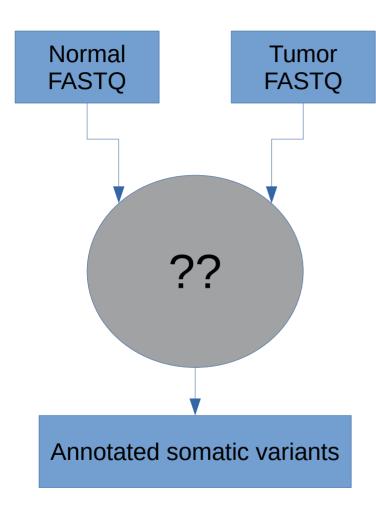
# Somatic mutation calling pipelin for a matched sample pair

Willie Yu 17 Jan 2022 willie.yu@duke-nus.edu.sg

# Given

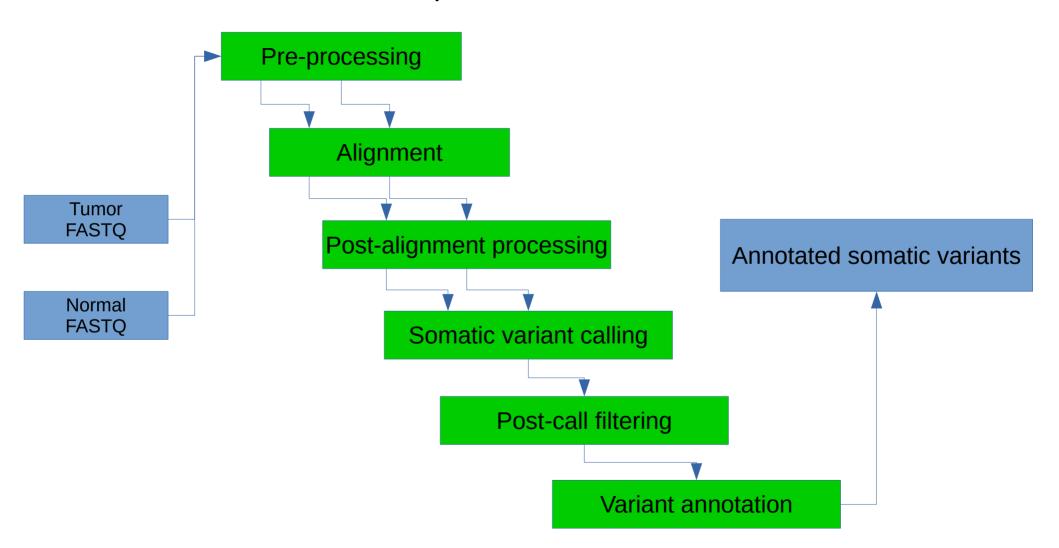
- Short read DNA sequences in FASTQ format from
  - Patient Tumor
  - Patient Matched Normal

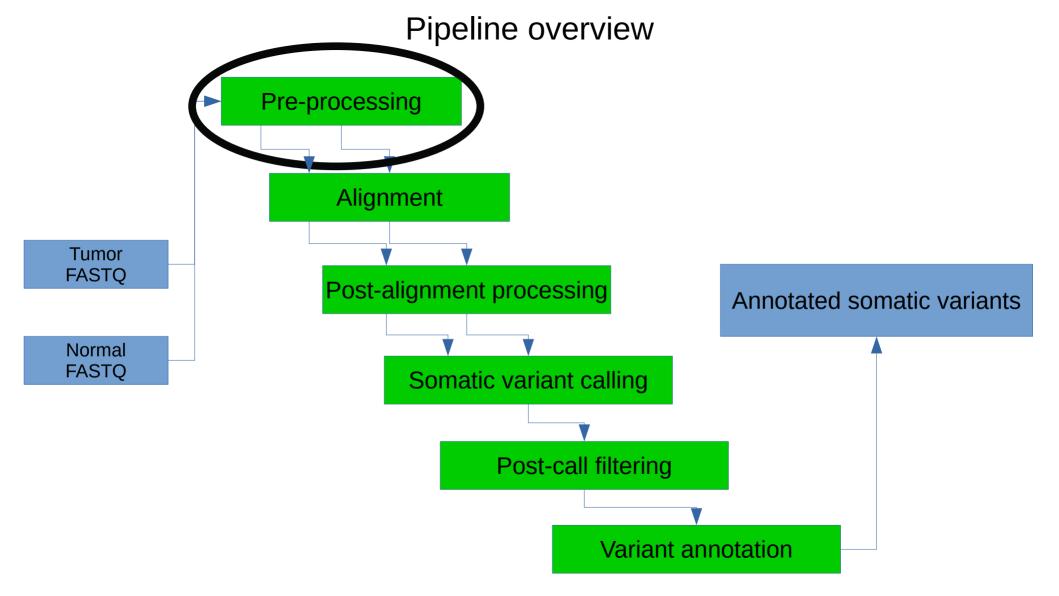
```
@A00609:341:HK33WDSX2:3:1101:1235:1000 1:N:0:GTAGCTGA+AGTAGGTC
NGCTCCTTTTTGTCCTCTTTTGCTCTGGTGTGACATGCCTCTTCTGCAACTGCTGAAGTAATTCACCGCACGTTTTCACTCTTTTCCTCACTTTTTTCCACCTGGCAGAAGCGTGAAGAAGATTACAGACACATGGGATACCTCTG
@A00609:341:HK33WDSX2:3:1101:2392:1000 1:N:0:GTAGCTGA+AGTAGGTC
NCCAGCCTGGCCAACAAGGTGAAACCCCATCTCTACTACAAATACAAAAACTAGCCAGGTGTGGTGGCACACGCCTGTAATCCCAGCTACTAGGGGAGGCTGAGGCAAGAGAATCGCTTGAACCTGGGAGGCAGAGGTTGAAGTGAGCTG
@A00609:341:HK33WDSX2:3:1101:2627:1000 1:N:0:GTAGCTGA+AGTAGGTC
NAAAGCTGGATGGAGAATGACTTTGACGAGCTAAGAGGAGAAGGCTTCAGACGATCAAATTACTCTGAGCTACAGGAGGACATTCAAACCAAAGGCAAACAAGTTGAAAAAAATTTAGAAGAATGTATAACTAGAATAAC
@A00609:341:HK33WDSX2:3:1101:3242:1000 1:N:0:GTAGCTGA+AGTAGGTC
@A00609:341:HK33WDSX2:3:1101:3477:1000 1:N:0:GTAGCTGA+AGTAGGTC
@A00609:341:HK33WDSX2:3:1101:3857:1000 1:N:0:GTAGCTGA+AGTAGGTC
NAACTGAATTTATTCTCAGCATAGATGAGCCTGGTGCCACCTTGAATATCCACCATTCACAGACTGCCACTTTGGTCATGGGAACTCACATGTGCAACTTGTTTCCATGACAGCCTGTAAATTACAGGTCCCAAAGTCAGCTGCAAGGAA
@A00609:341:HK33WDSX2:3:1101:3965:1000 1:N:0:GTAGCTGA+AGTAGGTC
NTTAATTTATTAAAGATAATTTTTAAAAACTGAGGCAAATCAGTCATGGTGTTAAATTTATGTGACAAAAAATATTTTGTCAGTATTATTTGAAGTATTTTCTAATTTTATTGATATAAAATCAAACTTATATTGGTAAATTCTCTTTC
```



CTCCTTTTTGCCCCTTTTGCTTGCTTGGTGTGACATGCCTCTTCTGCAACTGCTGAAGTAATTCACCGCACGTTTTCACTCTTTTTCCTCATTTTTTCCACCTGGCAGAAGC<u>TGAAGAAAGATTACAGACACATGGGATACCTCT</u> CCAGCCTGGCCAACAAGGTGAAACCCCATCTCTACTACAAATACAAAAACTAGCCAGGTGTGGTGGCACACGCCTGTAATCCCAGCTACTAGGGGAGGCTGAGGCAAGAGAATCGCTTGAACCTGGGAGGCAGAGGTTGAAGTTGAGCTG .00609:341:HK33WDSX2:3:1101:2627:1000 1:N:0:GTAGCTGA+AGTAGGTC AAGCTGGATGGAGAATGACTTTGACGAGCTAAGAGGAGAAAGGCTTCAGACGATCAAATTACTCTGAGCTACAGGAGGACATTCAAACCAAAGGCAAACAAGTTGAAAAACTTTGAAAAAAATTTAGAAGAATGTATAACTAGAATAACC 00609:341:HK33WDSX2:3:1101:3242:1000 1:N:0:GTAGCTGA+AGTAGGTC A00609:341:HK33WDSX2:3:1101:3477:1000 1:N:0:GTAGCTGA+AGTAGGTC A00609:341:HK33WDSX2:3:1101:3857:1000 1:N:0:GTAGCTGA+AGTAGGTC A00609:341:HK33WDSX2:3:1101:3965:1000 1:N:0:GTAGCTGA+AGTAGGTC . VITAATITATITAAAGATAATITITAAAAACTGAGGCAAATCAGTCATGGTGTTAAATITATGTGACAAAAAATATITTGTCAGTATTATTTGAAGTATTITCTAATITTATTGATATAAAATCAAACTTATATTGGTAAATTCCCTTTC .

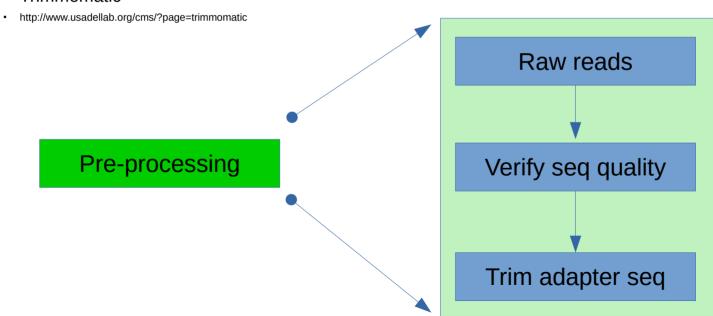
Chr	Start	End	Ref	Alt	Func.refGene	Gene.refGene	GeneDetail.refGene
10	63211352	63211352	G	Т	intronic	JMJD1C	
10	111455769	111455769	Т	С	intergenic	HEAT2;GPAM	dist=337173;dist=694096
11	4130486	4130486	Α	G	intronic	RRM1	
12	120420659	120420659	С	Α	intergenic	MSI1;COX6A1	dist=51495;dist=17454
16	2226050	2226050	С	G	intronic	E4F1	
16	85499571	85499571	Α	G	intergenic	MIR5093;GSE1	dist=193246;dist=111838
17	82626591	82626591	С	Α	intronic	WDR45B	
19	6797171	6797171	G	Α	intronic	VAV1	
19	13950460	13950460	G	Α	intronic	PODNL1	
3	73247693	73247693	С	Т	intergenic	PPP4R2;PDZRN3	dist=178492;dist=134737



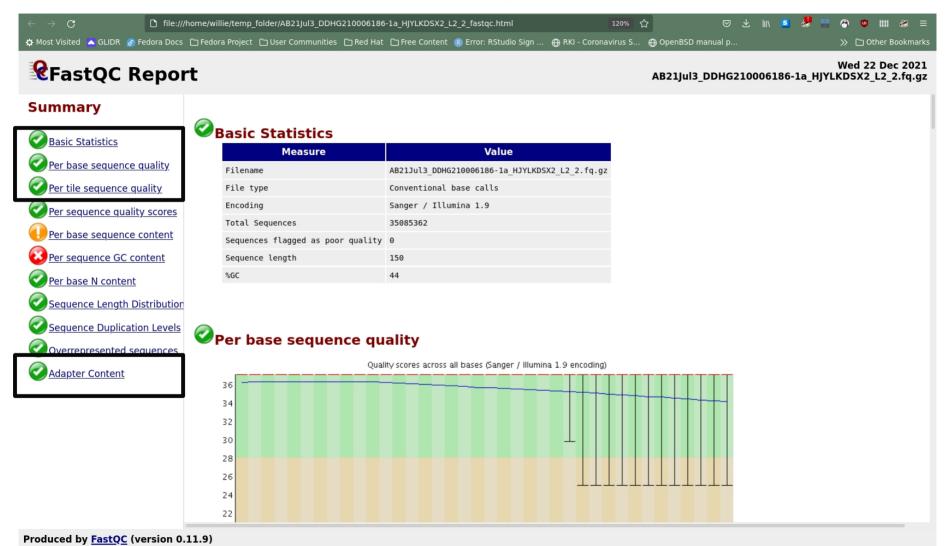


# Pre-processing

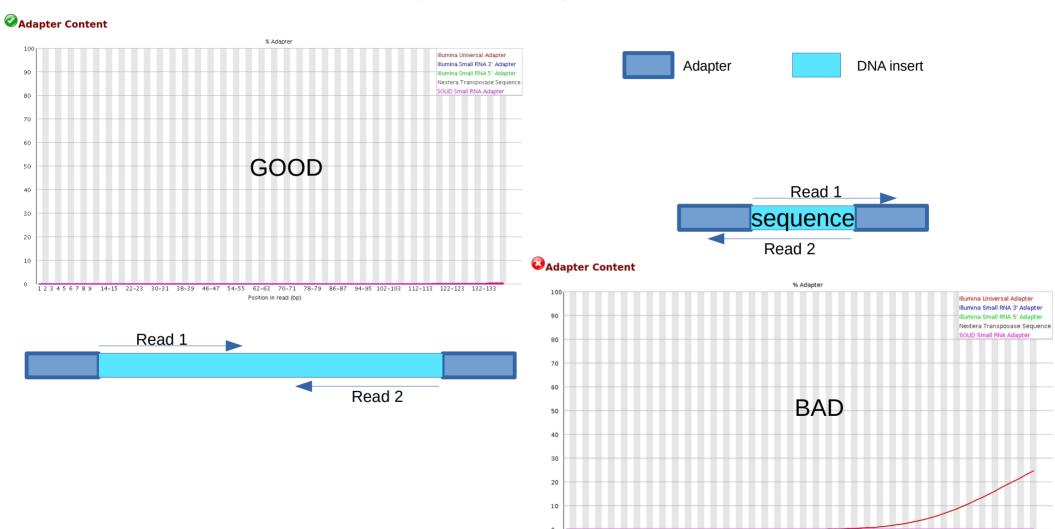
- Purpose
  - Check and verify sequence qualities
    - FastQC
    - https://www.bioinformatics.babraham.ac.uk/projects/fastgc/
  - Trimming of adapter sequences
    - Trimmomatic



# Pre-processing: FastQC



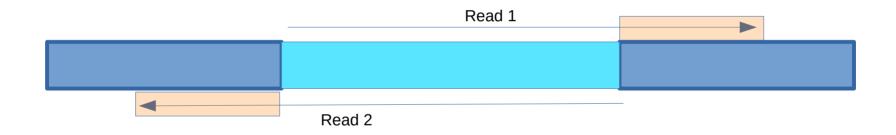
# Pre-processing: FastQC



1 2 3 4 5 6 7 8 9 12-13 18-19 24-25 30-31 36-37 42-43 48-49 54-55 60-61 66-67 72-73 78-79 84-85 90-91 96-97 104-105 114-115 124-125 134-135 Position in read (bp)

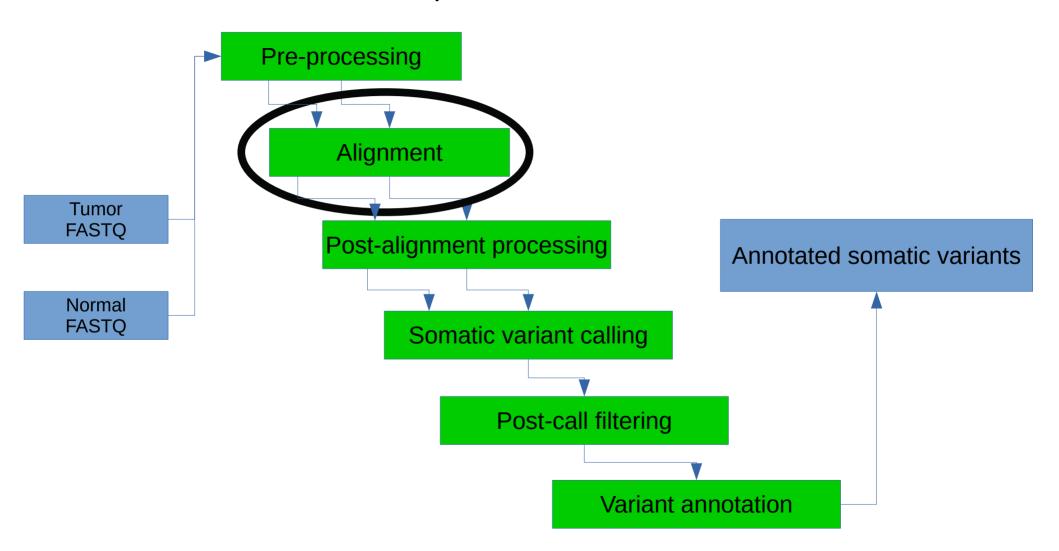
# Adapter trimming

Cut out the part of the DNA insert matching the adapter sequence



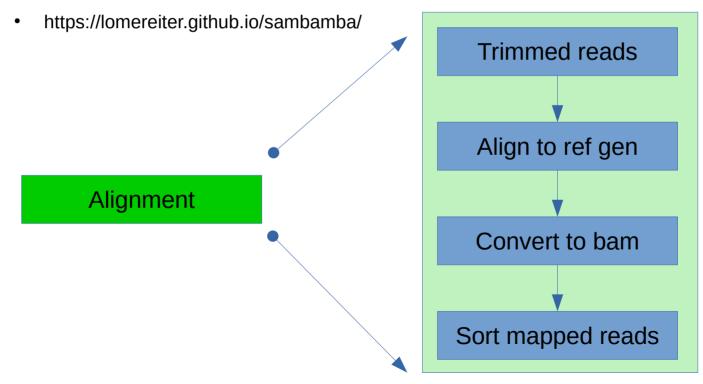
**Trimmomatic** 

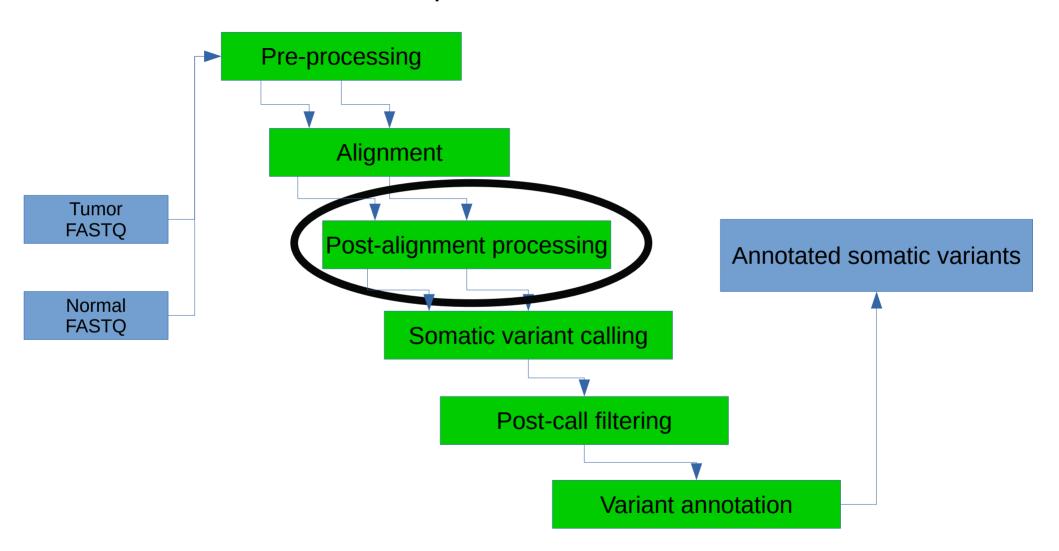
http://www.usadellab.org/cms/?page=trimmomatic



# Alignment

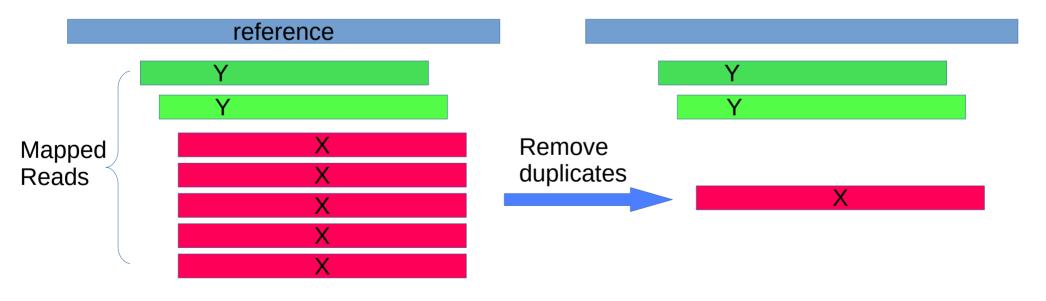
- Goal: Mapping each DNA insert against a reference genome
  - BWA-MEM2: mapping DNA sequences against a large reference genome
    - https://github.com/bwa-mem2/bwa-mem2
  - Sambamba:convert aligned output into bam format and sort mapped reads by chr

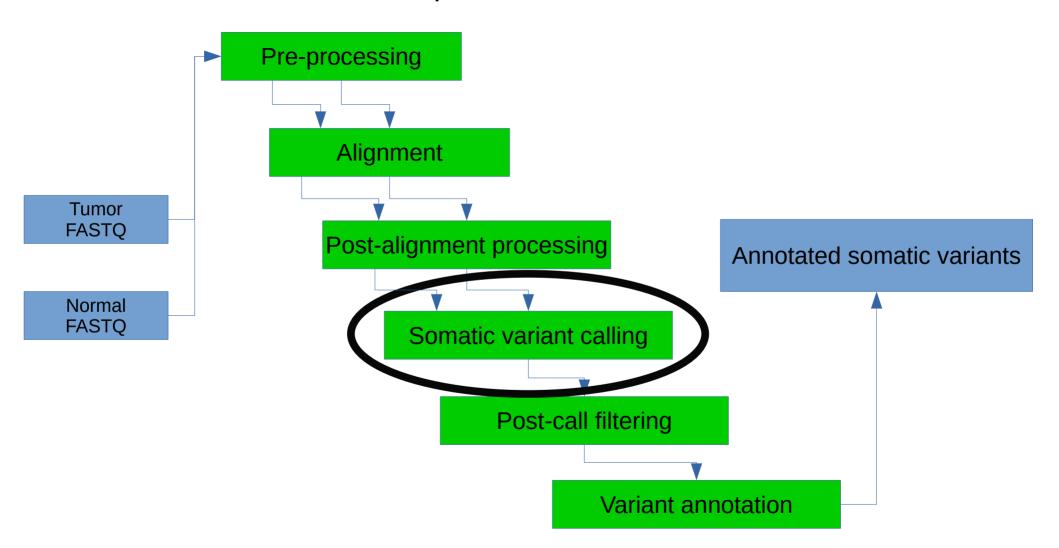




# Post-alignment processing

- PCR duplicate removal:
  - Sambamba: duplicate removal
- Why duplicate removal?
  - Minimize sequencing error propagation
  - Reduce false positives in variant calling

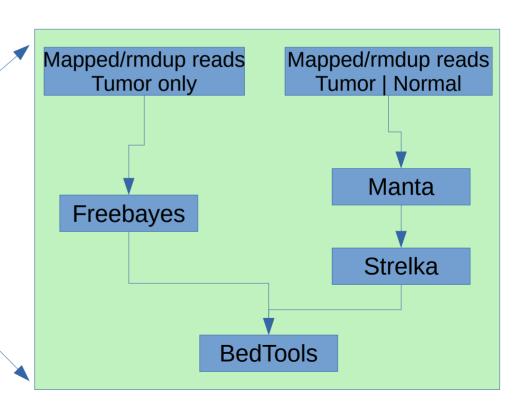


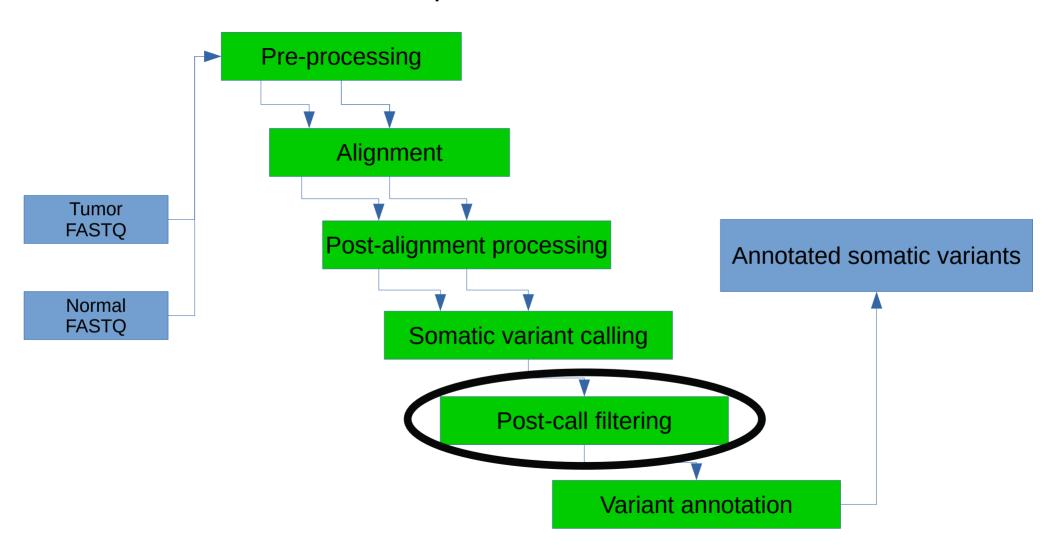


# Somatic variant calling

- All variant callers generates artifacts/false positive calls
  - Solution: Intersect variants with at least 2 callers
- Variant callers used
  - Freebayes: call all variants
    - https://github.com/freebayes/freebayes
  - Manta/Strelka: call somatic variants only
    - https://github.com/Illumina/manta
    - https://github.com/Illumina/strelka
- Intersect tool used
  - BedTools
    - https://bedtools.readthedocs.io/en/latest/

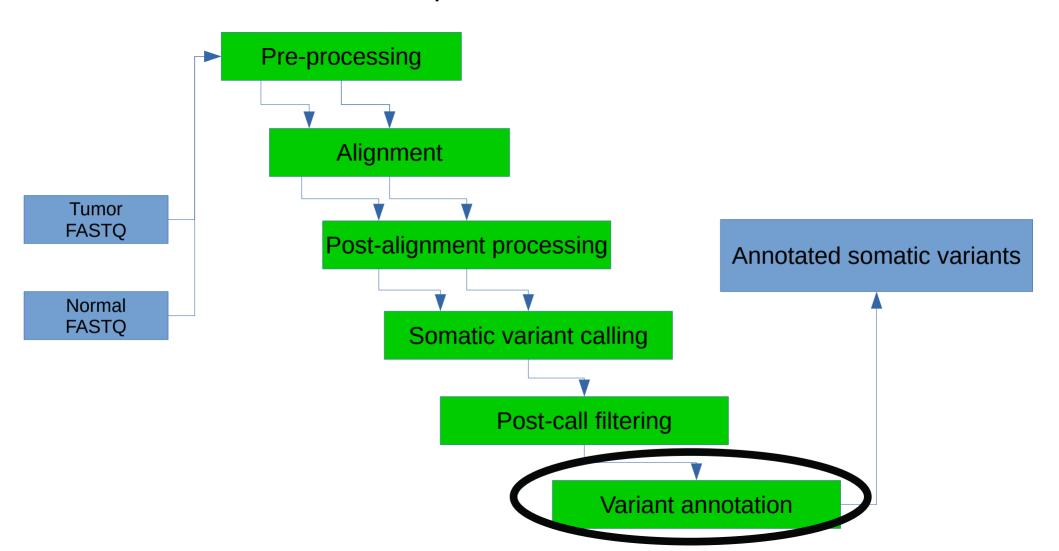
Somatic variant calling





# Post-call filtering

- Caller specific artifacts removal still insufficient
- False positive calls still in high numbers due to
  - Germline variants leak
  - Variant calls in highly variable / repetitive regions of the genome
- Solution
  - Create blacklists of genomic positions / regions
  - Use BedTools to filter against called variants



#### **Variant Annotation**

#### Filtered somatic variant list





#### Annotated somatic variant list

#CHRON	POS II	) RE	F AL	Γ QUA	L FILTER	INF0	FORMAT NORMAL	TUMOR
1	1591986 .					SOMATIC	;QSS=23;TQSS=1;	NT=ref;QSS_NT=23;
,0:19,	19:0,0							
1	3605430 .				PASS	SOMATIC	;QSS=19;TQSS=1;	NT=ref;QSS_NT=19;
,0:0,0	9:17,17							
1	3827839 .				PASS	SOMATIC	;QSS=20;TQSS=1;	NT=ref;QSS_NT=20;
:0,0	15:0:0:0							
1	5491320 .					SOMATIC	;QSS=36;TQSS=1;	NT=ref;QSS_NT=36;
,0:24,	24:0,0							
1	7985092 .				PASS	SOMATIC	;QSS=16;TQSS=1;	NT=ref;QSS_NT=16;
:0,0	21:0:0:0							
1	17562546		Α	T		PASS	SOMATIC;QSS=41	;TQSS=1;NT=ref;Q
:0:34,	35:0,0:0,0:0	9,0	32:0:0:0	:22,22:0,	0:0,0:10,10			
1	18141927		G			PASS	SOMATIC;QSS=19	;TQSS=1;NT=ref;Q
:0:0,0	0:0,0:18,19:0							
1	21200.00					PASS	SOMATIC;QSS=16	;TQSS=1;NT=ref;Q
:0:0,0	0:0,0:0,0:15,	. 15	23:0:0:0	:0,0:7,7:	0,0:16,16			
1	24992764			Α		PASS	SOMATIC;QSS=31	;TQSS=1;NT=ref;Q
	9:0,0:22,22:0	9,0	21:0:0:0	:9,9:0,0:	12,12:0,0			
	25949960			A		PASS	SOMATIC;QSS=18	;TQSS=1;NT=ref;Q
,0:15,			0:7,7:0,0		Θ			
1	26044713		С			PASS	SOMATIC;QSS=15	;TQSS=1;NT=ref;Q
	0:16,16:0,0:0				9:0,0:12,12			
	27747855			Α		PASS	SOMATIC;QSS=31	;TQSS=1;NT=ref;Q
:0:0,0	0:0,0:22,22:0				0:14,14:0,0			
1	28029437					PASS	SOMATIC;QSS=34	;TQSS=1;NT=ref;Q
:0:23,	24:0,0:0,0:0	9,0	25:0:0:0		0:10,10:0,0			
1	28393932					PASS	SOMATIC;QSS=33	;TQSS=1;NT=ref;Q
:0:0,0	0:0,0:0,0:23,	. 23	27:0:0:0	:0,0:0,0:	13,13:14,14			

Çhr	Start	End	Ref	Alt	Func_refGene	Gene refGene	GeneDetail refGene
1	1591986	1591986	G	Α	intergenic	SSU72;FNDC10	dist=17123;dist=6026
1	3605430	3605430	T	G	intronic	MEGF6	
1	3827839	3827839	G	Α	intronic	CEP104	
1	5491320	5491320	G	С	intergenic	AJAP1;MIR4689	dist=698798;dist=371352
1	7985092	7985092	G	Α	UTR3	PARK7	NM_001123377:c.*38G>A;NM_007262:c.*38G>A
1	17562546	17562546	Α	T	intronic	ARHGEF10L	_
1	18141927	18141927	G	Α	intronic	IGSF21	
1	21283788	21283788	T	С	intronic	ECE1	
1	24992764	24992764	G	Α	intergenic	RUNX3;MIR4425	dist=27754;dist=30739
1	25949960	25949960	G	Α	intergenic	STMN1;PAFAH2	dist=43083;dist=9807
1	26044713	26044713	С	T	intronic	SLC30A2	
1	27747855	27747855	G	Α	intronic	FAM76A	
1	28029437	28029437	Α	G	intronic	EYA3	
1	28393932	28393932	T	G	intronic	PHACTR4	

Solution: ANNOVAR https://annovar.openbioinformatics.org/en/latest/

