

Nonsynonymous mutation and deletion analysis: A fast and accurate pipeline

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1. Pyfasta (<https://github.com/brentp/pyfasta>): For large dataset (more than 10,000 sequences), we need to differentiate the data into two or more dataset as MAFFT alignment tools has a maximum limit $\leq 10,000$.

Usages (Linux):

2. MAFFT online alignment tools (<https://mafft.cbrc.jp/alignment/server/>): MAFFT is a very user friendly multiple alignment program for amino acid or nucleotide sequences. For SARS-CoV-2, specialize version (https://mafft.cbrc.jp/alignment/server/add_fragments.html?frommanual) has been launched where the maximum limit was 10,000. Here, existing alignment or reference sequences can be selected in a menu and other menu also available for new alignment. Here, in my experiment, I have used first as reference and the second one were for aligning sequence upload.

MAFFT version 7

Multiple alignment program for amino acid or nucleotide sequences



[Download version](#)

[Mac OS X](#)

[Windows](#)

[Linux](#)

[Source](#)

[Online version](#)

[Alignment](#)

[mafft --add](#)

[Merge](#)

[Phylogeny](#)

[Rough tree](#)

[Merits / limitations](#)

[Algorithms](#)

[Tips](#)

[Benchmarks](#)

[Feedback](#)

Add **fragmentary sequence(s)** to existing alignment or sequence [Help](#) [Help for closely-related long data](#)

Existing alignment: [Example](#)

Gaps (-) will be preserved.

or upload a plain text file: No file chosen

[Clear](#)

Zipped file is acceptable.

Fragmentary sequence(s) to be added to the above alignment: [Example](#)

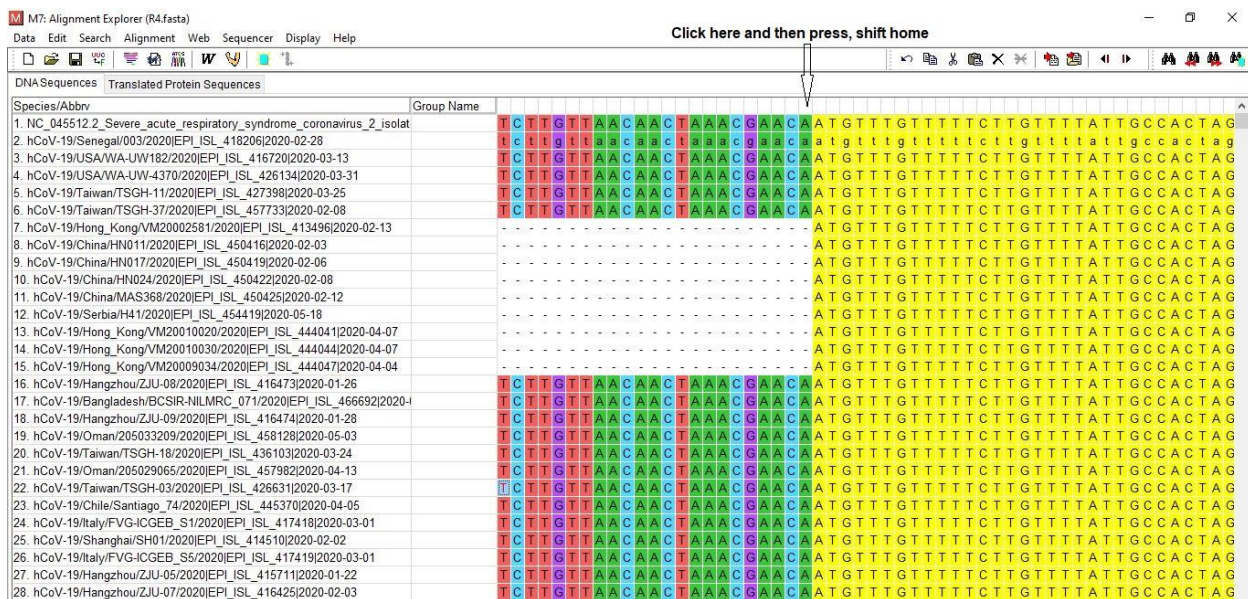
Gaps (if any) will be removed.

or upload a plain text file: No file chosen

[Clear](#)

Zipped file is acceptable.

3. Separate ORFs (S for SARS-CoV-2) from the alignment: I have downloaded the complete S (for example) from reference (<https://www.ncbi.nlm.nih.gov/nuccore/1798174254>) from NCBI and open it in a text editor (Notepad/Notepad++). Open the alignment in MEGA 7 or other version. Copy the first few nucleotide (at least 20) from S and then go to MEGA. Press, ctrl F, then ctrl V and finally press enter. In this, we can find the first section of S Protein in the alignment. Click on the upstream of the S starting and press, shift home, this will select the upstream part of S protein, then delete the upstream part. This will remove the upstream of the S protein. For downstream removal do the same but press, shift end, then delete the downstream part. Insertional sequence must be checked and delete them. If it has importance, preserve the sequence for further analysis.

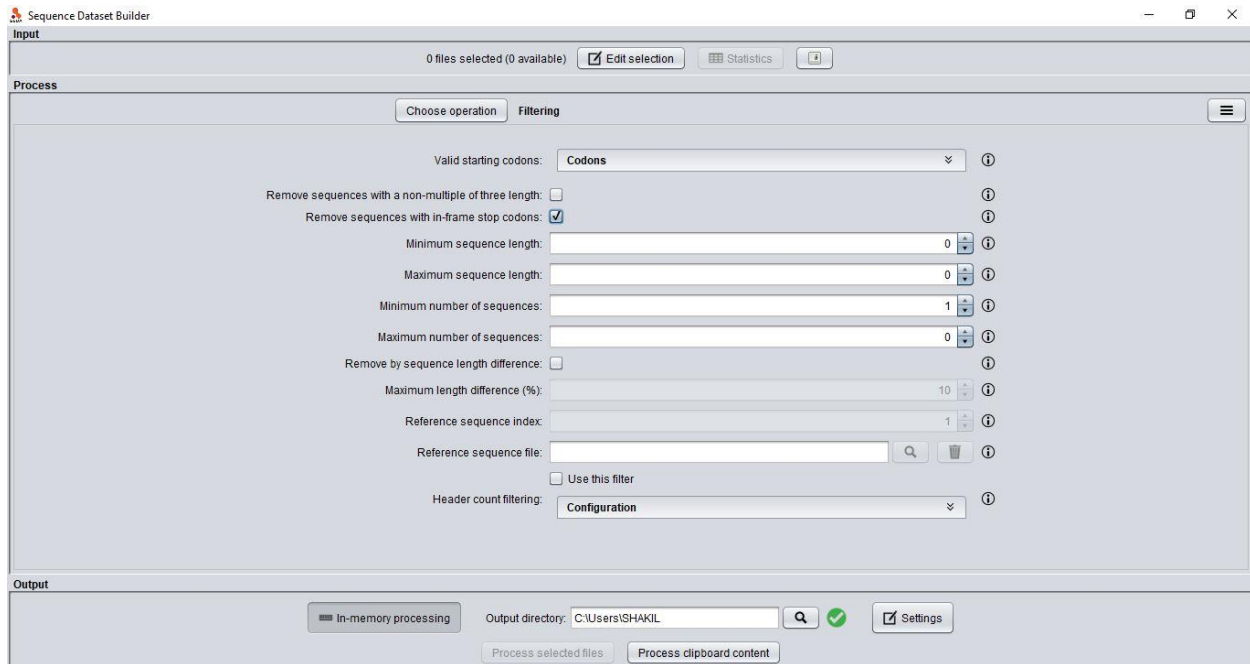


4. Sequence cleaner (<https://github.com/metageni/Sequence-Cleaner>): Sequence cleaner will assist to remove all sequences which contain ambiguous ('M', 'D', 'R', 'N', 'K', 'Y', 'S', 'B', 'H', '-', 'V', 'W') characters. -ml 3822 means minimum length less than remove and -mn 0 means remove all N containing sequences.

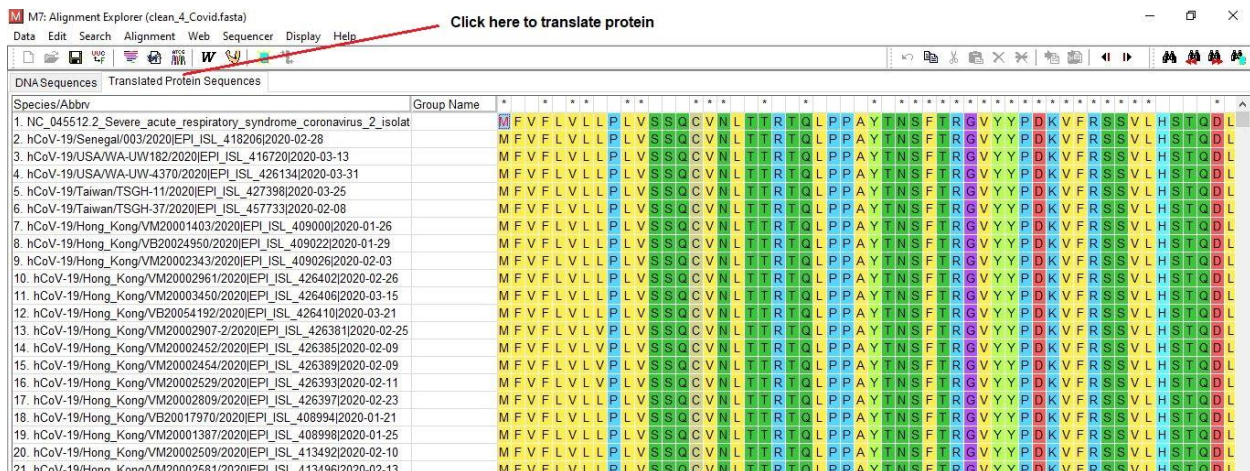
Usages (Linux)

5. Remove internal stop codon using SEDA (<http://www.sing-group.org/seda/download.html>):

Internal stop codon must remove before mutation analysis and SEDA v1.1 (<https://www.sing-group.org/seda/manual/operations.html#id5>) will assist to remove this. Windows 10 defender takes the file as corrupted, so before using it just make sure off the real-time protection.



6. DNA to Protein in MEGA (https://www.megasoftware.net/dload_win_gui): After removal of all low quality and in frame stop codon sequences, we need to translate the DNA to protein. MEGA 7 or other version will aid us to do it. Just open the multiple sequence alignment and press the translate to protein options. Then export the protein alignment to fasta format.



7. Pairwise mutation analysis (<https://github.com/SShaminur/Mutation-Analysis>): Here the mutation results come from pairwise alignment with respect to reference genome (reference amino acid:position:strain amino acid). Before running, we must ensure that there are no stop codon (*) in the last position of the ORF. We must delete the last stop codon in MEGA analysis.

Multiple sequence alignment (Top one will be selected as reference)

		1	2	4	6	8	10	13							
A	[1	M	D	I	I	F	W	V	S	T	F	V	L	F
B	[1	M	D	I	E	F	W	F	S	T	F	V	L	F
C	[1	M	D	I	I	F	W	F	S	T	W	V	L	F

Results (Reference A)

```
B I4E
B V7F
C V7F
C F10W
```

Usages (Linux): (Requirements: Python ≥ 3.7 , Biopython)

8. Deletion analysis (<https://bioinf.shenwei.me/seqkit/usage/>): SeqKit tools used to arrest all the sequences containing gap (-). From there in frame deletion should be carefully find out. To ensure the in frame deletion with MEGA, remove the reference strain, then remove the gap of the strain/s and translate it to protein. Export the protein sequence/s in fasta format. Usually this translation finds the stop codon in last position. If not, then there are sequencing error (if virus not evolve too much). Then again align the protein sequence to the reference genome protein sequences. We can find the deleted amino acid position. Triplet codon deletion should be screen for deletion analysis. Then finally visualize the deletion data in a suitable software (Jalview, Unipro-UGENE, BioEdit etc.)

Usages (Linux):

DNA deletion

M7: Alignment Explorer (198_all.fas)

Data Edit Search Alignment Web Sequencer Display Help

DNA Sequences Translated Protein Sequences

Species/Abbrev	Group Name	
1. NC_045512.2_Severe_acute_respiratory_syndrome_coronavirus_2_isolat		TGGTTCATGCTATACATGTCCTCTGCGACCAATGCTACTAAGAGCTTTGAT
2. New[hCoV-19/England/SHEF-C43D2/2020]EPI_ISL_453709 2020-04-20		TGGTTCATGCTA-----TCTCTGGGACCAATGGTACTAAGAGGTTTGTAT
3. hCoV-19/Australia/VIC1606/2020[EPI_ISL_456517 2020-05-12		TGGTTCATGCTA-----TCTCTGGGACCAATGGTACTAAGAGGTTTGTAT
4. hCoV-19/Thailand/NIH-15/2020[EPI_ISL_434692 2020-01-05		TGGTTCATGCTA-----CTAAGAGGTTTGTAT

Respective protein deletion

M7: Alignment Explorer (S_ref_protein.fas)

Data Edit Search Alignment Web Sequencer Display Help

Protein Sequences

Species/Abbrev	Group Name	
1. NC_045512.2_Severe_acute_respiratory_syndrome_coronavirus_2_isolat		FSNVTWFHAIHVSCTNGTKRFDNPVLPFNDGVYFASTEKSNII
2. New[hCoV-19/England/SHEF-C43D2/2020]EPI_ISL_453709 2020-04-20		FSNVTWFHAI--SGTNGTKRFDNPVLPFNDGVYFASTEKSNII
3. hCoV-19/Australia/VIC1606/2020[EPI_ISL_456517 2020-05-12		FSNVTWFHAI--SGTNGTKRFDNPVLPFNDGVYFASTEKSNII
4. hCoV-19/Thailand/NIH-15/2020[EPI_ISL_434692 2020-01-05		FSNVTWFHAI-----TKRFDNPVLPFNDGVYFASTEKSNII

Mutational analysis using Microsoft Excel:

- I. **Text to column:** Select the column, Then, **Data >Text to Columns > Delimited > Next > Space, Other (/) > Next and Finish.** This will differentiate country name Mutation and others. If we want to separate accession number, the just put Other (/).

PIPLINE - Excel

File Home Insert Page Layout Formulas Data Review View Kutools™ Kutools Plus Foxit PDF Tell me what you want to do... Sign in Share

Get External Data New Query Recent Sources Get & Transform

Connections Refresh All Edit Links

Sort Filter Advanced

Flash Fill Remove Duplicates Consolidate Relationships Manage Data Model What-If Analysis Forecast Sheet Outline Kutools

Convert Text to Columns Wizard - Step 2 of 3

This screen lets you set the delimiters your data contains. You can see how your text is affected in the preview below.

Delimiters

☒ Tab ☐ Semicolon ☐ Treat consecutive delimiters as one

☐ Comma ☐ Space

☒ Other: /

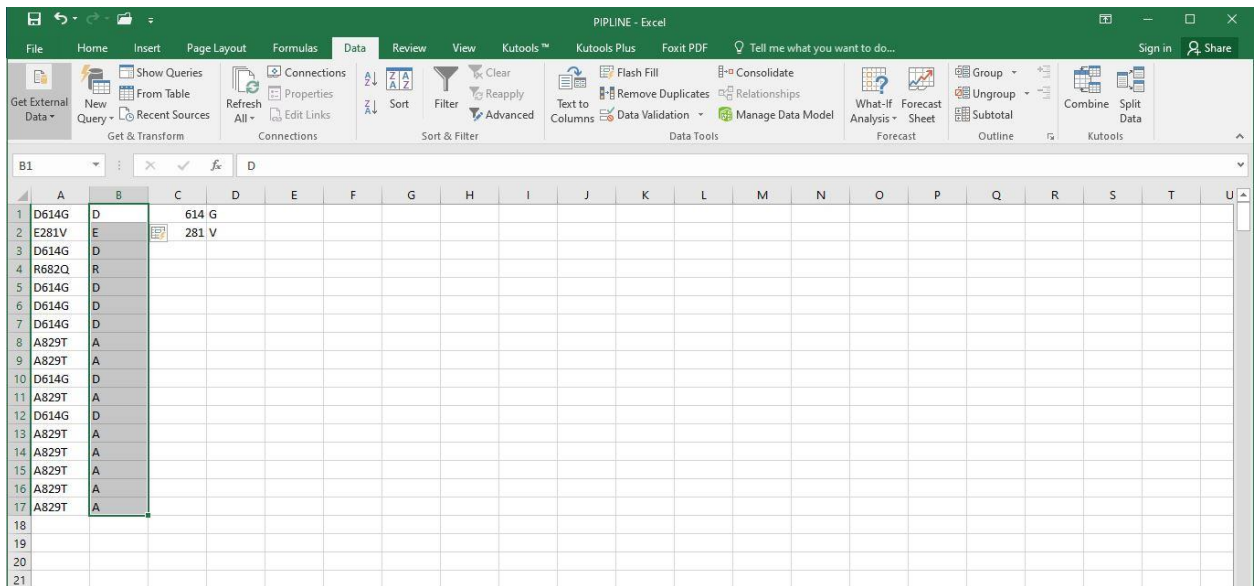
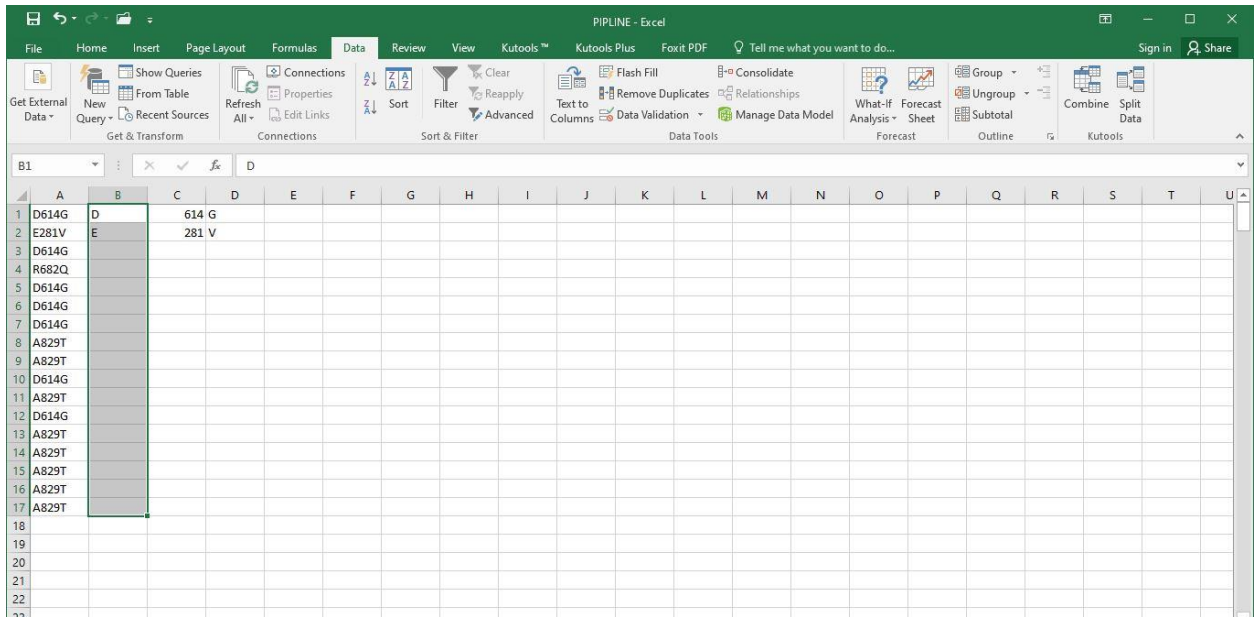
Text qualifier: "

Data preview

hCoV-19	Saudi_Arabia	KAUST-Makkah165	2020 EPI_ISL_437478 2020-04
hCoV-19	Senegal	020	2020 EPI_ISL_418208 2020-03
hCoV-19	Taiwan	TSGH-07	2020 EPI_ISL_427394 2020-03
hCoV-19	Taiwan	TSGH-07	2020 EPI_ISL_427394 2020-03
hCoV-19	Hangzhou	ZJU-01	2020 EPI_ISL_415709 2020-01

Cancel < Back Next > Finish

- II. Flash Fill:** This will separate the mutation (ref:position:strain) into three different columns. From there, we can sort largest to smallest or vice versa. For Flash Fill, first we need to fill up at least two rows then, **Data** > select the 1st desired column > **Flash Fill**. For rest of the two columns, do the same thing.



- III. Remove Duplicates:** Unique mutation, Unique position mutation can be found by removing duplicates. Select the column (where duplicates need to remove) then, **Data** > **Remove Duplicates**.

- IV. Frequency count:** For frequency count, 1st column contains the original data and second column contains the duplicate removal data. Then select a cell in third column and then, =COUNTIFS(A1:A17,B1).

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U
1	D614G	D614G	7																		
2	E281V	E281V	1																		
3	D614G	R682Q	1																		
4	R682Q	A829T	8																		
5	D614G																				
6	D614G																				
7	D614G																				
8	A829T																				
9	A829T																				
10	D614G																				
11	A829T																				
12	D614G																				
13	A829T																				
14	A829T																				
15	A829T																				
16	A829T																				
17	A829T																				
18																					

10. Arrest sequence through sequence ID:

Usages (Linux):