

# NSCLC\_Salcher2022\_process

December 25, 2025

## 1 load data

```
[ ]: obj.NSCLC <- readRDS('/project/sex_cancer/data/NSCLC_Salcher2022/  
  ↪NSCLC_Salcher2022_coreAtlas.rds')  
obj.NSCLC <- UpdateSeuratObject(obj.NSCLC)
```

## 2 modify meta.data

```
[ ]: obj.NSCLC@meta.data <- obj.NSCLC@meta.data %>%  
  dplyr::rename(c("Sex" = "sex")) %>%  
  transform(barcode = rownames(.))
```

## 3 filter sample

```
[ ]: obj.NSCLC <- obj.NSCLC %>%  
  subset(Sex %in% c('female', 'male')) %>%  
  subset(tissue == 'lung') %>%  
  subset(origin != 'nan') %>%  
  subset(platform == '10x') %>%  
  subset(disease != 'chronic obstructive pulmonary disease')  
length(unique(obj.NSCLC$sample))  
table(obj.NSCLC$study)
```

```
[ ]: ## de-factor  
obj.NSCLC@meta.data <- obj.NSCLC@meta.data %>%  
  mutate_if(~ !is.numeric(.), ~ ext_list(.)) %>%  
  dplyr::rename(c('SampleID' = 'sample', 'Chemistry' =  
  ↪'assay', 'DonorID' = 'donor_id')) %>%  
  transform(Sex = ifelse(Sex == 'female', 'F', 'M')) %>%  
  mutate(SampleType = case_when(origin == 'normal' ~  
  ↪'normal',  
                                     origin == 'normal_adjacent' ~  
  ↪~ 'normal_adjacent',  
                                     origin == 'tumor_primary' ~  
  ↪'tumor',  
                                     origin == 'tumor_secondary' ~  
  ↪'tumor'))
```

```

      TRUE ~ 'Others')) %>%
  transform(Cohort = 'NSCLC_Salcher2022')

```

## 4 trans ENSG to symbol

```

[ ]: table(rownames(obj.NSCLC@assays$RNA@counts) == rownames(obj.
  ↪NSCLC@assays$RNA@data))

trans <- obj.NSCLC@assays$RNA@meta.features %>%
  dplyr::select(c('feature_type', 'feature_name')) %>%
  .[rownames(obj.NSCLC@assays$RNA@counts),] %>%
  rownames_to_column('feature')

trans %>% head(n = 2)

## rename counts
rownames(obj.NSCLC@assays$RNA@counts) <- trans$feature_name
## rename data
rownames(obj.NSCLC@assays$RNA@data) <- trans$feature_name
## rename meta.feature
obj.NSCLC@assays$RNA@meta.features <- trans %>% ↪
  ↪column_to_rownames('feature_name')

```

## 5 cell type annotation

### 5.1 assign oCT

```

[ ]: obj.NSCLC@meta.data <- obj.NSCLC@meta.data %>%
  mutate(oCT = cell_type_major)

```

### 5.2 check annotation

check cell type annotation provided in the original research via COSG

```

[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.NSCLC
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.
  ↪factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
        mu = 10, ## The penalty factor to penalize gene expression in ↪
  ↪cells not belonging to the cluster of interest

```

```

n_genes_user = 50, # Number of top ranked genes returned in the result
remove_lowly_expressed=T, # If TRUE, genes that express a percentage of target cells smaller than a specific value (expressed_pct) are not considered as marker genes for the target cells. The default value is TRUE.
expressed_pct=0.1) # If TRUE, genes that express a percentage of target cells smaller than a specific value (expressed_pct) are not considered as marker genes for the target cells.
marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::rename(c("oCT" = "variable", "marker" = "value")),
marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::select(-"variable") %>% dplyr::rename(c("COSGscore" = "value")))) %>% mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
  check <- oCT_marker %>% subset(oCT == x & marker %in% marker_annotation[[x]])
  ifelse(nrow(check) == 0, print(x), return(check))
})

```

### 5.3 assign mCT

```

[ ]: obj.NSCLC@meta.data <- obj.NSCLC@meta.data %>%
  mutate(mCT = case_when(cell_type_major %in% c('Tumor cells') ~ 'Tumor',
                         cell_type_major %in% c('Alveolar cell type 1', 'Ciliated', 'Club', 'transitional club/AT2', 'Alveolar cell type 2') ~ 'Epi',
                         cell_type_major %in% c('T cell CD8') ~ 'CD8T',
                         cell_type_major %in% c('T cell CD4') ~ 'CD4T',
                         cell_type_major %in% c('T cell regulatory') ~ 'Treg',
                         cell_type_major %in% c('NK cell') ~ 'NK',
                         cell_type_major %in% c('B cell', 'Plasma cell') ~ 'B',
                         c('Monocyte') ~ 'Mono',

```

```

            cell_type_major %in% ↵
↳ c('Macrophage alveolar', 'Macrophage') ~ 'Mph',
            cell_type_major %in% c('cDC2', ↵
↳ 'cDC1', 'pDC', 'DC mature') ~ 'DC',
            cell_type_major %in% ↵
↳ c('Neutrophils') ~ 'Neu',
            cell_type_major %in% c('Mast', ↵
↳ 'cell') ~ 'Mast',
            cell_type_major %in% ↵
↳ c('Endothelial cell') ~ 'Endo',
            cell_type_tumor %in% ↵
↳ c('Fibroblast adventitial', 'Fibroblast alveolar', 'Fibroblast', ↵
↳ 'peribronchial') ~ 'Fibro',
            cell_type_tumor %in% ↵
↳ c('Pericyte') ~ 'Pericyte',
            cell_type_tumor %in% ↵
↳ c('Mesothelial') ~ 'Mesothelial',
            cell_type_tumor %in% c('Smooth', ↵
↳ 'muscle cell') ~ 'SMC',
            cell_type_major %in% c('other') ↵
↳ ~ 'Others',
            ))
head(obj.NSCLC@meta.data, n = 2)

```

## 5.4 assign gCT

```

[ ]: obj.NSCLC@meta.data <- obj.NSCLC@meta.data %>%
            mutate(gCT = case_when(mCT %in% c('Tumor', 'Epi') ~ ↵
↳ 'Tumor',
            mCT %in% c('Neu', 'Mast', ↵
↳ 'CD8T', 'CD4T', 'NK', 'DC', 'B', 'Treg', 'Mono', 'Mph') ~ 'Immune',
            mCT %in% c('Pericyte', 'SMC', ↵
↳ 'Mesothelial', 'Fibro', 'Endo') ~ 'Stromal',
            TRUE ~ 'Others',
            ))
head(obj.NSCLC@meta.data, n = 2)

```

## 5.5 discard unannotated cells

```

[ ]: obj.NSCLC <- obj.NSCLC %>% subset(gCT != "Others")
obj.NSCLC

```

```
[ ]: table(obj.NSCLC$oCT, obj.NSCLC$gCT, useNA = "ifany")
```

## 6 save

```
[ ]: saveRDS(obj.NSCLC, 'obj.NSCLC.use.rds')
```