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vscode  
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home.html  
result.html  
app.py  
server.py  
BloodCell.h5  
BloodCellClassifier.ipynb

app.py BloodCellClassifier.ipynb X Home.html BloodCell.h5

BloodCellClassifier.ipynb Import tensorflow as tf  
Generate Code + Markdown Run All

```
import tensorflow as tf
print("tensorflow version:", tf.__version__)
```

tensorflow version: 2.19.0

```
!pip install seaborn
```

Requirement already satisfied: seaborn in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (0.13.2)  
Requirement already satisfied: numpy!=1.24.0,>=1.20 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from seaborn) (2.1.3)  
Requirement already satisfied: pandas>=1.2 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from seaborn) (2.3.0)  
Requirement already satisfied: matplotlib=3.6.1,>=3.4 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from seaborn) (3.10.3)  
Requirement already satisfied: contourpy>=1.0.1 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (1.3.0)  
Requirement already satisfied: cycler>=0.10 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (0.12.1)  
Requirement already satisfied: fonttools>=4.22.0 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (4.53.0)  
Requirement already satisfied: kiwisolver>=1.3.1 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (1.4.6)  
Requirement already satisfied: packaging>=20.0 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (25.0)  
Requirement already satisfied: pillow>=8 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (11.2.1)  
Requirement already satisfied: pyparsing>=2.3.1 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (3.2.0)  
Requirement already satisfied: python-dateutil>=2.7 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from matplotlib=3.6.1,>=3.4->seaborn) (2.9.0.post0)  
Requirement already satisfied: pytz>=2020.1 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from pandas>=1.2->seaborn) (2025.2)  
Requirement already satisfied: tzdata>=2022.7 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from pandas>=1.2->seaborn) (2025.2)  
Requirement already satisfied: six>=1.5 in c:\users\balas\onedrive\desktop\hematovision\venv\lib\site-packages (from python-dateutil>=2.7->matplotlib=3.6.1,>=3.4->seaborn) (1.17.0)

[notice] A new release of pip is available: 24.3.1 -> 25.1.1  
[notice] To update, run: python.exe -m pip install --upgrade pip

```
import os
import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
import cv2
import numpy as np
```

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Python

```
# Define the TRAIN folder path
data_dir = 'dataset2-master/images/TRAIN'

# Define the class labels
class_labels = ['eosinophil', 'lymphocyte', 'monocyte', 'neutrophil']

# Store file paths and labels
filepaths = []
labels = []

# Loop through each class folder
for label in class_labels:
    class_folder = os.path.join(data_dir, label)
    for file in os.listdir(class_folder):
        if file.endswith('.jpeg') or file.endswith('.png'):
            filepaths.append(os.path.join(class_folder, file))
            labels.append(label)

# Create a DataFrame
bloodCell_df = pd.DataFrame({
    'filepaths': filepaths,
    'labels': labels
})

# Shuffle the DataFrame
bloodCell_df = bloodCell_df.sample(frac=1).reset_index(drop=True)

# Show top 5 entries
bloodCell_df.head()
```

filepaths labels

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249/249 1560s 6s/step - accuracy: 0.6952 - loss: 0.7573 - val\_accuracy: 0.7582 - val\_loss: 0.6187  
EPOCH 10/10

```
model.save("BloodCell.h5")
```

WARNING:absl:You are saving your model as an HDF5 file via `model.save()` or `keras.saving.save\_model(model)`. This file format is considered legacy. We recommend

# Evaluation: Confusion Matrix + Classification Report + Accuracy/Loss Plots

```
from sklearn.metrics import confusion_matrix, classification_report
import seaborn as sns

# Step 1: Reset the validation generator
val_generator.reset()

# Step 2: Predict labels
predictions = model.predict(val_generator)
pred_labels = np.argmax(predictions, axis=1)
true_labels = val_generator.classes

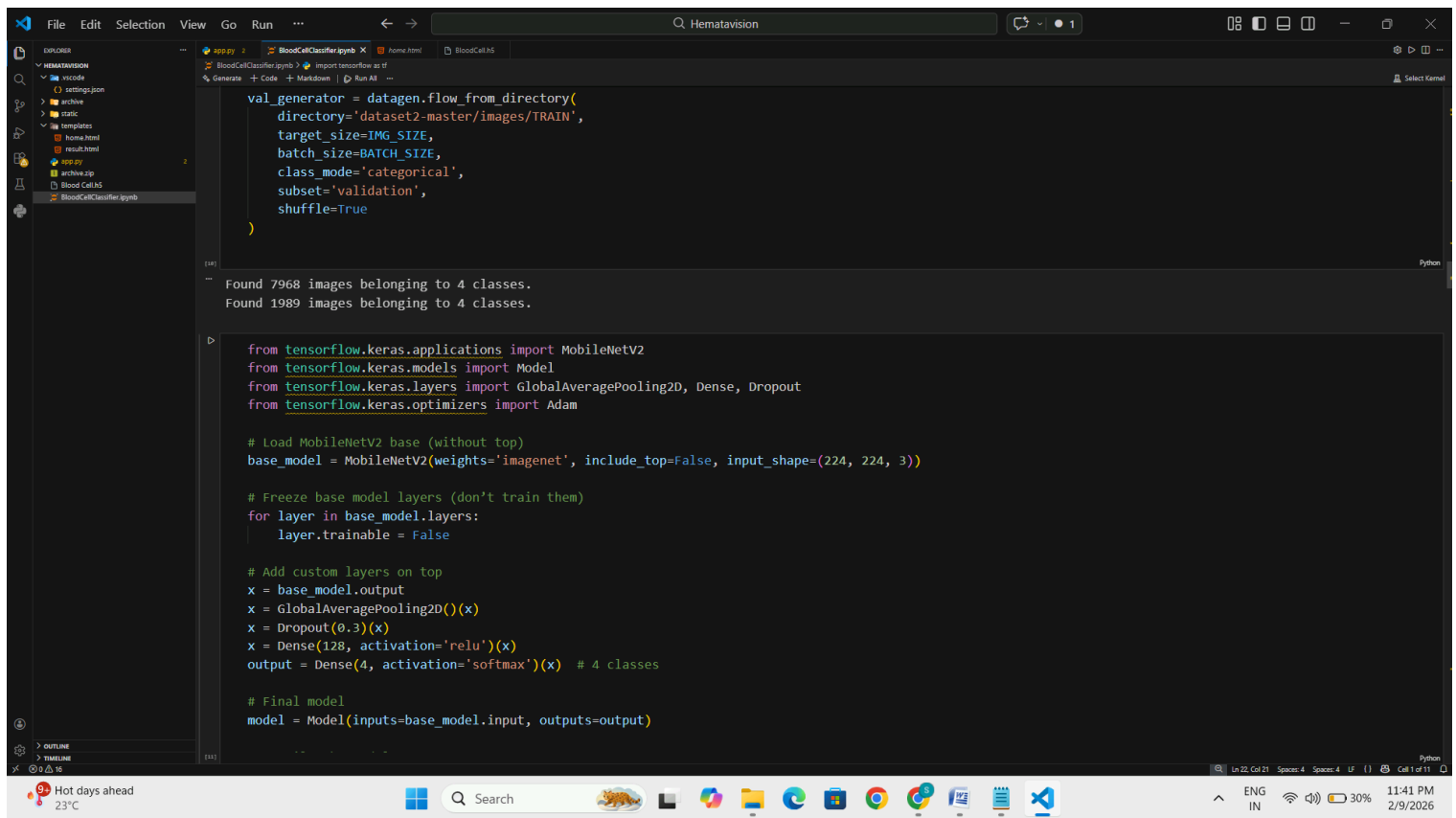
# Step 3: Classification report
print("\nClassification Report:\n")
print(classification_report(true_labels, pred_labels, target_names=class_labels))

# Step 4: Confusion matrix
cm = confusion_matrix(true_labels, pred_labels)
plt.figure(figsize=(8, 6))
sns.heatmap(cm, annot=True, fmt='d', cmap='Blues', xticklabels=class_labels, yticklabels=class_labels)
plt.xlabel('Predicted')
plt.ylabel('Actual')
plt.title('Confusion Matrix')
plt.show()
```

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filepaths

labels

0 dataset2-master/images/TRAIN\lymphocyte\1\_59... lymphocyte

1 dataset2-master/images/TRAIN\neutrophil\37\_68... neutrophil

2 dataset2-master/images/TRAIN\lymphocyte\15\_69... lymphocyte

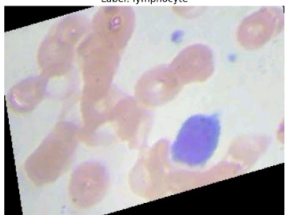
3 dataset2-master/images/TRAIN\eosinophil\36\_46... eosinophil

4 dataset2-master/images/TRAIN\lymphocyte\18\_17... lymphocyte

```
import matplotlib.pyplot as plt
import random
from PIL import Image

# Show 5 random images
for i in range(5):
    img_path = random.choice(bloodCell_df['filepaths'])
    img = Image.open(img_path)
    plt.imshow(img)
    plt.title(f"Label: {bloodCell_df[bloodCell_df['filepaths'] == img_path]['labels'].values[0]}")
    plt.axis('off')
    plt.show()
```

Label: lymphocyte



OUTLINE

FILE

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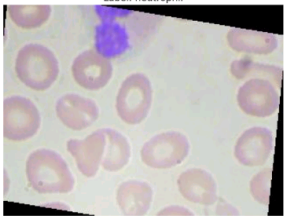
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Label: neutrophil



Label: neutrophil




Label: eosinophil

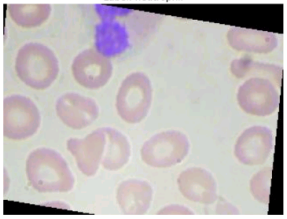


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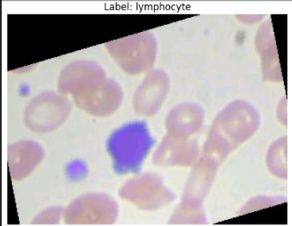


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Label: lymphocyte



```
from tensorflow.keras.preprocessing.image import ImageDataGenerator
from tensorflow.keras.applications.mobilenet_v2 import preprocess_input

# Create training and validation generators with augmentation
datagen = ImageDataGenerator(
    preprocessing_function=preprocess_input,
    validation_split=0.2, # 80% training, 20% validation
    rotation_range=15,
    zoom_range=0.1,
    width_shift_range=0.1,
    height_shift_range=0.1,
    shear_range=0.1,
    horizontal_flip=True
)

# Define image size and batch
IMG_SIZE = (224, 224)
BATCH_SIZE = 32

# Load training data
train_generator = datagen.flow_from_directory(
    directory='dataset2-master/images/TRAIN',
```

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Python

model.summary()

[110]

Model: "functional"

Layer (type)	Output Shape	Param #	Connected to
input_layer (InputLayer)	(None, 224, 224, 3)	0	-
Conv1 (conv2D)	(None, 112, 112, 32)	864	input_layer[0][0]
bn_Conv1 (BatchNormalizati...	(None, 112, 112, 32)	128	Conv1[0][0]
Conv1_relu (ReLU)	(None, 112, 112, 32)	0	bn_Conv1[0][0]
expanded_conv_dep... (Depthwiseconv2D)	(None, 112, 112, 32)	288	Conv1_relu[0][0]
expanded_conv_dep... (BatchNormalizati...	(None, 112, 112, 32)	128	expanded_conv_d...
expanded_conv_dep... (ReLU)	(None, 112, 112, 32)	0	expanded_conv_d...
expanded_conv_pro... (conv2D)	(None, 112, 112, 16)	512	expanded_conv_d...
expanded_conv_pro... (BatchNormalizati...	(None, 112, 112, 16)	64	expanded_conv_p...
block_1_expand (conv2D)	(None, 112, 112, 96)	1,536	expanded_conv_p...

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The screenshot displays a Jupyter Notebook interface with a Keras model architecture for BloodCellClassifier. The model is a sequential model with 11 layers. The layers are: block\_4\_expand (Conv2D), block\_4\_expand\_BN (Batch Normalization), block\_4\_expand\_re (ReLU), block\_4\_depthwise (Depthwise Conv2D), block\_4\_depthwise (Batch Normalization), block\_4\_depthwise (ReLU), block\_4\_project (Conv2D), block\_4\_project\_BN (Batch Normalization), block\_4\_add (Add), block\_5\_expand (Conv2D), block\_5\_expand\_BN (Batch Normalization), block\_5\_expand\_re (ReLU), block\_5\_depthwise (Depthwise Conv2D), and block\_5\_depthwise (Batch Normalization). The model has 6,144 parameters. The output is a single unit.

Layer	Config	Params	Next Layer
block_4_expand	(Conv2D)	6,144	block_3_project...
block_4_expand_BN	(Batch Normalizati...	768	block_4_expand[...
block_4_expand_re	(ReLU)	0	block_4_expand_...
block_4_depthwise	(DepthwiseConv2D)	1,728	block_4_expand_...
block_4_depthwise	(Batch Normalizati...	768	block_4_depthwi...
block_4_depthwise	(ReLU)	0	block_4_depthwi...
block_4_project	(Conv2D)	6,144	block_4_depthwi...
block_4_project_BN	(Batch Normalizati...	128	block_4_project...
block_4_add	(Add)	0	block_3_project... block_4_project...
block_5_expand	(Conv2D)	6,144	block_4_add[...][...
block_5_expand_BN	(Batch Normalizati...	768	block_5_expand[...
block_5_expand_re	(ReLU)	0	block_5_expand_...
block_5_depthwise	(DepthwiseConv2D)	1,728	block_5_expand_...
block_5_depthwise	(Batch Normalizati...	768	block_5_depthwi...



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Classification Report:

	precision	recall	f1-score	support
eosinophil	0.22	0.20	0.21	499
lymphocyte	0.23	0.25	0.24	496
monocyte	0.25	0.30	0.27	495
neutrophil	0.24	0.21	0.23	499
accuracy			0.24	1989
macro avg	0.24	0.24	0.24	1989
weighted avg	0.24	0.24	0.24	1989

Confusion Matrix

	eosinophil	lymphocyte	monocyte	neutrophil
eosinophil	98	136	158	107
lymphocyte	121	122	131	122
monocyte	99	141	148	107
neutrophil	125	123	145	106

Model Accuracy

Epoch	Train Accuracy	Validation Accuracy
1	0.60	0.60
2	0.62	0.65
3	0.64	0.68
4	0.66	0.70
5	0.68	0.72
6	0.69	0.74
7	0.70	0.75
8	0.70	0.74
9	0.70	0.75
10	0.70	0.75

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