SMAP: A Pipeline for Sample Matching in Proteogenomics

Version 1.0.0

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Contents

1. Introduction

SMAP is a pipeline designed for verifying and correcting sample identity for a large mass spectrometry (MS)-based proteomics project. SMAP takes a variant peptide data that can be generated using the proteogenomics approach. The program then infers allelic information for each sample based on its expression level of the variant peptides. The program finally aligns the MS-based proteomic samples with genomic information (i.e., genotypic data) by using two discriminant scores.

1.1 Software requirement

SMAP has both standard alone and cloud-based versions. The standard alone version supports all 64-bit operating systems. The program is written by a combination of Perl and R. The minimum required Perl version should be Perl 5.6 or R 3.1.0.

1.2 Contact information

For any questions, please contact Xusheng Wang (xusheng.wang@und.edu)

1.3 License

2. How to run SMAP (standard alone version)

2.1 Download the pipeline

The pipeline could be downloaded from *https://github.com/UND-Wanglab/SMAP*

2.2 Run SMAP program

After installing SMAP program, you can run the program using the following command.

perl SMAP.pl -vf variant_peptide_table[file] -g genotype[file] -o result[file]

--variant_peptide,-vf	*(A file containing quantitative values of variant peptides; required)*
--genotype, -g	*(A genotype file used sample verification; required)*
--output, -o	*(An output filename; required)*
--plex, -p	*(Multiplex number of the isobaric labeling approach)*
--fold_change, -fc	*(Signal to Noise ratio (optional; default is 3))*
--noise_level, -nl	*(The upper threshold of a noise level)*
--version, -h	*(Print version)*
--help, -h	*(Print help)*
--licence, -l	*(Print licencejump –s (search))*

2.3 Input data

2.3.1 A variant peptide table

The variant peptide table uses the following format:

Column 1: Peptide ID

Column 2: Gene/Protein

Column 3: Peptide Spectrum Match (PSM)

Column 4: SNP ID **MUST MATCH GENOTYPE SNP ID

Column 5-N: Sample Peptide Quantification (One column per sample)

An example of the variant peptide table

Peptide	Gene	PSM	SNP	2015-1341	…	2016-965	Internal standard
VSNEEKVR	CAPZA1	b20_f39.15855.1.3	chr1:113162494:G:A	53788.04	…	83146.90	46477.36
HWQQFYFLSTR	FBXO2	b20_f36.35042.1.3	chr1:11710561:T:G	25447.82	…	15590.47	19626.55
SIEDLLR	PDE4DIP	b20_f22.28382.1.2	chr1:144877111:G:T	13161.86	…	10127.43	8410.05

2.3.2 A genotype in VCF format

SMAP also takes a genotype in VCF format.

An example of the genotype data

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	2014-2194	2014-2195	2014-2196
1	949608	chr1:949608:G:A	G	A	.	.	PR	GT	0/1	0/1	0/1
1	2441358	chr1:2441358:T:C	T	C	.	.	PR	GT	0/0	0/0	0/0
10	115644040	chr10:115644040:G:A	G	A	.	.	.	GT	0/1	0/0	0/1

2.3.3 Output files

An example of the final report

Sample ID	Inferred ID	CSore	DeltaCScore
2015-1341	2015-1341	4.22	0.70
2015-737	2015-737	4.03	0.56
2015-804	2015-804	3.70	0.59
2015-42	2015-37	3.14	0.51
2015-1555	2015-1555	2.91	0.54
2015-244	2015-244	2.62	0.44
2015-735	2015-735	2.53	0.43
2014-2200	2015-857	2.52	0.48
2016-958	2016-958	1.39	0.03
2016-965	2016-965	1.27	0.03
Internal standard	2015-1339	1.71	0.00

SMAP generates a final report and several intermediate results.

The final report contains four columns, including Sample ID, Inferred ID, CScore and DeltaCScore.

In addition, the program also generates three intermediate files, including sample-specific genotypes and inferred genotypes.

An example of sample-specific genotype

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	2014-2194	2014-2195	2014-2196
1	949608	chr1:949608:G:A	G	A	.	.	PR	GT	H	H	H
1	2441358	chr1:2441358:T:C	T	C	.	.	PR	GT	T	T	T
10	115644040	chr10:115644040:G:A	G	A	.	.	.	GT	C	H	H

An example of inferred genotypes

SNP	2015-1341	2015-737	2015-804	2015-42	2015-1555	2015-244	2015-735	2014-2200	2016-958	2016-965	Internal standard
chr11:75298468:A:C	A	C	A	A	A	A	A	A	A	A	A
chr5:140503474:C:G	H	H	H	H	H	C	H	H	C	C	C
chr19:40408821:C:G	C	C	C	C	C	H	G	C	C	C	C

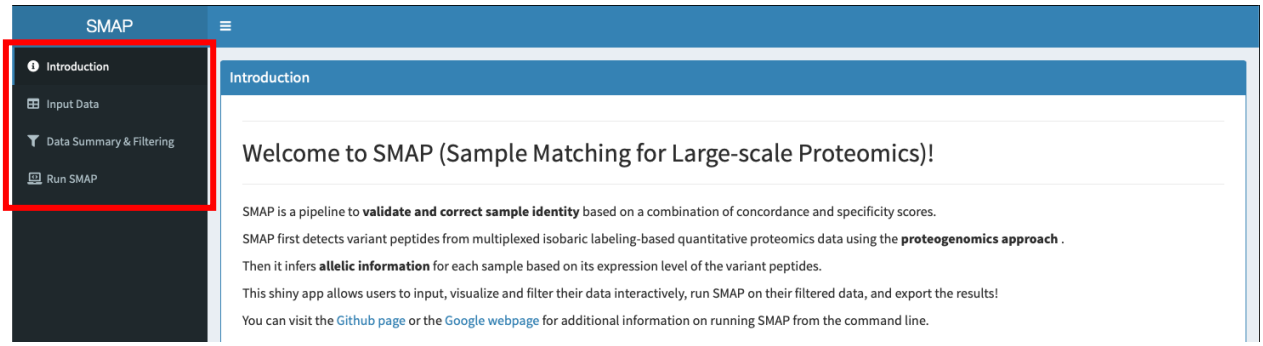
3 Cloud-based SMAP

The cloud-based SMAP is built with R shiny. It can be found at:

https://smap.shinyapps.io/smap/

3.1 Introduction

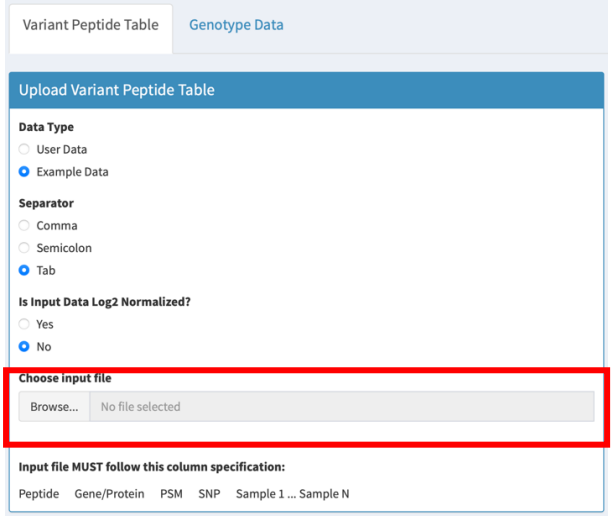
Navigation through the webpage is done by clicking on any of the four tabs at the left.

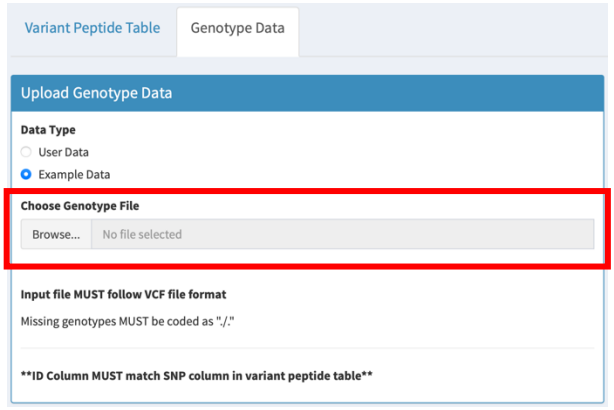


3.2 Input data

User can upload data using “Browse” buttons in **“Variant Peptide Table”** and “**Genotype Data”** menus. The format of both files can be found in the section 2.3.

- The cloud based SMAP application accepts .vcf files with any length of meta-information (including none).
- Variant peptide data will be converted into log2 scale if it is not already.





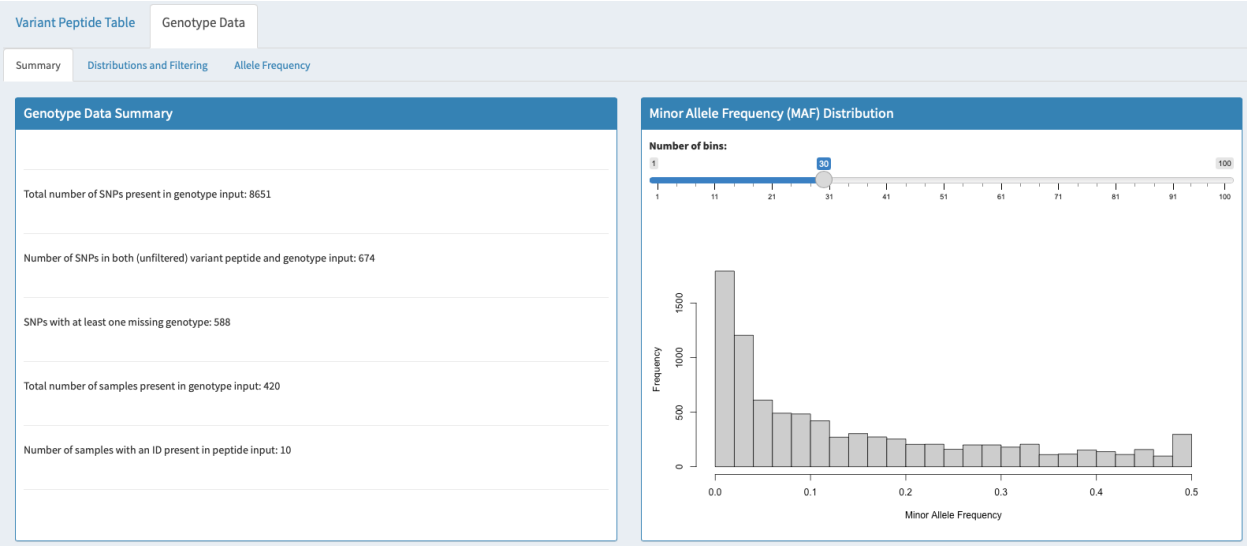
3.3 Data Summary & Filtering

For both variant peptide and genotype data, SMAP provides summary values and relevant distributions for the input files.

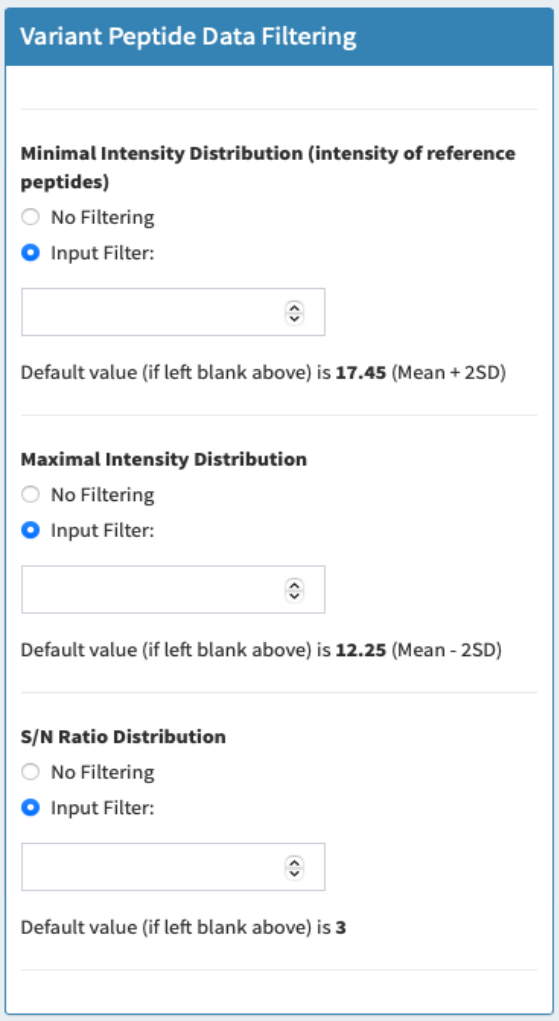
- Large genotype files may take a few moments to load

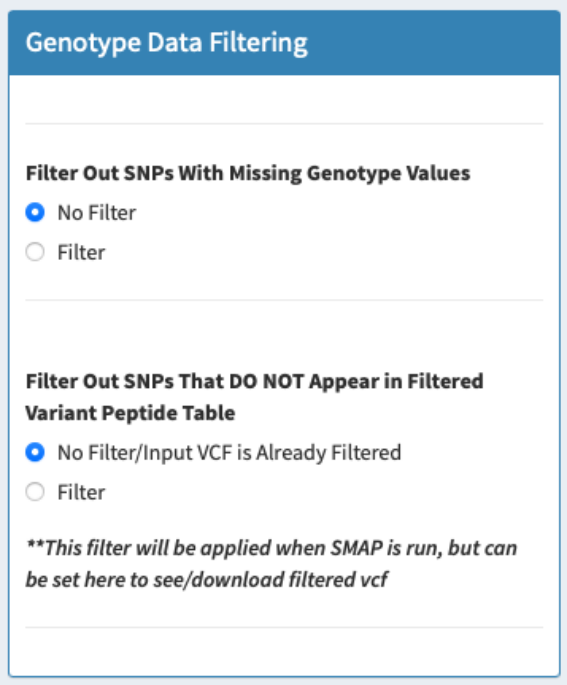
Variant Peptide Data Summary (Example):

Genotype Data Summary (Example):



Users have the options to set each variant peptide filtering parameter (minimal intensity, maximal intensity, signal/noise ratio) and the genotype filtering parameter (number of missing genotypes tolerated per SNP) based on the data distributions.



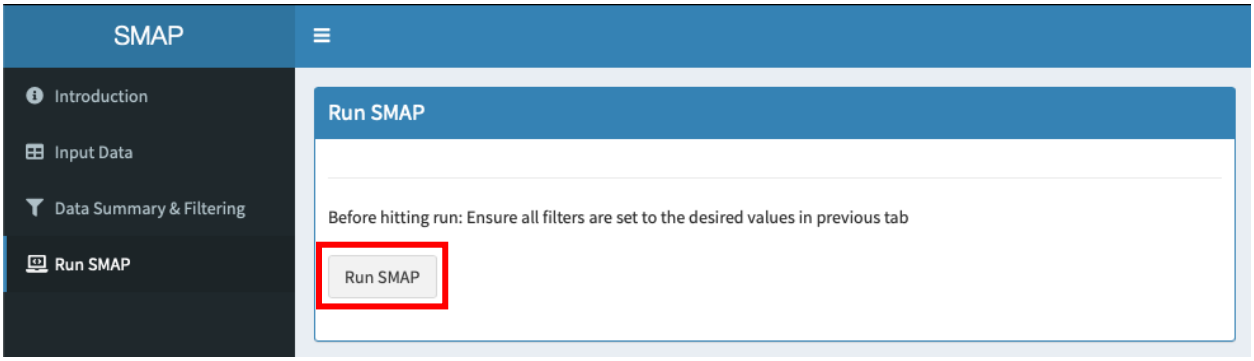


Default parameters are set (and selected if no user input is given) as follows:

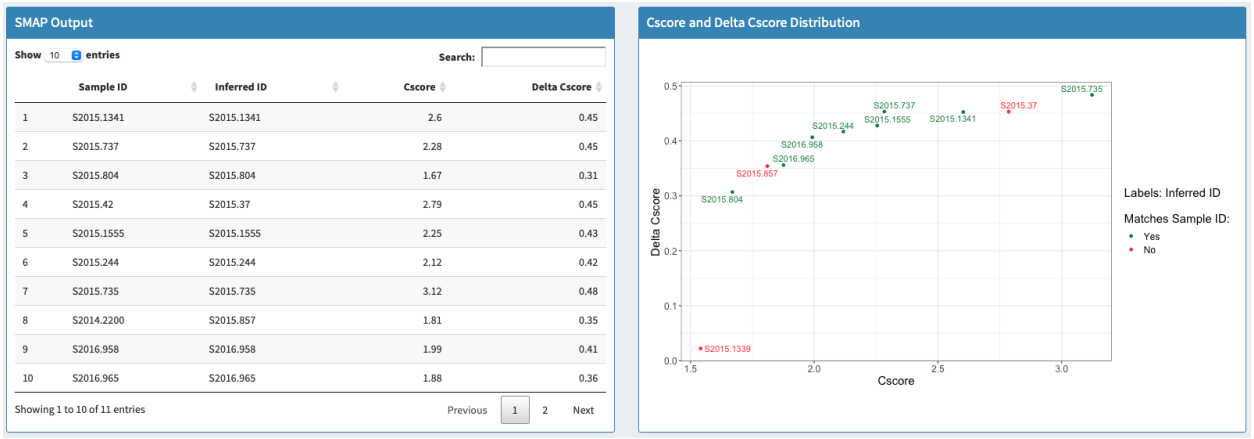
- Minimal and maximal intensity filters are set based on the means and standard deviations of the minimal and maximal peptide distributions.
- Signal/noise ratio is always set at the default of 3.
- Number of missing genotypes tolerated filter is OFF at default, but the user can set this filter if they have a large amount of missing data.

3.4 Run SMAP

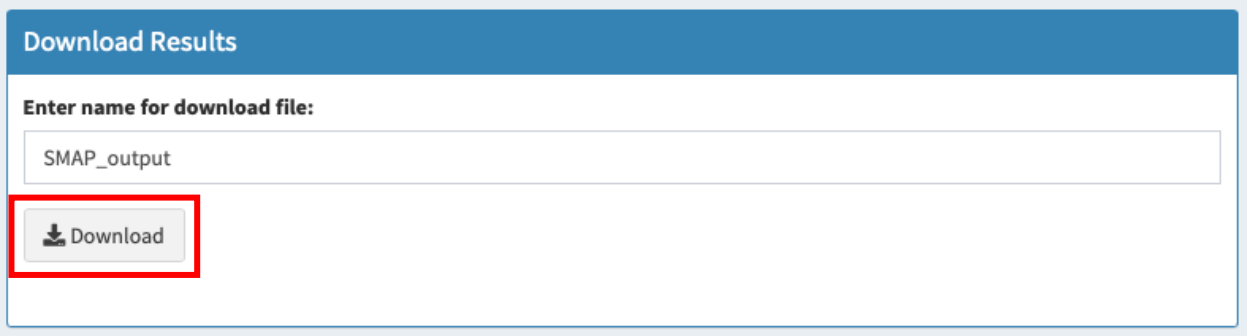
After selecting the desired filters, SMAP is by clicking the “Run SMAP” button.



After running SMAP, a table output will be generated displaying the variant peptide sample IDs and their matched genotype IDs. The Cscore and Delta Cscore for each match is also reported and graphed.



Users can download the results table by entering a desired file name and clicking download at the top right:



4 References

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