

# PDAC\_Hwang2022\_process

December 25, 2025

## 1 Of note

content in obj.PDAC@assays\$RNA@counts: “UMI counts were normalized by the total number of UMIs per nucleus and converted to transcripts-per-10,000 (TP10K) as the final expression unit”

## 2 load data

```
[ ]: ## gene info
feature <- read.delim('/project/sex_cancer/data/PDAC_Hwang2022/Group1/genes.
  ↳txt', header = F, row.names = NULL)

## obj1
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group1/
  ↳Exp_data_TP10K_1.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group1/Cells1.csv')_
  ↳%>% transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj1 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")

## obj2
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group2/
  ↳Exp_data_TP10K_2.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group2/Cells2.csv')_
  ↳%>% transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj2 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")

## obj2
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group3/
  ↳Exp_data_TP10K_3.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group3/Cells3.csv')_
  ↳%>% transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj3 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")
```

```
## all merge into 1
obj.PDAC <- merge(obj1, c(obj2, obj3))
```

```
[ ]: info <- read_xlsx('/project/sex_cancer/data/PDAC_Hwang2022/
  ↳ PDAC_WilliamL2022_PatientInfo.xlsx', skip = 1)
info1 <- info[grepl('PDAC_', info$ID),]

meta <- obj.PDAC@meta.data %>% transform(barcode2 = rownames(.))
sample=apply(info1$ID, function(x){
  parts=strsplit(x, split = '_')[[1]]
  new=paste(parts[2:3], collapse = '')
  return(new)
})
info2 <- cbind(sample, info1) %>%
  .[which(.$sample %in% meta$sample),]
obj.PDAC@meta.data <- merge(meta, info2, by = 'sample', all = TRUE) %>%
  ↳ column_to_rownames('barcode2') %>% .[colnames(obj.PDAC),]
```

### 3 modify meta.data

```
[ ]: ## remove useless meta.data
obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
  dplyr::select(-c('orig.ident', 'complexity', 'umap1',
  ↳ 'umap2', 'g1s_score', 'g2m_score', 'cell_cycle_phase', 'nFeature_RNA',
  ↳ 'mp_top', 'mp_top_score',
  ↳ 'mp_assignment', 'mCT', 'disease'))

[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
  dplyr::rename(c('Chemistry' = '10x Chemistry',
  ↳ 'SampleID' = 'sample', 'Disease' = 'source')) %>%
  mutate(Chemistry = case_when(Chemistry == 'v2' ~ "10x
  ↳ v2",
  ↳ Chemistry == 'v3' ~ "10x
  ↳ v3",
  ↳ TRUE ~ 'Others')) %>%
  mutate(Sex = case_when(Sex == 'Female' ~ 'F', Sex ==
  ↳ 'Male' ~ 'M', TRUE ~ 'Others')) %>%
  transform(Cohort = 'PDAC_Hwang2022')
```

## 4 filter sample

```
[ ]: obj.PDAC <- obj.PDAC %>% subset(Neoadjuvant == 'None')
obj.PDAC@meta.data <- obj.PDAC@meta.data %>% transform(SampleType = 'tumor')
```

## 5 UMAP visualization

```
[ ]: obj.h5ad <- anndata::read_h5ad('/project/sex_cancer/data/PDAC_Hwang2022/
  ↪GSE202051_totaldata-final-toshare.h5ad')
umap_emb <- obj.h5ad$obs$X_umap %>% as.data.frame() %>%
  `rownames<-`(rownames(obj.h5ad$X)) %>% `colnames<-`(c('umap_1',
  ↪'umap_2'))
umap_emb <- umap_emb %>% .[colnames(obj.PDAC),] %>% as.matrix()
## add UMAP embeddings
Idents(obj.PDAC) <- obj.PDAC$barcode
obj.PDAC[['umap']] <- CreateDimReducObject(embeddings = umap_emb[colnames(obj.
  ↪PDAC),] , key = 'umap_', assay = 'RNA')
```

## 6 cell type annotation

### 6.1 assign oCT

```
[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
  mutate(oCT = cell_subtype) %>%
  mutate(dCT = case_when(cell_subtype %in% c('Ductal',
  ↪(atypical)', 'Ductal', 'Acinar', 'ADM') ~ 'Epi',
  cell_subtype %in% c('Malignant') ~
  ↪'Epi',
  cell_subtype %in% c('CD8+ T') ~
  ↪'CD8T',
  cell_subtype %in% c('CD4+ T') ~
  ↪'CD4T',
  cell_subtype %in% c('Treg') ~
  ↪'Treg',
  cell_subtype %in% c('NK_cell') ~
  ↪'NK',
  cell_subtype %in% c('B_cell') ~
  ↪'B',
  cell_subtype %in% c('Plasma') ~
  ↪'Plasma',
  cell_subtype %in% c('Macrophage') ~
  ↪~ 'Mph',
  cell_subtype %in% c('Dendritic') ~
  ↪'DC',
```

```

cell_subtype %in% c('Neutrophil')
↪ '~ 'Neu',
cell_subtype %in% c('Mast') ~
↪ 'Mast',
cell_subtype %in% c('myCAF',
↪ 'CAF') ~ 'Fibro',
cell_subtype %in% c('Pericyte') ~
↪ 'Pericyte',
cell_subtype %in% c('Vascular',
↪ 'Lymphatic') ~ 'Endo',
cell_subtype %in% c('Schwann') ~
↪ 'Schwann',
cell_subtype %in% c('Alpha',
↪ 'Beta', 'Delta', 'Epsilon', 'Gamma', 'Hormone-negative neuroendocrine',
↪ 'Intra-pancreatic neurons') ~ 'Neuron',
cell_subtype %in% c('Adipocyte') ~
↪ 'Adipocyte',
cell_subtype %in% c('Vascular
↪ smooth muscle') ~ 'VSMC',
TRUE ~ 'Others'))

```

## 6.2 check annotation

check cell type annotation provided in the original research via COSG

```

[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.PDAC
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.
↪ factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
  mu = 10, ## The penalty factor to penalize gene expression in
↪ cells not belonging to the cluster of interest
  n_genes_user = 50, # Number of top ranked genes returned in the
↪ result
  remove_lowly_expressed=T, # If TRUE, genes that express a
↪ percentage of target cells smaller than a specific value (expressed_pct) are
↪ not considered as marker genes for the target cells. The default value is
↪ TRUE.

```

```

        expressed_pct=0.1) # If TRUE, genes that express a percentage of
        ↪target cells smaller than a specific value (expressed_pct) are not
        ↪considered as marker genes for the target cells.
marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
        ↪rename(c("oCT" = "variable", "marker" = "value")),
        marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
        ↪select(-"variable") %>% dplyr::rename(c("COSGscore" = "value"))) %>%
        mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
        ↪ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
        check <- oCT_marker %>% subset(oCT == x & marker %in%
        ↪marker_annotation[[x]])
        ifelse(nrow(check) == 0, print(x), return(check))
})

```

### 6.3 assign mCT

```

[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
        mutate(mCT = case_when(cell_subtype %in% c('Ductal
        ↪(atypical)', 'Ductal', 'Acinar', 'ADM') ~ 'Epi',
        cell_subtype %in% c('Malignant') ~
        ↪'Epi',

        cell_subtype %in% c('CD8+ T') ~
        ↪'CD8T',
        cell_subtype %in% c('CD4+ T') ~
        ↪'CD4T',
        cell_subtype %in% c('Treg') ~
        ↪'Treg',
        cell_subtype %in% c('NK_cell') ~
        ↪'NK',
        cell_subtype %in% c('B_cell',
        ↪'Plasma') ~ 'B',

        cell_subtype %in% c('Macrophage')
        ↪~ 'Mph',
        cell_subtype %in% c('Dendritic') ~
        ↪'DC',
        cell_subtype %in% c('Neutrophil')
        ↪~ 'Neu',
        cell_subtype %in% c('Mast') ~
        ↪'Mast',

```

```

↪ 'CAF') ~ 'Fibro',
cell_subtype %in% c('myCAF',
↪ 'Pericyte',
cell_subtype %in% c('Pericyte') ~
↪ 'Lymphatic') ~ 'Endo',
cell_subtype %in% c('Vascular',
↪ 'Schwann',
cell_subtype %in% c('Schwann') ~
↪ 'Beta', 'Delta', 'Epsilon', 'Gamma', 'Hormone-negative neuroendocrine',
↪ 'Intra-pancreatic neurons') ~ 'Neuron',
cell_subtype %in% c('Alpha',
↪ 'Adipocyte',
cell_subtype %in% c('Adipocyte') ~
↪ smooth muscle') ~ 'SMC',
cell_subtype %in% c('Vascular',
TRUE ~ 'Others'))

```

## 6.4 assign gCT

```

[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
      mutate(gCT = case_when(mCT %in% c('Epi') ~ 'Tumor',
                             mCT %in% c('CD8T', 'CD4T', 'Treg',
↪ 'NK', 'B', 'Mph', 'DC', 'Neu', 'Mast') ~ 'Immune',
                             mCT %in% c('Fibro', 'Pericyte',
↪ 'Endo', 'Schwann', 'Neuron', 'Adipocyte', 'SMC') ~ 'Stromal',
                             TRUE ~ 'Others'))

```

## 7 save

```

[ ]: saveRDS(obj.PDAC, 'obj.PDAC.use.rds')

```