

ESCA_Zhang2021_process

December 25, 2025

1 load data

“All patients were not treated with chemotherapy or radiotherapy before tumor resection”

```
[ ]: exp1 <- fread('/project/sex_cancer/data/ESCC_Zhang2021/GSE160269_CD45neg_UMIs.  
    ↴txt') %>% column_to_rownames('V1')  
exp2 <- fread('/project/sex_cancer/data/ESCC_Zhang2021/GSE160269_CD45pos_UMIs.  
    ↴txt') %>% column_to_rownames('V1')  
  
all_genes <- unique(c(rownames(exp1), rownames(exp2)))  
# create merged df  
exp_merge <- data.frame(matrix(0, nrow = length(all_genes), ncol = ncol(exp1) +  
    ↴ncol(exp2)))  
rownames(exp_merge) <- all_genes  
colnames(exp_merge) <- c(colnames(exp1), colnames(exp2))  
# load data  
exp_merge[rownames(exp1), colnames(exp1)] <- exp1  
exp_merge[rownames(exp2), colnames(exp2)] <- exp2
```



```
[ ]: metaList <- list.files('/project/sex_cancer/data/ESCC_Zhang2021', pattern =  
    ↴'GSE160269_*cells', full.names = TRUE)  
meta <- lapply(metaList, function(x){fread(x)}) %>%  
    do.call(rbind, .) %>%  
    transform(DonorID = gsub('T|N', '', sample), barcode = rownames(.))  
  
info <- read_xlsx('./ESCC_Zhang2021/ESCC_Zhang2021_PatientInfo.xlsx', skip = 1)  
names(info)[1]='DonorID'  
meta <- merge(meta, info, by = 'DonorID', all = TRUE) %>%  
    ↴column_to_rownames('barcode') %>% mutate_if(~!is.numeric(.), ext_list) %>% .  
    ↴[colnames(exp_merge),]
```



```
[ ]: obj.ESCA <- CreateSeuratObject(counts = exp_merge, meta.data = meta, min.cells=  
    ↴= 0,min.features = 0)  
obj.ESCA
```

2 modify meta.data

```
[ ]: ## 60patient ~ 64 samples (4 normal_adjacent
obj.ESCA@meta.data <- obj.ESCA@meta.data %>%
  dplyr::rename(c('SampleID' = 'sample')) %>%
  transform(Cohort = 'ESCA_Zhang2021') %>%
  mutate(Sex = case_when(Sex == 'Female' ~ 'F', Sex == 'Male' ~ 'M', TRUE ~ 'Others')) %>%
  mutate(SampleType = case_when(SampleID %in% c('P126N', 'P127N', 'P128N', 'P130N') ~ 'normal_adjacent', TRUE ~ 'tumor'))
obj.ESCA@meta.data %>% head(n = 2)

[ ]: obj.ESCA@meta.data[,c('SampleID', 'SampleType', 'Sex')] %>% .[!duplicated(.,
  .$SampleID),] %% table(.$.SampleType, .$.Sex)
obj.ESCA@meta.data %>% subset(SampleType == 'tumor') %>% nrow()
```

3 cell type annotation

3.1 assign oCT

```
[ ]: obj.ESCA@meta.data <- obj.ESCA@meta.data %>%
  mutate(oCT = annotated_type) %>%
  mutate(dCT = case_when(oCT %in% c('Epithelial') ~ 'Epi',
    oCT %in% c('Bcell') ~ 'B/Plasma',
    oCT %in% c('Endothelial') ~ 'Endo',
    oCT %in% c('Fibroblast') ~ 'Fibro',
    oCT %in% c('Tcell') ~ 'NK/T',
    oCT %in% c('Pericytes') ~ 'Pericyte',
    oCT %in% c('FRC') ~ 'FRC', ## TRUE ~ oCT))
  ↵Fibroblastic Reticular Cells
```

3.2 check annotation

check cell type annotation provided in the original research via COSG

```
[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.ESCA
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)
```

```

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
        mu = 10, ## The penalty factor to penalize gene expression in
  ↵cells not belonging to the cluster of interest
  n_genes_user = 50, # Number of top ranked genes returned in the
  ↵result
  remove_lowly_expressed=T, # If TRUE, genes that express a
  ↵percentage of target cells smaller than a specific value (expressed_pct) are
  ↵not considered as marker genes for the target cells. The default value is
  ↵TRUE.
  expressed_pct=0.1) # If TRUE, genes that express a percentage of
  ↵target cells smaller than a specific value (expressed_pct) are not
  ↵considered as marker genes for the target cells.

marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
  ↵rename(c("oCT" = "variable", "marker" = "value")),
  marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
  ↵select(-"variable") %>% dplyr::rename(c("COSGscore" = "value")))) %>%
  mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
  ↵ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
  check <- oCT_marker %>% subset(oCT == x & marker %in% 
  ↵marker_annotation[[x]])
  ifelse(nrow(check) == 0, print(x), return(check))
})

```

3.3 assign NK/T

```

[ ]: obj.ESCA.nkt <- obj.ESCA %>% subset(dCT == 'NK/T')
obj.ESCA.nkt

obj.ESCA.nkt <- obj.ESCA.nkt %>%
  NormalizeData(normalization.method = "LogNormalize", scale.
  ↵factor = 10000, verbose = F) %>%
  FindVariableFeatures(selection.method = "vst", nfeatures =
  ↵1000, verbose = F) %>%
  ScaleData(vars.to.regress = c("nCount_RNA"), verbose = F) %>%
  RunPCA(verbose = F) %>%
  RunHarmony(group.by.vars = "SampleID", plot_convergence = TRUE)

## cluster
nPC <- min(PC_selection_harmony(obj.ESCA.nkt)$PCselect)
obj.ESCA.nkt <- obj.ESCA.nkt %>%
  RunUMAP(reduction = "harmony", dims = 1:nPC, umap.method =
  ↵"uwot") %>%

```

```

    FindNeighbors(reduction = "harmony", dims = 1:nPC) %>%
    FindClusters(resolution=0.1) %>% FindClusters(resolution=0.2) %>%
    %>%>% FindClusters(resolution=0.3) %>%>% FindClusters(resolution=0.4)
colnames(obj.ESCA.nkt@meta.data) <- gsub("RNA_snn_res.0.", "r0", colnames(obj.
    ESCA.nkt@meta.data))

```

```

[ ]: ## view marker
marker_list <- c('CD3D', 'CD3E', 'CD3G', 'TRDC',
                 'CD4', 'FOXP3', 'CTLA4',
                 'CD8A', 'CD8B', 'CD28', 'GZMA', 'GZMH',
                 'TIGIT', 'PDCD1', 'TCF7',
                 'GNLY', 'NKG7', 'KLRD1', 'NCAM1', 'FCGR3A', 'PRF1',
                 'MKI67', 'TOP2A', 'STMN1', 'TOX')

options(repr.plot.height = 6, repr.plot.width = 12)
VlnPlot(obj.ESCA.nkt, group.by = 'r03', features = marker_list, pt.size = 0,
        cols = pal_igv("default")(51),
        slot = 'data', assay = 'RNA', raster=FALSE, stack = TRUE, flip = TRUE) +
theme(legend.position = 'none') |
DimPlot_scCustom(obj.ESCA.nkt, pt.size = 1, group.by = "r03", label = TRUE,
                  label.size = 8, colors_use = pal_igv("default")(51))

```

```

[ ]: Idents(obj.ESCA.nkt) <- factor(obj.ESCA.nkt$r03, levels = 9:0)
cluster_deg <- FindAllMarkers(obj.ESCA.nkt, assay = "RNA", slot = "data",
                               logfc.threshold = 0.25, min.pct = 0.1, test.use =
                               "wilcox")
rownames(cluster_deg) <- NULL

```

```

[ ]: obj.ESCA.nkt@meta.data <- obj.ESCA.nkt@meta.data %>%
        mutate(dCT = case_when(r03 %in% c('0') ~ 'Treg',
                               r03 %in% c('5') ~ 'NK',
                               r03 %in% c('8') ~ 'NKT',
                               r03 %in% c('1', '2', '7') ~
                               'CD8T',
                               r03 %in% c('6') ~
                               'CD8T_proliferation',
                               r03 %in% c('3') ~ 'CD4Tnaive',
                               r03 %in% c('4') ~ 'CD4Tfh',
                               r03 %in% c('9') ~ 'CD4T',
                               TRUE ~ 'Others'
        )) %>%
        mutate(mCT = case_when(r03 %in% c('0') ~ 'Treg',
                               r03 %in% c('5') ~ 'NK',
                               r03 %in% c('8') ~ 'NKT',
                               r03 %in% c('1', '2', '6', '7') ~
                               'CD8T',
                               ~ 'CD8T',
                               r03 %in% c('3') ~ 'CD4Tnaive',
                               r03 %in% c('4') ~ 'CD4Tfh',
                               r03 %in% c('9') ~ 'CD4T',
                               TRUE ~ 'Others'
        ))

```

```

r03 %in% c('3', '4', '9') ~_
↳ 'CD4T',
      TRUE ~ 'Others'
    ))
obj.ESCA.nkt@meta.data <- obj.ESCA.nkt@meta.data %>% dplyr::select(-c('r01',_
↳ 'r02', 'r03', 'r04', 'seurat_clusters'))

```

```

[ ]: options(repr.plot.height = 4, repr.plot.width = 16)
DimPlot_scCustom(obj.ESCA.nkt, pt.size = 1, group.by = "r03", label = TRUE,_
↳ label.size = 8, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA.nkt, pt.size = 1, group.by = "mCT", label = TRUE,_
↳ label.size = 8, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA.nkt, pt.size = 1, group.by = "dCT", label = TRUE,_
↳ label.size = 8, colors_use = pal_igv("default")(51))

```

3.4 assign B/Plasma

```

[ ]: obj.ESCA.bp <- obj.ESCA %>% subset(dCT == 'B/Plasma')
obj.ESCA.bp

obj.ESCA.bp <- obj.ESCA.bp %>%
  NormalizeData(normalization.method = "LogNormalize", scale.-
↳ factor = 10000, verbose = F) %>%
  FindVariableFeatures(selection.method = "vst", nfeatures =_
↳ 2000, verbose = F) %>%
  ScaleData(vars.to.regress = c("nCount_RNA"), verbose = F) %>%
  RunPCA(verbose = F) %>%
  RunHarmony(group.by.vars = "SampleID", plot_convergence = TRUE)
## cluster
nPC <- min(PC_selection_harmony(obj.ESCA.bp)$PCselect)
obj.ESCA.bp <- obj.ESCA.bp %>%
  RunUMAP(reduction = "harmony", dims = 1:nPC, umap.method =_
↳ "uwot") %>%
  FindNeighbors(reduction = "harmony", dims = 1:nPC) %>%
  FindClusters(resolution=0.1) %>% FindClusters(resolution=0.2)_
↳ %>% FindClusters(resolution=0.3)
colnames(obj.ESCA.bp@meta.data) <- gsub("RNA_snn_res.0.", "r0", colnames(obj.-
↳ ESCA.bp@meta.data))

```

```

[ ]: ## marker expression
marker_list <- c('PTPRC', 'CD19', 'CD79A', 'MS4A1', 'IGHD', 'IGHM', 'FCER2',_
↳ 'CD72',
  'JCHAIN', 'MZB1', 'IGHA1', 'IGHG1',
  'RGS13', 'MEF2B', 'BCL6', 'MKI67', 'TOP2A')

options(repr.plot.height = 4, repr.plot.width = 10)

```

```
VlnPlot(obj.ESCA.bp, group.by = 'r01', features = marker_list, pt.size = 0,
  ↪cols = pal_igv("default")(51),
    slot = 'data', assay = 'RNA', raster=FALSE, stack = TRUE, flip = TRUE) +
theme(legend.position = 'none') |
DimPlot_scCustom(obj.ESCA.bp, pt.size = 1, reduction = 'umap', group.by =
  ↪"r01", label = TRUE, label.size = 8, colors_use = pal_igv("default")(51))
```

```
[ ]: Idents(obj.ESCA.bp) <- factor(obj.ESCA.bp$r01, levels = 4:0)
cluster_deg <- FindAllMarkers(obj.ESCA.bp, assay = "RNA", slot = "data",
  logfc.threshold = 0.25, min.pct = 0.1, test.use =
  ↪"wilcox")
rownames(cluster_deg) <- NULL
```

```
[ ]: obj.ESCA.bp@meta.data <- obj.ESCA.bp@meta.data %>%
  mutate(dCT = case_when(r01 %in% c('0', '2') ~ 'B',
    r01 %in% c('1') ~ 'Plasma',
    r01 %in% c('3') ~
      ↪'B_GerminalCenter',
    r01 %in% c('4') ~ 'Others',
    TRUE ~ 'Others'
  )) %>%
  mutate(mCT = case_when(r01 %in% c('0', '2', '3') ~
    ↪'B',
    r01 %in% c('1', '4') ~
      ↪'Plasma',
    TRUE ~ 'Others'
  ))
obj.ESCA.bp@meta.data <- obj.ESCA.bp@meta.data %>% dplyr::select(-c('r02',
  ↪'r03', 'seurat_clusters'))
```

```
[ ]: options(repr.plot.height = 3, repr.plot.width = 12)
DimPlot_scCustom(obj.ESCA.bp, pt.size = 1, group.by = "r01", label = TRUE,
  ↪label.size = 5, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA.bp, pt.size = 1, group.by = "mCT", label = TRUE,
  ↪label.size = 5, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA.bp, pt.size = 1, group.by = "dCT", label = TRUE,
  ↪label.size = 5, colors_use = pal_igv("default")(51))
```

3.5 assign Myeloid

```
[ ]: obj.ESCA.mye <- obj.ESCA %>% subset(dCT == 'Myeloid')
obj.ESCA.mye

obj.ESCA.mye <- obj.ESCA.mye %>%
  NormalizeData(normalization.method = "LogNormalize", scale.
  ↪factor = 10000, verbose = F) %>%
```

```

    FindVariableFeatures(selection.method = "vst", nfeatures = □
    ↵3000, verbose = F) %>%
      ScaleData(vars.to.regress = c("nCount_RNA"), verbose = F) %>%
      RunPCA(verbose = F) %>%
      RunHarmony(group.by.vars = "SampleID", plot_convergence = TRUE)
## cluster
nPC <- min(PC_selection_harmony(obj.ESCA.mye)$PCselect)
obj.ESCA.mye <- obj.ESCA.mye %>%
  RunUMAP(reduction = "harmony", dims = 1:nPC, umap.method = □
  ↵"uwot") %>%
  FindNeighbors(reduction = "harmony", dims = 1:nPC) %>%
  FindClusters(resolution=0.1) %>% FindClusters(resolution=0.2) □
  ↵%>% FindClusters(resolution=0.3)
colnames(obj.ESCA.mye@meta.data) <- gsub("RNA_snn_res.0.", "r0", colnames(obj.
  ↵ESCA.mye@meta.data))

```

```

[ ]: ## marker expression
marker_list <- c('PTPRC', 'CD163', 'CD68', 'S100A8', 'C1QA', 'APOC1', □
  ↵'ITGAX', 'SPP1', 'MRC1', 'SLC40A1', ## Mph
  'CD14', 'FCGR3A', 'FCN1', 'VCAN', ## monocyte
  'CSF3R', 'S100A8', 'S100A9', ## neutrophil
  'MS4A2', 'TPSAB1', 'TPSB2', ## mast
  'CD1C', 'IDO1', 'CLEC4C', 'CSF2RA', 'LAMP3', 'CLEC10A', □
  ↵'CD1A', 'LILRA4' ## DC
  )

options(repr.plot.height = 6, repr.plot.width = 12)
VlnPlot(obj.ESCA.mye, group.by = 'r01', features = marker_list, pt.size = 0, □
  ↵cols = pal_igv("default")(51),
  slot = 'data', assay = 'RNA', raster=FALSE, stack = TRUE, flip = TRUE)+
theme(legend.position = 'none')|
DimPlot_scCustom(obj.ESCA.mye, pt.size = 1, reduction = 'umap', group.by = □
  ↵"r01", label = TRUE, label.size = 8, colors_use = pal_igv("default")(51))

```

```

[ ]: Idents(obj.ESCA.mye) <- factor(obj.ESCA.mye$r01, levels = 9:0)
cluster_deg <- FindAllMarkers(obj.ESCA.mye, assay = "RNA", slot = "data",
  logfc.threshold = 0.25, min.pct = 0.1, test.use = □
  ↵"wilcox")
rownames(cluster_deg) <- NULL

```

```

[ ]: obj.ESCA.mye@meta.data <- obj.ESCA.mye@meta.data %>%
  mutate(dCT = case_when(r01 %in% c('0', '9') ~ 'Mph',
  r01 %in% c('1') ~ 'Mono',
  r01 %in% c('2', '7', '8') ~ □
  ↵'cDC',
  r01 %in% c('3') ~ 'Mast',

```

```

r01 %in% c('4') ~ 'pDC',
r01 %in% c('5') ~ 'tDC', ##_
`->tolerogenic DC

`->'Mph_proliferation',
`->'Mph',
`->'8') ~ 'DC',
`->'9', '6') ~_
TRUE ~ 'Others'
)) %>%
mutate(mCT = case_when(r01 %in% c('0', '9', '6') ~_
r01 %in% c('1') ~ 'Mono',
r01 %in% c('2', '4', '5', '7',_
`->'8') ~ 'DC',
r01 %in% c('3') ~ 'Mast',
TRUE ~ 'Others'
))
obj.ESCA.mye@meta.data <- obj.ESCA.mye@meta.data %>% dplyr::select(-c('r02',_
`->'r03', 'seurat_clusters'))

```

```

[ ]: options(repr.plot.height = 4, repr.plot.width = 15)
DimPlot_scCustom(obj.ESCA.mye, pt.size = 1, group.by = "r01", label = TRUE,_
`->label.size = 5, colors_use = pal_igv("default")(51)|
DimPlot_scCustom(obj.ESCA.mye, pt.size = 1, group.by = "mCT", label = TRUE,_
`->label.size = 5, colors_use = pal_igv("default")(51)|
DimPlot_scCustom(obj.ESCA.mye, pt.size = 1, group.by = "dCT", label = TRUE,_
`->label.size = 5, colors_use = pal_igv("default")(51))

```

3.6 assign mCT

```

[ ]: obj.ESCA.nkt@meta.data <- obj.ESCA.nkt@meta.data %>% dplyr::select(-c('r03',_
`->'r04'))
obj.ESCA.bp@meta.data <- obj.ESCA.bp@meta.data %>% dplyr::select(-c('r01'))
obj.ESCA.mye@meta.data <- obj.ESCA.mye@meta.data %>% dplyr::select(-c('r01'))
table(names(obj.ESCA.nkt@meta.data) == names(obj.ESCA.bp@meta.data))
table(names(obj.ESCA.nkt@meta.data) == names(obj.ESCA.mye@meta.data))

```

```

[ ]: obj.ESCA.others <- obj.ESCA %>% subset(dCT %in% c('NK/T', 'B/Plasma',_
`->'Myeloid') == FALSE)
obj.ESCA.others@meta.data <- obj.ESCA.others@meta.data %>% transform(mCT = dCT)
table(names(obj.ESCA.others@meta.data) == names(obj.ESCA.nkt@meta.data))

```

```

[ ]: obj.ESCA <- merge(obj.ESCA.others, c(obj.ESCA.nkt, obj.ESCA.bp, obj.ESCA.mye))
obj.ESCA

```

3.7 assign gCT

```
[ ]: obj.ESCA@meta.data <- obj.ESCA@meta.data %>%
      mutate(gCT = case_when(mCT %in% c('Epi') ~ 'Tumor',
                             mCT %in% c('DC', 'Mono', 'Mph', 'Mast', 'CD8T', 'Treg', 'CD4T', 'NK', 'NKT', 'Plasma', 'B') ~ 'Immune',
                             mCT %in% c('Endo', 'Fibro', 'FRC', 'Pericyte') ~ 'Stromal',
                             TRUE ~ 'Others'
      ))
```

4 UMAP visualization

```
[ ]: obj.ESCA <- obj.ESCA %>%
      NormalizeData(normalization.method = "LogNormalize", scale.factor = 10000, verbose = F) %>%
      FindVariableFeatures(selection.method = "vst", nfeatures = 3000, verbose = F) %>%
      ScaleData(vars.to.regress = c("nCount_RNA"), verbose = F) %>%
      RunPCA(verbose = F) %>%
      RunHarmony(group.by.vars = "SampleID", plot_convergence = TRUE)
## cluster
nPC <- min(PC_selection_harmony(obj.ESCA)$PCselect)
obj.ESCA <- obj.ESCA %>%
      RunUMAP(reduction = "harmony", dims = 1:nPC, umap.method = "uwot")
```

```
[ ]: options(repr.plot.height = 5, repr.plot.width = 30)
select <- 'umap'
DimPlot_scCustom(obj.ESCA, pt.size = 1, reduction = select, group.by = "gCT", label = TRUE, label.size = 5, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA, pt.size = 1, reduction = select, group.by = "mCT", label = TRUE, label.size = 5, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA, pt.size = 1, reduction = select, group.by = "oCT", label = TRUE, label.size = 5, colors_use = pal_igv("default")(51)) |
DimPlot_scCustom(obj.ESCA, pt.size = 1, reduction = select, group.by = "SampleType", label = TRUE, label.size = 5, colors_use = pal_igv("default")(51))
```

```
[ ]: obj.ESCA@meta.data[,c('SampleID', 'SampleType', 'Sex')] %>% .[!duplicated(. $SampleID),] %$% table($.SampleType, .$Sex)
```

5 save

```
[ ]: saveRDS(obj.ESCA, 'obj.ESCA.use.rds')
```