

ccRCC_Hu2024_process

December 25, 2025

1 load data

```
[ ]: obj.ccRCC <- readRDS('/project/sex_cancer/data/ccRCC_Hu2024/snRNA.rds')
```

2 modify meta.data

```
[ ]: info <- read_xlsx('/project/sex_cancer/data/ccRCC_Hu2024/
  ↳ccRCC_Hu2024PatientInfo.xlsx', skip = 1)

names(info)[1] <- 'sample'
info1 <- info[info$sample%in%unique(obj.ccRCC@meta.data$sample),]
info1$Sex=gsub('male','M',info1$Sex)
info1$Sex=gsub('female','F',info1$Sex)

meta <- obj.ccRCC@meta.data %>% transform(barcode = rownames(.))
meta <- merge(meta, info1, by = 'sample',all = TRUE) %>%
  ↳column_to_rownames('barcode') %>% .[colnames(obj.ccRCC),]
```

```
[ ]: # modify meta.data
obj.ccRCC@meta.data <- meta@meta.data %>%
  transform(barcode = rownames(.), Cohort =
  ↳'ccRCC_Hu2024', orig.ident = sample) %>%
  dplyr::rename(c('SampleID' = 'sample', 'percent.mt' =
  ↳'percent.mito')) %>%
  dplyr::select(-c('nUMI', 'nGene', 'class'))
```

```
[ ]: ## add sample info
sample_info <- openxlsx::read.xlsx('/project/sex_cancer/data/ccRCC_Hu2024/
  ↳41588_2024_1662_MOESM4_ESM.xlsx', startRow = 2, rowNames = TRUE)
sample_info <- sample_info %>%
  transform(SampleID = rownames(.)) %>%
  subset(SampleID %in% obj.ccRCC$SampleID) %>%
  dplyr::rename(c('Sex' = 'sex'))

obj.ccRCC@meta.data <- obj.ccRCC@meta.data %>%
  merge(sample_info, by = 'SampleID', all = TRUE) %>%
```

```
transform(SampleType = 'tumor', DonorID = SampleID) %>%
  `rownames<-`(`.$barcode)
```

```
[ ]: options(repr.plot.height = 5, repr.plot.width = 25)
DimPlot_scCustom(obj.ccRCC, pt.size = 1, group.by = "celltype", reduction = "umap", label = T, label.size = 4, colors_use = pal_igv("default")(51)) |
  DimPlot_scCustom(obj.ccRCC, pt.size = 1, group.by = "celltype2", reduction = "umap", label = F, colors_use = pal_igv("default")(51))
```

3 cell type annotation

3.1 assign oCT

```
[ ]: obj.ccRCC@meta.data <- obj.ccRCC@meta.data %>%
  transform(oCT = celltype2)
```

3.2 check annotation

check cell type annotation provided in the original research via COSG

```
[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.ccRCC
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.
  factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
    mu = 10, ## The penalty factor to penalize gene expression in
    cells not belonging to the cluster of interest
    n_genes_user = 50, # Number of top ranked genes returned in the
    result
    remove_lowly_expressed=T, # If TRUE, genes that express a
    percentage of target cells smaller than a specific value (expressed_pct) are
    not considered as marker genes for the target cells. The default value is
    TRUE.
    expressed_pct=0.1) # If TRUE, genes that express a percentage of
    target cells smaller than a specific value (expressed_pct) are not
    considered as marker genes for the target cells.
marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
  rename(c("oCT" = "variable", "marker" = "value")),
  marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
  select(-"variable") %>% dplyr::rename(c("COSGscore" = "value"))) %>%
```

```

mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
↳ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
  check <- oCT_marker %>% subset(oCT == x & marker %in%
↳marker_annotation[[x]])
  ifelse(nrow(check) == 0, print(x), return(check))
})

```

3.3 assign mCT

```

[ ]: obj.ccRCC@meta.data <- obj.ccRCC@meta.data %>%
mutate(mCT = case_when(celltype %in% c('Malignant') ~
↳'Epi',
celltype2 %in% c('1_B',
↳'2_Plasma') ~ 'B',
celltype2 %in% c('1_CD4-Tcm/
↳Th', '1_CD4-Th') ~ 'CD4T',
celltype2 %in% c('1_CD4-Treg') ~
↳'Treg',
celltype2 %in%
↳c('2_CD8-cycling', '2_CD8-ITGAE', '2_CD8-Tex') ~ 'CD8T',
celltype2 %in% c('3_MAIT') ~
↳'MAIT',
celltype2 %in% c('4_NK-KLRC1',
↳'4_Temra/aNK') ~ 'NK',
celltype2 %in% c('4_Macro01',
↳'4_Macro02-IGF1', '4_Macro03-LILRB5', '4_Macro04-GPNMB',
'4_Macro05-NR4A3', '4_Macro06-C3', '4_Macro07-GBP1', '5_Macro-prolif') ~
↳'Mph',
celltype2 %in% c('6_DC-cDC1',
↳'6_DC-cDC2', '6_DC-pDC', '6_DC-mDC') ~ 'DC',
celltype2 %in%
↳c('3_Mono01-SMIM25', '3_Mono02-VCAN') ~ 'Mono',
celltype2 %in% c('7_Mast') ~
↳'Mast',
celltype2 %in% c('peri') ~
↳'Pericyte',
celltype2 %in% c('EC_Cap01-F8',
↳'EC_artery', 'EC_Cap04-NRP2', 'EC_low UMI', 'EC-cycling', 'EC_vein',

```

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↪ 'EC_Cap03-IGFBP5', 'EC_Cap02', 'EC_Cap07-DNASE1L3', 'EC_Cap05-KIT', ↪
↪ 'EC_Cap06-WT1', 'EC_Cap-CD36') ~ 'Endo',

celltype2 %in% c('Fib') ~ ↪
↪ 'Fibro',

celltype2 %in% c('SMC') ~ 'SMC',
TRUE ~ 'Others'
))

```

3.4 assign gCT

```

[ ]: obj.ccRCC@meta.data <- obj.ccRCC@meta.data %>%
      mutate(gCT = case_when(mCT %in% c('Tumor') ~ 'Tumor',
                             mCT %in% c('B', 'CD4T', 'Treg', ↪
                             ↪ 'CD8T', 'MAIT', 'NK', 'Mph', 'DC', 'Mono', 'Mast') ~ 'Immune',
                             mCT %in% c('Pericyte', 'Endo', ↪
                             ↪ 'Fibro', 'SMC') ~ 'Stromal',
                             TRUE ~ 'Others'
                             ))

```

```

[ ]: options(repr.plot.height = 5, repr.plot.width = 30)
DimPlot_scCustom(obj.ccRCC, pt.size = 1, group.by = "gCT", reduction = 'umap', ↪
↪ label = T, label.size = 4, colors_use = pal_igv("default")(51))|
DimPlot_scCustom(obj.ccRCC, pt.size = 1, group.by = "mCT", reduction = 'umap', ↪
↪ label = T, label.size = 4, colors_use = pal_igv("default")(51))|
DimPlot_scCustom(obj.ccRCC, pt.size = 1, group.by = "oCT", reduction = 'umap', ↪
↪ label = F, colors_use = pal_igv("default")(51))

```

```

[ ]: meta_now[,c('SampleType', 'Sex', 'SampleID')] %>% .[!duplicated(.$SampleID),] ↪
↪ %$% table(.$SampleType, .$Sex)
table(meta_now$SampleType)

```

4 save

```

[ ]: saveRDS(obj.ccRCC, 'obj.ccRCC.use.rds')

```