SML 201 – Week 9

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Inference on Binomial Data in R

OIS Exercise 6.10

The way a question is phrased can influence a person's response. For example, Pew Research Center conducted a survey with the following question:

"As you may know, by 2014 nearly all Americans will be required to have health insurance. [People who do not buy insurance will pay a penalty] while [People who cannot afford it will receive financial help from the government]. Do you approve or disapprove of this policy?"

For each randomly sampled respondent, the statements in brackets were randomized: either they were kept in the order given above, or the two statements were reversed.

The Data

Table 6.2 shows the results of this experiment, reproduced below.

2nd	Sample	% Approve	% Disapprove	% Other
Statement	Size	Law	Law	
"people who cannot afford it will receive financial help from the government" "people who do not buy it will pay a	771 732	47 34	49 63	3

Inference on the Difference

Create and interpret a 90% confidence interval of the difference in approval. Also perform a hyppthesis test that the approval rates are equal.

OIS 90% CI

The book OIS does a "by hand" calculation using the z-statistics and comes up with a similar answer (but not identical).

```
> p1.hat <- 0.47
> n1 <- 771
> p2.hat <- 0.34
> n2 <- 732</pre>
```

```
> stderr <- sqrt(p1.hat*(1-p1.hat)/n1 + p2.hat*(1-p2.hat)/n2)
> # the 90% CI
> (p1.hat - p2.hat) + c(-1,1)*abs(qnorm(0.05))*stderr
[1] 0.08872616 0.17127384
```

Inference on Poisson Data in R

poisson.test()

From the help:

Arguments

- x number of events. A vector of length one or two.
- T time base for event count. A vector of length one or two.
- r hypothesized rate or rate ratio

alternative indicates the alternative hypothesis and must be one of "two.sided", "greater" or "less". You can specify just the initial letter.

conf.level confidence level for the returned confidence interval.

Example: RNA-Seq

RNA-Seq gene expression was measured for p53 lung tissue in 12 healthy individuals and 14 individuals with lung cancer.

The counts were given as follows.

Healthy: 82 64 66 88 65 81 85 87 60 79 80 72

Cancer: 59 50 60 60 78 69 70 67 72 66 66 68 54 62

It is hypothesized that p53 expression is higher in healthy individuals. Test this hypothesis, and form a 99% CI.

```
H_1: \lambda_1 \neq \lambda_2
```

```
> cancer <- c(59, 50, 60, 60, 78, 69, 70, 67, 72, 66, 66, 68,
+ 54, 62)

> poisson.test(x=c(sum(healthy), sum(cancer)), T=c(12,14),
+ conf.level=0.99)

Comparison of Poisson rates

data: c(sum(healthy), sum(cancer)) time base: c(12, 14)
count1 = 909, expected count1 = 835.38, p-value =
0.0005739
alternative hypothesis: true rate ratio is not equal to 1
99 percent confidence interval:
1.041626 1.330051
sample estimates:
rate ratio
```

> healthy <- c(82, 64, 66, 88, 65, 81, 85, 87, 60, 79, 80, 72)

$H_1: \lambda_1 < \lambda_2$

1.177026

 $H_1: \lambda_1 > \lambda_2$

Question

Which analysis is the more informative and scientifically correct one, and why?

Modeling Relationships Among Variables

Rationale

One of the most important goals when analyzing data is to understand how variables relate to one another. This may include:

- Characterizing how variables covary
- Measuring and identifying associations between variables
- Explaining the variation of one variable in terms of others
- Predicting the outcome of a variable in terms of others

Strategies

We will consider both categorical and quantitative variables to achieve these goals. Over the next few weeks we will study:

- Analyzing two categorical variables
- Analyzing two quantitative variables
- Least squares linear regression to characterize variation of a quantitative variable in terms of other variables

- Logistic regression to characterize the probability distribution of a dichotomous variable in terms of other variables
- Predicting future values of a given variable based on measured values of other variables

Two Categorical Variables

Survey Data

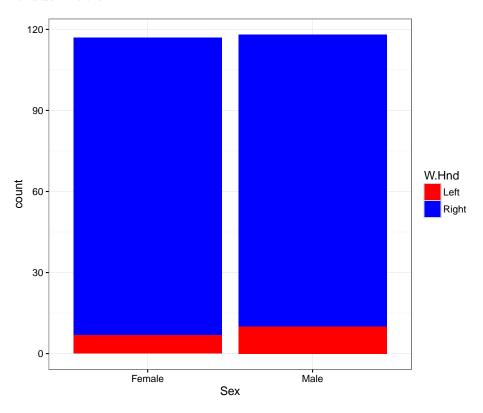
```
> library("MASS")
> data("survey", package="MASS")
> survey <- tbl_df(survey)
> head(survey)
Source: local data frame [6 x 12]
     Sex Wr.Hnd NW.Hnd W.Hnd
                                Fold Pulse
                                              Clap
                                                     Exer
  (fctr) (dbl) (dbl) (fctr)
                                            (fctr) (fctr)
                              (fctr) (int)
1 Female
         18.5
                18.0 Right R on L
                                        92
                                             Left
                                                     Some
                       Left R on L
   Male 19.5
                20.5
                                       104
                                              Left
                                                     None
3
   Male 18.0
                 13.3 Right L on R
                                       87 Neither
                                                    None
   Male 18.8
                 18.9 Right R on L
                                        NA Neither
                                                    None
   Male 20.0
                 20.0 Right Neither
                                        35
                                             Right
                                                     Some
                 17.7 Right L on R
6 Female
         18.0
                                             Right
                                                     Some
Variables not shown: Smoke (fctr), Height (dbl), M.I (fctr),
 Age (dbl)
```

2 x 2 Table

A contingency table:

Let's test the null hypothesis that sex and writing hand are independent vs. the alternative hypothesis that they are dependent.

Visualization



Pearson's Chi-Squared Test

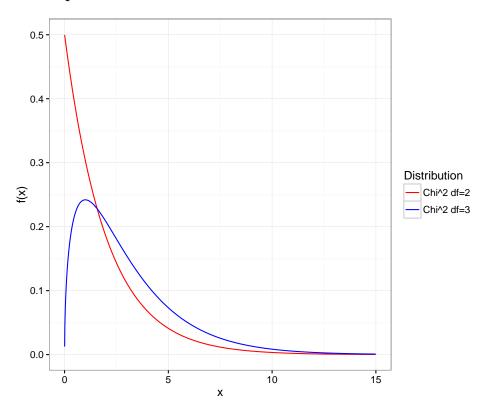
Chi-Squared Distribution

A χ^2 distribution with d degrees of freedom is equivalent to the sum of d independent Normal(0,1) random variables.

$$\chi_d^2 \sim Z_1^2 + Z_2^2 + \dots + Z_d^2$$

where Z_1, Z_2, \ldots, Z_d are iid Normal(0, 1).

Chi-Squared PDFs



Expected Counts

Observed counts:

```
> tbl

Left Right
Female 7 110
Male 10 108
```

Expected (under H_0) counts:

Chi-Squared Statistic

The chi-squared statistic is calculated as

$$X^2 = \sum \frac{(O-E)^2}{E}$$

where O is the observed count, E is the expected count, and the sum is taken over all cells in the table.

Calculate the Statistic

```
> X2 <- sum((tbl - expected)^2 / expected)
> X2
[1] 0.5435149
> chisq.test(tbl, correct=FALSE)$statistic # equals X2
X-squared
0.5435149
> chisq.test(tbl)$statistic # with continuity correction
X-squared
0.2356302
```

Calculate the P-value

The null distribution of X^2 is a χ^2 distribution with d degrees of freedom. We calculate d by d = (r-1)(c-1) where r is the number of rows and c is the number of columns.

```
> 1-pchisq(X2, df=1)
[1] 0.4609797
>
> chisq.test(tbl, correct=FALSE)$p.value
[1] 0.4609797
```

Derivation

The theoretical derivation of this test is beyond the scope of this course.

However, it is worth noting that it is related to the Z-statistic approximation from last week:

$$Z = \frac{\text{estimator} - \text{parameter}}{\text{standard error}} \sim \text{Normal}(0, 1).$$

Guidelines for Practice

- The total number of observations should be "large" so that \dots
- The expected number of counts per cell should be 10 or greater
- The observed number of counts per cell should be 5 or greater

When these are violated, continuity corrections and simulation based p-values can be used... or other tests can be used such as Fisher's Exact Test — see fisher.test().

Clapping and Writing Hand

Note that now one of the categorical variables takes three values.

Also note the existence of low cell counts.

Chi-Squared Test Via Simulation

```
> chisq.test(tbl)
Warning in chisq.test(tbl): Chi-squared approximation may be incorrect

Pearson's Chi-squared test

data: tbl
X-squared = 19.252, df = 2, p-value = 6.598e-05
```

We address this warning by simulating tables from the null hypothesis.

```
> chisq.test(tbl, simulate.p.value = TRUE, B=10000)

Pearson's Chi-squared test with simulated p-value
   (based on 10000 replicates)

data: tbl
X-squared = 19.252, df = NA, p-value = 9.999e-05
```

Exercise Vs. Writing Hand

```
> tbl = table(survey$Exer, survey$W.Hnd)
> tbl
      Left Right
       7 107
 Freq
 None
         3
              21
         8
              90
  Some
> chisq.test(tbl, simulate.p.value = TRUE, B=10000)
    Pearson's Chi-squared test with simulated p-value
    (based on 10000 replicates)
data: tbl
X-squared = 1.2065, df = NA, p-value = 0.5532
```

Smoking Vs. Exercise

```
> tbl = table(survey$Smoke, survey$Exer)
> tbl
        Freq None Some
 Heavy
          7
               1
 Never
          87
               18
                    84
  Occas
         12
                3
                     4
                     7
 Regul
          9
                1
> chisq.test(tbl, simulate.p.value = TRUE, B=10000)
    Pearson's Chi-squared test with simulated p-value
    (based on 10000 replicates)
data: tbl
X-squared = 5.4885, df = NA, p-value = 0.4794
```

What feature of the data is this test ignoring?

Goodness of Fit Tests

The chisq.tes() function also performs goodness of fit tests. These are goodness of fit tests of a set of probabilities, very related to our tests of proportions from last week.

For example, suppose we want to test whether a six-sided die is fair. We roll the die 100 times and record the frequency with which we observe each face.

```
> die
die
1 2 3 4 5 6
17 14 21 28 14 6
> chisq.test(x=die, p=rep(1/6, 6))

    Chi-squared test for given probabilities

data: die
X-squared = 16.52, df = 5, p-value = 0.005506
```

Two Quantitative Variables

Correlation

- It is often the case that two or more quantitative variables are measured on each unit of observation (such as an individual).
- We are then often interested in characterizing how pairs of variables are associated or how they vary together.
- A common measure that is used is called "correlation", which is most well suited for measuring linear associations

Sample Correlation

Suppose we observe n pairs of data $(x_1, y_1), (x_2, y_2), \dots, (x_n, y_n)$. Their sample correlation is

$$r_{xy} = \frac{\sum_{i=1}^{n} (x_i - \overline{x})(y_i - \overline{y})}{\sqrt{\sum_{i=1}^{n} (x_i - \overline{x})^2 \sum_{i=1}^{n} (y_i - \overline{y})^2}}$$

$$= \frac{\sum_{i=1}^{n} (x_i - \overline{x})(y_i - \overline{y})}{(n-1)s_x s_y}$$
(1)

$$= \frac{\sum_{i=1}^{n} (x_i - \overline{x})(y_i - \overline{y})}{(n-1)s_x s_y}$$
 (2)

where s_x and s_y are the sample standard deviations of each measured variable.

Ranked-Based Correlation

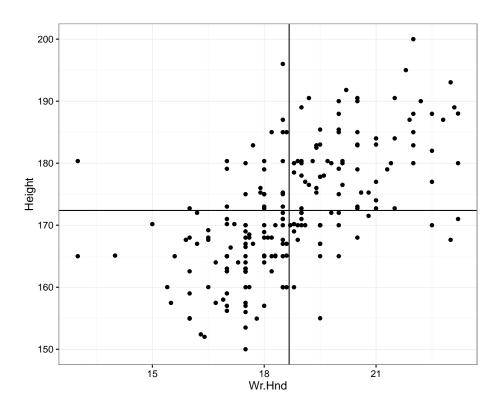
- There are other ways to measure correlation that are less reliant on linear trends in covariation and are also more robust to outliers.
- Specifically, one can convert each measured variable to ranks by size (1 for the smallest, n for the largest) and then use a formula for correlation designed for these ranks.
- One popular measure of rank-based correlation is the Spearman correlation.

Population Correlation

Suppose there are two random variables \$X and Y. Their population correlation is

$$\rho_{XY} = \frac{\mathrm{E}\left[(X - \mathrm{E}[X])(Y - \mathrm{E}[Y])\right]}{\sqrt{\mathrm{Var}(X)\mathrm{Var}(Y)}}$$

Hand Size Vs. Height



Calculating Correlation

Example Correlations

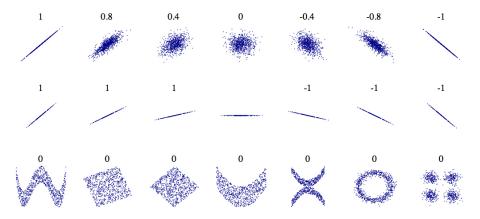


Image from Wikipedia.

HT of Correlation

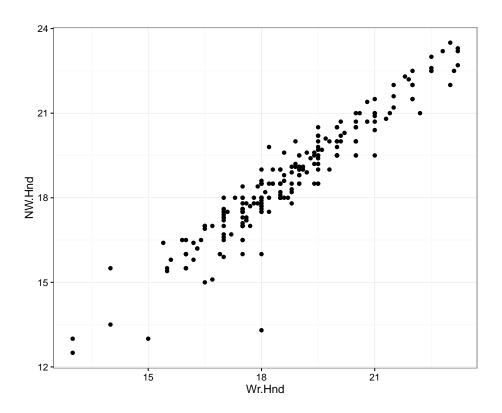
HT of Correlation

HT By Hand

Compare the following to the above output of cor.test().

Hand Sizes

```
> ggplot(data = survey) +
+ geom_point(aes(x=Wr.Hnd, y=NW.Hnd))
```



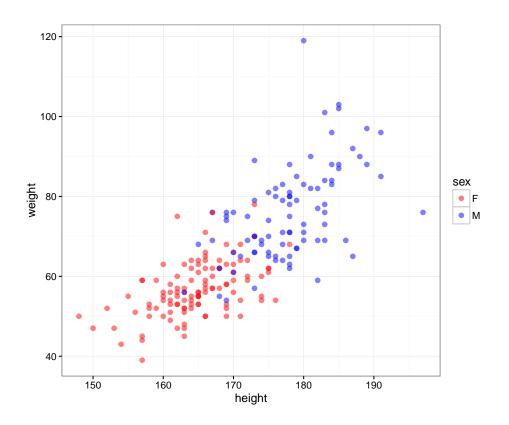
Correlation of Hand Sizes

Davis Data

```
> library("car")
> data("Davis", package="car")
> htwt <- tbl_df(Davis)</pre>
> htwt[12,c(2,3)] <- htwt[12,c(3,2)]
> head(htwt)
Source: local data frame [6 x 5]
     sex weight height repwt repht
  (fctr) (int) (int) (int)
1
      M
            77
                  182
                        77
                             180
2
      F
            58
                  161
                        51
                            159
      F
3
                161
                        54 158
            53
4
      M
            68
                  177
                        70 175
      F
5
            59
                  157
                        59
                            155
6
      Μ
            76
                170
                        76 165
```

Height and Weight

```
> ggplot(htwt) +
+ geom_point(aes(x=height, y=weight, color=sex), size=2, alpha=0.5) +
+ scale_color_manual(values=c("red", "blue"))
```



Correlation Test

Correlation Test with Outlier

Recall we had to fix an error in the data, which we noticed as an outlier in the scatterplot. Here is the effect of the outlier:

Correlation Test with Outlier

Let's use the Spearman rank-based correlation:

```
> cor.test(x=Davis$height, y=Davis$weight, method="spearman")
Warning in cor.test.default(x = Davis$height, y = Davis$weight,
method = "spearman"): Cannot compute exact p-value with ties

Spearman's rank correlation rho

data: Davis$height and Davis$weight
S = 308750, p-value < 2.2e-16
alternative hypothesis: true rho is not equal to 0
sample estimates:
    rho
0.7684305</pre>
```

Correlation Among Females

```
> htwt %>% filter(sex=="F") %>%
+ cor.test(~ height + weight, data = .)

Pearson's product-moment correlation

data: height and weight
t = 6.2801, df = 110, p-value = 6.922e-09
alternative hypothesis: true correlation is not equal to 0
95 percent confidence interval:
    0.3627531    0.6384268
sample estimates:
```

```
cor
0.5137293
```

Correlation Among Males

Why are the stratified correlations lower?

Least Squares Linear Regression

Rationale

- It is often the case that we would like to build a model that explains the variation of one variable in terms of other variables.
- Least squares linear regression is one of the simplest and most useful modeling systems for doing so.
- It is simple to fit, it satisfies some optimality criteria, and it is straightforward to check assumptions on the data so that statistical inference can be performed.

Setup

- Let's start with least squares linear regression of just two variables.
- Suppose that we have observed n pairs of data $(x_1, y_1), (x_2, y_2), \ldots, (x_n, y_n)$.
- Least squares linear regression models variation of y_i in terms of $\beta_0 + \beta_1 x_i$ where β_0 and β_1 are chosen to satisfy a least squares optimization.

Line that Minimizes the Squared Error

The least squares regression line is formed from the value of β_0 and β_1 that minimize:

$$\sum_{i=1}^{n} (y_i - \beta_0 - \beta_1 x_i)^2.$$

For a given set of data, there is a unique solution to this minimization as long as there are at least two unique values among x_1, x_2, \ldots, x_n .

Let $\hat{\beta}_0$ and $\hat{\beta}_1$ be the values that minimize this sum of squares.

Least Squares Solution

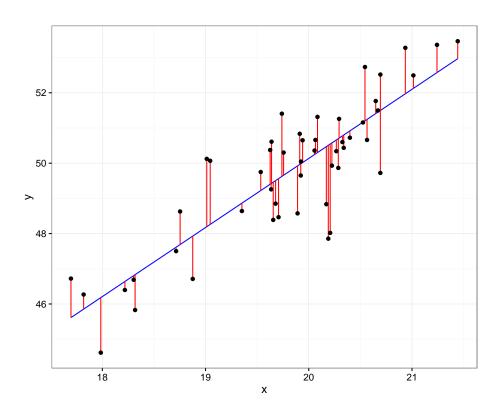
These values are:

$$\hat{\beta}_1 = r_{xy} \frac{s_y}{s_x}$$

$$\hat{\beta}_0 = \overline{y} - \hat{\beta}_1 \overline{x}$$

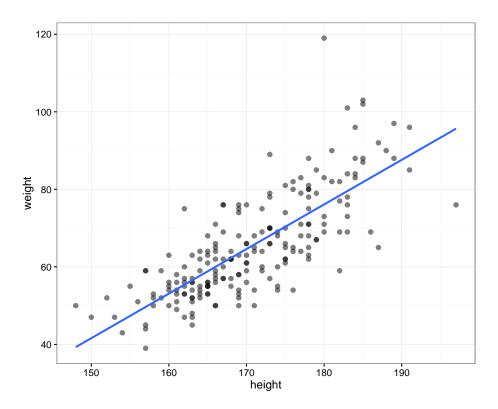
These values have a useful interpretation.

Visualizing Least Squares Line



Example: Height and Weight

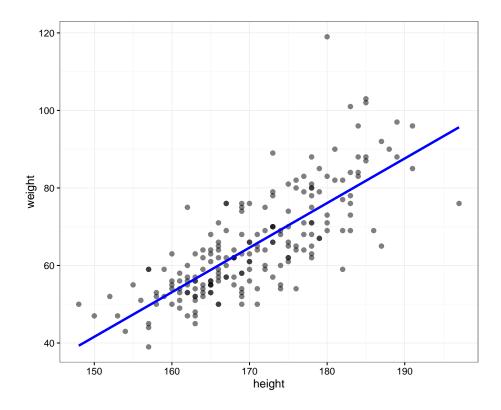
```
> ggplot(data=htwt, mapping=aes(x=height, y=weight)) +
+ geom_point(size=2, alpha=0.5) +
+ geom_smooth(method="lm", se=FALSE, formula=y~x)
```



Calculate the Line by Hand

Plot the Line

```
> df <- data.frame(htwt, yhat=yhat)
> ggplot(data=df) + geom_point(aes(x=height, y=weight), size=2, alpha=0.5) +
        geom_line(aes(x=height, y=yhat), color="blue", size=1.2)
```



Calculate the Line in R

```
> myfit <- lm(weight ~ height, data=htwt)
> myfit

Call:
lm(formula = weight ~ height, data = htwt)

Coefficients:
(Intercept) height
    -130.91 1.15
```

What's Next?

- Why minimize the sum of squares?
- What is the output provided by R?
- How do we access and interpret this output from R?
- What assumptions are required to use this machinery?

- How do we check these assumptions on data?
- How can we build more complex models?

Extras

License

https://github.com/SML201/lectures/blob/master/LICENSE.md

Source Code

https://github.com/SML201/lectures/tree/master/week9

Session Information

```
> sessionInfo()
R version 3.2.3 (2015-12-10)
Platform: x86_64-apple-darwin13.4.0 (64-bit)
Running under: OS X 10.11.3 (El Capitan)
locale:
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
attached base packages:
[1] stats
             graphics grDevices utils
                                            datasets methods
[7] base
other attached packages:
[1] car_2.1-1
                    MASS_7.3-45
                                    broom 0.4.0
                                    knitr_1.12.3
[4] dplyr_0.4.3
                    ggplot2_2.1.0
[7] magrittr_1.5
                   devtools_1.10.0
loaded via a namespace (and not attached):
 [1] Rcpp_0.12.3
                        nloptr_1.0.4
                                           formatR_1.2.1
 [4] plyr_1.8.3
                        tools_3.2.3
                                           digest_0.6.9
 [7] lme4_1.1-11
                        evaluate_0.8
                                           memoise_1.0.0
[10] nlme_3.1-125
                        gtable_0.2.0
                                           lattice_0.20-33
                        Matrix_1.2-3
                                           psych_1.5.8
[13] mgcv_1.8-11
[16] DBI_0.3.1
                        yaml_2.1.13
                                           parallel_3.2.3
[19] SparseM 1.7
                        stringr_1.0.0
                                           MatrixModels 0.4-1
[22] grid_3.2.3
                        nnet_7.3-12
                                           R6 2.1.2
[25] rmarkdown_0.9.5
                        minqa_1.2.4
                                           reshape2_1.4.1
```

```
[28] tidyr_0.4.1 scales_0.4.0 htmltools_0.3
[31] splines_3.2.3 assertthat_0.1 pbkrtest_0.4-6
[34] mnormt_1.5-3 colorspace_1.2-6 quantreg_5.21
[37] labeling_0.3 stringi_1.0-1 lazyeval_0.1.10
[40] munsell_0.4.3
```