Analysing Twitter for Ubisoft

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8.1 Analysing the Relationship Between Friends and Followers for Twitter Users

8.1.1 Retrieve the posts from Twitter

relevant posts can be retrieved from twitter by utilising the rtweet package, packages can be loaded for use in **R** thusly:

```
# Load Packages
   setwd("~/Dropbox/Notes/DataSci/Social_Web_Analytics/SWA-Project/scripts_
   if (require("pacman")) {
     library(pacman)
  } else{
     install.packages("pacman")
     library(pacman)
9
10
   pacman::p_load(xts, sp, gstat, ggplot2, rmarkdown, reshape2,
11
                  ggmap, parallel, dplyr, plotly, tidyverse,
12
                  reticulate, UsingR, Rmpfr, swirl, corrplot,
13
                   gridExtra, mise, latex2exp, tree, rpart,
14
                  lattice, coin, primes, epitools, maps, clipr,
15
                   ggmap, twitteR, ROAuth, tm, rtweet, base64enc,
16
                  httpuv, SnowballC, RColorBrewer, wordcloud,
17
                   ggwordcloud, tidyverse, boot)
```

Listing 1: Load the Packages for *R*

The rtweet API will search for tweets that contain all the words of a query regardless of uppercase or lowercase usage [5].

In order to leverage the *Twitter* API it is necessary to use tokens provided through a *Twitter* developer account:

and hence all tweets containing a mention of *Ubisoft* can be returned and saved to disk as shown in listing 3:

```
# Set up Tokens
 options(RCurlOptions = list(
  verbose = FALSE,
  capath = system.file("CurlSSL", "cacert.pem", package = "RCurl"),
  ssl.verifypeer = FALSE
 ))
 setup_twitter_oauth(
  consumer_secret =
  12
  access secret = "*******************************
13
 )
14
15
 # rtweet
16
   ______
 tk <-
     rtweet::create_token(
  app = "SWA",
18
          = "************************
  consumer_key
19
  consumer secret =
20
  access_token
^{21}
  access_secret
  set_renv
           = FALSE
23
```

Listing 2: Import the twitter tokens (redacted)

Listing 3: Save the Tweets to the HDD as an rdata file

8.2.2 Count of Followers and Friends

In order to identify the number of users that are contained in the *tweets* the unique() function can be used to return a vector of names which can then be passed as an index to the vector of counts as shown in listing 4, this provides that 81.7% of the tweets are by unique users.

```
1 (users <- unique(tweets.company$name)) %>% length()
2 x <- tweets.company$followers_count[duplicated(tweets.company$name)]
3 y <- tweets.company$friends_count[duplicated(tweets.company$name)]
4
5 ## > [1] 817
```

Listing 4: Return follower count of twitter posts

8.1.3 Summary Statistics

The average number of friends and followers from users who posted tweets mentioning *Ubisoft* can be returned using the mean() as shown in listing 5 this provides that on average each user has 586 friends and 63.620 followers.

```
1 x <- rnorm(090)
2 y <- rnorm(090)
3 (xbar <- mean(x))
4 (ybar <- mean(y))
5
6 ## > [1] 4295.195
7 ## > [1] 435.9449
```

Listing 5: Determine the average number of friends and followers

8.1.4 Above Average Followers

Each user can be compared to the average number of followers, by using a logical operator on the vector (e.g. y > ybar), this will return an output of logical values. R will coerce logicals into 1/0 values meaning that the mean value will return the proportion of TRUE responses as shown in listing 6. This provides that:

- 2.4% of the have identified have an above average **number of followers**.
- 20.6% of the users identified have an above average **number of friends**.

```
1  (px_hat <- mean(x>xbar))
2  (py_hat <- mean(y>ybar))
3
4  ## > [1] 0.0244798
5  ## > [1] 0.2729498
```

Listing 6: Calculate the proportion of users with above average follower counts

8.1.5 Bootstrap confidence intervals

a/b.) Generate a bootsrap distribution

A bootstrap assumes that the population is an infinitely large repetition of the sample and may be produces with respect to follower counts by resampling with replacement/repetition and plotted using the ggplot2 library as deomonstrated in listings 7 and .1 and shown in figure 1.

This shows that the population follower counts is a non-normal skew-right distribution, which is expected because the number of friends is an integer value bound by zero [6].

```
1 ## Resample the Data
2 (bt_pop <- sample(x, size = 10^6, replace = TRUE)) %>% head()
3
4 ## > [1] 7 515 262 309 186 166
```

Listing 7: Bootstrapping a population from the sample.

```
## Make the Population
bt_pop_data <- tibble("Followers" = bt_pop)
ggplot(data = bt_pop_data, aes(x = Followers)) +

geom_histogram(aes(y = ..density..), fill = "lightblue", bins = 35,

col = "pink") +

geom_density(col = "violetred2") +

scale_x_continuous(limits = c(1, 800)) +

theme_bw() +

labs(x = "Number of Followers", y = "Density",

title = "Bootstrapped population of Follower Numbers")</pre>
```

c.) Estimate a Confidence Interval for the population mean Follower Counts

In order to perform a bootrap for the population mean value of follower counts it is necessary to:

1. Resample the data with replacement

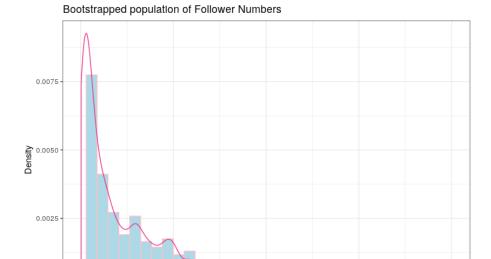


Figure 1: Histogram of the bootrapped population of follower counts

Number of Followers

- i.e. randomly select values from the sample allowing for repetition
- 2. Measure the statistic of concern

0.0000

- 3. Replicate this a sufficient number of times
 - i.e. Greater than or equal to 1000 times [2, Ch. 5]

This is equivalent to drawing a sample from a population that is infinitely large and constructed of repetitions of the sample. This can be performed in \mathbf{R} as shown in listing 8.

```
1 xbar_boot_loop <- replicate(10^3, {
2    s <- sample(x, replace = TRUE)
3    mean(s)
4    })
5 quantile(xbar_boot_loop, c((1-0.97)/2, (1+0.97)/2))
6
7 ##    1.5%    98.5%
8 ##    588.4189 10228.7352</pre>
```

Listing 8: Confidence Interval of Mean Follower Count in Population

A 97% probability interval is such that a sample drawn from a population will contain the population mean in that interval 97% of the time, this means that it may be concluded with a high degree of certainty that the true population mean lies between 588 and 10228.

- 1. Alternative Approaches If this data was normally distributed it may have been appropriate to consider bootstrapping the standard error and using a t distribution, however it is more appropriate to use a percentile interval for skewed data such as this, in saying that however this method is not considered to be very accurate in the literature and is often too narrow. [3, Section 4.1]
 - It's worth noting that the normal t value bootstrap offers no advantage over using a t distribution (other than being illustrative of bootstrapping generally) [3, Section 4.1]

The boot package is a bootstrapping library common among authors in the data science sphere [4, p. 295] [8, p. 237] that implements confidence intervals consistent with work by Davison and Hinkley [7] in there texbook *Bootstrap Methods and their Application*. In this work it is provided that the BC_a method of constructing confidence intervals is superior to mere percentile methods in terms of accuracy [2, Ch. 5], a sentiment echoed in the literature. [1, 2, Ch. 5]

Such methods can be implemented in $\it R$ by passing a function to the the boot call as shown in listing 9. This provides a broader interval, providing that the true confidence interval could lie between 1079 and 16227 followers.

```
xbar_boot <- boot(data = x, statistic = mean_val, R = 10^3)</pre>
   boot.ci(xbar_boot, conf = 0.97, type = "bca", index = 1)
   ## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
   ## Based on 1000 bootstrap replicates
   ## CALL :
   ## boot.ci(boot.out = xbar_boot, conf = 0.97, type = "bca", index = 1)
   ## Intervals :
10
   ## Level
                  BCa
11
   ## 97%
            ( 1079, 16227 )
12
   ## Calculations and Intervals on Original Scale
13
   ## Warning : BCa Intervals used Extreme Quantiles
14
   ## Some BCa intervals may be unstable
   ## Warning message:
   ## In norm.inter(t, adj.alpha) : extreme order statistics used as
       endpoints
```

Listing 9: Bootstrap of population mean follower count implementing the BC_a method

references

d.) Estimate a Confidence Interval for the population mean Friend Counts

A Confidence interval for the population mean friend counts may be constructed in a like wise fashion as shown in listings 10. This provides that the 97% confidence interval for the population mean friend count is between 384 and 502 (or 387 and 496 if the BC_a method used, they're quite close and so the more conservative percentile method will be accepted).

```
# d.) Estimate a Confidence Interval for the populattion mean Friend
   \hookrightarrow Count ===
  # Using a Percentile Method
   ybar_boot_loop <- replicate(10^3, {</pre>
     s <- sample(y, replace = TRUE)</pre>
    mean(s)
  quantile(ybar_boot_loop, c(0.015, 0.985)
  # Using BCA Method
   mean_val <- function(data, index) {</pre>
    X = data[index]
    return(mean(X))
12
  }
13
  xbar_boot <- boot(data = y, statistic = mean_val, R = 10^3)</pre>
15
  boot.ci(xbar_boot, conf = 0.97, type = "bca", index = 1)
17
18
         1.5%
                 98.5%
19
  ## 383.7619 501.5903
20
  ##
21
  ## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
22
  ## Based on 1000 bootstrap replicates
23
24
  ##
  ## CALL :
25
  ## boot.ci(boot.out = xbar_boot, conf = 0.97, type = "bca", index = 1)
26
27
  ## Intervals :
28
  ## Level
                BCa
29
          (386.8, 496.7)
  ## 97%
30
  ## Calculations and Intervals on Original Scale
31
  ## Some BCa intervals may be unstable
```

Listing 10: Bootstrap of population mean follower count

FIXME 8.1.6 Estimate a 97% Confidence Interval for the High Friend Count Proportion

In order to bootstrap a confidence interval for the proportion of users with above average follower counts, repeteadly draw random samples from an infinitely large population composed entirely of the sample, and record the sampled proportion. this can be acheived by resampling the observations of above and below as shown in listing 11.

This provides that:

- The 97% confidence interval for the population proportion of users that have an above average number of friends is between 0.24 and 0.31.
 - i.e. The probability of any given sample containing the population mean within this interval would be 97%, although that doesn't however mean that there is a 97% probability that this interval contains the value, merely that we may be 97% confident

8.1.7 Is the Number of Friends Independent to the Number of Followers

One method to determine whether or not the number of followers is independent of the number of friends is to bin the counts and determine whether or not the distribution of users across those counts is consistent with the hypothesis of independence.

Bin the Follower and Friend Categories

The counts may be binned by performing a logical interval test as shown in listing 12.

Find the Group frequency

These values may be tabluated in order to count the occurrence of users among these categories as shown in listing 13 and table 1.

Table 1: Table of Binned Friend and Follower counts, transposed relative to code.

	Followers	Friends
Tens	421	262
Hundreds	317	476
1 - Thousands	39	47
2 - Thousands	11	15
3 - Thousands	9	6
4 - Thousands	2	9
5 Thousand or More	18	2

Find the Expected Counts under each group and test for independence

The expected count of each cell, under the assumption that the two metrics are independent, will be the proportion users per bracket multiplied by the number of users in that status group. This implies that any cell will be:

```
1 # 8.1.6 High Friend Count Proportion
   prop <- factor(c("Below", "Above"))</pre>
  ## 1 is above average, 2 is below
   py_hat_bt <- replicate(10^3, {</pre>
           <- sample(c("Below", "Above"),</pre>
                       size = length(y),
                       prob = c(py_hat, 1-py_hat),
                       replace = TRUE)
   isabove <- rs == "Above"</pre>
   mean(isabove)
10
11
12
   quantile(py_hat_bt, c(0.015, 0.985))
13
14
           1.5%
                    98.5%
  ##
  ## 0.2399021 0.3072215
16
  ## > > > . + > > >
   ## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
  ## Based on 1000 bootstrap replicates
19
20
  ## CALL :
21
  ## boot.ci(boot.out = py_hat_boot, conf = 0.97, type = "bca")
22
  ## Intervals :
25
  ## Level
                 BCa
  ## 97% ( 0.2399,  0.3072 )
26
   ## Calculations and Intervals on Original Scale
27
```

Listing 11: Bootstrap of Proportion of Friends above average

```
## Assign Categories
2 x_df <- data.frame(x)</pre>
                    <= x_df$x & x_df$x < 100] <- "Tens"</pre>
з x_df$cat[0
4 x_df$cat[100
                    <= x_df$x & x_df$x < 1000] <- "Hundreds"</pre>
5 x_df$cat[1000
                    <= x_df$x & x_df$x < 2000] <- "1Thousands"</pre>
6 x_df$cat[2000
                    <= x_df$x & x_df$x < 3000] <- "2Thousands"
  x_df$cat[3000
                    <= x_df$x & x_df$x < 4000] <- "3Thousands"
                  <= x_df$x & x_df$x < 5000] <- "4Thousands"</pre>
  x_dfcat [4000
  x_dfcat [5000
                    <= x_df$x & x_df$x < Inf] <- "5ThousandOrMore"</pre>
10
11
  ### Make a factor
  x_df$cat <- factor(x_df$cat, levels = var_levels, ordered = TRUE)</pre>
12
13
  ### Determine Frequencies
  (x_freq <- table(x_df$cat) %>% as.matrix())
  ## ** b) Find the Friend Count Frequency
17
   →
  ## Assign Categories
  y_df <- data.frame(y)</pre>
                    <= y_df$y & y_df$y < 100] <- "Tens"
  y_df$cat[0
20
  y_df$cat[100
                    <= y_df$y & y_df$y < 1000] <- "Hundreds"</pre>
  y_df$cat[1000
                    <= y_df$y & y_df$y < 2000] <- "1Thousands"</pre>
22
                    <= y_df$y & y_df$y < 3000] <- "2Thousands"
23
  y_df$cat[2000
  y_df$cat[3000
                    <= y_df$y & y_df$y < 4000] <- "3Thousands"</pre>
^{24}
  y_df$cat[4000
                    <= y_df$y & y_df$y < 5000] <- "4Thousands"
25
  y_df$cat[5000
                    <= y_df$y & y_df$y < Inf] <- "5ThousandOrMore"</pre>
26
27
  ### Make a factor
28
  y_df$cat <- factor(y_df$cat, levels = var_levels, ordered = TRUE)</pre>
29
30
  ### Determine Frequencies
31
  (y_freq <- table(y_df$cat) %>% as.matrix())
```

Listing 12: Use Logical Test to Assign observations into bins

```
vals <- t(cbind(x_freq, y_freq))</pre>
  rownames(vals) <- c("Followers.x", "followers.y")</pre>
   vals
                   Tens Hundreds 1Thousands 2Thousands 3Thousands 4Thousands
   ## Followers.x
                                           39
                   421
                              317
                                                       11
                                                      15
                                                                               9
   ## followers.y 262
                             476
                   5ThousandOrMore
   ## Followers.x
                                 18
9
   ## followers.y
                                  2
```

Listing 13: Tabulate the binned counts for the distribution of users among amount and status.

• the product of the row sum, multiplied by the column sum divided by the number of counts.

This can be equivalently expressed as an outer product as shown in equation (1), in R this operation is denoted by the %0% operator, which is shorthand for the outer() function, this and other summary statistics may be evaluated as shown in listing 14.

The outer product is such that:

$$\mathbf{u} \otimes \mathbf{v} = \mathbf{u} \mathbf{v}^{\mathsf{T}} = \begin{bmatrix} u_1 \\ u_2 \\ u_3 \\ u_4 \end{bmatrix} \begin{bmatrix} v_1 & v_2 & v_3 \end{bmatrix} = \begin{bmatrix} u_1 v_1 & u_1 v_2 & u_1 v_3 \\ u_2 v_1 & u_2 v_2 & u_2 v_3 \\ u_3 v_1 & u_3 v_2 & u_3 v_3 \\ u_4 v_1 & u_4 v_2 & u_4 v_3 \end{bmatrix}.$$

This means the matrix of expected frequencies can be expressed as an outer product thusly:

$$\tilde{\mathbf{e}} = \frac{1}{n} \times \begin{bmatrix} \sum_{j=1}^{n} [o_{1j}] \\ \sum_{j=1}^{n} [o_{2j}] \\ \sum_{j=1}^{n} [o_{3j}] \\ \sum_{j=1}^{n} [o_{4j}] \\ \vdots \\ \sum_{j=1}^{n} [o_{nj}] \end{bmatrix} \begin{bmatrix} \sum_{j=1}^{n} [o_{i1}] \\ \sum_{j=1}^{n} [o_{i2}] \\ \sum_{j=1}^{n} [o_{i3}] \\ \vdots \\ \sum_{j=1}^{n} [o_{nj}] \end{bmatrix}^{T}$$

$$(1)$$

```
## ***** Calculate Summary Stats
n <- sum(vals)
bracket_prop <- colSums(vals) / n
metric_prop <- rowSums(vals) / n
o <- vals
e e <- rowSums(vals) %o% colSums(vals) / n
chi_obs <- sum((e-o)^2/e)</pre>
```

Listing 14: Calculate Expected frequency of values under the assumption of independence.

1. Testing Independence In order to test whether or not the distribution of users among brackets is independent of being a follower or friend a χ^2 test may be used, this can be evaluated from a model or simulated, in $\textbf{\textit{R}}$, the simulated test is shown in listing 15, this provides a p-value < 0.0005, which means that the hypothesis of independence may be rejected with a high degree of certainty.

```
chisq.test(vals, simulate.p.value = TRUE)

## ^IPearson's Chi-squared test with simulated p-value (based on 2000)

## ^Ireplicates)

## data: vals

## X-squared = 88.109, df = NA, p-value = 0.0004998
```

Listing 15: Chi-Square testing for independence between friend and follower bin categories.

(a) From First Principles The χ^2 statistic may be performed from first principles by randomly sampling the values at the rate at which they occured, tabulating those counts, measuring the χ^2 -value and then repeating this many times.

Because the samples are random they must be independent and average number of positives is hence an estimate for the FPR, which is in turn an estimate for the p-value. This technique is demonstrated in listing 16, the p-value being returned as 0.0004, this value is consistent with the value produced by R's built in chisq.test function and so is accepted.

FIXME Conclusion

The p-value measures the probability of rejecting the null hypothesis when it is true, i.e. the probability of a detecting a *false positive*, a very small p-value is hence good evidence that the null hypothesis should be rejected (because doing so would unlikely to be a mistake).

In saying that however the p-value is distinct from the *power* statistic, which is a measure of /the probability of accepting the alternative hypothesis when it is true, a low p-value is not a measurement of the probability of being correct.

Hence me way conclude, with a high degree of certainty, that the follower and friend counts are not independent of one another.

8.2 Finding Themes in tweets

8.2.8 Find Users with Above Average Friend Counts

Users with Above average Friend Counts can be identified by filtering the tweets data frame for two conditions:

1. non-duplicated user-id

```
## **** Create Vectors of factor levels
   brackets <- unique(x_df$cat)</pre>
   metrics <- c("follower", "friend")</pre>
   ## **** Simulate the data Assuming H_O
   ## I.e. assuming that the null hypothesis is true in that
   ## the brackets assigned to followers are independent of the friends
   ## (this is a symmetric relation)
   s <- replicate(10^4,{</pre>
10
     ## Sample the set of Metrics
11
     m <- sample(metrics, size = n, replace = TRUE, prob = metric_prop)</pre>
12
13
     ## Sample the set of Brackets (i.e. which performance bracket the
14
      \hookrightarrow user falls in)
     b <- sample(brackets, size = n, replace = TRUE, prob = bracket_prop)</pre>
15
16
     ## Make a table of results
17
     o <- table(m, b)
18
     0
19
20
     ## Find What the expected value would be
^{21}
     e_sim <- t(colSums(e) %o% rowSums(e) / n)</pre>
22
23
     ## Calculate the Chi Stat
^{24}
     chi_sim <- sum((e_sim-o)^2/e_sim)</pre>
25
     chi_sim
26
27
     ## Is this more extreme, i.e. would we reject null hypothesis?
28
29
     chi_sim > chi_obs
30
   })
31
32
   mean(s)
33
34
   ## [1] 4e-04
35
```

Listing 16: Performing a χ^2 statistic from first principles

2. friend_count greater than average

This can be acheived easily using the dplyr package as shown in 17, the top 20 of these users are shown in table 3 of the appendix

```
select <- dplyr::select
filter <- dplyr::filter</pre>
   interested_vars <- c("user_id", "friends_count")</pre>
   (friend counts <- tweets.company %>%
     select(interested_vars) %>%
     filter(!duplicated(user_id)))
6
   (high_friends <- friend_counts %>%
     filter(friends_count > mean(friends_count, na.rm = TRUE)))
   ## Export Friends List
   write.csv(high_friends[order(
12
13
     high_friends$friends_count,
     decreasing = TRUE),], file = "/tmp/highfriend.csv")
14
```

Listing 17: Use dplyr to Filter for Users with a high Friend Count

8.2.9 Find Users with Below Average Friend Counts

Users with high friends may be determined by a similar method (or by taking the complement of the high friends) as shown in listing 18, the lowest 20 of these users are shown in table 4 of the appendix.

```
(low_friends <- friend_counts %>%
filter(friends_count <= mean(friends_count, na.rm = TRUE)))

low_friends <- low_friends[order(
    low_friends$friends_count,
    decreasing = TRUE),]

## Export Users
write.csv(low_friends[order(
    low_friends$friends_count,
    decreasing = FALSE),], file = "/tmp/lowfriend.csv")</pre>
```

Listing 18: Use dplyr to Filter for Users with a low Friend Count

8.2.10 Find the *Tweets* corresponding to users with high or low friend counts

The tweets corresponding to users with high and low friend counts can be identified by filtering the dataframe based on the friend count and using that to the index the tweets from the data frame ¹, alternatively it is possible to test whether or not the ID of a user appears in the high or low vector set using the %in% operator as shown in listing 19.

```
## Method 1
2 friend_test <- tweets.company$friends_count >

→ mean(tweets.company$friends_count)
  tweets_high <- tweets.company$text[friend_test]</pre>
   tweets_low <- tweets.company$text[!friend_test]</pre>
  ## Method 2
            :15b5a74:
  tweets high <- tweets.company$text[tweets.company$user id %in%
    → high_friends$user_id]
  tweets_low <- tweets.company$text[tweets.company$user_id %in%</pre>
    → low_friends$user_id]
   tweets <- c(tweets_high, tweets_low)</pre>
   ## Mark as High or Low
  tweets_low <- cbind(tweets_low, rep("Low_Friend", length(tweets_low)))</pre>
  tweets_high <- cbind(tweets_high, rep("High_Friend",</pre>
   → length(tweets_high)))
tweets <- as.data.frame(rbind(tweets_high, tweets_low))</pre>
  tweets$Friend_Status <- factor(tweets$Friend_Status)</pre>
```

Listing 19: Identify tweets corresponding to users with high and low friend counts

8.2.11 Clean the tweets

Create a Corpus Object

In order to clean the tweets it is necessary to create a corpus object as shown in listing 20, it is possible to pass a dataframe source in order to include the user ID, this isn't strictly necessary however because the tm package preserves order when performing transformations.

¹This works because the tm package preserves the order of the data, this can be confirmed by using a dataframe source as opposed to a vector source (e.g. in listing 20) and comparing the ID's before/after transformation.

```
tweet_source <- tm::VectorSource(tweets$text)
tweet_corpus <- tm::Corpus(x = tweet_source)</pre>
```

 $Listing\ 20:$ Create a Corpus from the tweets

Next it is necessary to choose an enoding, a primary consideration of this is whether or not the use of *emoji* characters will influence the model performance. There is research to suggest that Emoji's can be used as predictive features [lecompte2017] and that they can improve *sentiment analysis* models [shiha2017] that implement a *bag of words* approach. For these reasons *emoji* characters will be preserved and UTF-8 implemented.

In order to encode the data as UTF-8, the iconv function can be used as shown in listing 21.

```
1 encode <- function(x) {
2    iconv(x, to = "UTF-8")
3 # iconv(x, to = "latin1")
4  # iconv(x, to = "ASCII")
5 }
6
7 tweet_corpus <- tm_map(x = tweet_corpus, FUN = encode)
8 tweet_corpus_raw <- tweet_corpus</pre>
```

Listing 21: Encode the Data as UTF-8

Process the tweets

Before analysis the tweets should be modified to remove characters that may interfere with categorising words, this is referred to as cleaning, in particular the following should be implemented:

- 1. Remove URL's
- 2. Remove Usernames
- 3. remove numbers
- 4. remove punctuation
- 5. remove whitespace
- 6. case fold all characters to lower case
- 7. remove a set of stop words
- 8. reduce each word to its stem

In particular it is important to reduce words to lower case before removing stop words otherwise an unorthodox use of capitalisation may prevent the word from being removed throughout.

The stop word ubisoft will also be used, this was the query term so it's expected to turn up at a very high frequency, the words can and 's also occured quite frequently and so were removed.

The cleaning can be implemented by mapping functions over the corpus, which is fundamentally a list, this can be performed via the tm_map function as shown in listing 22.

```
mystop <- c(stopwords(), "s", "can", "ubisoft", "@ubisoft",</pre>
    → "#ubisoft")# <<stphere>>
   clean_corp <- function(corpus) {</pre>
      ## Remove URL's
      corpus <- tm_map(corpus,content_transformer(function(x))</pre>

    gsub("(f|ht)tp(s?)://\\S+","",x)))

      ## Remove Usernames
      corpus <- tm_map(corpus,content_transformer(function(x)</pre>

    gsub("@\\w+","",x)))

      ## Misc
     corpus <- tm map(corpus, FUN = removeNumbers)</pre>
      corpus <- tm_map(corpus, FUN = removePunctuation)</pre>
     corpus <- tm map(corpus, FUN = stripWhitespace)</pre>
     corpus <- tm_map(corpus, FUN = tolower)</pre>
12
      corpus <- tm_map(corpus, FUN = removeWords, mystop)</pre>
13
14
      ## stopwords() returns characters and is fead as second argument
      corpus <- tm_map(corpus, FUN = stemDocument)</pre>
15
      return(corpus)
16
   }
17
18
  tweet_corpus_clean <- clean_corp(tweet_corpus)</pre>
```

Listing 22: Use the tm_map function to clean the tweets

8.2.12 Display the first two tweets before/after processing

The tweets can be viewed from inside the corpus by selecting with the [function 2 as demonstrated in listing 23, the first *tweet* was rendered empty by the processing and the following two tweets were:

Pre-Processing

- "Today was the first time in over a month that I have gone 24 hours without checking the coronavirus death toll. Thanks @Ubisoft."

²The [function is actually shorthand for Extract(), most things in R are functions, this is similar to LISP and has to do with the origins of the language, e.g. sum(1:10) = (sum (1:10)), also relevant see the relevant xkcd in figure 5.

- "@btwimskrank @TheDivisionGame @UbiMassive @Ubisoft @jgerighty @hamishbode @Tideman92 @janeyo_jane @slimjd Very odd... I'll even post a video about it."

Post-Processing

- "today first time month gone hour without check coronavirus death toll thank"
- "odd ill even post video"

```
tweet_corpus_raw[[1]]$content
tweet_corpus_clean[[1]]$content
tweet_corpus_raw[[2]]$content
tweet_corpus_clean[[2]]$content
tweet_corpus_raw[[3]]$content
tweet_corpus_clean[[3]]$content
tweet_corpus_clean[[3]]$content
```

Listing 23: Load the Packages for *R*

8.2.13 Create a Term Document Matrix

Apply Weighting Manually

1. Create Term Documen Matrix A term Document matrix (and it's transpose) can be constructed from a corpus using the tm:TermDocumentMatrix function as shown in listing 24.

```
tweet_matrix_tdm <- TermDocumentMatrix(tweet_corpus_clean)
tweet_matrix_dtm <- DocumentTermMatrix(tweet_corpus_clean)</pre>
```

Listing 24: Load the Packages for **R**

- 2. Apply TF-IDF Weighting Weighted term frequency is defined as shown in equation (2), where:
 - ullet $f_{d,t}$ is the frequency of a given term t in the a document d
 - $w_{d,t}$ is the weight of a given term t in the a document d
 - ullet N is the number of documents
 - f_t is the number of documents containing t

$$w_{d,t} = \text{TF}_t \times \text{IDF}_{d,t}$$

$$= \log_e (f_{d,t} + 1) \log_e \left(\frac{N}{f_t}\right)$$
(2)

This would require multiplying each term of each row of the TF_t matrix by the corresponding vector element of $\mathrm{IDF}_{d,t}$, this can be implemented by taking the matrix product of a diagonalised matrix, this is shown in listing 25.

Listing 25: Apply TF-IDF Weigting

There is however a function built in to the tm package that will weight term document matrices and this will instead be implented to analyse the data because it will produce more maintainable code.

Apply Weighting with Built in Function

In order to create a term document matrix (and its transpose) with TF-IDF weighted values, the weighting argument may be specified as weightTfIdf by passing an appropriate list to the control argument of the TermDocumentMatrix, as shown in listing 26

Listing 26: Create a Document Term Matrix by transforming a Term Document Matrix

Remove Empty Documents

Empty Documents may be removed from the matrix by a logical test as shown in listing 27^{-3} this provides that 328 documents were empty following the processing. A summary of the first rows and columns of this matrix, following the removal of empty documents, is provided in table 5 of the appendix.

³It is important not to filter based the logic of an empty vector, because otherwise an empty vector will returned, hence the if statement in listing 27.

```
null = which(rowSums(as.matrix(tweet_weighted_dtm)) == 0)
rowSums(as.matrix(tweet_weighted_dtm)==0)

if(length(null)!=0){
   tweet_weighted_dtm = tweet_weighted_dtm[-null,]
}

length(null)
## [1] 328
```

Listing 27: Load the Packages for R

8.2.14 How many Clusters are there

Use Cosine Distance

In order to consider clustering, it can be more effective to consider the distance between the weighted documents in terms of cosine distance, the cosine distance can be calculated from the euclidean distance using the identity shown in (11), and this can be performed in R by taking the matrix product of a diagonalised matrix as shown in listing 28.

$$dist(\mathbf{X}, \mathbf{Y}) = ||\mathbf{X} - \mathbf{Y}|| \tag{3}$$

$$=\sqrt{\sum_{i=1}^{n}\left[\left(x_{i}-y_{i}\right)^{2}\right]}\tag{4}$$

$$\operatorname{dist}(\mathbf{X}, \mathbf{Y})^2 = \sum_{i=1}^n \left[(x_i - y_i)^2 \right]$$
 (5)

$$= \sum_{i=1}^{n} (x^{2}) + \sum_{i=1}^{n} (y_{i}^{2}) + 2 \sum_{i=1}^{n} (x_{i} y_{i})$$
 (6)

$$=1+1+2\times\frac{\sum_{i=1}^{n}(x_{i}y_{i})}{(1)}\tag{7}$$

$$=2+2\times\frac{\sum_{i=1}^{n}(x_{i}y_{i})}{||\mathbf{X}||\times||\mathbf{Y}||}\tag{8}$$

$$=2+2\cos\left(\mathbf{X},\mathbf{Y}\right)\tag{9}$$

(10)

$$\implies (1 - \cos(\mathbf{X}, \mathbf{Y})) = \frac{\operatorname{dist}(\mathbf{X}, \mathbf{Y})}{2} \tag{11}$$

Listing 28: Load the Packages for *R*

Project into Euclidean Space

The cosine distance however is not appropriate to perform clustering on and so instead should be projected back into euclidean space, this can be acheived using *Multi-Dimensional Scaling* via the cmdscale functionas shown in listing 29. The distance is a measure of between document distance so the number of dimensions should correspond to the number of documents, however, if there are zero-value eigenvalues, these dimensions won't help explain the data in the projection, hence the number of eigenvalues has been used as the dimension of projection in this case.

Listing 29: Load the Packages for **R**

Measure Within Cluster Variance

In order to determine the appropriate number of clusters, the within cluster variance can be measurd, the number of clusters at which this value ceases to decrease is indicative of a potentially appropriate number of clusters. This is implemented in listing 30 and shown in figure 2.

Figure 2 Indicates a sudden stop of decrease in variance at 7 clusters and following that the within cluster variance begins to decrease at a slightly slower rate. For this reason 7 could be an appropriate candidate for the number of clusters, however the minimal amount of change in the within-cluster variance indicates that the data is most likely not clustered at all.

8.2.15 Find the Number of tweets in each cluster

Moving forward we'll use 3 clusters, 7 is too large and a smaller number will likely be more effective at categorising the data (particularly given that the stratification of the data appears to be quite limited from figure 2). The number of tweets in a cluster may be measured by using the table function as shown in listing 31 and table 2.

```
set.seed(271)
   n = 15 # Assume it bends at 7 clusters
   SSW = rep(0, n)
   for (a in 1:n) {
     K = kmeans(mds.tweet_weighted_dtm, a, nstart = 20)
     SSW[a] = K$tot.withinss
     paste(a*100/n, "%") %>% print()
   }
   SSW
11
   SSW_tb <- tibble::enframe(SSW)</pre>
12
   ggplot(SSW_tb, aes(x = name, y = value)) +
13
     geom_point(col = "#Cd5b45", size = 5) +
14
     geom_line(col = "#Da70d6") +
15
     geom_vline(xintercept = 7, lty = 3, col = "blue") +
     theme_bw() +
17
     labs(x = "Number of Clusters",
          y = "Within Cluster Sum of Square Distance",
19
          title = "Within Cluster Variance across Clusters")
20
```

Listing 30: Use a loop to evaluate the performace of various cluster models, plot this with ggplot2

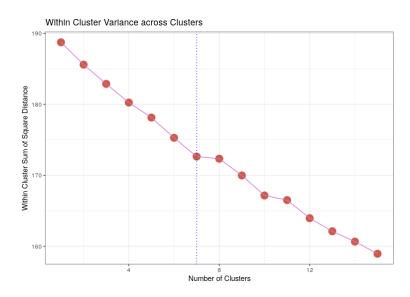


Figure 2: Plot of the Within Cluster Variance of the tweets using cosine distance (projected into Euclidean Space)

Table 2: Number of tweets in each cluster identified by k means clustering

```
1 K = kmeans(mds.tweet_weighted_dtm, 3, nstart = 20)
2 table(K$cluster)
```

Listing 31: The table function can count the number of tweets per cluster.

8.2.16 Visualise the Clusters

The clusters can be projected into 2D *Euclidean-Space* using *Multi-Dimensional Scaling*, these dimensions would represent the first two principle components of the cosine distance of the weighted tweets. This is demonstrated in listings 32. A plot of the friend countstaus mapped to shape and clusters mapped to colour is demonstrated in listing 34 shown in 4, In order to get a better understanding of the distribution of friend counts a plot of the friend count status mapped to colour is demonstrated in listing 33 and shown in figure 4.

Listing 32: Use Multi-Dimensional scaling to project the data into 2 dimensions

Listing 33: Create a plot of the distribution of Friends among Clusters as shown in figure 3

Listing 34: Create a plot of the distribution of Friends Count Status as shown in figure 4

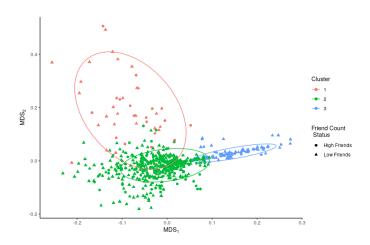


Figure 3: Distribution of Friend Count Status over Clusters

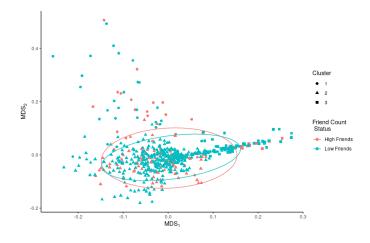


Figure 4: Distribution of Friend Count Status

Appendix

Users with High Friend Count

Table 3: User ID and Friend Count of users with above highest friend count in sample

User ID	Friend Count		
274488119	8752		
743771665	5002		
1036014247	4999		
2281452613	4992		
1554453560	4958		
981233818408570880	4944		
931765564388921344	4836		
807405140	4710		
1112579152970842112	4514		
2441577446	4322		
552692862	4229		
956297007127252992	3976		
22493896	3675		
255922782	3500		
1067409881332936709	3312		
27998570	3210		
715118521555017728	3099		
2356170174	2885		
2372688230	2880		
1868357425	2719		

Users with Low Friend Count

TF-IDF Matrix

Relevant XKCD

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Table 4: User ID and Friend Count of users with above highest friend count in sample

User ID Friend Count

User ID	Friend Count
1254280995592966145	0
875126772978913280	0
1254256124217319425	0
1250219450210480128	0
1214921087328411648	0
1254115699628421120	0
1217600080376520704	0
1253480062453600257	0
1254178435502571521	0
1251955545092718592	0
1106864828700712960	0
1160744587620524032	0
1254256536710504448	1
1129040408384868352	1
1254121201871589376	1
1248687797755658243	2
1210265263867932675	2
3380784928	3
1177274165239275520	3
54645521	3

Table 5: Document Term Matrix of first 6 documents and first 6 Words check coronavirus death first gone hour

check	coronavirus	death	first	gone	hour
0.615	0.747	0.830	0.596	0.747	0.698
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0
0	0	0	0	0	0

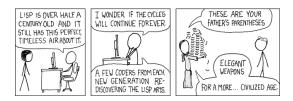


Figure 5: *xkcd* # 297

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