**LEISHMANIA DISEASE DETECTION**

A PROJECT REPORT

***Submitted by***

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**Abstract:**

Leishmaniasis is a parasitic disease caused by the Leishmania parasite, which is transmitted through the bite of infected sandflies. Early and accurate detection of Leishmaniasis is crucial for effective treatment and control of the disease. In this study, we propose a method for Leishmaniasis detection using DNA sequences.

The approach involves the analysis of DNA sequences to identify the presence of Leishmania parasites. We start by collecting DNA sequences from Leishmania-positive and Leishmania-negative samples. The sequences are processed and pre-processed to extract relevant features. These features may include nucleotide composition, GC content, motifs, or other sequence characteristics.

Next, we employ machine learning techniques such as K-Nearest Neighbor (KNN) , Support Vector Machines (SVM) to train models using the extracted features. The trained models are then used to predict the presence of Leishmaniasis in unseen DNA sequences. To evaluate the performance of the proposed method, we employ metrics such as accuracy, precision, recall, and F1-score. Additionally, we utilize techniques like cross-validation and confusion matrix analysis to assess the robustness and reliability of the models.The results of our study demonstrate the effectiveness of the proposed approach in detecting Leishmaniasis using DNA sequences. By accurately identifying infected samples, this method can aid in early diagnosis, prompt treatment, and effective control of Leishmaniasis.In conclusion, our study presents a novel approach for Leishmaniasis detection through the analysis of DNA sequences. The method combines bioinformatics techniques, machine learning algorithms, to provide accurate and efficient identification of the disease. This approach has the potential to contribute significantly to the field of Leishmaniasis diagnosis and improve public health outcomes.

**CHAPTER - 1**

**INTRODUCTION**

**Introduction to Leishmanisis:**

Leishmaniasis is a vector-borne parasitic disease caused by the Leishmania parasite. It is transmitted to humans through the bites of infected female sandflies, which are small insects found in tropical and subtropical regions. Leishmaniasis is considered a neglected tropical disease and affects millions of people worldwide, particularly in regions with poor socio-economic conditions.

The disease has a wide spectrum of clinical manifestations, ranging from mild skin lesions to severe systemic infections. There are three main forms of leishmaniasis: cutaneous, mucocutaneous, and visceral. Each form is caused by different species of Leishmania parasites and presents with distinct symptoms.

**Cutaneous Leishmaniasis**: This is the most common form of the disease. It is characterized by skin lesions at the site of the sandfly bite. The lesions may appear as painless, raised bumps or ulcers with a central crater. They can be single or multiple and may heal on their own over several months, leaving scars.

**Mucocutaneous Leishmaniasis**: This form affects the mucous membranes of the nose, mouth, and throat in addition to the skin. It typically occurs as a complication of untreated cutaneous leishmaniasis. Mucocutaneous leishmaniasis can lead to severe damage and disfigurement of the affected areas, including the destruction of nasal and oral tissues.

**Visceral Leishmaniasis** (also known as Kala-azar): This is the most severe form of leishmaniasis. It affects multiple organs, primarily the spleen, liver, and bone marrow. Visceral leishmaniasis can cause prolonged fever, weight loss, fatigue, anemia, and enlargement of the spleen and liver. If left untreated, it can be fatal.

Diagnosis of leishmaniasis involves clinical evaluation, microscopic examination of tissue samples or fluid aspirates, and laboratory tests such as serological assays or molecular methods. Treatment options include antiparasitic drugs, with the choice of medication depending on the form and severity of the disease.Prevention and control strategies for leishmaniasis focus on vector control measures, such as insecticide-treated nets, environmental management, and personal protective measures. Additionally, research efforts are underway to develop vaccines against the disease.

**Motivation:**

The motivation for Leishmaniasis disease detection stems from the urgent need to diagnose and treat the infection promptly. The Motivations are

**Early Diagnosis:** Early detection of Leishmaniasis is crucial for initiating timely treatment. Detecting the infection at an early stage can help prevent the progression of the disease, reduce the severity of symptoms, and minimize complications. Timely detection also enables the implementation of appropriate control measures to prevent the spread of the disease.

**Improved Patient Outcomes:** Accurate and timely detection of Leishmaniasis can significantly improve patient outcomes. It allows healthcare providers to prescribe the appropriate treatment regimen tailored to the specific form and severity of the disease. Early intervention can help prevent long-term complications and reduce morbidity and mortality associated with Leishmaniasis.

**Public Health Impact:** Effective detection of Leishmaniasis plays a crucial role in public health interventions. It allows for the identification of high-risk areas and populations, facilitating targeted control measures. Disease surveillance and monitoring through detection methods help in tracking the prevalence and incidence of Leishmaniasis, which is essential for planning and implementing effective prevention and control strategies.

**CHAPTER – 2**

**LITERATURE SURVEY**

**Paper - [1]: A two-step approach-machine learning, variational autoencoder, and weighted gene co-expression network analysis identify key signature genes and pathways implicated in active visceral leishmaniasis.**

In this study, they aimed to detect key signature genes and pathways implicated in active visceral leishmaniasis. We have fivegenes IFNG, SC5D, LSM1, CMC2, and SAR1B, with higher interamodular connectivity, as the key signature genes. These genes were validated using a machine learning algorithm by creating new gene features by applying a variational autoencoder. These genes are involved in processes like cytokine-cytokine receptor interaction, IL-17 signaling pathway, T-cell receptor signaling pathway, and Th1 and Th2 cell differentiation. Besides, hsa-miR-340-5p, hsa-miR-325-3p, hsa-miR-182-5p, hsa-miR-1271-5p/hsamiR-96-5p miRNAs were found to target key signature genes.

Now we are going to Detect the Leishmania species in DNA through a different approach. We are going to use Bio-informatics analysis and Machine learning techniques to detect the Leishmaniasis in the given DNA sequence accurately and Correctly.

**Paper - [2]: Molecular identification of Leishmania RNA virus in cutaneous**

**leishmaniasis patients.**

This research is the first molecular and phylogenetic study based on the LRV2 strains from rodent reservoirs and CL patients in Isfahan province. This study is the first comprehensive epidemiological study in the Isfahan province region regarding LRV2 in Leishmania species. The findings of this study were consistent with the results of previous studies and Leishmania isolates in the present study were from CL patients and rodent reservoirs in Isfahan province so could not be generalized to other geographical zones of Iran and the around countries.

Now we are going to Detect the Leishmania species in DNA through a different approach. We are going to use Bio-informatics analysis and Machine learning techniques to detect the Leishmaniasis in the given DNA sequence accurately and Correctly.

**CHAPTER – 3**

**SYSTEM DESIGN & IMPLEMENTATION**

**Brief:**

**Leishmaniasis:**

Leishmaniasis disease detection using DNA/RNA analysis involves the identification and characterization of the Leishmania parasite's genetic material in patient samples. DNA/RNA-based techniques provide a sensitive and specific approach to diagnose Leishmaniasis, particularly in cases where other diagnostic methods may yield inconclusive results. Here is an overview of some commonly used DNA/RNA-based methods for Leishmaniasis detection:

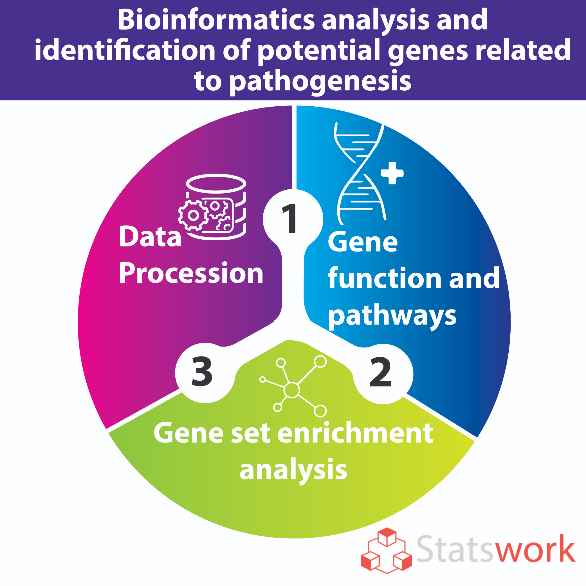
1. **Polymerase Chain Reaction (PCR):** PCR is a widely used technique that amplifies specific DNA/RNA sequences of the Leishmania parasite in patient samples. It involves the use of specific primers that target conserved regions of the Leishmania genome. By amplifying the target DNA/RNA, PCR enables the detection and quantification of the parasite. Different PCR variants, such as conventional PCR, real-time PCR, and nested PCR, can be employed depending on the specific requirements of the diagnostic assay.
2. **-Mediated Isothermal Amplification (LAMP):** LAMP is an isothermal amplification method that can rapidly and specifically amplify DNA/RNA sequences. It offers advantages over PCR, such as simplicity, rapidity, and robustness, as it does not require sophisticated thermal cycling equipment. LAMP has been successfully utilized for the detection of Leishmania DNA/RNA in patient samples.
3. **Bio-Informatics Analysis**
4. **K – Nearest Neighbour (Machine Learning)**
5. **Support Vector Machines(Machine Learning)**

In the above 5 techniques we are going to use Bio-Informatics Analysis , K – Nearest Neighbour (Machine Learning) , Support Vector Machines(Machine Learning).

1. **Bio-Informatics Analysis:**

Detecting Leishmania using bioinformatics analysis involves several steps, including data retrieval, sequence alignment, and identification. Here's an outline of the process:

1. **Data Retrieval:** Obtain the genetic sequence data for Leishmania from public databases like GenBank or other relevant sources. You can search for specific Leishmania species or use broader queries to retrieve a diverse set of sequences.
2. **Sequence Alignment:** Align the obtained Leishmania sequences with reference sequences to identify similarities and variations. Multiple sequence alignment tools like ClustalW, MAFFT, or Muscle can be used for this step. The alignment helps identify conserved regions across different Leishmania strains.
3. **Feature Extraction**: Extract relevant features from the aligned sequences, such as conserved motifs, protein domains, or regions of interest. This information can be useful for designing specific primers or probes for targeted Leishmania detection.
4. **Data Analysis and Interpretation**: Analyze the experimental results and interpret them in the context of Leishmania detection. Compare the obtained sequences with reference sequences, evaluate the amplification products, and assess the sensitivity and specificity of the assay.



**Leishmania DNA taken at the start of the Analysis it is of the length 2400 Nucleotides and Human sequence of 2500 length is taken and both are compared with Bio informatics analysis.** **If the Leishmania matches with Human DNA it return True that person have Leishmania.**

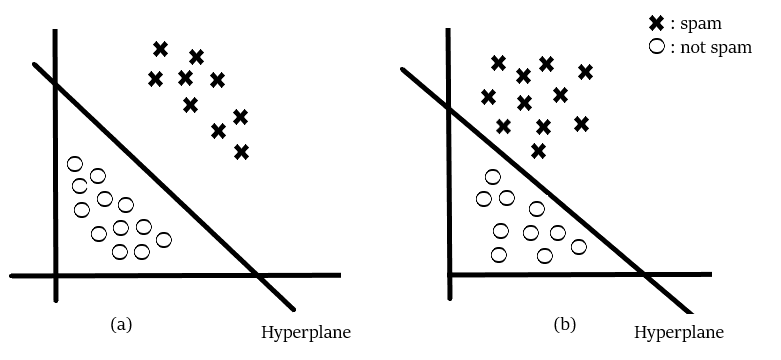
1. **K-Nearest Neighbor:**

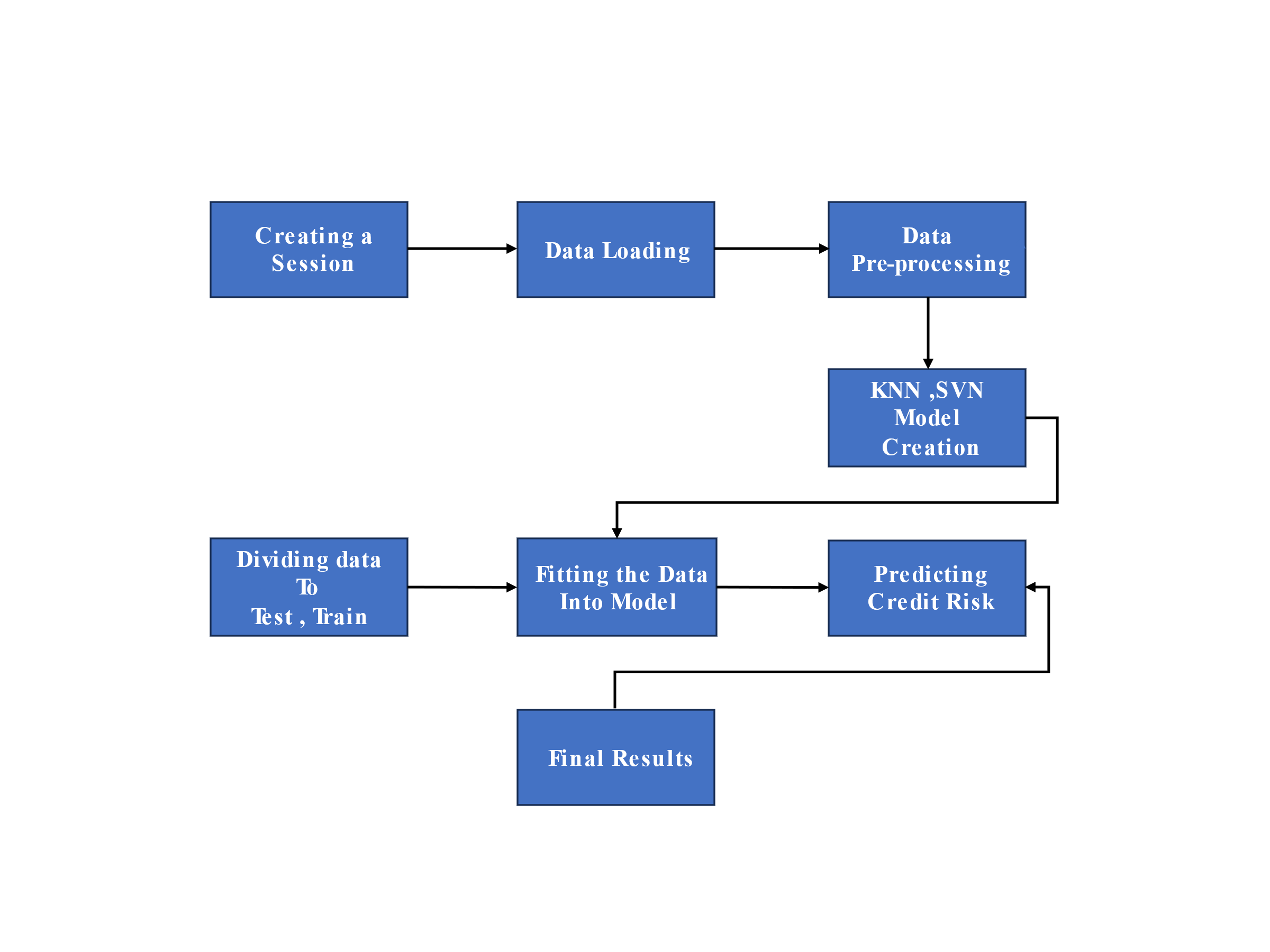
* From the given Data set find dependent and independent Variables.
* Divide the variables to training and testing data, it means Data pre-processing.
* Fit the train and test data.
* Using the KNN Classifier taking n neighbors find the predicted for test data
* From predicted data find Accuracy, Precision, Recall, F1-Score etc.
* Specifically, four different distance functions, which are Euclidean distance, cosine similarity measure, correlation, and Chi square, are used in the k-NN classifier respectively

*Fig-1*

1. **Support Vector Machines:**

* From the given Data set find dependent and independent Variables.
* Divide the variables to training and testing data, it means Data pre-processing.
* Fit the train and test data.
* Using the SVC Classifier taking path as linear, curve find the predicted for test data
* From predicted data find Accuracy, Precision, Recall, F1-Score etc.

Three different types of SVM-Kernels are displayed below. The polynomial and RBF are especially useful when the data-points are not linearly separable

**Design****:**

**Input Data**

**Training Data**

**Training for ML Algorithm**

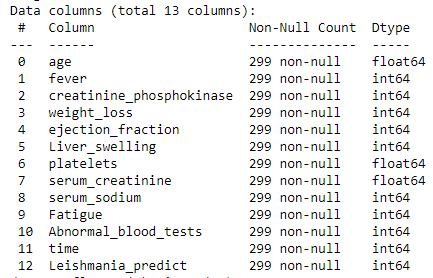
**Model Input**

**ML Algorithm**

**Successful Model**

**Prediction**

**Implementation**

1. **Data Set Loading:**
2. **For KNN & SVM:**
3. **For Bio-Informatics Analysis:**

**Lesishmania.fasta file:**

GATTTAAGTGAATAGCTTGGCTATCTCACTTCCCCTCGTTCTCTTGCAGAACTTTGATTTTAACGAACTT

AAATAAAAGCCCTGTTGTTTAGCGTATCGTTGCACTTGTCTGGTGGGATTGTGGCATTAATTTGCCTGCT

CATCTAGGCAGTGGACATATGCTCAACACTGGGTATAATTCTAATTGAATACTATTTTTCAGTTAGAGCG

TCGTGTCTCTTGTACGTCTCGGTCACAATACACGGTTTCGTCCGGTGCGTGGCAATTCGGGGCACATCAT

GTCTTTCGTGGCTGGTGTGACCGCGCAAGGTGCGCGCGGTACGTATCGAGCAGCGCTCAACTCTGAAAAA

CATCAAGACCATGTGTCTCTAACTGTGCCACTCTGTGGTTCAGGAAACCTGGTTGAAAAACTTTCACCAT

GGTTCATGGATGGCGAAAATGCCTATGAAGTGGTGAAGGCCATGTTACTTAAAAAGGAGCCACTTCTCTA

TGTGCCCATCCGGCTGGCTGGACACACTAGACACCTCCCAGGTCCTCGTGTGTACCTGGTTGAGAGGCTC

ATTGCTTGTGAAAATCCATTCATGGTTAACCAATTGGCTTATAGCTCTAGTGCAAATGGCAGCCTGGTTG

GCACAACTTTGCAGGGCAAGCCTATTGGTATGTTCTTCCCTTATGACATCGAACTTGTCACAGGAAAGCA

AAATATTCTCCTGCGCAAGTATGGCCGTGGTGGTTATCACTACACCCCATTCCACTATGAGCGAGACAAC

ACCTCTTGCCCTGAGTGGATGGACGATTTTGAGGCGGATCCTAAAGGCAAATATGCCCAGAATCTGCTTA

AGAAGTTGATTGGCGGTGATGTCACTCCAGTTGACCAATACATGTGTGGCGTTGATGGAAAACCCATTAG

TGCCTACGCATTTTTAATGGCCAAGGATGGAATAACCAAACTGGCTGATGTTGAAGCGGACGTCGCAGCA

CGTGCTGATGACGAAGGCTTCATCACATTAAAGAACAATCTATATAGATTGGTTTGGCATGTTGAGCGTA

AAGACGTTCCATATCCTAAGCAATCTATTTTTACTATTAATAGTGTGGTCCAAAAGGATGGTGTTGAAAA

CACTCCTCCTCACTATTTTACTCTTGGATGCAAAATTTTAACGCTCACCCCACGCAACAAGTGGAGTGGC

GTTTCTGACTTGTCCCTCAAACAAAAACTCCTTTACACCTTCTATGGTAAGGAGTCACTTGAGAACCCAA

CCTACATTTACCACTCCGCATTCATTGAGTGTGGAAGTTGTGGTAATGATTCCTGGCTTACAGGGAATGC

TATCCAAGGGTTTGCCTGTGGATGTGGGGCATCATATACAGCTAATGATGTCGAAGTCCAATCATCTGGC

ATGATTAAGCCAAATGCTCTTCTTTGTGCTACTTGCCCCTTTGCTAAGGGTGATAGCTGTTCTTCTAATT

GCAAACATTCAGTTGCTCAGTTGGTTAGTTACCTTTCTGAACGCTGTAATGTTATTGCTGATTCTAAGTC

CTTCACACTTATCTTTGGTGGCGTAGCTTACGCCTACTTTGGATGTGAGGAAGGTACTATGTACTTTGTG

1. **Preprocessing:**

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1. **Correlation Matrix:**

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**CHAPTER – 4**

**RESULTS AND ANALYSIS**

**Result:**

1. **Bioinformatics:**

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1. **KNN & SVM:**

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**Analysis:**

In the abpve 2 techniques SVM is showing accuracy more and it has the more predicting power.

**CHAPTER – 5**

**CONCLUSION**

**Conclusion:**

In core research paper they have used genes for predicting the Leishmania using Auto Encoders and weighted gene co-expression network analysis. But we have used only Symptoms of Leishmaniasis and with those symptoms we Combined with K-Nearest Neighbour (KNN) & Support vector Machines (SVM) ML Algorithms and we got an efficient Output.

We got an accuracy, precision & Recall as follows:

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And also we have taken 2 sequences (one with Leishmaniasis sequence & other is Human DNA sequence) Both sequences are analysed with the Bio Informatics Analysis, we have taken a sample of 2400 length fasta file of leishmaniasis and 2556 length of human DNA sequence and both are compared. The result is the algorithm calculates the GC content in the to be tested Sequence and also calculates the score to compare with Leishmania sequence.

We have taken 2 human sequences and one got having leishmania and other don’t have leishmaniasis.

**CHAPTER – 5**

**REFERENCES**

1.WHO: Leishmaniasis Fact Sheet - Google Scholar <https://www.who.int/news-room/fact-sheets/detail/leishmaniasis>

2. Croft, S. L., Sundar, S., & Fairlamb, A. H. (2006). Drug Resistance in Leishmaniasis. Clinical Microbiology Reviews, 19(1), 111–

126. <https://doi.org/10.1128/CMR.19.1.111-126.2006>

3. Ma, H., & Zhao, H. (2013). Drug target inference through pathway analysis of genomics data. Advanced Drug Delivery

Reviews, 65(7), 966–972. <https://doi.org/10.1016/j.addr.2012.12.004>

4. Rabinowitz, J. D., Purdy, J. G., Vastag, L., Shenk, T., & Koyuncu, E. (2011). Metabolomics in Drug Target Discovery. Cold Spring

Harbor Symposia on Quantitative Biology, 76, 235–246. <https://doi.org/10.1101/sqb.2011.76.010694>

5. Zinzalla, G., & Thurston, D. E. (2009). Targeting protein–protein interactions for therapeutic intervention: A challenge for the

future. Future Medicinal Chemistry, 1(1), 65–93. <https://doi.org/10.4155/fmc.09.12>