

Effect of Genotype, Life-Stage and Temperature on Oxidative Stress

Introduction

The Overarching Problem and Broad Biological Context

Oxidative stress, an imbalance between reactive oxygen species (ROS) production and antioxidant defenses, presents a significant threat to cellular integrity by damaging essential macromolecules like DNA, proteins, and lipids. ROS are generated as byproducts of normal cellular respiration, and while cells have evolved mechanisms to neutralize these molecules, conditions of redox imbalance such as changes in metabolic state or environmental factors like temperature can lead to an overproduction of ROS and subsequently, oxidative stress. Catalase, an antioxidant enzyme, is crucial in protecting cells by converting hydrogen peroxide (H_2O_2) into water and oxygen, thereby mitigating the potential harm of ROS accumulation. In *Drosophila melanogaster*, a well-established model organism for oxidative stress research, catalase activity is influenced by both genetic variation and environmental conditions, offering a suitable framework for investigating the adaptive mechanisms underlying oxidative stress resilience.

The Specific Dataset and Methods Used

In this study, we measure catalase activity across various *Drosophila* genotypes and life stages under heat shock conditions to assess the influence of genetic and environmental factors on oxidative resilience. Heat shock, a well-known environmental stressor, increases ROS production, requiring an enhanced antioxidant response, which makes it a suitable context for examining catalase activity. The method employed in this experiment is a spectrophotometric assay, as outlined by Hadwan (2018). This assay measures the formation of a carbonato-cobaltate (III) complex, which changes color upon oxidation of cobalt (II) by H_2O_2 , providing a stable colorimetric response that allows precise quantification of catalase activity. By recording absorbance at 440 nm, we can infer catalase levels in the samples based on the reduction of H_2O_2 in the reaction mixture. This approach enables reliable comparisons of catalase activity across different genetic and environmental conditions.

The Gap in Knowledge this Experiment Addresses

Although previous research has provided insights into the genetic basis of oxidative stress response in *Drosophila*, the interplay between genetic factors and environmental stressors in regulating catalase activity remains poorly understood. Studies such as those by Weber et al. (2012), which identified single nucleotide polymorphisms (SNPs) associated with oxidative stress resistance, and Guio et al. (2016), which demonstrated that the Bari-Jheh transposable element enhances oxidative resilience

by upregulating antioxidant genes, have established the importance of genetic variation in modulating antioxidant defenses. However, environmental factors, such as hypoxic conditions that exacerbate oxidative stress, as demonstrated by Habib et al. (2021), and combined thermal and electromagnetic stress, as reported by Zhang et al. (2016), also influence catalase activity by increasing ROS production and the need for cellular defenses. Despite these findings, the specific ways in which genetic predispositions and environmental challenges interact to shape antioxidant responses are not fully understood. This experiment aims to address this gap by investigating how catalase activity is modulated across genotypes and life stages in response to heat shock.

The Question and Hypothesis the Experiment is Designed to Address

This study investigates two primary questions: (1) How does catalase activity vary across genotypes and life stages in *Drosophila melanogaster*, and (2) how does heat shock influence catalase activity within these groups? Based on previous research indicating that adult *Drosophila* typically exhibit heightened physiological responses to environmental stressors, we hypothesize that catalase activity will be highest in heat-shocked adult flies. This is thought to be due to the increased need for resilience against oxidative stress in mature life stages, particularly under stress-inducing conditions.

To address these questions, we quantified catalase activity across different *Drosophila* genotypes and life stages using a spectrophotometric assay that measures hydrogen peroxide breakdown through cobalt oxidation. Experimental groups included both control and heat-shocked individuals to assess temperature effects. By measuring absorbance changes at 440 nm, catalase levels were inferred across developmental stages and genotypes.

The results indicated that life stage significantly influenced catalase activity, with adult flies showing higher activity than pupae. Additionally, while there was no significant effect of genotype on catalase activity, the analysis revealed that heat shock did not significantly impact catalase levels across groups. These findings support our hypothesis regarding life stage but suggest that heat shock may not universally elevate catalase activity under the tested conditions. This study thus underscores the role of developmental stage in antioxidant responses and highlights the complex interactions between genotype, life stage, and environmental stressors in shaping oxidative stress resilience in *Drosophila melanogaster*.

Literature Cited

- Weber, A. L., Khan, G. F., Magwire, M. M., Tabor, C. L., Mackay, T. F. C., & Anholt, R. R. H. (2012). Genome-wide association analysis of oxidative stress resistance in *Drosophila melanogaster*. *PLOS ONE*, 7(4), e34745. <https://doi.org/10.1371/journal.pone.0034745>
- Guio, L. (2016). Stress affects the epigenetic marks added by Bari-Jeh: A natural insertion associated with two adaptive phenotypes in *Drosophila*. *bioRxiv*. <https://doi.org/10.1101/037598>
- Habib, S., Lwin, Y. Y., & Li, N. (2021). Down-regulation of *SIGRAS10* in tomato confers abiotic stress tolerance. *Genes*, 12(5), 623. <https://doi.org/10.3390/genes12050623>
- Zhang, B., Peng, Y., Zheng, J., Liang, L., Hoffmann, A. A., & Ma, C.-S. (2016). Response of heat shock protein genes of the oriental fruit moth under diapause and thermal stress reveals multiple patterns dependent on the nature of stress exposure. *Cell Stress and Chaperones*, 21(4), 653–663. <https://doi.org/10.1007/s12192-016-0690-8>
- Hadwan, M. H. (2018). Simple spectrophotometric assay for measuring catalase activity in biological tissues. *BMC Biochemistry*, 19(1), 7. <https://doi.org/10.1186/s12858-018-0097-5>

Supplemental Information

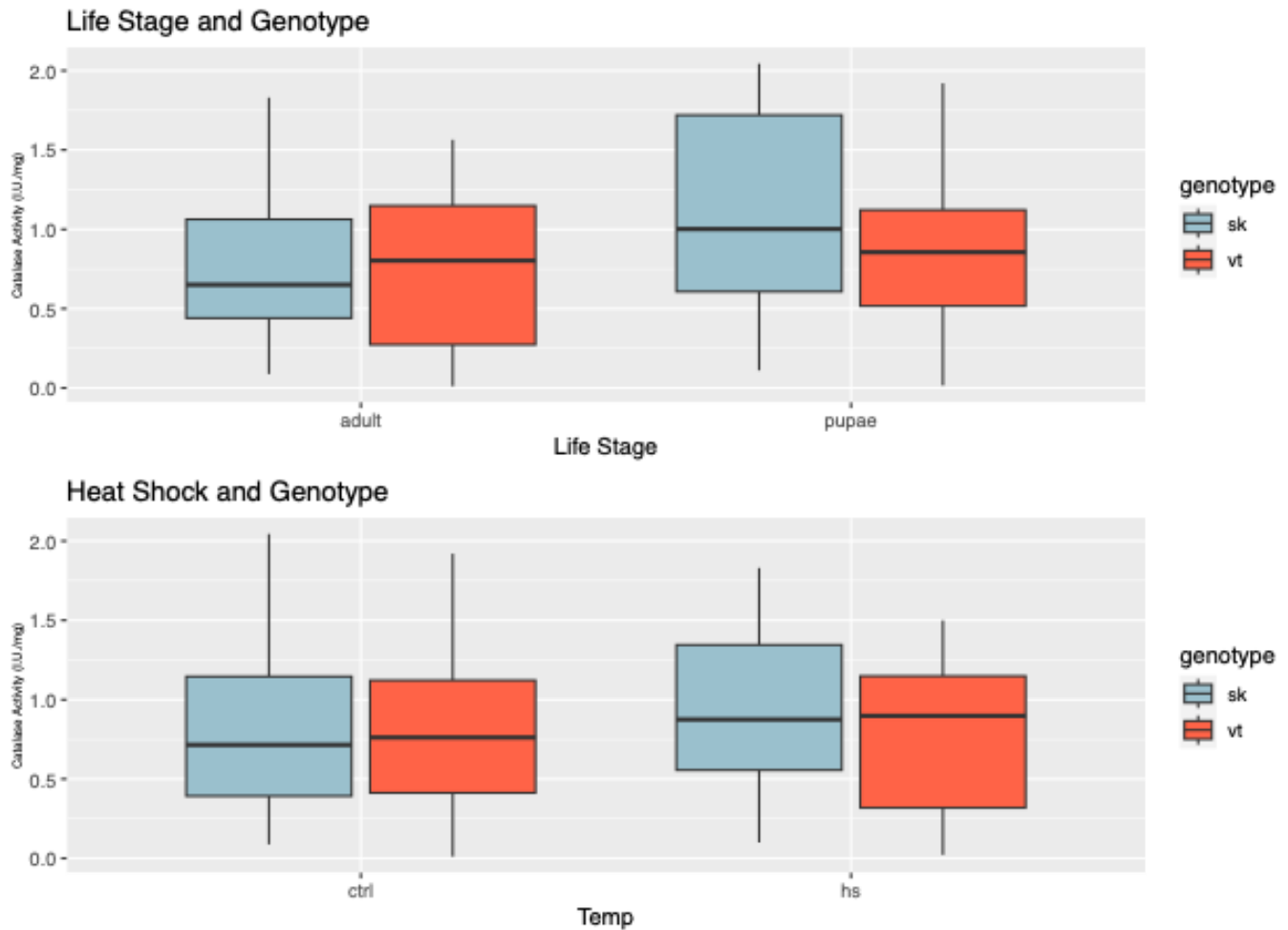


Figure 1: Box plots showing catalase activity (I.U./mg) in different experimental groups based on life stage, genotype, and temperature treatments. In the first plot, a significant effect of life stage on catalase activity was observed ($p = 0.020$), with higher activity in one stage over the other, as indicated by the ANOVA. Genotype and the interaction between life stage and genotype were not significant ($p > 0.05$). In the second plot, neither temperature treatment nor genotype showed significant effects on catalase activity.

Life Stage Analysis

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Life S.	1	1.38	1.3752	5.54	0.02
Genotype	1	0.23	0.2274	0.916	0.34
Life S. ~ Genotype	1	0.42	0.4228	1.703	0.194
Residuals	140	34.75	0.2482		

Table 1: Analysis of Variance (ANOVA) summary for the effects of life stage, genotype, and their interaction on the response variable. Degrees of freedom (Df), sum of squares (Sum Sq), mean squares (Mean Sq), F-values, and p-values (Pr(>F)) are shown for each factor and interaction term. A statistically significant effect of life stage was observed at the 0.05 significance level, with a p-value of 0.020. No significant effects were found for genotype or the life stage and genotype interaction.

Heat Shock Analysis

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Temp	1	0.08	0.07977	0.307	0.581
Genotype	1	0.23	0.22745	0.874	0.351
Temp ~ Genotype	1	0.05	0.05366	0.206	0.65
Residuals	140	36.42	0.26013		

Table 2: Analysis of Variance (ANOVA) summary for the effects of temperature (temp), genotype, and their interaction on the response variable. Degrees of freedom (Df), sum of squares (Sum Sq), mean squares (Mean Sq), F-values, and p-values (Pr(>F)) are presented for each factor and interaction term. No statistically significant effects were observed at a 0.05 significance level.