

# **STA 426: Diffusion maps for high-dimensional single-cell analysis of differentiation data**

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# Motivation

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# Single Cell Variability and Differentiation

- Variability between cells and measurement error – Noise
- Differential expression of genes steering differentiation – Signal
- Continuous vs. abrupt, switch like gene expression change

# Questions

- Which genes change in their expression due to differentiation?
- Where along the differentiation trajectory does a cell lie?

# Statistical Model

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- Let  $n$  be the number of all cells and  $G$  the number of all genes measured
- $\mathbf{x}$  is the position of the cell in the multi-dimensional space  $\mathbb{R}^G$
- Isotropic Gaussian wave function:

$$\mathbf{Y}_{\mathbf{x}}(\mathbf{x}') = \left(\frac{2}{\pi\sigma^2}\right)^{1/4} \exp\left(-\frac{\|\mathbf{x}' - \mathbf{x}\|^2}{\sigma^2}\right)$$

# Diffusion Transition Probability Matrix

- Interference between two cells  $\mathbf{x}$  and  $\mathbf{y}$ :

$$\int_{-\infty}^{\infty} \mathbf{Y}_{\mathbf{x}}(\mathbf{x}') \mathbf{Y}_{\mathbf{y}}(\mathbf{x}') d\mathbf{x}' = \exp\left(-\frac{\|\mathbf{y} - \mathbf{x}\|^2}{2\sigma^2}\right)$$

- Transition probability:

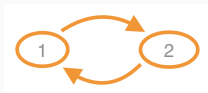
$$P_{xy} = \frac{1}{Z(\mathbf{x})} \exp\left(-\frac{\|\mathbf{y} - \mathbf{x}\|^2}{2\sigma^2}\right),$$

$$\text{where } Z(\mathbf{x}) = \sum_{i=1}^n \exp\left(-\frac{\|\mathbf{y}_i - \mathbf{x}\|^2}{2\sigma^2}\right)$$

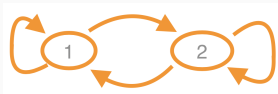
# Markov Chain Transition Probability Matrix

Two state Markov chain:

$$\begin{pmatrix} 0 & 1 \\ 1 & 0 \end{pmatrix}$$



$$\begin{pmatrix} 0.3 & 0.7 \\ 0.2 & 0.8 \end{pmatrix}$$





# Diffusion Maps – Quantities

- Diffusion components
- Diffusion coefficients
- Assumption: Diffusion coefficients drop to a noise level after the first few components

# Missing and Censored Values

- Decompose the kernel of the Gaussian wave into  $G$  components
- Replace missing/censored values with hypothetical distribution

# Determination of the Gaussian Kernel Width

- The average dimensionality of the manifold is equal to

$$\mathbb{E}_{\mathbf{x}}[d(\sigma)] = \frac{\partial \mathbb{E}_{\mathbf{x}}[\log(Z(\mathbf{x}))]}{\partial \log(\sigma)}$$

- Choose  $\sigma$  such that  $\mathbb{E}_{\mathbf{x}}[d(\sigma)]$  is maximized
- Possibly multiple maxima

## Comparison with Other Methods

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## Principal Component Analysis (PCA)

- Orthogonal transformation of the data
- Works best for linear data subspace
- Non-linear extensions (Kernel-PCA)

# Dimension Reduction Methods for Single-Cell Omics Data

## Principal Component Analysis (PCA)

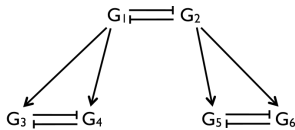
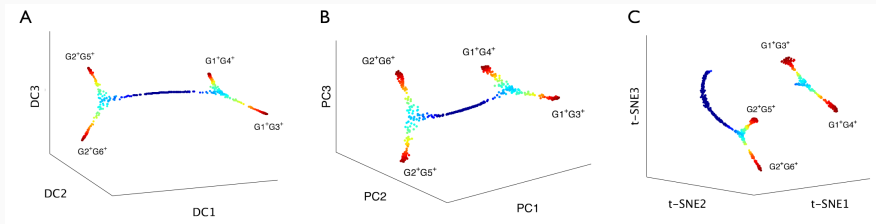
- Orthogonal transformation of the data
- Works best for linear data subspace
- Non-linear extensions (Kernel-PCA)

## t-distributed Stochastic Neighbor Embedding (t-SNE)

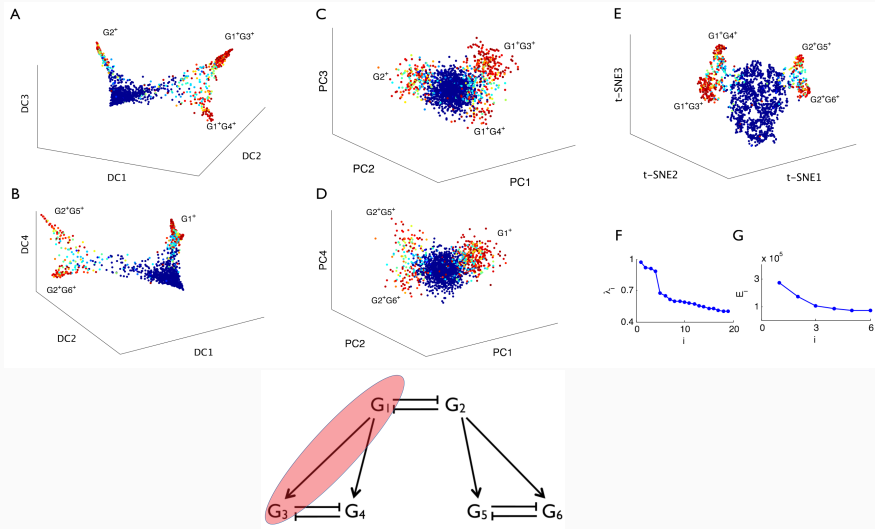
- Probabilistic, non-linear method
- Very robust to noise / density heterogeneities
- Tuning parameter perplexity

Many more methods exist!

# Simulated Data – Balanced

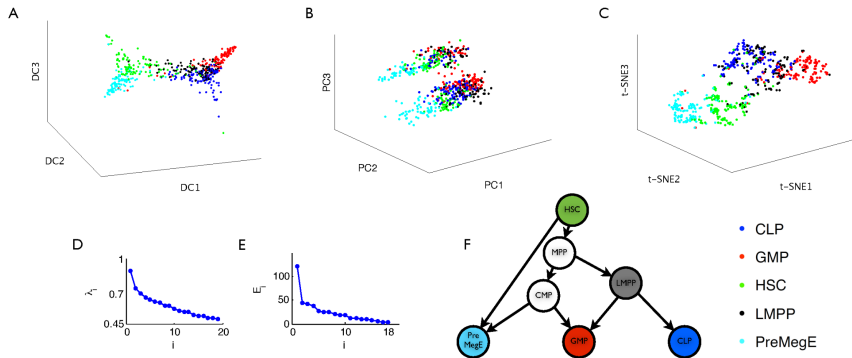


# Simulated Data – Imbalanced + Noise





# qPCR Data – Haematopoietic Stem Cells



# Results

- + Diffusion maps outperform other methods in finding cell differentiation trajectories
- + Diffusion maps are robust to sampling density heterogeneities and noise

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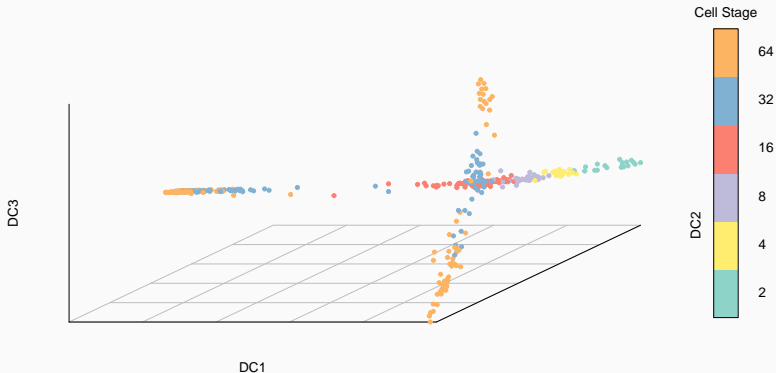
- + Diffusion maps outperform other methods in finding cell differentiation trajectories
- + Diffusion maps are robust to sampling density heterogeneities and noise
  - In all examples  $\sigma^2$  was finetuned
  - Unclear if finetuning perplexity parameter in *t*-SNE method would lead to similar results
  - Number of diffusion components needed for visualization not known in advance

# R Demonstration

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# destiny Package

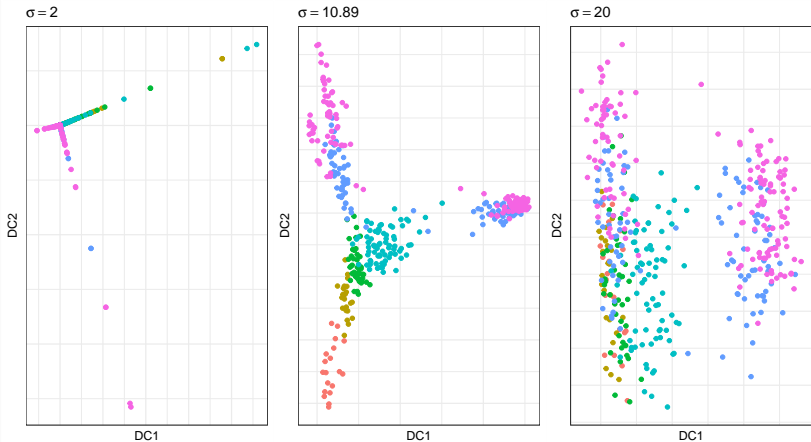
```
set.seed(12)
library(destiny)
data("guo_norm")
dm_guo <- DiffusionMap(guo_norm)
plot(dm_guo, pch = 20, col_by = "num_cells", legend_main = "Cell Stage")
```



# Choice of $\sigma^2$

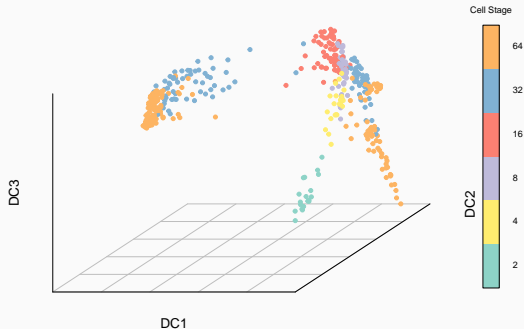
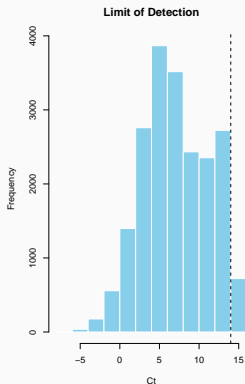
```
sigmas <- find_sigmas(guo_norm, verbose = FALSE)
optimal_sigma(sigmas)
```

```
## [1] 10.8946
```



# Censored/Missing Values

```
layout(matrix(c(1, 2, 2), ncol = 3))
exprs(guo_missing)[sample(length(exprs(guo_missing)), 100)] <- NA
dm_guo_missing <- DiffusionMap(guo_missing, verbose = FALSE,
                              censor_val = 14, censor_range = c(14, 40),
                              missing_range = c(-5, 10))
hist(exprs(guo_norm), main = "Limit of Detection", xlab = "Ct", col = "skyblue", border = "white")
abline(v = 14, lty = 2)
plot(dm_guo_missing, pch = 20, cex.symbols = 1.5, col_by = "num_cells", legend_main = "Cell Stage")
```



## Conclusions

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- + Handle high noise levels, missing data, sampling heterogenitites
- + Diffusion distance is a biologically relevant distance metric
- + Capture nonlinear/complex differentiation dynamics

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  - Number of significant dimension not determinable in advance
  - Finetuning of  $\sigma^2$  required (but there is a proposed criterion)
  - $n^2 \times G$  computation time (but there are approximate versions)

## Discussion

- + Handle high noise levels, missing data, sampling heterogenitites
  - + Diffusion distance is a biologically relevant distance metric
  - + Capture nonlinear/complex differentiation dynamics
    - Number of significant dimension not determinable in advance
    - Finetuning of  $\sigma^2$  required (but there is a proposed criterion)
    - $n^2 \times G$  computation time (but there are approximate versions)
- Powerful dimension reduction tool for single cell differentiation data

- Angerer, P., Haghverdi, L., Büttner, M., Theis, F., Marr, C., and Büttner, F. (2015). *destiny*: diffusion maps for large-scale single-cell data in R. *Bioinformatics*, 32(8):1241–1243.
- Coifman, R. R. and Lafon, S. (2006). Diffusion maps. *Applied and Computational Harmonic Analysis*, 21:5–30.
- Haghverdi, L., Büttner, F., and Theis, F. J. (2015). Diffusion maps for high-dimensional single-cell analysis of differentiation data. *Bioinformatics*, 31(18):2989–2998.