# SMAP: A Pipeline for Sample Matching in Proteogenomics

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# **Contents**

1. Introduction	. 3
1.1 Software requirement	. 3
1.2 Contact information	. 3
1.3 License	. 3
2. How to run SMAP (standard alone version)	. 4
2.1 Download the pipeline	. 4
2.2 Run SMAP program	. 4
2.3 Input data	. 4
2.3.1 A variant peptide table	. 4
2.3.2 A genotype in VCF format	. 5
2.3.3 Output files	. 5
2.4 Cloud-based SMAP	. 7
2.4.1 Introduction	. 7
2.4.1 Input data	. 7
2.4.2 Data Summary & Filtering	. 8
2.4.2.1 summary of variant peptide data	. 8
2.4.3.3 Distributions and filtering	. 9
2.4.3 sample cluster	11
2.4.4 summary of Genotype data	12
2.4.5 Filtering of Genotype data	13
2.4.6 Allele frequency	15
2.4.7 Run SMAP	15
3 References	17

#### 1. Introduction

SMAP is a pipeline designed for verifying and correcting sample identity for a large mass spectrometry (MS)-based proteomics project. SMAP takes a variant peptide data that can be generated using the proteogenomics approach. The program then infers allelic information for each sample based on its expression level of the variant peptides. The program finally aligns the MS-based proteomic samples with genomic information (i.e., genotypic data) by using two discriminant scores.

#### 1.1 Software requirement

SMAP has both standard alone and cloud-based versions. The standard alone version supports all 64-bit operating systems. The program is written by a combination of Perl and R. The minimum required Perl version should be Perl 5.6 or R 3.1.0.

#### 1.2 Contact information

For any questions, please contact Xusheng Wang (xusheng.wang@und.edu)

#### 1.3 License

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#### 2. How to run SMAP (standard alone version)

#### 2.1 Download the pipeline

The pipeline could be downloaded from <a href="https://github.com/XWangLab/SMAP">https://github.com/XWangLab/SMAP</a>

#### 2.2 Run SMAP program

After installing SMAP program, you can run the program using the following command.

perl SMAP.pl -vf variant\_peptide\_table[file] -g genotype[file] -o result[file]

--variant\_peptide,-vf (A file containing quantitative values of variant peptides; required)

--genotype, -g (A genotype file used sample verification; required)

--output, -o (An output filename; required)

--plex, -p (Multiplex number of the isobaric labeling approach)

--fold\_change, -fc (Signal to Noise ratio (optional; default is 3))
--noise\_level, -nl (The upper threshold of a noise level)

--version, -h (Print version)

--help, -h (Print help)

--licence, -I (Print licencejump –s (search))

#### 2.3 Input data

#### 2.3.1 A variant peptide table

The variant peptide table uses the following format:

Column 1: Peptide ID

Column 2: Gene/Protein

Column 3: Peptide Spectrum Match (PSM)

Column 4: SNP ID \*\*MUST MATCH GENOTYPE SNP ID

Column 5-N: Sample Peptide Quantification (One column per sample)

An example of the variant peptide table

Peptide	Gene	PSM	SNP	2015- 1341	 2016- 965	Internal standard	group
VSNEEKVR	CAPZA1	b20_f39.15855.1.3	chr1:113162494:G:A	53788.04	 83146.90	46477.36	nonzero
HWQQFYFLSTR	FBXO2	b20_f36.35042.1.3	chr1:11710561:T:G	25447.82	 15590.47	19626.55	nonzero
SIEDLLR	PDE4DIP	b20_f22.28382.1.2	chr1:144877111:G:T	13161.86	 10127.43	8410.05	nonzero

## 2.3.2 A genotype in VCF format

SMAP also takes a genotype in VCF format.

#### An example of the genotype data

									2014-	2014-	2014-
#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	2194	2195	2196
1	949608	chr1:949608:G:A	G	Α	•		PR	GT	0/1	0/1	0/1
1	2441358	chr1:2441358:T:C	Т	С			PR	GT	0/0	0/0	0/0
10	115644040	chr10:115644040:G:A	G	Α				GT	0/1	0/0	0/1

#### 2.3.3 Output files

SMAP generates a final report and several intermediate results.

The final report contains four columns, including Sample ID, Inferred ID, CScore and DeltaCScore.

An example of the final report

Sample ID	Inferred ID	CSore	DeltaCScore
2015-1341	2015-1341	4.22	0.70
2015-737	2015-737	4.03	0.56
2015-804	2015-804	3.70	0.59
2015-42	2015-37	3.14	0.51
2015-1555	2015-1555	2.91	0.54
2015-244	2015-244	2.62	0.44
2015-735	2015-735	2.53	0.43
2014-2200	2015-857	2.52	0.48
2016-958	2016-958	1.39	0.03
2016-965	2016-965	1.27	0.03
Internal standard	2015-1339	1.71	0.00

In addition, the program also generates three intermediate files, including samplespecific genotypes and inferred genotypes.

# An example of sample-specific genotype

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	2014- 2194	2014- 2195	2014- 2196
1	949608	chr1:949608:G:A	G	Α			PR	GT	Н	Н	Н
1	2441358	chr1:2441358:T:C	Т	С			PR	GT	Т	Т	Т
10	115644040	chr10:115644040:G:A	G	Α				GT	С	Н	Н

# An example of inferred genotypes

SNP	2015- 1341	2015- 737	2015- 804	2015- 42	2015- 1555	2015- 244	2015- 735	2014- 2200	2016- 958	2016- 965	Internal standard
chr11:75298468:A:C	Α	С	Α	Α	Α	Α	Α	Α	Α	Α	Α
chr5:140503474:C:G	Н	Н	Н	Н	Н	С	Н	Н	С	С	С
chr19:40408821:C:G	С	С	С	С	С	Н	G	С	С	С	С

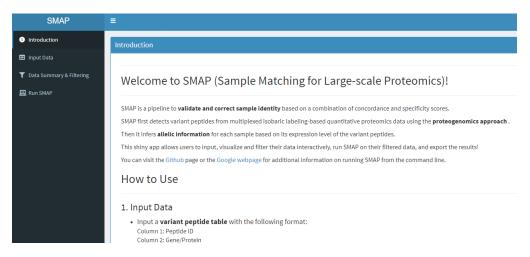
#### 2.4 Cloud-based SMAP

The cloud-based SMAP is built with R shiny. It can be found:

https://smap.shinyapps.io/smap/

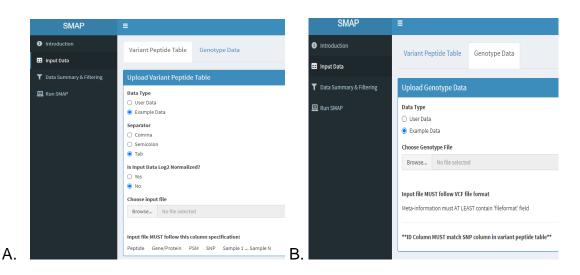
Once the website is loaded, the main page of SMAP is shown as below:

#### 2.4.1 Introduction



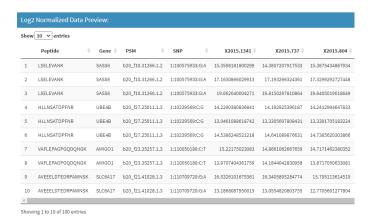
#### 2.4.1 Input data

User can upload data using "Browse" buttons in variant peptide table and genotype data menus. The format of both files can be found in the section 2.3.



For the variant peptide table, the data will be converted into log2 scale if the data is not log2 transformed.

#### Log2 transformed data (Preview)

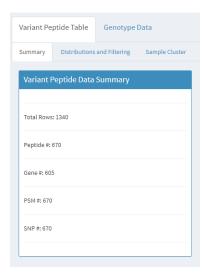


#### 2.4.2 Data Summary & Filtering

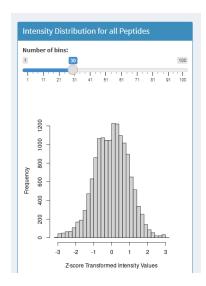
#### 2.4.2.1 summary of variant peptide data

- 1. Select Data Summary & Filter tab on left.
- Select Variant peptide table then select Summary, set the number of groups in your dataset. Default is 30, then the window will expand to show intensity distribution for all peptides, other parameters will be listed in the left, such as number of rows, number of peptides, number of genes, number of PSMs, and number of SNP.

#### Variant peptide data summary (Preview)



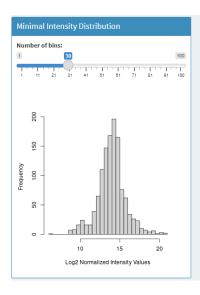
## Intensity distribution for all peptides (Preview)



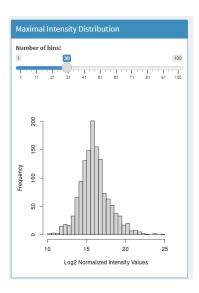
#### 2.4.3.3 Distributions and filtering

The parameters are included the minimal expression value in variant peptide (default is 30); the maximal expression value in variant peptide (default is 30); and the ration between Maximal and minimal values (default is 30).

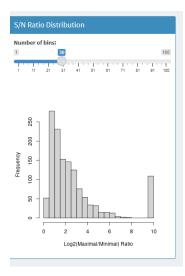
## **Minimal intensity distribution (Preview)**



## **Maximal intensity distribution (Preview)**



#### S/N ratio distribution (Preview)



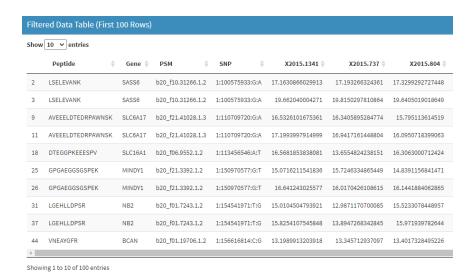
## Variant peptide data filtering

User could modify the parameters, such as the minimal expression value in variant, the maximal expression value in variant peptide; and the ration between Maximal and minimal values. Then download the filtered data.

#### Filtering parameters and summary (Preview)



#### Filtered data table (Preview)



#### 2.4.3 sample cluster

- 1. Select Data Summary & Filter tab on left.
- Select Variant peptide table then select Sample Cluster. Then the window will explore PCA plot for the test samples.

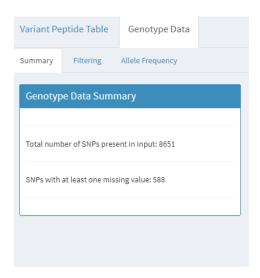
#### **PCA plot (Preview)**



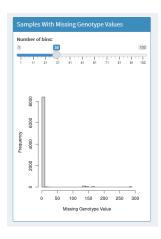
#### 2.4.4 summary of Genotype data

- 1. Select Data Summary & Filter tab on left.
- 2. Select Genotype Data then select Summary, the default parameter is 30. Then the window will explore a plot for distribution of allele frequency. The basic information will also be listed in the left, such as total number of SNP present in input, SNPs with at least one missing value; average MAF(minor allele frequency).

## **Geno type data summary (Preview)**



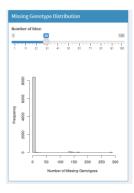
# Samples with missing genotype values (Preview)



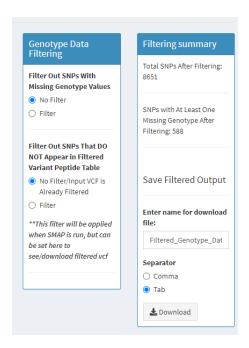
#### 2.4.5 Filtering of Genotype data

- 1. Select Data Summary & Filter tab on left.
- 2. Select Genotype Data then select Filtering, input the number of missing genotypes. Then the window will explore the filtered VCF data and the number of total SNPs after filtering and number of SNPs with at least one missing genotype after filtering.

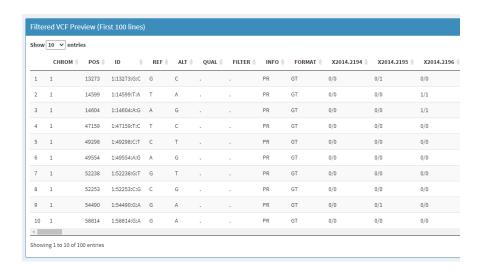
#### Missing genotype distribution (Preview)



## Genotype filtering and summary (Preview)

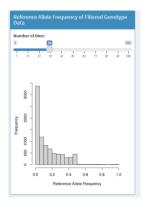


## Filtered VCF preview (Preview)

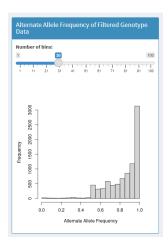


## 2.4.6 Allele frequency

#### Reference allele frequency of filtered genotype data (Preview)



#### Alternate allele frequency of filtered genotype data (Preview)

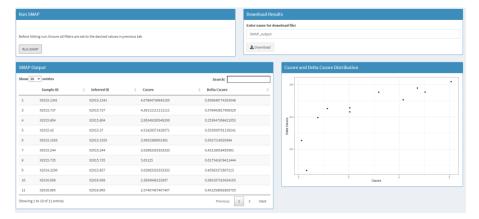


#### **2.4.7 Run SMAP**

- 1. Select Run SMAP on the left.
- 2. Note: before hitting run, ensure all filters are set to the desired values in previous tab.



After running SMAP, user could find the score table in the left and a plot in the right to distribute all of scores. User could download the score table using the tab in the upright.



#### 3. References

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- 2. Li, Y., Wang, X., Cho, J.H., Shaw, T.I., Wu, Z., Bai, B., Wang, H., Zhou, S., Beach, T.G., Wu, G.\*, et al.\* (2016). JUMPg: An Integrative Proteogenomics Pipeline Identifying Unannotated Proteins in Human Brain and Cancer Cells. J Proteome Res \*15\*, 2309-2320.
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