

CRC_Pelka2021_process

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1 load data

```
[ ]: data1 <- load3Cadata('/project/sex_cancer/data/CRC_Pelka2021/Group1')
data2 <- load3Cadata('/project/sex_cancer/data/CRC_Pelka2021/Group2')
data3 <- load3Cadata('/project/sex_cancer/data/CRC_Pelka2021/Group3')
data4 <- load3Cadata('/project/sex_cancer/data/CRC_Pelka2021/Group4')
obj.CRC <- merge(data1, c(data2, data3, data4)) %>% UpdateSeuratObject()
```

```
[ ]: ## add sample info
sample_info <- read.csv('/project/sex_cancer/data/CRC_Pelka2021/
↳GSE178341_crc10x_full_c295v4_submit_metatables.csv')
```

```
[ ]: ## add cell type annotation
cell_anno <- read.csv('/project/sex_cancer/data/CRC_Pelka2021/
↳crc10x_full_c295v4_submit_cluster.csv')
nrow(cell_anno)
cell_anno %>% head(n = 2)
```

2 modify meta.data

```
[ ]: meta_data <- cbind(sample_info %>% transform(barcode = cellID) %>%
↳column_to_rownames('cellID'),
                        cell_anno %>% column_to_rownames('sampleID')) %>%
dplyr::rename(c('SampleID' = 'PatientTypeID', 'DonorID' = 'PID'))
↳%>%
transform(SampleType = ifelse(SPECIMEN_TYPE == 'T', 'tumor',
↳'normal_adjacent')) %>%
transform(Cohort = 'CRC_Pelka2021') %>%
mutate(Chemistry = case_when(SINGLECELL_TYPE == 'SC3Pv2' ~ "10x 3'
↳v2",
                              SINGLECELL_TYPE == 'SC3Pv3' ~ "10x 3'
↳v3",
                              TRUE ~ 'unknown')) %>%
.[colnames(obj.CRC),]
obj.CRC@meta.data <- meta_data
```

```
[ ]: obj.CRC@meta.data[,c('SampleType', 'SampleID', 'Sex', 'MetastasisStatus')] %>% .
  ↪[!duplicated(.$SampleID),] %$% table(.$Sex, .$SampleType, useNA = 'ifany')
obj.CRC@meta.data %$% table(.$SampleType)
```

3 cell type annotation

3.1 assign oCT

```
[ ]: obj.CRC@meta.data <- obj.CRC@meta.data %>%
  dplyr::rename(c('oCT' = 'c1MidwayPr', 'dCT' =
  ↪'c1295v11SubFull'))
```

3.2 check annotation

check cell type annotation provided in the original research via COSG

```
[ ]: ## check marker expression
marker_annotation <- readRDS("marker_annotation.rds")

obj <- obj.CRC
DefaultAssay(obj) <- "RNA"
obj <- obj %>% NormalizeData(normalization.method = "LogNormalize", scale.
  ↪factor = 10000, verbose = F)
Idents(obj) <- ext_list(obj$oCT)

marker_oCT <- obj %>%
  cosg(groups = "all", assay = "RNA", slot = "data",
  mu = 10, ## The penalty factor to penalize gene expression in
  ↪cells not belonging to the cluster of interest
  n_genes_user = 50, # Number of top ranked genes returned in the
  ↪result
  remove_lowly_expressed=T, # If TRUE, genes that express a
  ↪percentage of target cells smaller than a specific value (expressed_pct) are
  ↪not considered as marker genes for the target cells. The default value is
  ↪TRUE.
  expressed_pct=0.1) # If TRUE, genes that express a percentage of
  ↪target cells smaller than a specific value (expressed_pct) are not
  ↪considered as marker genes for the target cells.
marker_oCT <- cbind(marker_oCT[[1]] %>% melt(id.vars = NULL) %>% dplyr::
  ↪rename(c("oCT" = "variable", "marker" = "value")),
  marker_oCT[[2]] %>% melt(id.vars = NULL) %>% dplyr::
  ↪select(-"variable") %>% dplyr::rename(c("COSGscore" = "value"))) %>%
  mutate(Cohort = unique(obj$Cohort)) %>% mutate(oCT =
  ↪ext_list(oCT))

oCT_marker <- marker_oCT
```

```

oCT_list <- unique(oCT_marker$oCT)
lapply(oCT_list, function(x){
  check <- oCT_marker %>% subset(oCT == x & marker %in%
  ↪ marker_annotation[[x]])
  ifelse(nrow(check) == 0, print(x), return(check))
})

```

3.3 assign mCT

```

[ ]: obj.CRC@meta.data <- obj.CRC@meta.data %>%
  mutate(mCT = case_when(oCT %in% c('Epi') ~ 'Epi',
    oCT %in% c('TCD8') ~ 'CD8T',
    (oCT == 'TCD4' & dCT %in% c("cTNIO8",
    ↪ (CD4+ Treg)", "cTNIO9 (CD4+ Treg prolifer)" == FALSE) ~ 'CD4T',
    dCT %in% c("cTNIO8 (CD4+ Treg)",
    ↪ "cTNIO9 (CD4+ Treg prolifer)" ~ 'Treg',
    oCT %in% c('TZBTB16') ~ 'T_ZBTB16',
    oCT %in% c('Tgd') ~ 'T',
    oCT %in% c('Macro') ~ 'Mph',
    oCT %in% c('Granulo') ~ 'Neu',
    oCT %in% c('SmoothMuscle') ~ 'SMC',
    oCT %in% c('Peri') ~ 'Pericyte',
    oCT %in% c('Plasma', 'B') ~ 'B',
    TRUE ~ oCT
  ))
unique(obj.CRC$mCT)

```

3.4 assign gCT

```

[ ]: obj.CRC@meta.data <- obj.CRC@meta.data %>%
  mutate(gCT = case_when(mCT %in% c('Epi') ~ 'Tumor',
    mCT %in% c('CD4T', 'Mph', 'Treg',
    ↪ 'DC', 'NK', 'Mono', 'CD8T', 'B', 'Mast', 'T_ZBTB16', 'T', 'Neu', 'ILC') ~
    ↪ 'Immune',
    mCT %in% c('Pericyte', 'Endo',
    ↪ 'Fibro', 'SMC', 'Schwann') ~ 'Stromal',
    TRUE ~ 'Others'
  ))
table(obj.CRC$mCT, obj.CRC$gCT, useNA = 'ifany')

```

4 UMAP visualization

```
[ ]: # add nCount
nCount_RNA <- colSums(x = obj.CRC, slot = "counts")
obj.CRC <- obj.CRC %>% AddMetaData(metadata = nCount_RNA, col.name = "nCount_RNA")
obj.CRC@meta.data %>% head(n = 2)

[ ]: # run harmony
obj.CRC <- obj.CRC %>%
  NormalizeData(normalization.method = "LogNormalize", scale.factor = 10000, verbose = F) %>%
  FindVariableFeatures(selection.method = "vst", nfeatures = 3000, verbose = F) %>%
  ScaleData(vars.to.regress = c("nCount_RNA"), verbose = F) %>%
  RunPCA(verbose = F) %>%
  RunHarmony(group.by.vars = "SampleID", plot_convergence = TRUE)

[ ]: # run UMAP
nPC <- min(PC_selection_harmony(obj.CRC)$PCselect)
obj.CRC <- obj.CRC %>%
  RunUMAP(reduction = "harmony", dims = 1:nPC, umap.method = "uwot") %>%
  RunTSNE(reduction = "harmony", dims = 1:nPC)

[ ]: options(repr.plot.height = 5, repr.plot.width = 25)
DimPlot_scCustom(obj.CRC, pt.size = 1, group.by = "gCT", label = TRUE, label.size = 4, colors_use = pal_igv("default")(51))|
DimPlot_scCustom(obj.CRC, pt.size = 1, group.by = "mCT", label = TRUE, label.size = 4, colors_use = pal_igv("default")(51))|
DimPlot_scCustom(obj.CRC, pt.size = 1, group.by = "oCT", label = TRUE, label.size = 4, colors_use = paletteer::paletteer_d('palettesForR:Web', 100))
```

5 save

```
[ ]: saveRDS(obj.CRC, "obj.CRC.use.rds")
```