# CGS698C, Assignment 04

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## Part 1: A simple linear regression: Power posing and testosterone

Download the file df\_powerpose.csv and load the following data set:

```
df_powerpose <- read.table("df_powerpose.csv",header=T,sep=",")
head(df_powerpose)

##    X id hptreat female age testm1    testm2
## 1 2 29    High    Male    19 38.725 62.375
## 2 3 30    Low Female    20 32.770    29.235
## 3 4 31    High Female    20 32.320    27.510
## 4 5 32    Low Female    18 17.995    28.655
## 5 7 34    Low Female    21 73.580    44.670
## 6 8 35    High Female    20 80.695 105.485</pre>
```

The data set, which was originally published in Carney, Cuddy, and Yap (2010) but released in modified form by Fosse (2016), shows the testosterone levels of 39 different individuals, before and after treatment, where treatment refers to each individual being assigned to a high power pose or a low power pose. In the original paper by Carney, Cuddy, and Yap (2010), the unit given for testosterone measurement (estimated from saliva samples) was picograms per milliliter (pg/ml). One picogram per milliliter is 0.001 nanogram per milliliter (ng/ml).

The research hypothesis is that on average, assigning a subject a high power pose vs. a low power pose will lead to higher testosterone levels after treatment.

Investigate this claim using a linear model and weakly informative priors<sup>1</sup> in brms. You'll need to <sup>1</sup>You can also use default processimate the effect of the variable that encodes the change in testosterone.

### Part 2: Poisson regression models and hypothesis testing

Human language has an interesting, empirical property: When dependency arcs are drawn between syntactically-related words, they rarely cross each other. You can ignore this sentence if you do not have any background in Linguistics. I am just setting a motivation for the problem.

salivary measurement. But I would suggest you try to come up with

Any sentence from a human language would contain some easy structures and some difficult structures. A crossing dependency is a type of structure that is arguably difficult to process and is rarely found in natural languages.

The number of crossing dependencies in a sentence can be given by a Poisson distribution

$$N_i \sim Poisson(\lambda_i)$$
 (1)

where  $N_i$  is the number of crossing dependencies in the sentence i;  $\lambda_i$  is rate parameter indicating the expected rate of crossing dependencies in the sentence i, such that

ryou can also use default priors of brms assuming you know nothing about typical ranges of testosterone using salivary measurement. But I would suggest you try to come up with weakly informative priors. Feel free to do a prior predictive check if you are unsure.

$$\log \lambda_i = \alpha + \beta L_i \tag{2}$$

where  $L_i$  is the length of the sentence i,  $\alpha$  is the expected rate of crossings in a sentence of average **length** (say 11) and  $\beta$  is the change in rate of crossings as a function of sentence length.

(Note: Sentence length means the number of words in a sentence. For simplicity, assume that sentence lengths can range from 2 to 20.)

Exercise 2.1 Implement the model in R or Python such that the function gives the number of crossings as the outcome, and takes sentence length,  $\alpha$ , and  $\beta$  as its arguments.

Exercise 2.2 Generate prior predictions of the model for sentences of length 4 under the following prior assumptions

$$\alpha \sim Normal_{lb=0}(0.15, 0.1) \tag{3}$$

$$\beta \sim Normal_{lb=0}(0.25, 0.05)$$
 (4)

Exercise 2.3 Consider a dataset of crossing dependencies from English and German corpora, "crossing.csv". This dataset contains number of crossings for each sentence from each language. Fit the following two models, M1 and M2, to the given data.

#### Model M1

Assumption: The rate of crossings is only a function of sentence length and it remains exactly the same in English and German.}

This assumption can be represented by the following regression.

$$N_{i,i} \sim Poisson(\lambda_{i,i})$$
 (5)

where  $N_{i,j}$  is the number of crossing dependencies in sentence i in language j;  $\lambda_{i,j}$  is rate parameter indicating the expected rate of crossing dependencies in sentence i in language j, such that

$$\log \lambda_{i,i} = \alpha + \beta L_{i,i} \tag{6}$$

where  $L_{i,j}$  is the length of sentence i of language j.

The above model implies the average rate of crossings depends only on the sentence length.

#### Model M2

Assumption: As sentence length increases, the number crossings grows at a different rate in English vs. German.}

$$N_{i,j} \sim Poisson(\lambda_{i,j})$$
 (7)

where  $N_{i,j}$  is the number of crossing dependencies in sentence i in language j;  $\lambda_{i,j}$  is rate parameter indicating the expected rate of crossing dependencies in sentence i in language j, such that

$$\log \lambda_{i,j} = \alpha + \beta L_{i,j} + \beta_{language} R_j + \beta_{interact} L_{i,j} * R_j$$
(8)

where  $L_{i,j}$  is the length of sentence i of language j,  $R_i$  is the indicator variable such that  $R_i = 0$  if the language is English and  $R_i = 1$  if the language is German.

The above model implies that the average rate of crossings depends on the sentence length as well the language (English vs German).

Load the data file "crossings.csv" and fit the above two models to the data and estimate all the parameters. You can use "brms" to fit the above models, you will have to use Poisson family for the likelihood. Use the following priors:

$$\alpha \sim Normal(0.15, 0.1) \tag{9}$$

$$\beta \sim Normal(0, 0.15) \tag{10}$$

$$\beta_{language} \sim Normal(0, 0.15)$$
 (11)

$$\beta_{interact} \sim Normal(0, 0.15)$$
 (12)

Exercise 2.4 Quantify evidence for the models M1 and M2 using k-fold cross-validation.

Here is a sample code to do the same.

```
observed <- read.table("crossings.csv",sep=",",header=T)</pre>
# Visualize average rate of crossings
observed %>% group_by(Language,s.length) %>%
  summarise(mean.crossings=mean(nCross)) %>%
  ggplot(aes(x=s.length,y=mean.crossings,
             group=Language, color=Language))+
  geom_point()+geom_line()
# Code/center the predictors
observed$s.length <- observed$s.length - mean(observed$s.length)</pre>
observed$lang <- ifelse(observed$Language=="German",1,0)</pre>
# These two vectors will store log predictive desnities
# in each fold
lpds.m1 <- c()
lpds.m2 <- c()
untested <- observed
for(k in 1:5){
  # Prepare test data and training data
  ytest <- sample_n(untested, size=nrow(observed)/5)</pre>
  ytrain <- setdiff(observed,ytest)</pre>
```

```
untested <- setdiff(untested,ytest)</pre>
  # Fit the models M1 and M2 on training data
  fit.m1 <-
    brm(nCross ~ 1 + s.length, data=ytrain,
        family = poisson(link = "log"),
        prior = c(prior(normal(0.15, 0.1), class = Intercept),
                   prior(normal(0, 0.15), class = b)),
        cores=4)
  fit.m2 <-
    brm(nCross ~ 1 + s.length + lang + s.length*lang,
        data=ytrain,
        family = poisson(link = "log"),
        prior = c(prior(normal(0.15, 0.1), class = Intercept),
                   prior(normal(0, 0.15), class = b)),
        cores=4)
  # retrieve posterior samples
  post.m1 <- posterior_samples(fit.m1)</pre>
  post.m2 <- posterior_samples(fit.m2)</pre>
  # Calculate log pointwise predcitive density using test data
  lppd.m1 <- 0
  lppd.m2 <- 0
  for(i in 1:nrow(ytest)){
    lpd_im1 <- log(mean(dpois(ytest[i,]$nCross,</pre>
                 lambda=exp(post.m1[,1]+
                 post.m1[,2]*ytest[i,]$s.length))))
    lppd.m1 <- lppd.m1 + lpd_im1</pre>
    lpd_im2 <- log(mean(dpois(ytest[i,]$nCross,</pre>
                 lambda=exp(post.m2[,1]+
                 post.m2[,2]*ytest[i,]$s.length+
                   post.m2[,3]*ytest[i,]$lang+
                   post.m2[,4]*ytest[i,]$s.length*ytest[i,]$lang)
                   )))
    lppd.m2 <- lppd.m2 + lpd_im2</pre>
  lpds.m1 <- c(lpds.m1,lppd.m1)</pre>
  lpds.m2 <- c(lpds.m2,lppd.m2)</pre>
# Predictive accuracy of model M1
elpd.m1 <- sum(lpds.m1)</pre>
```

}

```
# Predictive accuracy of model M2
elpd.m2 <- sum(lpds.m2)</pre>
```

# Evidence in favor of M2 over M1 difference\_elpd <- elpd.m2-elpd.m1</pre>