1. Prerequisites -

Install miniconda, qiime2=2023.7 and usearch. For details to install qiime2=2023.7, check <u>giime2 documentation</u>,

Follow this tutorial for detailed guidance - https://currentprotocols.onlinelibrary.wiley.com/doi/full/10.1002/cpbi.100

2. Paired-end fastq files merged using usearch

Example command -

```
usearch11.0.667_i86linux32 -fastq_mergepairs
raw_seq_dir/001AU46701-16S_V1-V3_L6RGF_GATCGTCGCG-CTGGATATGT_L001_R1.fastq
-reverse
raw_seq_dir/001AU46701-16S_V1-V3_L6RGF_GATCGTCGCG-CTGGATATGT_L001_R2.fastq
-fastqout 001AU46701-16S_V1-V3_L6RGF_GATCGTCGCG-CTGGATATGT_L001_R1R2.fq
```

3. Creating metadata file from Raw Reads

```
echo -e "sample-id\tabsolute-filepath" > manifest.tsv

for f in *R1R2.fq; do
   n=$(basename "$f")
   echo -e "${n}\t$PWD/$f"

done >> manifest.tsv
```

4. Using manifest file to import raw reads into QIIME 2 artifacts

```
qiime tools import \
    --input-path manifest.tsv \
    --type 'SampleData[SequencesWithQuality]' \
     --input-format SingleEndFastqManifestPhred33V2 \
     --output-path paired_end-demux.qza
```

5. Creating a summary of the demultiplexed artifact to interactively explore the metadata (Optional Commands)

Summarizing manifest.tsv

COLUMNS: 1

| <pre>qiime metadata tabulate \ m-input-file manifest.tsv \ o-visualization metadata-summary.qzv</pre> | | | | | | |
|---|--|-------------------------------|---------------|--------------------|---------|--|
| dime2view | File: met | adata-summary.qzv | Visualization | Details Provenance | | |
| Download metadata TSV file | | | | | | |
| This file won't necessarily reflect dynamic sorting or filtering options based on the interact | tive table below. | | | | Search: | |
| sample-id #q2rtypes | 17 | absolute-filepath categorical | | | 11 | |
| 001AU27472-16S_V1-V3-L6RGF-GCACCACCAA-TGGTACTGAT_L001_R1R2.fq | /home/salmaneurus27/001AU27472-16S_V1-V3-L6RGF-GCACCACCAA-TGGTACTGAT_L001_R1R2.fq | | | | | |
| 001AU33237-16S_V1-V3_L6RGF_GTTCGGAGTT-CGCCTTCTGA_L001_R1R2.fq | /home/salmaneunus/27/001AU33237-16S_V1-V3_L6RGF_GTTCGGAGTT-CGCCTTCTGA_L001_R1R2.fq | | | | | |
| 001AU46701-16S_V1-V3_L6RGF_GATCGTCGCG-CTGGATATGT_L001_R1R2.fq | /home/salmaneunus27/001AU46701-16S_V1-V3_L6RGF_GATCGTCGCG-CTGGATATGT_L001_R1R2 fq | | | | | |
| To get some basic info. | | | | | | |
| qiime tools inspec t-metadata manifest.tsv | | | | | | |
| COLUMN NAME | | 27@3acillai1. 3 | | | | |
| absolute-filepath | | | | | | |
| TDS: | 3 | | | | | |

Summarizing paired-end reads



6. Sequence Quality Control & Feature Table Construction

Initial filtering based on quality scores

```
qiime quality-filter q-score \
--i-demux paired_end-demux.qza \
--o-filtered-sequences pe-filtered.qza \
--o-filter-stats pe-filter-stats.qza
```

Clustering sequences into OTUs Using q2-vsearch

Reference - https://github.com/qiime2/docs/blob/master/source/tutorials/otu-clustering.rst

Using the filtered sequences from the previous step, we will dereplicate the sequences to form a FeatureTable[Frequency] artifact and a FeatureData[Sequence] artifact

qiime vsearch dereplicate-sequences \

- --i-sequences pe-filtered.qza \
- --o-dereplicated-table feature-table.qza \
- --o-dereplicated-sequences rep-seqs.qza

Closed-Reference Clustering of the OTU table

Clustering is performed at 85% identity against the Greengenes 13_8 85% OTUs reference database.

Downloading greengenes OTUs reference database

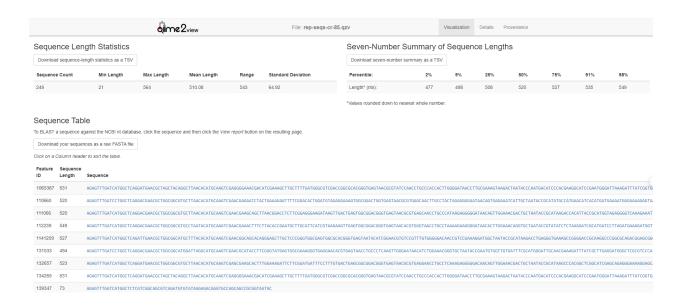
wget https://data.qiime2.org/2023.9/tutorials/otu-clustering/85_otus.qza

qiime vsearch cluster-features-closed-reference --i-table feature-table.qza

- --i-sequences rep-seqs.gza --i-reference-sequences 85 otus.gza --p-perc-identity 0.85
- --o-clustered-table table-cr-85.qza --o-clustered-sequences rep-seqs-cr-85.qza
- --o-unmatched-sequences unmatched-cr-85.qza

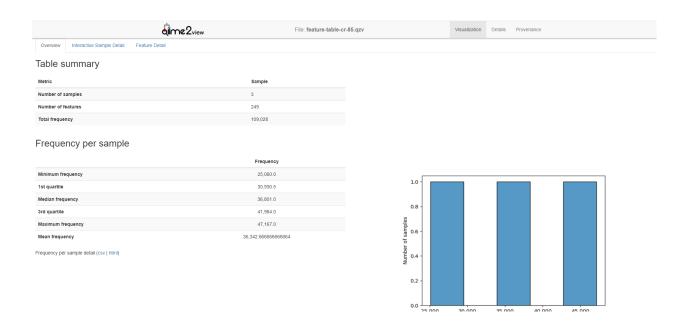
Visualizing the representative sequences after Closed-Reference Clustering

qiime feature-table tabulate-seqs --i-data rep-seqs-cr-85.qza --o-visualization rep-seqs-cr-85.qzv



Visualizing feature table

qiime feature-table summarize --i-table table-cr-85.qza --m-sample-metadata-file manifest.tsv --o-visualization feature-table-cr-85.qzv



7. Generating Feature table in BIOM and CSV format

qiime tools export --input-path table-cr-85.qza --output-path exported-feature-table

| | 134265 | 1065387 | 551008 | 587572 | 215040 | 552456 | 185940 | 4308184 | 183870 | 53032 |
|---|---------|---------|--------|--------|--------|--------|--------|---------|--------|-------|
| 001AU27472-16S_V1-V3- L6RGF-GCACCACCAA- TGGTACTGAT_L001_R1R2.fq | 35662.0 | 1400.0 | 4804.0 | 1.0 | 611.0 | 0.0 | 480.0 | 3.0 | 327.0 | 0.0 |
| 001AU33237-16S_V1- V3_L6RGF_GTTCGGAGTT- CGCCTTCTGA_L001_R1R2.fq | 15282.0 | 2273.0 | 2396.0 | 2.0 | 75.0 | 0.0 | 1181.0 | 2.0 | 17.0 | 6.0 |
| 001AU46701-16S_V1- V3_L6RGF_GATCGTCGCG- CTGGATATGT_L001_R1R2.fq | 9094.0 | 1532.0 | 3943.0 | 440.0 | 804.0 | 346.0 | 2390.0 | 13.0 | 453.0 | 718.0 |

See details here to read .biom file and convert from .biom to .csv - https://colab.research.google.com/drive/1LcprGCrPnvrzujMZH37NToIMIUAvToWx?usp=sharing

All OTU_ids matched with previous otu_taxonomy table - https://colab.research.google.com/drive/1ivNYwkXi9f4EqGbRMzc-6w2U1frx59 k?usp=sharing

8. Taxonomic Classification

Downloading 3 required files from the inventory

waet

https://github.com/BenKaehler/readytowear/raw/master/data/gg_13_8/515f-806r/human-stool.qza

wget

https://github.com/BenKaehler/readytowear/raw/master/data/gg_13_8/515f-806r/ref-seqs.qza

wget https://github.com/BenKaehler/readytowear/raw/master/data/gg_13_8/515f-806r/ref-tax.qza

Training classifiers using these files

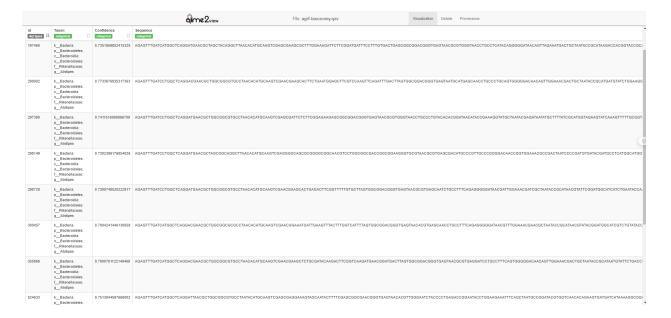
qiime feature-classifier fit-classifier-naive-bayes --i-reference-reads ref-seqs.qza --i-reference-taxonomy ref-tax.qza --i-class-weight human-stool.qza --o-classifier human gut classifier.gza

Assign taxonomy to our representative sequences using our newly trained classifier:

qiime feature-classifier classify-sklearn \
--i-reads rep-seqs-cr-85.qza \
--i-classifier human_gut_classifier.qza \
--o-classification agrf-taxonomy.qza

Now, we will visualizing the taxonomy:

qiime metadata tabulate \
--m-input-file agrf-taxonomy.qza \
--m-input-file rep-seqs-cr-85.qza \
--o-visualization agrf-taxonomy.qzv



| | id | Taxon | Confidence | Sequer |
|---|-----------|--|--------------------|--|
| 0 | | | antagarinal | · |
| U | #q2:types | categorical | categorical | categori |
| 1 | 1065387 | k_Bacteria; p_Firmicutes; c_Clostridia; o | 0.7176739941419537 | AGAGTTTGATCATGGCTCAGGATGAACGCTAGCTACAGGCTTAAC, |
| 2 | 110660 | kBacteria | 0.998410293516579 | AGAGTTTGATCCTGGCTCAGGACGAACGCTGGCGGCGTGCTTAAC |
| 3 | 111066 | k_Bacteria; p_Firmicutes; c_Clostridia; o | 0.7497422366344827 | AGAGTTTGATCATGGCTCAGGACGAACGCTGGCGGCGTGCTTAAC |
| 4 | 112239 | kBacteria | 0.9980215995275771 | AGAGTTTGATCATGGCTCAGGACGAACGCTGGCGGCGTGCCTAATA |
| 5 | 1141209 | kBacteria; pFirmicutes | 0.712636651358992 | AGAGTTTGATCATGGCTCAGATTGAACGCTGGCGGCATGCTTTAC |
| 6 | 131533 | k_Bacteria; p_Firmicutes; | 0.8163357852277051 | AGAGTTTGATCCTGGCTCAGGACGAACGCTGGCGGCATGGATTAGG |

Taxonomy Based Filtering of Data

Excluding sequences that are unexpected such as those from chloroplasts or mitochondria

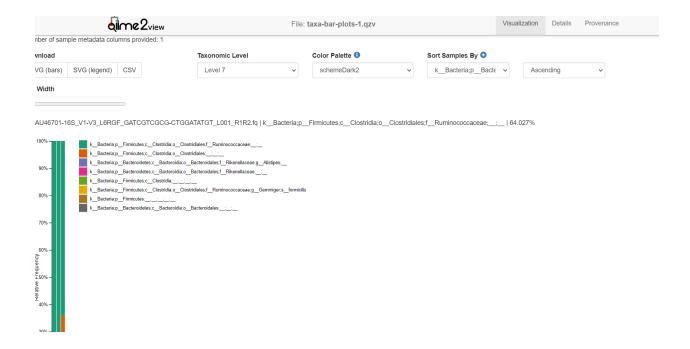
qiime taxa filter-table --i-table table-cr-85.qza --i-taxonomy agrf-taxonomy.qza --p-mode contains --p-include p__ --p-exclude 'p__;,Chloroplast,Mitochondria' --o-filtered-table filtered-table-3.qza

In this step we are removing those features to save computational time

qiime feature-table filter-seqs --i-data rep-seqs-cr-85.qza --i-table filtered-table-3.qza --o-filtered-data filtered-sequences-2.qza

Visualizing the filtered taxonomy

qiime taxa barplot --i-table filtered-table-3.qza --i-taxonomy agrf-taxonomy.qza --m-metadata-file manifest.tsv --o-visualization taxa-bar-plots-1.qzv



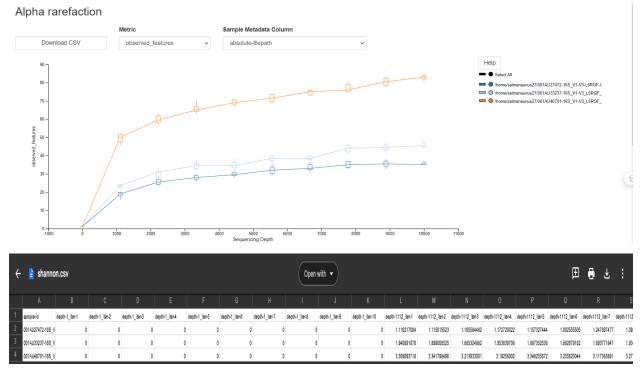
9. Generating a phylogenetic tree to understand evolutionary relationship between DNA and sequences

Creating an insertion tree

qiime phylogeny align-to-tree-mafft-fasttree --i-sequences filtered-sequences-2.qza --output-dir phylogeny-align-to-tree-mafft-fasttree

10. Diversity Plots

qiime diversity alpha-rarefaction --i-table filtered-table-3.qza --i-phylogeny phylogeny-align-to-tree-mafft-fasttree/rooted_tree.qza --p-max-depth 10000 --m-metadata-file manifest.tsv --o-visualization child-alpha-rarefaction.qzv



More csv output files can be found here - drive link

Alpha & Beta diversity

qiime diversity core-metrics-phylogenetic --i-table filtered-table-3.qza --i-phylogeny phylogeny-align-to-tree-mafft-fasttree/rooted_tree.qza --p-sampling-depth 3400 --m-metadata-file manifest.tsv --p-n-jobs-or-threads 1 --output-dir child-norep-co re-metrics-results

Weighted Unifrac distance



Statistical tests on diversity