

PDAC_Hwang2022_process

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1 Of note

content in obj.PDAC@assays\$RNA@counts: “UMI counts were normalized by the total number of UMIs per nucleus and converted to transcripts-per-10,000 (TP10K) as the final expression unit”

2 load data

```
[ ]: ## gene info
feature <- read.delim('/project/sex_cancer/data/PDAC_Hwang2022/Group1/genes.
˓→txt', header = F, row.names = NULL)

## obj1
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group1/
˓→Exp_data_TP10K_1.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group1/Cells1.csv') %>%
  transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj1 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")

## obj2
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group2/
˓→Exp_data_TP10K_2.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group2/Cells2.csv') %>%
  transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj2 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")

## obj3
exp <- Matrix::readMM('/project/sex_cancer/data/PDAC_Hwang2022/Group3/
˓→Exp_data_TP10K_3.mtx')
meta <- read.csv('/project/sex_cancer/data/PDAC_Hwang2022/Group3/Cells3.csv') %>%
  transform(barcode = cell_name) %>% column_to_rownames('cell_name')
colnames(exp) <- rownames(meta)
rownames(exp) <- feature$V1
obj3 <- CreateSeuratObject(counts = exp, meta.data = meta, project = "PDAC")
```

```

## all merge into 1
obj.PDAC <- merge(obj1, c(obj2, obj3))

[ ]: info <- read_xlsx('/project/sex_cancer/data/PDAC_Hwang2022/
    ↪PDAC_WilliamL2022_PatientInfo.xlsx',skip = 1)
info1 <- info[grep('PDAC_',info$ID),]

meta <- obj.PDAC@meta.data %>% transform(barcode2 = rownames(.))
sample=sapply(info1$ID,function(x){
    parts=strsplit(x,split = '_')[[1]]
    new=paste(parts[2:3],collapse = '')
    return(new)
})
info2 <- cbind(sample,info1) %>%
    .[which(.sample %in% meta$sample),]
obj.PDAC@meta.data <- merge(meta, info2, by = 'sample',all = TRUE) %>%
    ↪column_to_rownames('barcode2') %>% .[colnames(obj.PDAC),]

```

3 modify meta.data

```

[ ]: ## remove useless meta.data
obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
    dplyr::select(-c('orig.ident', 'complexity', 'umap1',
    ↪'umap2', 'g1s_score', 'g2m_score', 'cell_cycle_phase', 'nFeature_RNA',
    'mp_top', 'mp_top_score',
    ↪'mp_assignment', 'mCT', 'disease'))

[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
    dplyr::rename(c('Chemistry' = '10x Chemistry',
    ↪'SampleID' = 'sample', 'Disease' = 'source')) %>%
    mutate(Chemistry = case_when(Chemistry == 'v2' ~ "10x",
    ↪3' v2",
    Chemistry == 'v3' ~ "10x",
    ↪3' v3",
    TRUE ~ 'Others')) %>%
    mutate(Sex = case_when(Sex == 'Female' ~ 'F', Sex ==
    ↪'Male' ~ 'M', TRUE ~ 'Others')) %>%
    transform(Cohort = 'PDAC_Hwang2022')

```

4 filter sample

```
[ ]: obj.PDAC <- obj.PDAC %>% subset(Neoadjuvant == 'None')
obj.PDAC@meta.data <- obj.PDAC@meta.data %>% transform(SampleType = 'tumor')
```

5 UMAP visualization

```
[ ]: obj.h5ad <- anndata::read_h5ad('/project/sex_cancer/data/PDAC_Hwang2022/
  ↪GSE202051_totaldata-final-toshare.h5ad')
umap_emb <- obj.h5ad$obsm$X_umap %>% as.data.frame() %>%
  `rownames<-`(`rownames(obj.h5ad$X)` %>% `colnames<-`(`c('umap_1', ↪
  ↪'umap_2')))
umap_emb <- umap_emb %>% .[colnames(obj.PDAC),] %>% as.matrix()
## add UMAP embeddings
Idents(obj.PDAC) <- obj.PDAC$barcode
obj.PDAC[['umap']] <- CreateDimReducObject(embeddings = umap_emb[colnames(obj.
  ↪PDAC),], key = 'umap_', assay = 'RNA')
```

6 cell type annotation

6.1 assign oCT

```
[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
  mutate(oCT = cell_subtype) %>%
  mutate(dCT = case_when(cell_subtype %in% c('Ductal',
  ↪(atypical)', 'Ductal', 'Acinar', 'ADM') ~ 'Epi',
  cell_subtype %in% c('Malignant') ~
  ↪'Epi',
  cell_subtype %in% c('CD8+ T') ~
  ↪'CD8T',
  cell_subtype %in% c('CD4+ T') ~
  ↪'CD4T',
  cell_subtype %in% c('Treg') ~
  ↪'Treg',
  cell_subtype %in% c('NK_cell') ~
  ↪'NK',
  cell_subtype %in% c('B_cell') ~
  ↪'B',
  cell_subtype %in% c('Plasma') ~
  ↪'Plasma',
  cell_subtype %in% c('Macrophage') ~
  ↪~ 'Mph',
  cell_subtype %in% c('Dendritic') ~
  ↪'DC',
```

```

    cell_subtype %in% c('Neutrophil') ~
    ↵~ 'Neu',
    ↵ 'Mast',
    ↵ 'CAF') ~ 'Fibro',
    ↵ 'Pericyte',
    ↵ 'Lymphatic') ~ 'Endo',
    ↵ 'Schwann',
    ↵ 'Beta', 'Delta', 'Epsilon', 'Gamma', 'Hormone-negative neuroendocrine',
    ↵ 'Intra-pancreatic neurons') ~ 'Neuron',
    ↵ 'Adipocyte',
    ↵smooth muscle') ~ 'VSMC',
    TRUE ~ 'Others'))

```

6.2 check annotation

check cell type annotation provided in the original research via COSG

```

[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.PDAC
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale =
  ↵factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
        mu = 10, ## The penalty factor to penalize gene expression in
        ↵cells not belonging to the cluster of interest
        n_genes_user = 50, # Number of top ranked genes returned in the
        ↵result
        remove_lowly_expressed=T, # If TRUE, genes that express a
        ↵percentage of target cells smaller than a specific value (expressed_pct) are
        ↵not considered as marker genes for the target cells. The default value is
        ↵TRUE.

```

```

    expressed_pct=0.1) # If TRUE, genes that express a percentage of
    ↵target cells smaller than a specific value (expressed_pct) are not
    ↵considered as marker genes for the target cells.

marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
    ↵rename(c("oCT" = "variable", "marker" = "value")),
            marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
    ↵select(-"variable") %>% dplyr::rename(c("COSGscore" = "value"))) %>%
        mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
    ↵ext_list(oCT))

oCT_marker <- marker_oCT
oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
    check <- oCT_marker %>% subset(oCT == x & marker %in% ↵
    ↵marker_annotation[[x]])
    ifelse(nrow(check) == 0, print(x), return(check))
})

```

6.3 assign mCT

```
[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
  mutate(mCT = case_when(cell_subtype %in% c('Ductal',
  ~ (atypical)', 'Ductal', 'Acinar', 'ADM') ~ 'Epi',
  cell_subtype %in% c('Malignant') ~
  ~ 'Epi',
  cell_subtype %in% c('CD8+ T') ~
  ~ 'CD8T',
  cell_subtype %in% c('CD4+ T') ~
  ~ 'CD4T',
  cell_subtype %in% c('Treg') ~
  ~ 'Treg',
  cell_subtype %in% c('NK_cell') ~
  ~ 'NK',
  cell_subtype %in% c('B_cell', 'B',
  ~ 'Plasma') ~ 'B',
  cell_subtype %in% c('Macrophage') ~
  ~ 'Mph',
  cell_subtype %in% c('Dendritic') ~
  ~ 'DC',
  cell_subtype %in% c('Neutrophil') ~
  ~ 'Neu',
  cell_subtype %in% c('Mast') ~
  ~ 'Mast')
```

```

    cell_subtype %in% c('myCAF', □
    ↵'CAF') ~ 'Fibro',
    cell_subtype %in% c('Pericyte') ~ □
    ↵'Pericyte',
    cell_subtype %in% c('Vascular', □
    ↵'Lymphatic') ~ 'Endo',
    cell_subtype %in% c('Schwann') ~ □
    ↵'Schwann',
    cell_subtype %in% c('Alpha', □
    ↵'Beta', 'Delta', 'Epsilon', 'Gamma', 'Hormone-negative neuroendocrine', □
    ↵'Intra-pancreatic neurons') ~ 'Neuron',
    cell_subtype %in% c('Adipocyte') ~ □
    ↵'Adipocyte',
    cell_subtype %in% c('Vascular', □
    ↵smooth muscle') ~ 'SMC',
    TRUE ~ 'Others'))

```

6.4 assign gCT

```
[ ]: obj.PDAC@meta.data <- obj.PDAC@meta.data %>%
      mutate(gCT = case_when(mCT %in% c('Epi') ~ 'Tumor',
                             mCT %in% c('CD8T', 'CD4T', 'Treg', □
                             ↵'NK', 'B', 'Mph', 'DC', 'Neu', 'Mast') ~ 'Immune',
                             mCT %in% c('Fibro', 'Pericyte', □
                             ↵'Endo', 'Schwann', 'Neuron', 'Adipocyte', 'SMC') ~ 'Stromal',
                             TRUE ~ 'Others'))
```

7 save

```
[ ]: saveRDS(obj.PDAC, 'obj.PDAC.use.rds')
```