

# Package ‘MMGS’

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**Type** Package

**Title** Multi-Methods for Multi-Envs Genomic Selection Pipeline

**Version** 1.0.0

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**Description** Package for Multi-  
envs Genomic Selection, contains Inbred Lines, CUBIC Lines and Light-cross Lines.

**License** GPL-3

**Encoding** UTF-8

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.2.3

**Imports** BGLR, e1071, randomForest, corrgram, rrBLUP, lemon, glmnet,  
lightgbm, tidyr, stringi, reshape2, grDevices, dplyr, stats,  
colorspace

**Depends** ggplot2, R (>= 2.10)

**LazyData** true

**Suggests** knitr, rmarkdown, testthat (>= 3.0.0)

**Config/testthat/edition** 3

**NeedsCompilation** no

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---

envMeanPara

*envMeanPara*


---

## Description

Calculate environmental mean parameters for a specific trait within a specified time range. This time range is calculated by the Exhaustive\_search.

## Usage

```
envMeanPara(data, env_paras,
  maxR_dap1 = NULL, maxR_dap2 = NULL,
  Paras = NULL)
```

## Arguments

data	Data frame containing trait mean values within different environments.
env_paras	Data frame containing environmental parameter information.
maxR_dap1	The starting day for calculating environmental mean parameters. Default is 18, which is used for the test data.
maxR_dap2	The ending day for calculating environmental mean parameters. Default is 43, which is used for the test data.
Paras	Vector containing the names of environmental parameters to be calculated.

## Value

A data frame containing trait mean values and calculated environmental mean parameters.

## Examples

```
## Not run:
envMeanPara<-envMeanPara(data=env_trait,env_paras=PTT_PTR)

## End(Not run)
```

---

```
envMeanPara_plotter
      envMeanPara_plotter
```

---

## Description

Generate a plot of environmental parameter means against the overall mean.

## Usage

```
envMeanPara_plotter(
  data,
  size = NULL,
  shape = NULL,
  method = NULL,
  Paras = NULL,
  env_code = NULL,
  linewidth = NULL,
  alpha = NULL,
  linetype = NULL,
  linecolor = NULL
)
```

## Arguments

data	An environmental parameter mean data frame.
size	Point size for scatter plot.
shape	Point shape for scatter plot.
method	Regression method for the smoothing line.
Paras	Vector containing the names of environmental parameters to be considered.
env_code	Point color for the scatter plot by env_code.Default is env_code.
linewidth	Line width for the smoothing line.
alpha	Alpha value for points in the scatter plot.
linetype	Line type for the smoothing line.
linecolor	Line color for the smoothing line.

## Value

A plotter displaying the relationship between environmental parameter means and the overall mean.

## Examples

```
## Not run: envMeanPara_plotter(envMeanPara,
Paras=c('DL', 'GDD', 'PTT', 'PTR', 'PTS'))
## End(Not run)
```

---

env\_info

*env\_info Dataset*


---

### Description

This dataset contains information about environmental factors for various environments.

### Usage

```
data(env_info)
```

### Format

A data frame with the following columns:

**env\_code** Environmental code assigned to each environment.

**env\_notes** Environmental note assigned to each environment.

**lat** Latitude of the environment.

**lon** Longitude of the environment.

**Location** Location of the environment.

**PlantingDate** Planting Date of lines.

**TrialYear** Planting Year of lines.

### Source

This dataset originates from the article and provides essential information for environmental analysis.

### Examples

```
data(env_info)
```

---

env\_trait\_calculate

*env\_trait\_calculate*


---

### Description

Calculate the mean and optional quantiles for a trait across different environments.

### Usage

```
env_trait_calculate(data, trait, env, q25_75 = TRUE)
```

**Arguments**

<code>data</code>	Input data frame containing trait values and environmental information.
<code>trait</code>	The name of the trait for which mean and optional quantiles will be calculated.
<code>env</code>	The column name in the data frame representing different environments.
<code>q25_75</code>	Logical, indicating whether to calculate the 25th and 75th quantiles. Default is TRUE

**Value**

Data frame containing the mean trait values for each environment. If `q25_75` is TRUE, it also includes columns for the 25th(`q25`), 75th(`q75`) quantiles, as well as the number of observations (`n_obs`).

**Examples**

```
env_trait<-env_trait_calculate(data=trait,trait="FTgdd",env="env_code")

# Access the result, e.g., mean trait values
env_trait$mean

# Access the result, e.g., 25th quantile values
env_trait$q25

# Access the result, e.g., 75th quantile values
env_trait$q75

# Access the result, e.g., number of observations
env_trait$n_obs
```

---

etl\_plotter

etl\_plotter

---

**Description**

Generate a plot of environmental trait lines with mean, q25, and q75 areas.

**Usage**

```
etl_plotter(
  data,
  trait,
  env_cols = NULL,
  shape = NULL,
  size = NULL,
  linewidth = NULL,
  area_color = NULL,
  area_alpha = NULL
)
```

**Arguments**

<code>data</code>	An etl input data frame.
<code>trait</code>	An env_trait data frame containing mean, q25, and q75 values.
<code>env_cols</code>	Environmental colors gradient; if NULL, it generates colors using rainbow_hcl.
<code>shape</code>	Point shape, similar to ggplot2.
<code>size</code>	Point size, similar to ggplot2.
<code>linewidth</code>	Line width, similar to ggplot2.
<code>area_color</code>	Fill color for the area between q25 and q75.
<code>area_alpha</code>	Fill color alpha for the area between q25 and q75.

**Value**

A plot of etl, filling the area between env-trait q25 to q75, with a red line representing the env-trait mean.

**Examples**

```
env_trait<-env_trait_calculate(data=trait,trait="FTgdd",env="env_code")
LbyE<-LbyE_calculate(trait,trait="FTgdd",env="env_code",line="line_code")
etl<-LbyE_Reshape(data=env_trait,LbyE=LbyE,env="env_code")
#Plot
etl_plotter(data=etl,trait=env_trait)
```

---

Exhaustive\_plotter *Exhaustive\_plotter*

---

**Description**

Generate a combined plot for pairwise correlations using pop\_cor data.

**Usage**

```
Exhaustive_plotter(Correlation,
  dap_x, dap_y, p = NULL,
  Paras, Cor = NULL)
```

**Arguments**

<code>Correlation</code>	A pop_cor data frame containing correlation values for different pairs.
<code>dap_x</code>	Specifies the column for the x-axis variable.
<code>dap_y</code>	Specifies the column for the y-axis variable.
<code>p</code>	A parameter with a default value of 1.
<code>Paras</code>	Vector containing the names of environmental parameters to be considered.
<code>Cor</code>	Assigning variable column names.Default is Cor.

**Value**

A combined pop\_cor plot.

**Examples**

```
## Not run: Exhaustive_plotter(Correlation,dap_x,dap_y,Paras)
```

---

Exhaustive\_search    *Exhaustive\_search*

---

**Description**

Perform exhaustive search for population correlation matrix based on environmental parameters.

**Usage**

```
Exhaustive_search(
  data,
  env_paras,
  searching_daps,
  p = NULL,
  dap_x = NULL,
  dap_y = NULL,
  LOO = NULL,
  Paras = NULL
)
```

**Arguments**

data	Data frame containing environmental trait data.
env_paras	Data frame containing environmental parameter information.
searching_daps	The total number of days to search for, typically set based on your data. Default is 122.
p	The parameter for controlling the shape of the search window. Default is 1.
dap_x	The number of days for the x-axis of the search window. Default is the same as searching_daps.
dap_y	The number of days for the y-axis of the search window. Default is the same as searching_daps.
LOO	Leave-One-Out cross-validation flag. If LOO is 1,
Paras	Vector containing the names of environmental parameters to be considered.

**Value**

A population correlation matrix based on the exhaustive search.

**Examples**

```
#Input from function : env_trait_calculate
env_trait<-env_trait_calculate(data=trait,trait="FTgdd",env="env_code")
#Run
## Not run: Exhaustive_search(denv_trait,PTT_PTR)
```

---

geno	<i>geno Dataset</i>
------	---------------------

---

**Description**

This dataset contains genetic information for various lines.

**Usage**

```
data(geno)
```

**Format**

An object of class `data.frame` with 237 rows and 1463 columns.

**Source**

This dataset originates from the article and provides genetic data for analysis.

**Examples**

```
data(geno)
```

---

LbyE_calculate	<i>LbyE_calculate</i>
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---

**Description**

Calculate the lines in all environments.

**Usage**

```
LbyE_calculate(data, trait, env, line)
```

**Arguments**

<code>data</code>	Input data frame containing trait values, line information, and environmental information.
<code>trait</code>	The trait for which the lines within different environments will be calculated.
<code>env</code>	Column name representing different environments in the input data frame.
<code>line</code>	Column name representing different lines in the input data frame.



**Value**

A data frame containing line values within different environments for the specified trait.

**Examples**

```
# Calculate lines within different environments for the trait "FTgdd"
LbyE_calculate(trait, "FTgdd", "env_code", "line_code")
```

---

LbyE_corrplot	<i>LbyE_corrplot</i>
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---

**Description**

Plot the correlation between lines and environments.

**Usage**

```
LbyE_corrplot(LbyE, cor_type = NULL, color = NULL)
```

**Arguments**

LbyE	A data frame calculated by LbyE_calculate. Column names represent Lines, and row names represent Environments.
cor_type	Graphic format of the correlation map. The default format is "heatmap". Other option is "pie".
color	Set the color gradient for the correlation map. It is a vector of three colors representing low, mid, and high values.

**Value**

A correlation plot between lines and environments in the specified format.

**Examples**

```
LbyE<-LbyE_calculate(trait, "FTgdd", "env_code", "line_code")
# Generate a heatmap correlation plot for Lines and Environments
colors=c("blue", "white", "red")
LbyE_corrplot(LbyE, cor_type="heatmap", color=colors)
# Generate a pie chart correlation plot for Lines and Environments
LbyE_corrplot(LbyE, cor_type="pie", color=colors)
```

---

LbyE_Reshape	<i>LbyE_Reshape</i>
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---

**Description**

Reshape trait values within specified environments and lines.

**Usage**

```
LbyE_Reshape(data, LbyE, env)
```

**Arguments**

data	Data frame containing environmental information from env_trait_calculate.
LbyE	A data frame calculated by LbyE_calculate. Column names represent Lines, and row names represent Environments.
env	Column name representing different environments in the data frame.

**Value**

Data frame containing trait values within specified environments and lines.

**Examples**

```
env_trait<-env_trait_calculate(trait, "FTgdd", "env_code")
LbyE<-LbyE_calculate(trait, "FTgdd", "env_code", "line_code")
#Run:
etl<-LbyE_Reshape(env_trait, LbyE, "env_code")
```

---

line_trait_mean	<i>line_trait_mean</i>
-----------------	------------------------

---

**Description**

Calculate the mean of residuals for each line after fitting a linear model.

**Usage**

```
line_trait_mean(data, LbyE, mean, trait, row)
```

**Arguments**

data	Input data frame containing trait values, line information, and environmental information.
LbyE	Data frame calculated by LbyE_calculate, containing line values within different environments.
mean	Data frame containing mean trait values within different environments. This data frame get from the function of env_mean_calculate.
trait	The trait for which the residuals will be calculated.
row	The minimum number of rows required for fitting a linear model for each line.

**Value**

A list containing two elements:

1. `data`: Data frame with columns `env_code`, `errors` (mean of residuals), and additional columns from `env_trait`.
2. `lm_residuals`: Data frame containing the residuals squared for each line and environment.

**Examples**

```
LbyE<-LbyE_calculate(trait,trait="FTgdd",env="env_code",line="line_code")
env_trait<-env_trait_calculate(data=trait,trait="FTgdd",env="env_code")
#Run
result<-line_trait_mean(data=trait,LbyE=LbyE,
                        mean = env_trait,trait="FTgdd",row=2)
print(result[[1]])
```

---

Mean_trait_plot	<i>Mean_trait_plot</i>
-----------------	------------------------

---

**Description**

Generate a plot displaying the relationship between the mean trait values and the environmental parameter mean.

**Usage**

```
Mean_trait_plot(
  Reg,
  MSE,
  point_size = NULL,
  color = NULL,
  point_shape = NULL,
  linewidth = NULL,
  linetype = NULL
)
```

**Arguments**

<code>Reg</code>	Regression result data frame.
<code>MSE</code>	Mean Squared Error (MSE) result data frame.
<code>point_size</code>	Size of points in the plot.
<code>color</code>	color for the points and the dash area.
<code>point_shape</code>	Shape of points in the plot.
<code>linewidth</code>	Line width for the smoothing line.
<code>linetype</code>	Line type for the smoothing line.

**Value**

A plot showing the relationship between mean trait values and the environmental parameter mean, with MSE information.

**Examples**

```
## Not run: Mean_trait_plot(Reg, MSE)
```

---

MMGP

---

*Cross Validation and Genomic Prediction Cross environments*


---

**Description**

MMGP (Multi-environment Multi-Genomic Prediction) is a function for genomic prediction that incorporates cross-validation and cross-environment prediction. It utilizes both genetic and environmental information to predict phenotypic values. The function supports various genomic prediction methods and models, allowing users to choose the most suitable approach based on data characteristics and preferences.

**Usage**

```
MMGP (
  pheno,
  geno,
  env,
  para,
  Para_Name,
  model,
  depend = NULL,
  fold = NULL,
  reshuffle = NULL,
  methods = NULL,
  ENalpha = NULL,
  SVM_cost = NULL,
  gamma = NULL,
  kernel = NULL,
  nIter = NULL,
  burnIn = NULL,
  thin = NULL,
  GBM_params = NULL,
  GBM_rounds = NULL,
  Envs = NULL,
  Obs = NULL,
  line_code = NULL,
  ms1 = NULL,
  ms2 = NULL
)
```

**Arguments**

pheno	Vector (n x j) of "phenotypes," i.e., observations or pre-processed, corrected values.
geno	Matrix (n x m) of genotypes for the training population: n lines with m markers. Genotypes should be coded -1, 0, 1. Missing data are not allowed.
env	Data.frame (j x l) of "environmental information," i.e., environmental conditions corresponding to phenotypes.
para	Data.frame returned from envMeanPara function, providing information about environmental means and parameters.
Para_Name	The most relevant environmental covariates to the subject's phenotype, obtained after stepwise correlation calculations.
model	The options for genomic breeding cross validation methods.
depend	The options for genomic breeding within different environment. The available options are: 1.Norm(Reaction Norm Model) 2.PEI(Polygenic Environment Interaction Model)
fold	Number of folds for cross-validation. Smallest recommended value is fold = 2.
reshuffle	Number of independent replicates for cross-validation. Smallest recommended value is reshuffle = 5.
methods	A vector specifying the prediction methods to be used.
ENalpha	Used for Elastic Net prediction model, with a value range from 0 to 1.
SVM_cost	Cost of constraints violation for SVM (default: 10).
gamma	Parameter needed for all kernels except linear (default: 0.001).
kernel	the kernel used in training and predicting. You might consider changing some of the following parameters, depending on the kernel type. linear, polynomial, radial basis, sigmoid.
nIter	Number of iterations for the Bayesian model.
burnIn	Number of burn-in iterations for the Bayesian model.
thin	Thinning parameter for the Bayesian model.
GBM_params	A list of parameters for LightGBM. See LightGBM documentation for details.
GBM_rounds	Number of training rounds for LightGBM.
Envs	Assign variable column names of Envs. Default is Envs.
Obs	Assign variable names of Observed values. Default is Obs.
line_code	Assign variable column names of line_code. Default is line_code.
ms1	Remove the line for which the number of missing environments > ms1.
ms2	Remove the line for which the number of missing environments > ms2.

**Value**

The class MMGP returns a list containing: The Observed and Predicted Values of each line in Multiple Environments. The Predictive accuracy in Multiple Environments. Calculated by Pearson Cor. The Predictive accuracy for All Environments. (Cross R2)

## Examples

```
## Not run:
out<-MMGP(pheno=pheno,geno=geno,env=env_info,
          para=envMeanPara,Para_Name="PTT",
          depend="Norm",fold=2,reshuffle=5,methods="RM.G",
          ms1=ms1,ms2=ms2)

## End(Not run)
```

---

MMPrdM

*MMPrdM*


---

## Description

MMPrdM (Multi-environment Multi-marker Prediction Model) is a function for genomic prediction that includes cross-environment prediction. It leverages genetic and environmental information to predict phenotype. The function supports various genomic prediction methods and models, providing flexibility in choosing the most suitable approach based on the data characteristics and user preferences.

## Usage

```
MMPrdM(
  pheno,
  geno,
  env,
  para,
  Para_Name,
  model,
  depend = NULL,
  reshuffle = NULL,
  methods = NULL,
  ENalpha = NULL,
  SVM_cost = NULL,
  gamma = NULL,
  kernel = NULL,
  Envs = NULL,
  line_code = NULL,
  nIter = NULL,
  burnIn = NULL,
  thin = NULL,
  GBM_params = NULL,
  GBM_rounds = NULL
)
```

## Arguments

pheno	Vector (n x j) of "phenotypes", i.e. observations or pre-processed, corrected values.
geno	Matrix (n x m) of genotypes for the training population: n lines with m markers. Genotypes should be coded -1, 0, 1. Missing data are not allowed.

env	Data.frame (j x l) of "environmental information", i.e.
para	Data.frame returned from envMeanPara function.
Para_Name	The most relevant environmental silvers to the subject's phenotype, obtained after stepwise correlation calculations, are referred to for more details:
model	The options for genomic breeding cross vadiation methods. The available options are: 1.GBLUP: performs G-BLUP using a marker-based relationship matrix, implemented through BGLR R-library. Equivalent to ridge regression (RR-BLUP) of marker effects. 2.RR: ridge regression, using package glmnet. In theory, strictly equivalent to gblup. 3.LASSO: Least Absolute Shrinkage and Selection Operator is another penalized regression methods which yield more shrinked estimates than RR.Run by glmnet library. 4.EN: Elastic Net (Zou and Hastie, 2005), which is a weighted combination of RR and LASSO, using glmnet library 5.rrBLUP:the RR-BLUP mixed model(Endelman ,2011). One application is to estimate marker effects by ridge regression; alternatively, BLUPs can be calculated based on an additive relationship matrix or a Gaussian kernel. Several Bayesian methods, using the BGLR library: 1.BRR: Bayesian ridge regression: same as rr-blup, but bayesian resolution. Induces homogeneous shrinkage of all markers effects towards zero with Gaussian distribution (de los Campos et al, 2013) 2.BL: Bayesian LASSO: uses an exponential prior on marker variances priors, leading to double exponential distribution of marker effects (Park & Casella 2008) 3.BA: uses a scaled-t prior distribution of marker effects. (Meuwissen et al 2001). 4.BB: Bayes B, uses a mixture of distribution with a point mass at zero and with a slab of non-zero marker effects with a scaled-t distribution (Habier et al 2011). 5.BC: Bayes C same as Bayes B with a slab with Gaussian distribution. A more detailed description of these methods can be found in Perez & de los Campos 2014 ( <a href="http://genomics.cimmyt.org/BGLR-extdoc.pdf">http://genomics.cimmyt.org/BGLR-extdoc.pdf</a> ). Four semi-parametric methods: 1.RKHS: reproductive kernel Hilbert space and multiple kernel MRKHS, using BGLR (Gianola and van Kaam 2008). Based on genetic distance and a kernel function to regulate the distribution of marker effects. This methods is claimed to be effective for detecting non additive effects. 2.RF: Random forest regression, using randomForest library (Breiman, 2001, Breiman and Cutler 2013). This methods uses regression models on tree nodes which are rooted in bootstrapping data. Supposed to be able to capture interactions between markers 3.SVM: support vector machine, run by e1071 library. For details, see Chang, Chih-Chung and Lin, Chih-Jen: LIBSVM: a library for Support Vector Machines <a href="http://www.csie.ntu.edu.tw/~cjlin/libsvm">http://www.csie.ntu.edu.tw/~cjlin/libsvm</a> 4.LightGBM: Light Gradient Boosting Machine, run by LightGBM library developed by Microsoft. For details, see Jun Yan, Yuetong Xu, at al: LightGBM: accelerated genomically designed crop breeding through ensemble learning
depend	The options for genomic breeding within different environment.The available options are: 1.Reaction Norm: 2.PEI:
reshuffle	Number of independent replicates for the Predict. Smallest value recommended is reshuffle = 3.
methods	RM.G RM.GE RM.E
ENalpha	used for EN predict model,the value range from 0 to 1.
SVM_cost	Cost of constraints violation for SVM (default: 10).
gamma	Parameter needed for all kernels except linear (default: 0.001).
kernel	the kernel used in training and predicting. You might consider changing some of the following parameters, depending on the kernel type. linear, polynomial, radial basis, sigmoid.

Envs	Assign variable column names of Envs. Default is Envs.
line_code	Assign variable column names of line_code Default is line_Code.
nIter	Number of iterations for the Bayesian model.
burnIn	Number of burn-in for the Bayesian model.
thin	Number of thins for the Bayesian model.
GBM_params	A list of parameters for LightGBM. See LightGBM documentation for details.
GBM_rounds	Number of training rounds for LightGBM.

**Value**

The class MMPrdM returns a list containing: The Predicted Values of each line in Multiple Environments.

**Examples**

```
## Not run:
out<-MMPrdM(pheno=pheno,geno=geno,env=env_info,para=envMeanPara)

## End(Not run)
```

---

mse_plotter	<i>mse_plotter</i>
-------------	--------------------

---

**Description**

Generate a plot for Mean Squared Errors (MSE) data analyzed by line\_trait\_mean.

**Usage**

```
mse_plotter(
  data,
  point_size = NULL,
  text_size = NULL,
  point_shape = NULL)
```

**Arguments**

data	Dataframe containing MSE data analyzed by line_trait_mean.
point_size	Size of points in the plot.
text_size	Size of text labels in the plot.
point_shape	Shape of the points in the plot.

**Value**

A plot illustrating Mean Squared Errors (MSE) data.

**Examples**

```
## Not run: mse_plotter(MSE)
```



---

PTT\_PTR*PTT\_PTR Dataset*

---

**Description**

Dataset contains information about environmental parameters related to PTT (Photothermal Time) and PTR (Photothermal Ratio) and others.

**Usage**

```
data (PTT_PTR)
```

**Format**

A data frame representing environmental parameters.

**Date** Date of the environmental information.

**env\_code** Environmental code assigned to each environment.

**PTS** Environmental parameter related to Photothermal spectroscopy.

**PTT** Environmental parameter related to Photothermal Time.

**PTR** Environmental parameter related to Photothermal Ratio.

**GDD** Environmental parameter related to Growing Degree Days.

**Tmax** Environmental parameter related to the maximum temperature.

**Tmin** Environmental parameter related to the minimum temperature.

**DL** Environmental parameter related to Day Length.

**Source**

This dataset originates from the article and provides environmental parameters for analysis.

**Examples**

```
data (PTT_PTR)
```

---

Reg*Regression*

---

**Description**

Calculate regression results between traits and environmental means for each line.

**Usage**

```
Reg(LbyE, env_trait, filter_num = NULL)
```

**Arguments**

LbyE	Data frame containing lines and their corresponding trait values within environments. It is from the function of LbyE_calculate.
env_trait	Data frame containing environmental means for different traits within environments. It is from the function of envV_mean_calculate.
filter_num	Minimum number of observations required to perform regression. Default is 4.

**Value**

A data frame containing regression results for each line.

**Examples**

```
#Get Input
LbyE<-LbyE_calculate(trait,trait="FTgdd",env="env_code",line="line_code")
env_trait<-env_trait_calculate(data=trait,trait="FTgdd",env="env_code")
#Calculate
Regression<-Reg(LbyE=LbyE,env_trait=env_trait)
print(head(Regression))
```

Reg\_plotter

*Reg\_plotter***Description**

Generate a regression plot between the mean and trait, showing the relationship between these two variables.

**Usage**

```
Reg_plotter(
  Reg = Reg,
  size = NULL,
  method = NULL,
  color = NULL,
  alpha = NULL)
```

**Arguments**

Reg	Input data frame containing regression results. param x Variable to be plotted on the x-axis. param y Variable to be plotted on the y-axis. param group Variable used for grouping the data.
size	Size of points in the plot.
method	Smoothing method for the regression line.
color	Vector containing colors for regression line and points.
alpha	Vector containing alpha values for regression line and points.

**Value**

A plot for the regression relationship between mean and trait.

### Examples

```
#' #Get Input
LbyE<-LbyE_calculate(trait,"FTgdd","env_code","line_code")
env_trait<-env_trait_calculate(trait,"FTgdd","env_code")
Regression<-Reg(LbyE = LbyE, env_trait = env_trait)
Reg_plotter(Reg = Regression)
```

---

trait	<i>trait Dataset</i>
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### Description

This dataset contains information about phenotypic traits for various lines and environments.

### Usage

```
data(trait)
```

### Format

A data frame representing trait parameters.

**line\_code** Line code assigned to each line.

**env\_code** Environmental code assigned to each environment.

**env\_note** Environmental note assigned to each environment.

**FTdap** Name of the phenotypic trait measured, related to FT.

**FTgdd** Name of the phenotypic trait measured, related to FT.

**pop\_code** Other columns present additional trait-related information

### Source

This dataset originates from the article and provides trait data for analysis.

### Examples

```
data(trait)
```