

Jupyter Notebook Tutorial

1. INTRODUCTION

This tutorial explains the basic data reduction process for the SOAR Adaptive-Module Optical Spectrograph (SAMOS). The accompanying jupyter notebook is aptly named ‘`jupyter_SAMOS`’.

2. STARTING UP

As of now, the pipeline uses PyRAF for spectral line identification, which requires Python2. The environment file `EnvSAMOS.yml` is located in the main working directory. The user may copy the environment to their own computer by executing ‘`conda env create --file EnvSAMOS.yml`’ in their terminal.

The user’s working directory should contain the following:

- `SAMOS-working-dir/:`
 - `cleaner.sh`
 - `Readme.md`
 - `Docs/`
 - `jupyterNB_tutorial/` **you are here**
 - `LDSS3/`
 - `helper_files/`
 - `linelists/`
 - `py_mods/`
- `SAMOS-working-dir/py_mods/:`

- `CreateFuelStructure.py`
- `CreateInput.py`
- `CreateSlitStructure.py`
- `FlatNorm.py`
- `InitializeSAMOS.py`
- `NormDivFlats.py`
- `OutlineSlits.py`
- `Overscan.py`
- `OverscanAndTrim.py`
- `SAMOSHelpers.py`
- `SlitCutout.py`
- `SlitID.py`
- `PyrafIdentify.py`
- `PyrafTransform.py`
- `iraf_steps.py`
- `iraf_params.py`
- `__init__.py`

- **SAMOS-working-dir/helper_files:**

- `LMask1.SMF`
- `LMask2.SMF`
- `LMask1_ycoords.c1.txt`
- `LMask2_ycoords.c1.txt`
- `LMask2_ycoords_red.c1.txt`

Note that the helper files are specifically for reducing the LDSS3 test data. These files will be obsoleted when we have sufficient data from the SAMOS instrument. When that happens, I will be adjusting the pipeline according to the instrument data. It should therefore be noted that parts of the pipeline that call these helper files employ less robust data reduction methods.

Eventually, the pipeline will be used to reduce multi-object spectroscopy data taken with the SOAR Adaptive-Module Optical Spectrograph (SAMOS). For information on the instrument, see (Robberto et al. 2016). In order to avoid reinventing the wheel, this pipeline is heavily based on the Flame DRP written by Belli, Contursi, and Davies (2017). Flame is written in IDL, while the SAMOS pipeline will ultimately be written in Python2.

3. NOTEBOOK WALKTHROUGH

3.1. *initialize_SAMOS*

The first cell of notebook sets up the paths to the various modules needed to run the pipeline so the notebook can find and use them.

```
In [1]: import sys
import os
#insert path to modules: WORKING_DIR/py_mods/
base_dir = os.path.split(os.getcwd())[0]
mods_dir = base_dir + '/py_mods/'
sys.path.insert(0,mods_dir)
print sys.path[0]
from InitializeSAMOS import initialize_SAMOS
from OverscanAndTrim import overscan_and_trim
from NormDivFlats import norm_div_flats
from OutlineSlits import outline_slits
from PyrafIdentify import pyraf_identify
import PyrafTransform
#the above are the main modules and functions called by the pipeline.
```

Figure 1. Cell initialization for a new data reduction session.

First, the pipeline runs the function `initialize_SAMOS`, which the FITS headers creates the basic data structure, separating the calibration files from the science files.

The output while the script is running will give the names of files blah.

Each instance of the fuel structure returned by `initialize_SAMOS` is organized as follows:

```
In [2]: ##INITIALIZATION##

data_dir = '2017-11-30'
mask = 'LMask2'
fuel_init = initialize_SAMOS(data_dir,mask)

Scanning                                SAMOS working dir /LDSS3/2017-11-30

Found field image:
['ccd8152c1.fits', 'ccd8152c2.fits']
Found flats:
['ccd8161c1.fits', 'ccd8161c2.fits', 'ccd8162c1.fits', 'ccd8162c2.fits', 'ccd8163c1.fits', 'ccd8163c2.fits']
Found lamps:
['ccd8159c1.fits', 'ccd8159c2.fits', 'ccd8160c1.fits', 'ccd8160c2.fits']
Found science:
['ccd8157c1.fits', 'ccd8157c2.fits', 'ccd8158c1.fits', 'ccd8158c2.fits']
With exposure times:
[900. 900. 900. 900.]
Keeping science:
['ccd8157c1.fits', 'ccd8157c2.fits', 'ccd8158c1.fits', 'ccd8158c2.fits']
4 science frames read
```

Figure 2. output from running the first step of the code.

```
util: ['science', 'intermediate_dir', 'arc', 'field', 'slitflat']

util.science: ['transf_files', 'master_file', 'raw_files', 'corr_files', 'n_frames']

util.arc: ['transf_files', 'master_file', 'raw_files', 'corr_files', 'n_frames']

util.field: ['transf_files', 'master_file', 'raw_files', 'corr_files', 'n_frames']

util.slitflat: ['transf_files', 'master_file', 'raw_files', 'corr_files', 'n_frames']

input: ['field_filelist', 'arc_filelist', 'working_dir', 'mask_SMF', 'slit_position_file', 'slit_mask', 'flat_filelist', 'db_file', 'science_filelist']

slits: []

identify: ['linelist', 'lgfile', 'db', 'linelist_ext']

instrument: ['default_dark', 'default_pixelflat', 'gr_angle', 'dewar', 'linear_disp', 'd_alignrot', 'linearity_correction', 'dref', 'instrument', 'epoch', 'camera', 'scale_arc_per_mm', 'default_badpixel_mask', 'ns', 'resolution_slitlarcsec', 'central_wavelength', 'default_arc', 'pixel_scale', 'dewoff', 'default_illumflat', 'date', 'celestial_dec', 'hangle', 'temp', 'gr_order', 'mask', 'filter', 'mode', 'trim_edges', 'readnoise', 'resolution', 'grism', 'celestial_ra', 'gain']
```

Figure 3. Base data structure created upon initialization.

The pipeline gets most of the initial information from header files and creates empty files and directories for future use, but slit locations for the current test data were created manually and stored in one of the helper files. The SAMOS instrument is a Digital Micromirror Device (DMD)

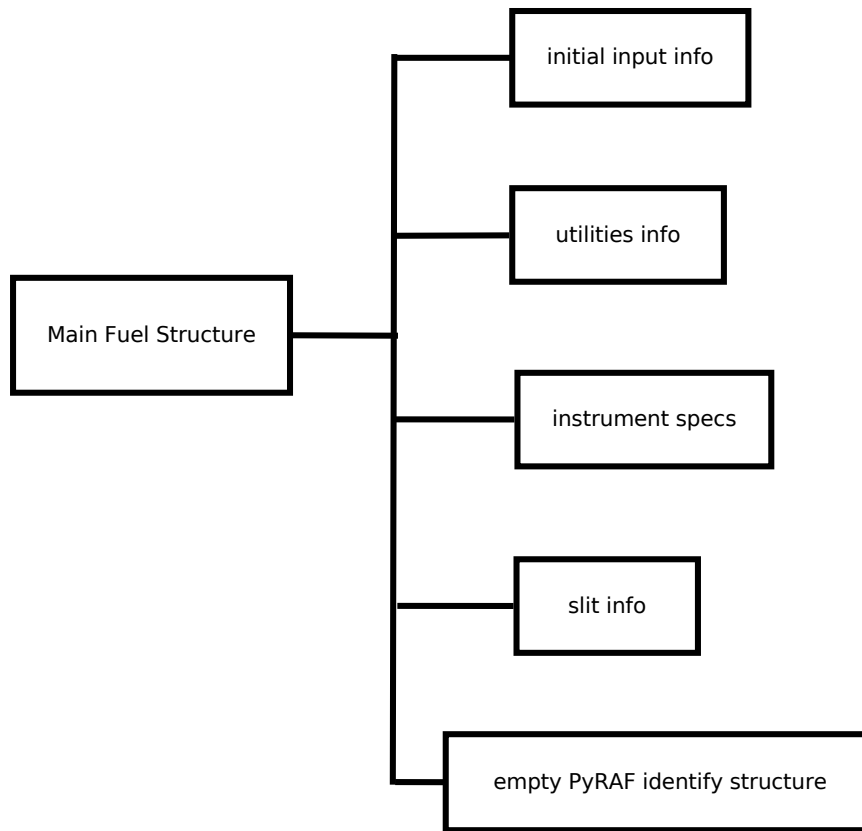


Figure 4. Base data structure flowchart.(Until I make more nice flow charts that better demonstrate the data structures, I will include screenshots of printed instance attributes when relevant.)

([Smee et al. 2018](#)), allowing for multiple slit configurations in a single observing run. Each exposure will have the slit configurations stored in the FITS headers so the pipeline will be able to access them easily and have them stored upon instantiation. For now, the slit position file is stored as `fuel_init.input.slit_position_file`.

3.2. *overscan_and_trim*

Next, we have to determine the overscan and trim the edges of the fits data. This code reads the database file and creates arrays which separate the data into flats, comparison lamps, and science data.

The main part of `overscan_and_trim` is the calling of the module `Overscan.py`. This module parses the fits data array into regions of bias and target data by reading the fits headers `BIASSEC` and `DATASEC` respectively. There is a function in `Overscan.py` called `FieldTrim(...)` that trims the field mask to the correct size.

The bias is due to a combination of the camera noise from the readout process and electric “pre-charge” on a CCD chip by the electronics. The `BIASSEC` header gives the region of overscan, which contains information for this bias. An array of this overscan region is made by grabbing the rows and columns from the data array (`d = f[0].data.astype('f')`) \rightarrow `overscan = d[biassec rows, biassec columns]`). The function then takes the median (or mean) of the overscan regions along the rows (`axis=1`), and subtracts these values from each data value in their respective data rows. The main procedure is shown in the figure below.

$$\begin{pmatrix} \text{cropped} & \text{and} \\ \text{bias} & \text{subtracted} \\ \text{data} & \text{matrix} \end{pmatrix} = \begin{pmatrix} d_{11} & d_{12} & d_{13} & \dots & d_{1m} \\ d_{21} & d_{22} & \dots & \dots & d_{2m} \\ \dots & \dots & \dots & \dots & \dots \\ d_{n1} & \dots & \dots & \dots & d_{nm} \end{pmatrix} - \begin{pmatrix} [b1] \\ [b2] \\ [b3] \\ [\dots] \\ [bn] \end{pmatrix}$$

During this task, the pipeline outputs its progress for the user.

REFERENCES

Smee, S. A., Barkhouser, R., Harding, A., et al.
 2018, Ground-based and Airborne
 Instrumentation for Astronomy VII, 107021O