

# BLCA\_Juric2025\_process

December 25, 2025

## 1 load data

```
[ ]: obj.BLCA <- readRDS("/project/sex_cancer/data/BLCA_Juric2025/  
  ↪GSE269877_dta_cancer.submission.rds") ## sex information not provided in the_  
  ↪normal sample object, making it not included in SexTumorDB  
obj.BLCA  
obj.BLCA@meta.data %>% head(n = 2)  
  
[ ]: obj.BLCA@meta.data %>%  
dplyr::select(c("orig.ident", "Tumor.Category", "Groups", "Sex", "Age")) %>%  
  .[!duplicated(.$orig.ident),] %>% subset(Groups != "Recurrent") %$%  
table(.$Sex)  
  
[ ]: obj.BLCA <- RenameAssays(obj.BLCA, RNACleaned = "RNA")  
obj.BLCA@assays$RNA@data <- obj.BLCA@assays$RNA@counts
```

## 2 filter samples

```
[ ]: obj.BLCA <- obj.BLCA %>%  
  subset(Groups %in% c("Naive_wo", "Naive_w")) ## drop samples_  
  ↪collected after BCG treatment (treated samples)  
length(unique(obj.BLCA$orig.ident))
```

## 3 modify meta.data

```
[ ]: rownames(obj.BLCA) %>% .[grepl("^MT-", .)] ## MT genes are removed in the_  
  ↪original dataset, seems to be set <30  
  
[ ]: obj.BLCA@meta.data <- obj.BLCA@meta.data %>%  
  transform(barcode = rownames(.)) %>%  
  dplyr::select(-c("nCount_RNACleaned", "  
  ↪nFeature_RNACleaned")) %>%  
  dplyr::rename(c("DonorID" = "Patient")) %>%  
  transform(SampleID = orig.ident) %>%
```

```

transform(Cohort = "BLCA_Juric2025", Chemistry = "10x 3'
↪v3", Tissue = "Bladder", SampleType = "tumor") %>%
  dplyr::select(-c("Exome.seq", "cell.names.CD6.status",
↪"percent.rps", "percent.rpl", "Tumor.Category", "Stage", "Grade")) %>%
  .[colnames(obj.BLCA),]
obj.BLCA@meta.data %>% head(n = 2)

```

```

[ ]: options(repr.plot.height = 4, repr.plot.width = 20)
DimPlot_scCustom(obj.BLCA, pt.size = 1, reduction = "umap", group.by = "Sex",
↪label = F, label.size = 8, colors_use = pal_igv("default")(51), raster = F)|
DimPlot_scCustom(obj.BLCA, pt.size = 1, reduction = "umap", group.by =
↪"low_res_compartment", label = F, label.size = 8, colors_use =
↪pal_igv("default")(51), raster = F)|
DimPlot_scCustom(obj.BLCA, pt.size = 1, reduction = "umap", group.by = "cell.
↪names", label = F, label.size = 8, colors_use = pal_igv("default")(51),
↪raster = F)

```

## 4 cell type annotation

### 4.1 assign oCT

```

[ ]: obj.BLCA@meta.data <- obj.BLCA@meta.data %>%
  dplyr::rename(c("oCT" = "cell.names"))

```

### 4.2 check annotation

check cell type annotation provided in the original research via COSG

```

[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.BLCA
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.
↪factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
  mu = 10, ## The penalty factor to penalize gene expression in
↪cells not belonging to the cluster of interest
  n_genes_user = 50, # Number of top ranked genes returned in the
↪result

```

```

remove_lowly_expressed=T, # If TRUE, genes that express a
percentage of target cells smaller than a specific value (expressed_pct) are
not considered as marker genes for the target cells. The default value is
TRUE.

expressed_pct=0.1) # If TRUE, genes that express a percentage of
target cells smaller than a specific value (expressed_pct) are not
considered as marker genes for the target cells.
marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
rename(c("oCT" = "variable", "marker" = "value")),
marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
select(-"variable") %>% dplyr::rename(c("COSGscore" = "value"))) %>%
mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
  check <- oCT_marker %>% subset(oCT == x & marker %in%
marker_annotation[[x]])
  ifelse(nrow(check) == 0, print(x), return(check))
})

```

### 4.3 assign mCT

```

[ ]: obj.BLCA@meta.data <- obj.BLCA@meta.data %>%
mutate(mCT = case_when(oCT %in% c("Proliferating
uroepithelial", "Intermediate A", "Intermediate B", "Intermediate C",
"Intermediate D", "Intermediate E", "Umbrella", "Basal") ~ "Epi",
oCT %in% c("Endothelial arterial",
"Enodothelial venous", "Activated endothelial") ~ "Endo",
oCT %in% c("WNThi periurothelial
fibroblast", "Myofibroblast", "Periurothelial fibroblast", "Interstitial
fibroblast") ~ "Fibro",
oCT %in% c("B") ~ "B",
oCT %in% c("Plasma") ~ "Plasma",
oCT %in% c("Treg") ~ "Treg",
oCT %in% c("CD4 T central memory
1", "CD4 T central memory 2", "CD4 T effector memory", "CD4 T exhausted") ~
"CD4T",
oCT %in% c("CD8 T 2", "CD8 T
effector", "CD8 T resident memory") ~ "CD8T",
oCT %in% c("Proliferating
lymphocyte") ~ "T_proliferation",
oCT %in% c("NK CD56 bright", "NK
CD56 dim") ~ "NK",
oCT %in% c("MMDSC") ~ "Mono",

```

```

oCT %in% c("CCL17+ dendritic",
↪ "Immature dendritic", "Type 2 conventional dendritic", "pDC") ~ "DC",
oCT %in% c("Macrophage") ~ "Mph",
oCT %in% c("Mast") ~ "Mast",
TRUE ~ 'Others'))

obj.BLCA@meta.data %>% subset(mCT == "Others") %$% table(.$oCT, useNA= "ifany")
↪ ## all removed.during.subclustering

```

```

[ ]: ## filter out removed cells
obj.BLCA <- obj.BLCA %>% subset(mCT != "Others") ## discard 9766 cells
↪ annotated as "removed.during.subclustering" by the original research
obj.BLCA

```

#### 4.4 assign gCT

```

[ ]: unique(obj.BLCA$mCT)

[ ]: obj.BLCA@meta.data <- obj.BLCA@meta.data %>%
      mutate(gCT = case_when(mCT %in% c("Epi") ~ "Tumor",
                             mCT %in% c("DC", "CD8T", "Treg",
↪ "Mph", "T_proliferation", "CD4T", "NK", "Plasma", "Mono", "B", "Mast") ~
↪ "Immune",
                             mCT %in% c("Fibro", "Endo") ~
↪ "Stromal",
                             TRUE ~ 'Others'))
table(obj.BLCA$gCT)

```

#### 5 save

```

[ ]: saveRDS(obj.BLCA, 'obj.BLCA.final.use.rds')

```