## About the project

The end goal of this project is to classify patients with high protein concentration in urine and the healthy group based on SERS (Surface Enchanced Raman Spectroscopy) spectral data and biomedical data.

This project is to be released as a research paper later in 2022 or 2023. Some information is not fully shown here as a result.

The project is divided into several Jupyter notebooks with the following names: 1) Import raw urine spectra (part 1) 2) Spectra processing (part 2) 3) Classification of patients (part 3)

Author of all codes: Sultan Aitekenov, sultanaitekenov@gmail.com

Part of the upcoming abstract: Excessive protein excretion in human urine is an early and sensitive marker of diabetic nephropathy, primary and secondary renal disease. Kidney problems, particularly chronic kidney disease, remain among the few growing causes of mortality in the world. Therefore, it is important to develop efficient, expressive, and low-cost method for protein determination. Surface enhanced Raman spectroscopy (SERS) methods are potential candidates to achieve those criteria. In this paper, the SERS methods was developed to distinguish patients with proteinuria and the healthy group. Commercial gold nanoparticles with the diameter of 60 nm and 100 nm, and silver nanoparticles with the diameter of 100 nm were employed. Silver, gold, silicon and test slides covered with aluminium tape were utilized as substrates. Obtained spectra were analysed with several machine learning algorithms coupled with the PCA, ROC curve, and cross-validation methods.

# Import raw urine spectra (part 1)

The end goal of this notebook is to create a dictionary file that contains information about each patient (keys) and their respective spectra (values). This script runs for 181 sec.

#### Imports important modules

```
In [1]: # imports modules
  import pandas as pd
  import numpy as np
  import matplotlib.pyplot as plt
  import glob # imports to find nested files
  import os
  import re
```

#### Searches for file names and detects corrupted files

In [2]:

# search for file names

path = "Data raw urine spectra/"

substrate\_folders = glob.glob(path + "/\*\*NPs")

The spectral data is saved in hundreds of txt files. The txt files were generated by the software called LabSpecthe that operates a Raman microscope.

```
patients folders = glob.glob(path + "/**NPs/**[0-9]")
         txt naming = glob.glob(path + "/**NPs/**[0-9]/*.txt")
In [3]: # visualize txt naming, first 5 entries
        txt_naming[:5]
        ['Data raw urine spectra\\Ag_100nm_AgNPs\\1\\1-1.txt',
Out[3]:
         'Data raw urine spectra\\Ag_100nm_AgNPs\\1\\1-2.txt',
         'Data raw urine spectra\\Ag 100nm AgNPs\\1\\1-3.txt',
         'Data raw urine spectra\\Ag 100nm AgNPs\\1\\2-1.txt',
         'Data raw urine spectra\\Ag 100nm AgNPs\\1\\2-2.txt']
        From the output below, a reader can understand how the data is organized. The cell below finds
        "corrupted" files.
        # Finds corrupted files and save them as strings in a list. Some files are corrupted.
In [4]:
        # This block finds that files by bytes size. Corrupted files take more than 1 mb.
         corrupted files = []
         for i in range(0, len(txt_naming)):
             check size=os.stat(txt naming[i])
            size=check size.st size
            if size > 1000000:
                 corrupted_files.append(txt_naming[i])
         corrupted_files
        ['Data raw urine spectra\\Ag_100nm_AgNPs\\1\\3-1.txt',
Out[4]:
         'Data raw urine spectra\\Ag 100nm AgNPs\\107\\3-1.txt',
         'Data raw urine spectra\\Ag_100nm_AgNPs\\108\\1-1.txt',
          'Data raw urine spectra\\Ag_100nm_AgNPs\\116\\2-2.txt',
         'Data raw urine spectra\\Ag 100nm AgNPs\\62\\3-1.txt',
         'Data raw urine spectra\\Ag_100nm_AuNPs\\108\\2-1.txt',
         'Data raw urine spectra\\Ag_100nm_AuNPs\\110\\3-1.txt',
         'Data raw urine spectra\\Ag 100nm AuNPs\\115\\2-1.txt',
         'Data raw urine spectra\\Al tape 100nm AuNPs\\113\\1-1.txt',
          'Data raw urine spectra\\Al tape 60nm AuNPs\\104\\1-1.txt',
         'Data raw urine spectra\\Au_60nm_AuNPs\\36\\3-1.txt']
        # Remove corrupted files by finding their names (filepath).
In [5]:
        for i in range(0, len(corrupted_files)):
            txt naming.remove(corrupted files[i])
```

#### Create dictionaries: Keys - patients, Value - spectra

Firstly, a function that organizes data into a dictionary is created.

```
3-3.txt - contains spectral data
Input:
1) files_path
2) substrate - a string with only these arguments:
Ag_100nm_AgNPs,
Ag 100nm AuNPs,
Al_tape_60nm_AuNPs,
Al_tape_100nm_AuNPs,
Au 60nm AuNPs,
Au 100nm AuNPs,
Si 60nm AuNPs
Output:
dictionary with Keys - patients, and Values - spectra
#find a relevant list containing relevant paths
rel path = []
for file path in files path:
    a = re.search(substrate, file path) #finds patients within a single set of a s
    if a != None:
        rel path.append(file path)
rel_files_path = rel_path
# create empty dictionary with numeric key values to delete later
# our research group does not have ID higher than 200, so 300 would be more than e
dict raw spectra = {}
for i in range(0,300):
    dict_raw_spectra[i] = []
# create matrix from relevant path
for file path in rel files path:
    # keys in dict
    y = re.findall(r"\([0-9]*)\", file_path)
    key = int(y[0])
    # values in dict
    value = pd.read table(file path)
    value_sliced = np.array(value.iloc[:,2:])
    # converts to dt
    # if statement prevents files with incorrect numbers of row to pass
    if value sliced.shape[0] <= 100:</pre>
        dict_raw_spectra[key].append(value_sliced)
# delete keys without values, if keys have values, this script concatenates all va
for key in list(dict raw spectra.keys()):
    if len(dict_raw_spectra[key]) == 0:
        del dict raw spectra[key]
    else:
        value=np.concatenate(dict raw spectra[key])
        dict raw spectra[key]=value
return dict raw spectra
```

The cell below creates dictionary for each substrate. Since 7 experimental sets are available, each of them are saved into the single dictionary called 'raw\_urine\_spectra'

```
In [7]: # create a dictionaries for each substrate (experimental set)
```

### Saves output dictionaries into pickle

```
In [8]: # save the output dictionary into a pickle file
import pickle
with open("raw_urine_spectra.pkl","wb") as file:
    pickle.dump(raw_urine_spectra,file)
```