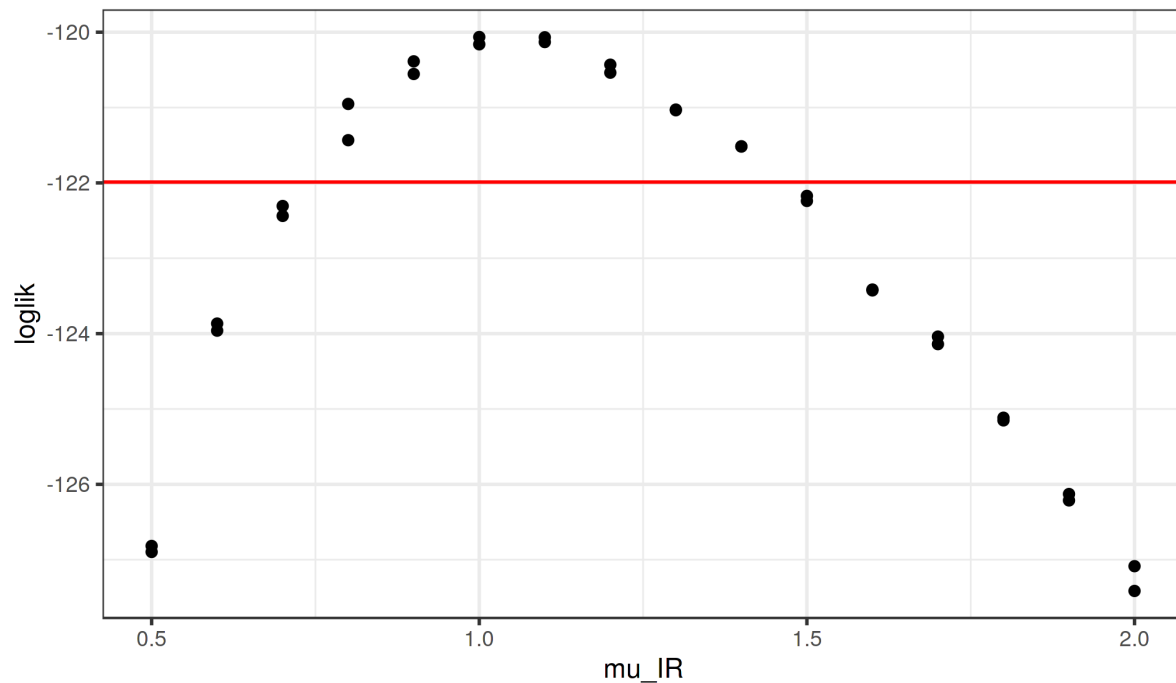
```

## Infectious period profile

```

results %>%
group_by(round(mu_IR,2)) %>%
filter(rank(-loglik)<3) %>%
ungroup() %>%
ggplot(aes(x=mu_IR,y=loglik))+
geom_point()+
geom_hline(
color="red",
yintercept=max(results$loglik)-0.5*qchisq(df=1,p=0.95)
)

```



- This suggests that  $\rho = 0.6$  is consistent only with smaller values of  $\mu_{IR}$ , and hence *longer* infectious periods than are possible if the duration of shedding is actually less than one week.
- Thus the model is incapable of reconciling both an infectious period of less than one week and a reporting rate of 60%.
- What structural changes to the model might we make to improve its ability to explain the data?

## 7 Exercises

### Exercise 4.1. Fitting the SEIR model

Following the template above, estimate the parameters and likelihood of the SEIR model you implemented in the earlier lessons. Specifically:

- (a) First conduct a local search and then a global search using the multi-stage, multi-start method displayed above.
- (b) How does the maximized likelihood compare with what we obtained for the SIR model?
- (c) How do the parameter estimates differ?

You will need to tailor the intensity of your search to the computational resources at your disposal. In particular, choose the number of starts, number of particles employed, and the number of IF2 iterations to perform in view of the size and speed of your machine.

### Exercise 4.2. Modify the measurement model

The binomial measurement model used here assumes that the reporting rate is fixed through time and that all infection reports are independent and equally likely. Commonly, it is advisable to consider measurement models with **overdispersion**, i.e., more variability than the binomial (or Poisson) allows. The negative binomial distribution, for instance, allows for overdispersed mean-variance relationships. Similarly, the beta-binomial distribution adds overdispersion to the binomial. Formulate an alternative measurement model and maximize the likelihood to compare it with existing results.

### Exercise 4.3. Construct a profile likelihood

How strong is the evidence about the contact rate,  $\beta$ , given this model and data? Use `mif2` to construct a profile likelihood. Due to time constraints, you may be able to compute only a preliminary version. It is also possible to profile over the basic reproduction number,  $R_0 = \beta/\mu_{IR}$ . Is this more or less well determined than  $\beta$  for this model and data?

### Exercise 4.4. Checking the source code

Check the source code for the `measSIR` pomp object. Does the code implement the model described? For various reasons, it can be surprisingly hard to make sure that the written equations and the code are perfectly matched. Here are some things to think about  
Papers should be written to be readable. Code must be written to run successfully. People rarely choose to clutter papers with numerical details which they hope and believe are scientifically irrelevant.

- (a) What problems can arise due to the conflict between readability and reproducibility?
- (b) What solutions are available?

Suppose that there is an error in the coding of `rprocess` and suppose that plug-and-play statistical methodology is used to infer parameters. As a conscientious researcher, you carry out a simulation study to check the soundness of your inference methodology on this model. To do this, you use `simulate` to generate realizations from the fitted model and checking that your parameter inference procedure recovers the known parameters, up to some statistical error.

- (a) Will this procedure help to identify the error in `rprocess`?
- (b) If not, how might you debug `rprocess`?
- (c) What research practices help minimize the risk of errors in simulation code?


### Exercise 4.5: Choosing the algorithmic settings for IF2

Have a look at [our advice on tuning IF2](#).

## References

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## License, acknowledgments, and links

- This lesson is prepared for the [Simulation-based Inference for Epidemiological Dynamics](#) module at the 2020 Summer Institute in Statistics and Modeling in Infectious Diseases, [SISMID 2020](#).
- The materials build on [previous versions of this course and related courses](#).
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- Produced with R version 4.0.2 and **pomp** version 3.1.1.1.
- Compiled on July 20, 2020.

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[R code for this lesson](#)