Application of Machine Learning Techniques to Next Generation Sequencing Quality Control

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Declaration

I certify that except where indicated, all material in this thesis is the result of my own investigation and references used in preparation of the text have been cited. The work has not previously been submitted as part of any other assessed module, or submitted for any other degree or diploma.

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Acknowledgements

Abstract

Over the past few years advances in genetic sequencing hardware have introduced the concept of massively parallel DNA sequencing, allowing potentially billions of chemical reactions to occur simultaneously, reducing both time and cost required to perform genetic analysis[1]. However, these "next-generation" processes are complex and open to error[2], thus quality control is an essential step to assure confidence in any downstream analyses performed.

This project investigates the accuracy of a range of decision tree classifiers for a series of quality control data sets and shows it is possible to generate minimal yet accurate models, creating trees which closely resemble the behaviour of an already existing QC system.

This project also introduces **Frontier**, a Python package which provides users with interfaces for the reading, storage and retrieval of large machine learning data sets; **Goldilocks**, a Python class which can be used to find a region on a genome that expresses a desired density of variants across different studies; and bug fixes to an internal tool used as the Sanger Institute and existing widely used open-source bioinformatics software.

Contents

Chapter 1

Introduction

Over the past few years advances in genetic sequencing hardware have introduced the concept of massively parallel DNA sequencing, allowing potentially billions of chemical reactions to occur simultaneously, reducing both time and cost required to perform genetic analysis[1]. However, these "next-generation" processes are complex and open to error[2], thus quality control is an essential step to assure confidence in any downstream analyses performed.

1.1 Project Aims

The project consists of two sub-projects:

- Analysis of a current quality control system in place

- Identification of quantifiable sample properties that affect downstream analysis

1.1.1 Analysis of Current System

With the support of the Wellcome Trust Sanger Institute in Cambridge, this project works with the Human Genetics Informatics team to investigate **auto_qc**[3], the institute's current automated quality control tool.

During genetic sequencing a large number of metrics are generated to determine the quality of the data read from the sequencing hardware itself. As part of the institute's vertebrate sequencing pipeline[4], **auto_qc** is responsible for applying quality control to samples within the pipeline by comparing a modest subset of these metrics to simple hard-coded thresholds; determining whether a particular sample has reached a level that requires a warning, or has exceeded the threshold and failed entirely. Whilst this catches most of the very poor quality outputs, a large number of samples are flagged for manual inspection at the warning level; a time consuming task which invites both inefficiency and error[5].

In practice most of these manual decisions are based on inspecting a range of diagnostic plots, which suggest that a machine learning classifier could potentially be trained on the combinations of quality control statistics available to make these conclusions without the need for much human intervention.

The first part of the project aims to apply machine learning techniques to replicate the current **auto_qc** rule set by training a decision tree classifier on a large set of these quality metrics. The idea is to investigate whether these simple threshold based rules can be recovered from such data, or whether a new classifier would produce different rules entirely. During this analysis it is hoped the classifier may be able to identify currently unused quality metrics that improve labelling accuracy. An investigation on the possibility of aggregating or otherwise reducing the dimensions of some of the more detailed quality statistics to create new parameters will also be conducted.

1.1.2   Identification of Properties that affect Downstream Analysis

The other half of this project is motivated by the question "What *is* good and bad in terms of quality?"

To be able to classify samples as a pass or a fail with understanding, we need an idea of what actually constitutes a good quality sample and must look at the effects quality has on analysis performed downstream from sequencing. An example of such is **variant calling** – the process of identifying differences between a DNA sample (such as your own) and a known reference sequence.

Given two high quality data sources where DNA sequences from individuals were identified in two different ways (one of which being next-generation sequencing) it would be possible to measure the difference between each corresponding pair. Using this, we could investigate the effect of leaving out part of the next-generation sample during the variant calling process. If we were to leave a part of a sample out of the variant calling pipeline would the variants found be more (or less) accurate than if it had been included? Would they agree more (or less) with the variants called after using the non next-generation sequencing method?

Having identified such sub-samples, can quality control metrics from the previous part be found in common? If so, such parameters would identify "good" or "bad" samples straight out of the machine. Samples that exhibit these quality variables will go on to improve or detriment analysis.

1.2   Project Method

1.2.1   Methodology

Clearly some team-based practices invited by agile methodologies – pair programming immediately comes to mind – are not applicable in a solo project. It is also unreasonable to expect an "on-site" customer for this particular project. In *The Case Against Extreme Programming*, Matt Stephens describes a "self referential safety net" where the perceived traps in each practice are supported and "made safe" by other extreme programming (XP) practices. This would rule out XP as a viable methodology for a solo project as cutting out some of the processes that allow this form of evolutionary design to work (and flatten that

cost-of-change curve) can introduce serious flaws to the management of a project and potentially result in failure. In the same breath it is important to remember that not all agile processes need be discarded just because XP seems incompatible. Indeed, some processes are common sense, for example: frequent refactoring, simple design, continuous integration and version control. Test driven development could also prove a useful process to consider as part of a methodology for this project as setting up a framework that allows for quick and frequent testing (before coding) and ensuring that any refactoring has a positive (or at least non-negative) effect on the system could be a worthwhile contribution to efficiency.

Could a more plan driven approach or form of agile-plan hybrid be considered appropriate here? In *Balancing Agility and Discipline*, Boehm and Turner introduce the idea of "homegrounds" for both agile and plan driven approaches; noting here that for projects that require high reliability and feature a non-collocated "CRACK customer" in fact align with some of these homegrounds for plan driven development. Combined with the thought that the project requirements will also be relatively stable it would seem that there may be no reason to switch to a more agile methodology as its primary feature is the welcoming of change that is not even needed? Perhaps this is the naivety of an optimist.

Personally I think I would approach this with a form of agile-plan hybrid; I like the idea of quick iterations and getting feedback as opposed to leaving acceptance testing until the end of the project, but I also want a somewhat detailed feedback process. In Neil Taylor's *Agile Methodologies* course it was suggested that it is dangerous to pick and choose processes (don't anger the Ring of Snakes) and also merely paying "lip service" to agile must be avoided (otherwise what's the point?), I feel that on this occasion it can be justified by the size of the project itself.

This project will consist of many research steps, each requiring some form of computational process to prepare the data for the next step. Whilst the implementations of the algorithms themselves pose computational complexity, there appears to be little challenge from a planning perspective and in fact a looser overall plan should be considered as we must account for unforeseen and unexpected outcomes from each research step.

The most important part of ensuring this project stays on track will be the development of a sensible testing methodology to ensure we are not only moving in the right direction in terms of which algorithm and parameters to use but also in terms of reliably measuring performance over time in a way that allows justification of such design choices.

Despite this trail of thought, given the research grounding this project entails it might be required to look beyond traditional and even modern software development methodologies and investigate a more scientific approach. A simple scientific method would involve establishing a null hypothesis that can be proven false by testing (*e.g.* "**auto_qc** classifier is more accurate than the new classifier") and executing experiments that attempt to prove this null hypothesis false in favour of an alternative hypothesis (typically the opposite, *e.g.* "The new classifier is more accurate than the old classifier"). This form of hypothesis testing could essentially become the project's acceptance tests (providing we have an empirical definition of what "more accurate" means in terms of this system) and any modification can be classed as an experiment ("Do these parameters allow us to reject the null hypothesis?"). Although care must be taken not to let this descend into unstructured cycles of mere hack-and-test, code-and-fix style programming.

Overall it is rather difficult to select a methodology for a project such as this as the research element makes it

almost impossible to draw on previous personal experience for ideas of what development processes would be effective.

1.2.2 Task Management

It is useful to be able to keep track of current tasks preferably via a medium that would allow some method of sorting and filtering. For various personal projects and the second year group project I have used **Redmine**, a Ruby-based web application designed for bug tracking. However over time I have come to find keeping the task information stored in Redmine a task in itself. Attempts to extend the platform to implement additional functionality have been fruitless.

Other alternatives included **TaskWarrior**, whose simple but effective command line interface was overshadowed by its occasional storage corruptions. For a lack of other options that met my requirements, I wrote my own open source web based task management application; **Triage**[6]. This will be useful in keeping track of current objectives and allow prioritisation in a quick and simple manner.

The list used to organise my project is also publicly available[7] for transparent progress tracking by my supervisor and those interested at the Sanger Institute.

1.2.3 Time Considerations

It must be remembered that this project needs to meet the requirements for a *minor* project and is to be completed alongside the study of several other modules which each have their own assignments and obligations. It would be easy to become overly ambitious and thus aims and goals will need to be revised as both obstacles and breakthroughs are encountered over the lifetime of the project.

Part I

Analysis of Current System

Chapter 2

Introduction and Background

2.1 Introduction

This part of the project can be outlined as follows:

- Collect data sets on which a machine learning classifier is to be trained

- Construct a program capable of processing and storing such data sets such that required subsets of the data can be quickly and easily returned for further analysis

- Select a suitable machine learning framework to handle the training and validation of a classifier

- Ensure a robust validation methodology exists for assuring quality of our own results

- Set up an environment capable of allowing results from such a classifier to be stored and compared

- Training a suitable classifier on the collected data sets

- Perform experiments by selecting subsets of the variables and observations and measure whether classification accuracy is improved

2.2 Concepts and Terminology

See Appendix A.1.

Chapter 3

Materials and Methods

3.1 Input Data and Format

3.1.1 "BAMcheckR'd" Data

As part of the project I have been granted access to significant data sets at the Sanger Institute, unlocking quality control data for two of the largest studies currently undergoing analysis. A wide array of quality metrics are available for each and every lanelet that forms part of either of the two studies, totalling 13,455 files: 9154 (68%) passes, 1542 (11%) fails and 2759 (21%) warnings.

The files are created by **samtools stats** – part of a collection of widely used open-source utilities for post-processing and manipulation of large alignments such as those produced by next-generation sequencers that are released under the umbrella name of "SAMtools"[8] (Sequence Alignment and Map Tools). **samtools stats** collects statistics from sequence data files and produces key-value summary numbers as well as more complex tab delimited dataframes tabulating several metrics over time.

The output of **samtools stats** is then parsed by an in-house tool called **bamcheckr**[1] which supplements the summary numbers section of the **samtools stats** output with additional metrics that are later used by **auto_qc** for classification. This process appends additional key-value pairs in the summary numbers section. A truncated example of a "bamcheckr'd" file can be found in Appendix B.1.

It is these summary numbers that will be the main focus of our learning task.

3.1.2 auto_qc Decision Data

To use these "bamcheckr'd" files for training and testing a machine learning classifier, it is necessary to map each file to a classification result from **auto_qc**. The one-to-one mapping between each input file and its label

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[1] Named such as **samtools stats** now incorporates **bamcheck** and the tool is written in R

are provided by the Sanger Institute in a separate file hereafter referred to as the *AQC Decision Matrix* or *AQC (Decision) File*.

A truncated example of such a file can be found in Appendix B.2. Only the first few columns are included – indeed we are only interested in the *lanelet* and *aqc* fields which provide an identifier that maps the row to a given input file and its classification by **auto_qc** respectively. Latter columns pertain to a breakdown of decisions made by **auto_qc** which are not included in the example for confidentiality (and brevity).

3.2 Development Environment

3.2.1 Language

Python was selected for the language of the program designed to handle this vast array of input data, more out of personal taste rather than a detailed analysis of required performance and features. From previous experience I was happy with the performance of Python when processing large datasets in terms of both file handling operations and storing the data in memory for later use. Python's generous choice of both built-in and third-party libraries have proven useful. Due to its concise and flexible nature it is possible to rapidly develop applications and its readability eases ongoing maintenance; useful given the short time-span allocated for this project and the possibility of others wishing to contribute to the project codebase after completion.

Whilst the choice was made primarily on preference, this is not to say other options were not considered: a highly popular Java-based collection of data mining tools, **WEKA**[9] would certainly have provided a framework for building decision tree classifiers but did not appear to offer any significant features that were unavailable elsewhere, whilst Java itself has the added constraint of requiring a virtual machine to be installed which could be undesirable from a performance or security standpoint when the application is deployed to servers at the Sanger Institute.

Difficulty was also encountered finding example implementations for **WEKA** with most documentation and tutorials providing information for performing analysis via the graphical "Explorer" interface instead, which would not be appropriate for quickly setting up and repeating experiments automatically.

Given the quality data we'll be using to train a machine learning classifier is output from the previously mentioned R script, **bamcheckr**, it was worth briefly investigating the options available for R itself as the potential of integrating the learning and predicting functions right in to the same process that outputs the data seemed convenient.

Whilst the **tree**[10] and **rpart**[11] packages are available for constructing decision trees in R (and actually **RWeka** provides an R interface to **WEKA**) neither appeared to be as robust as other more well-known frameworks. Also putting it politely, the programming paradigm of R[12] is rather different to other languages and can significantly increase development time (and frustration[13]) if one is not well versed in the patterns and grammar of the language and it seemed best to stick to one's comfort zone given the brief timescale for the project.

Had performance been a critical decision factor, lower level languages such as C, C++ or even Fortran could be

used. Briefly looking at frameworks available for C++ in particular, two popular solutions include: **dlib**[14], which did not support tree-based classifiers but did offer implementations of other potentially useful algorithms such as multi-class support vector machines; and **Shark**[15], which supported both decision tree and random forest classifiers. Both packages also provided a series of utility functions for performing mathematical or statistical operations on vectors of data.

3.2.2 Framework

Having studied the *Machine and Intelligent Learning* module in my final year, the prospect of getting stuck in to the deep of a machine learning algorithm was exciting. However the reality is a lot of cumulative time and effort has gone in to the creation and optimisation of a framework, which is unlikely to be surpassed successfully by a short-term one-person project. Thus utilisation of a third party machine learning library seems a wise investment for the project's codebase.

There are clearly numerous machine learning frameworks available in many languages, some of which were touched upon in the previous section and formed part of the development environment decisions. Whilst it is obviously unnecessary to select a framework which uses the same language as the project, it seemed counter-intuitive to select otherwise, for the establishing of additional arbitrary output and input steps to move data between the two environments could impede prompt experiment repeatability and introduce error.

A mixed bag of machine learning frameworks exist in Python[16], ranging from several highly active general purpose libraries to a multitude of smaller projects that focus on one specific learning task.

I investigate two of the larger libraries; **scikit-learn**[17] and **Orange**[18], primarily as their general purpose nature will allow exploration of various classifier solutions (given time) without the need to integrate many packages together, but also for their built-in functionality that aids the measuring of classifier performance and accuracy. These two packages were also selected for investigation on their recommendation from the project supervisor.

scikit-learn is somewhat the "new kid on the block" in terms of machine learning packages for Python, originally starting as a Google Summer of Code project in 2007 under the name of **scikits.learn**[2] the package was not officially published until 2010 where it quickly gained popularity and built an international team of contributors[19].

Arguably one of the most useful features of **scikit-learn** is its seamless integration with the "big names" of scientific computing in Python: **NumPy**[20] and **SciPy**[21], making heavy use of the optimised data structures and mathematical algorithms these packages offer, not only providing improved performance against packages that implement their own structures but also easing interaction by not requiring users to populate abstruse structures with their data or implement such algorithms themselves.

scikit-learn offers support for a wide range of algorithms covering many different machine learning tasks, including decision trees and random forests. Also of interest are the various utility subpackages, taking particular note of one for measuring the performance of the various classifiers; including functions for executing

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[2]Named as a **SciPy** Toolkit for Learning Tasks

cross-validation and generating confusion matrices; and another which assists with the selection of parameters (features) to use in a machine learning model.

The library boasts a growing community of users and a dedicated team of developers[3] who submit hundreds of commits to the project repository each month according to Github Pulse.

Orange, like **scikit-learn** is designed to be a general purpose machine learning framework and provide a wide array of tools to work with many different types of problems. Setting it apart from many of the other packages is the inclusion of a graphical user interface (GUI) that presents drag-and-drop-esque access to widgets from which users assemble a workflow to apply to their data. The GUI also allows visualisation of elements such as decision trees or confusion matrix plots without any effort from the end user.

At first glance **Orange** appears to offer a larger collection of functions to work specifically with classification tasks when compared to **scikit-learn**. Indeed it would later be discovered that it ships with features useful to the project that have not been implemented in **scikit-learn** including tree pruning and printing.

However it does not feature the same integration with Python's scientific computing packages; **NumPy** and **SciPy** and also requires that end users input their data in a structure defined by the framework rather than a generic data structure. Thus use of **Orange** would introduce more data wrangling and munging steps to ensure that data is represented in the correct format for a classifier to perform its learning and prediction role and also to interpret returned objects.

Although **Orange** affords a suitable (and in some cases better) range of functions to use for decision trees, I was concerned at the push to use its GUI as opposed to the package API, potentially impeding experiment repeatability by having to reset the graphical environment each time.

Despite the **scikit-learn** package not yet reaching a major version milestone, its feature set, interaction with **NumPy** and **SciPy** and useful built-in subpackages led me to choose **scikit-learn** as the machine learning framework for this part of the project.

3.2.3   Additional External Libraries

As discussed, the project will make use of the open source[4] **SciPy Stack**, consisting of several core packages including **NumPy**[20]: a package for efficient numerical computation which defines generic N-dimensional array and matrix data containers; and **SciPy**[21], which provides a multitude of optimised numerical routines.

Whilst the stack also includes **Matplotlib**, a highly popular graphing package, I typically prefer to use the R package **ggplot2**[24] for creating graphs. However it is useful to note that **Matplotlib** integrates easily with the data structures of **NumPy** (and thus **scikit-learn** too).

argparse is a third-party Python library[5] used for specifying the arguments and options of a command line interface via a simple API. **argparse** spares the developer from having to check the presence and validity of a

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[3]Some full time developer positions are funded by *INRIA*, the French Institute for Research in Computer Science and Automation
[4]Components of the **SciPy Stack** are distributed under the 3-clause Modified BSD License[22][23]
[5]Although since Python 2.7, **argparse** has been included as part of the standard library[25]

user's arguments to a Python program themselves whilst also automatically generating a friendly interface to the program for the end user based on the definitions provided by the developer.

3.2.4 Testing

As discussed in Chapter 1.2.1, testing forms a critical part of the project given the need to monitor the impact of changes to classification accuracy as well as to ensure the program is working correctly. Ideally, execution of a test suite should be simple and easily repeatable. Results that pertain to accuracy should also be stored for future reference to monitor ongoing performance of the classifier.

Such requirements could be fulfilled by a continuous integration platform – a server dedicated to the building and testing of the code contained in a centralised repository typically to which an entire team will have write access[26]. Whilst in this scenario there will be much less "risk" from integration issues due to the single person team size, the themes of automated building and self-testing code can still be taken on board.

Jenkins[27] is a highly popular[28] example of such a platform with which I am familiar. Although an out-of-the-box **Jenkins** instance is suitable for variety of software engineering projects, it would be necessary to invest some time to install and tweak plugins to perform actions on test results (such as failing a build that causes accuracy to decrease). Previous experience found that more specific tasks will often require a plugin to be authored to overcome limitations in the feature set of a more generic plugin, which given the intricacies of the **Jenkins** package layout could easily turn in to a project of its own. Unfortunately, other features that would be useful to the project including the indexing and searching of logs are somewhat lacking in **Jenkins**.

Online solutions such as **Travis**[29] and **Wercker**[30] are free for open source projects and could potentially offer a quicker set up, as both services merely require a small configuration file in the root of the repository and a hook to be registered (allowing builds to be triggered on a code push for example) before being able to deploy a build in the cloud.

However these online services would not have been able to handle the return of build artifacts (such as logs, graphs or dot files containing a decision tree) without some convoluted solution of uploading them to another service or committing them to a private code repository during execution of the job – the non-persistent nature of these nodes would cause any artifacts to be destroyed when the node is terminated at the end of the build.

Also, as these virtual nodes are isolated from any sort of persisting file system it would be necessary to upload large quantities of training data any time a build and test job is to be run. This sounds rather inefficient in terms of both bandwidth and time and would more than likely constitute unreasonable use in view of the platform's "fair use" terms and conditions too.

It would seem that none of the widely available solutions would provide an environment suitable for the testing of this project. I wanted to provide my own platform for this problem but given the time constraints of the project this was simply not practical and in the end we settled for well formatted log files which could be searched and processed with command line tools such as **awk**.

Chapter 4

Frontier

This chapter introduces **Frontier**: the main programming effort for this part of the project. **Frontier** is a Python package that serves as a data manager, providing both interfaces to read inputs into structures in memory and to retrieve them in formats acceptable to a machine learning framework.

Appendix D describes some analysis performed before implementation of **Frontier**.

4.1 Design

4.1.1 Purpose

Frontier's purpose is to supplement analysis with **scikit-learn** by allowing a user to read in and parameterise data in a format that can then be used for analysis by the **scikit-learn** library. **Frontier** is designed to simplify the process of setting up machine learning tasks and enable experiment repeatability by removing the need for users to spend time constructing classes and functions to parse their input data and to just get on with analysis using **scikit-learn**.

Initially **Frontier** was to act as a wrapper around **scikit-learn**, essentially removing the end user's interaction with the library and merely providing an interface for data to be passed in and some sort of classifier to be returned. However this quite clearly limited **Frontier**'s audience by tying it to a particular framework and would quickly become unmanageable in the task of providing wrappers for all aspects of an external library.

Instead is far better to allow **Frontier** to be used alongside a user's chosen machine learning framework, providing a useful API to parse and extract observations and variables from input data and arrange them in structures suitable for processing with **scikit-learn**.

If possible, **Frontier** could also handle any objects returned, displaying or logging any textual or graphical information pertaining to a returned classifier's accuracy to assist a user in the ongoing performance monitoring of changes to used data subsets or parameters.

4.1.2 Format

Frontier is designed as a Python package, allowing a user to import its functionality into other programs. The result of this project's technical output could almost be considered as two separate entities: **Frontier** itself, the package designed to ease user interaction with scikit-learn and **Front**, a Python script which implements **Frontier**'s functionality in order to interact with scikit-learn to conduct analysis on the **auto_qc** data.

4.1.3 Method

Frontier is the result of abstracting code and tools created during the use of **scikit-learn** for analysis of the current **auto_qc** system. Whilst scripts initially hard-coded to suit the specific needs of the project – with hard coded classes and encodings – **Frontier** followed an evolutionary design process and grew to become a more generic framework applicable to learning tasks outside of this study.

4.2 Concepts

Appendix A.2 introduces some ideas that will assist understanding of the application's purpose and terminology used in the following implementation section.

4.3   Implementation

This section investigates the implementation of **Frontier**'s major components:

- Informing the package of the problem domain...

- Interfaces provided for reading in data

- Storage of read data in memory

- Retrieval of stored data for interaction with **scikit-learn**

4.3.1   Class Definitions

Frontier is designed to support classification machine learning problems, to adequately support this task, the package must be aware of each of the possible classes in the problem space. Early versions of **Frontier** were specifically designed for training and testing data from **auto_qc** and could only support encoding and decoding of the pass, fail and warn classes.

However this implementation was clearly esoteric and held little to no further use outside the domain of the project's learning task. What would happen if a class label were to be added or removed in future? Most likely many lines of code would need to be re-written to handle such cases; the package was inflexible.

To counter this, **Frontier** was refactored to remove hard coded label definitions enabling its use as a more general purpose tool where users can specify the domain's classes and their associated labels and encodings. Listing 1 shows the definitions used by Front to work with the **auto_qc** data.

```
CLASSES = {
        "pass": {
            "names": ["pass", "passed"],
            "code": 1,
        },
        "fail": {
            "names": ["fail", "failed"],
            "code": -1,
        },
        "warn": {
            "names": ["warn", "warning"],
            "code": 0,
        },
}
```

Listing 1 : Class definitions for **auto_qc** as passed to **Frontier**

14

As per Listing 1, to define classes a user must provide a Python dict containing the following for each label:

- **class** *String*
  The dictionary key is used as the canonical name of the class label

- **names** *[String]*
  A list of labels that denote membership of this class

- **code** *Integer*
  The encoded representation of this class (See Chapter A.2.3)

- **_recode** *Boolean, Private*
  A flag to indicate whether an API action has changed the code of this class, typically used when treating all members of one class as a member of another – A user should never set this manually

- **_count** *Integer, Private*
  The number of observations with this label, typically used when calculating weightings based on the proportions of class sizes and outputting logging information – A user should not set this manually

This simple structure allows **Frontier** to be compatible with almost any classification learning task with minimum input from the end user. Additional utilities provided by the **Frontier** utils subpackage use this structure to automatically classify and encode labels without user intervention with the following functions:

- **classify_label**
  Attempt to classify a label by comparing a given string to each set of *names*, locating an exact match will return the relevant canonical class label

- **encode_class**
  Given a canonical class label, return its *code*

- **decode_class**
  Given a *code*, return the canonical class label unless *_recode* is True

- **count_class**
  Increment the *_count* for a particular class given its canonical label

These functions are put to use throughout **Frontier** and are essential for reader classes (detailed in the next chapter) to parse, classify and then encode observation targets automatically from relevant files.

4.3.2   Input Handling

Frontier's modular nature allows users to write their own Python classes to read data from any form of input file or stream. Two examples of which are the classes used to read from the "bamcheckr'd files" documented in Appendix B.1 and the **auto_qc** decisions matrix briefly demonstrated in Appendix B.2.

These classes are described as **Readers** and implement a common base class, **AbstractReader** which takes care of setting up the file handler, including functions to both close and iterate over the file's contents. It will

also automatically call its own **process_file** function that skips over any header lines before passing each line in the file stripped of any newline characters to **process_line**.

It is expected that derived classes will at least provide their own implementations for **process_line** and **get_data**. Failing to do so will cause **Frontier** to throw a **NotImplementedError** when attempting to use the class to read data.

process_line defines the line handling operations that extract and store desired data found in a given line. This responsibility includes returning None for lines that contain comments and irrelevant data and conducting any necessary sanitisation (the **BamcheckReader** for example makes use of a utility function to translate spaces, underscores and dots to hyphens).

get_data must return any read in data in a suitable structure for storage by **Frontier**. Typically this will be a Python dictionary using some unique identifier for each observation as a key, mapping to an arbitrary value or object containing that observation's parameters. Further discussion on **Frontier**'s storage of data and targets is to follow.

The **AbstractReader** class is designed to simplify the process of reading in observations and their targets for end users, however it is still up to the author of the derived class to set up any structures to store data (which cannot be done automatically without likely enforcing potentially unhelpful constraints) before initialisation of the inherited base class (via the call to **super**) as shown in Listing 2.

```python
class BamcheckReader(AbstractReader):
    [...]

    def __init__(self, filepath, CLASSES, auto_close=True):
        self.summary = SummaryNumbers()
        self.indel = IndelDistribution()
        super(BamcheckReader, self).__init__(filepath, CLASSES, auto_close, 0)

    [...]
```

Listing 2 : Extract from **BamcheckReader** class documenting initialisation of necessary data structures and calling for initialisation of its inherited base class

As shown in Listing 2, the initialisation of the **AbstractReader** allows four arguments:

- **filepath** *String*
  A relative or absolute path to a file from which to extract data or targets

- **CLASSES** *Dictionary*
  A dictionary of user specified class labels defined as described in the previous chapter

- **auto_close** *Boolean, Optional*
  Whether or not to close the file handle immediately after executing **process_file**, this is True by default to prevent either memory leaking when users are reading in a large number of files and are perhaps

unaware that they require closing or the throwing of an IOError caused by having too many file handles open at once

- **header** *Integer, Optional*
  The number of lines to ignore before the reader should begin passing stripped lines to **process_line**, defaults to 0

Currently it is also the responsibility of the author of a derived class to perform relevant sanity checking of any extracted data. For example the **BamcheckReader** class checks for the presence of multiple entries of a particular metric which will print a notice if found, unless the entries have differing values, upon which an exception is thrown and the process is halted.

Once a reader has been defined for a particular file format, a user need only provide a directory of files to be parsed and the name of the class designed to complete the parsing. **Frontier** will then take care of executing the parsing process on all files in the directory. After a derived reader has completed file handling, **Frontier** will call its **get_data** function to "move"[1] the extracted data to its own storage.

At this point **Frontier** will also check the integrity of the data, primarily that all parameters have a non-zero variance[2]. Users will be warned when this requirement is violated; parameters with no variance cannot provide much information for successful classification as their values are equal for all class labels!

In future it would be useful to investigate whether it would be feasible to perform such sanity checking in a generic manner to ensure it could be applied to a wide enough range of scenarios to make it worth including functionality in the **AbstractReader** directly.

With more time, future improvements could overhaul the reader interfaces to allow users to simply specify the format of a file in a string that can be parsed by **Frontier**'s IO subpackage, rather than having to write their own derived class. Classes could also list file extensions they are capable of processing which could potentially be used to automatically determine which readers to use without requiring the user to explicitly specify.

4.3.3 Storage

Frontier specifies a class called the **Statplexer**[3] which provides users with a single point of access to all read in data. The reader interfaces described in the previous chapter implement their own loading functions to populate the **_data** and **_targets** class members of the **Statplexer** object, both of which are Python dictionaries.

During the parsing of observation data with the relevant reader class, the reader is expected to locate an appropriate unique ID for each observation. In the case of processing of **auto_qc** data, this would be the lanelet's barcode which is collected from the filename of that particular lanelet's "bamcheckr'd" file. This identifier is then used as a key in both the **_data** and **_targets** dictionaries to map to a structure (typically another dictionary or an arbitrary class) that stores that observation's parameters and its known target (encoded class label), respectively.

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[1]Rather, a pointer to the address of the extracted data's dictionary hashmap will be copied to memory inside a **Frontier** class
[2]Using an algorithm for accurately computing running variance introduced in Donald Knuth's *Art of Computer Programming*[31]
[3]A somewhat contrived contraction of 'Statistics Multiplexer'

Although these attributes can be manipulated directly (and indeed they are for testing purposes) the leading underscore follows a popular convention defined in Python's style guideline, PEP8[32], where class members with leading underscores should be treated as non-public. Python doesn't have private variables such as those that may be found in other languages like Java, indeed the Python style guide points out that "no attribute is really private in Python"[32]. In an interesting StackOverflow answer on the subject, a user describes that this is "cultural"[33] and that Python programmers are trusted not to circumvent convention and "mess around with those private members". Users are therefore expected to use the functionality **Frontier** provides for controlled getting and setting of data stored in these pseudo-private **_data** and **_targets** members.

load_data, for example, should be used exclusively when populating the two dictionaries and is automatically called on construction of the **Statplexer** if the construction arguments are valid. **load_data** will automatically call other necessary (pseudo-private) functions of the class including **_test_variance** which checks that parameter variances are non-zero and also warns users if new observations are overwriting old ones, or if an observation does not appear to have a corresponding target.

The following section details the retrieval of data and targets from the **Statplexer** which are returned to the user in **NumPy** arrays. Why does **Frontier** not just store the read in data in such a structure to begin with?

This is primarily due to the underlying layout of a list structure as introduced in Appendix A.2.4. Given the nature of input data, the number of observations and parameters are unknown before the input data is actually read and so memory cannot be reserved to prevent these operations. The potential number of input files renders reading through the data once to reserve the right amount of memory for a second read-through impractical in terms of time.

It is arguable that despite this, the data could be unloaded from the **_data** and **_targets** dictionaries into some form of array at the end of the call to **load_data**. However as the **Statplexer** allows loading of additional data at any time, which would not only risk increasingly expensive resizing operations as the list continues to expand but the sanity checking that takes place in **load_data** involves checking for membership of a given ID in **_data** and **_targets** which is constant for dictionaries and significantly slower for large lists (see Appendix A.2.4).

It is possible that an implementation could use both a dictionary and an array; appending the observations to the array and entering a mapping between the observation ID and its index in the array. Though this would still not avoid the issue of relocating the array once it grows beyond its bound in memory.

Worse still, as results requested from the **Statplexer** API are returned to the user in a sorted matrix[4] (*i.e.* rows and columns are ordered alphanumerically by the observation ID and parameter name, respectively), if observations are not processed in a sorted order (which is not a requirement) then returning results will involve accessing the proposed **_data** and **_targets** arrays in a non-linear fashion, losing any efficiency that could be gained from using an array in the first place.

Whilst not entirely optimal[5], in terms of a compromise between memory complexity and avoiding expensive operations on data input and retrieval, dictionaries offer the best balance.

It should be noted that the **Statplexer** stores a copy of the user defined CLASSES dictionary (as the class

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[4]A two-dimensional **NumPy** array
[5]Dictionaries still require use of **sorted** when creating a result matrix via the API

member **_classes**), which is an argument to its construction, allowing the **Statplexer** to share this information with any readers or utility functions who require it.

4.3.4 Retrieval

Frontier's **Statplexer** is designed to provide functions to an end user for extracting desired data in a format suitable for passing as an argument to functions of an external framework or library, such as **scikit-learn**. As specified in **Frontier**'s purpose, the package must provide user-friendly functions to extract observations and parameters of interest from the internal **Statplexer** representation.

The previous section suggests that directly accessing elements of the **Statplexer**'s **_data** and **_targets** member dictionaries, whilst possible, would violate the psuedo-private nature of the variables and even so, this would hardly be a user friendly way in which to obtain data from the **Statplexer**.

As described in Section 4.1.3, development of **Frontier** was evolutionary, growing to meet the needs and requirements of the underlying machine learning problem that this project strives to solve. It became clear that in trying many combinations of observation and parameter subsets that **Frontier** would need to provide not just a function to return the contents of **_data** and **_targets** but to assist in retrieving similar subsets automatically with as little effort from the end user as possible.

But how can a user make an informed decision on parameters to subset? Though this is primarily the role of feature selection, which will be discussed later in this chapter, firstly a user will need to have a feeling for what parameters are actually present and ideally be offered functionality to allow for quick elementary exploration of that set. For this, **Frontier** can be used to inspect the parameters extracted from the observations via the functions:

- **list_parameters**
  Return a sorted list of all parameters

- **find_parameters**
  Given a list of input strings, return a list of parameters which contain any of those strings as a substring

- **exclude_parameters**
  Given a list of input strings, return a list of parameters which do not contain any of the input strings as a substring, or if needed an exact match

Once users have an idea of the parameters they wish to extract from each observation, **Frontier** maintains an additional set of functions that act as a form of API, allowing users to retrieve subsets of data based on both parameters and targets:

- **get_data_by_parameters**
  Return data for each observation, but only include columns for each parameter in the given list

- **get_data_by_target**
  Return data for each observation that have been classified in one of the targets specified and additionally only return columns for the parameters in the given list

19

The prior section first introduces the format in which data is returned, the two-dimensional **NumPy** array – referred to as the *Frontier Results Matrix* – in which a row represents a particular observation and each column represents a parameter (or feature). By default both dimensions are ordered alphanumerically; rows are sorted by their observation ID, columns by the parameter name[6]. Targets are also returned in a **NumPy** array, with each index of the target array mapping 1:1 with the observation row of the same index in the *Results Matrix*.

4.3.5 Logging

Section 3.2.4 concluded that time constraints would not allow significant effort to be invested in building a platform for cataloguing results from test runs of the classifier. I decided to settle for populating fields in a structured text file, an example of which can be found in Appendix C.1.

Frontier provides functions such as **write_log** to dump **Statplexer** parameters to a given file path along with cross-validation performance associated with a particular training and prediction run of a decision tree classifier. Such files can easily be manipulated with common command line tools such as **grep** and the text processing language, **awk**.

Given the time to create specialised software for storing and comparing different runs of the classifier in future, it would be trivial to import old **Frontier** log files.

---

[6]Typically the 'cleaned' or otherwise sanitised parameter name, as noted in Section 4.3.2 for example where the **BamcheckReader** class will automatically convert spaces and other predefined characters to hyphens.

4.4   Usage Example

```python
from Frontier import frontier
from Frontier.IO import DataReader, TargetReader

data_dir = "/home/sam/Projects/owl_classifier/data/"
target_path = "/home/sam/Projects/owl_classifier/targets.txt"

CLASSES = {
        "hoot": {
            "names": ["owl", "owls"],
            "code": 1,
        },
        "unhoot": {
            "names": ["cat", "dog", "pancake"],
            "code": 0,
        },
}

statplexer = frontier.Statplexer(data_dir,
                                 target_path,
                                 CLASSES,
                                 DataReader,
                                 TargetReader)
```

Listing 3 **Constructing a Statplexer**: Syntax for instantiation of **Frontier**'s **Statplexer**.

Constructing the Statplexer requires the following arguments:

- **data_dir** *String*

  Root data directory under which all files will be parsed for observation data

- **target_path** *String*

  Path to file to be parsed for observation targets

- **CLASSES** *Dictionary*

  A dictionary of user specified class labels defined as described in Chapter 4.3.1

- **DataReader** *Module*

  Module containing the class (of the same name) to be used to parse each file in the *data_dir* tree for observation data

- **TargetReader** *Module*

  Module containing the class (of the same name) to be used to parse the *target_path* file for target data

```
statplexer.find_regressors(["mean", "percent", "rate", "average"])
```

Listing 4 **Retrieving Data**: Example usage of **Frontier**'s **Statplexer** API.

4.5 Testing

Frontier is bundled with a small testing suite designed to flex the functionality of the two included readers (**AQCReader** and **BamcheckReader**) as well as both the **Frontier** utilities and the **Frontier** API itself. The suite consists of three Python scripts which utilise the **unittest**[34] package in the Python standard library. The scripts are located in the *tests* directory of the **Frontier** package.

4.5.1 AQCReader and BamcheckReader

Both of **Frontier**'s included readers are distributed with their own modest test suite, designed to test the particular functionality of that reader, including; cleaning or sanitisation of data, ensuring warnings or exceptions are thrown when duplicate observations are encountered as required and that the two readers are capable of returning data and labels that were expected for a given observation.

4.5.2 Frontier Utilities

Members of the utilities package responsible for encoding and decoding of class labels and class codes are checked to ensure that they return the expected values. The suite also checks whether relevant exceptions are raised when unknown class labels or codes are encountered.

4.5.3 Frontier API

Testing **Frontier** itself is significantly more complicated, to accomplish the feat, a test data set with various observations, parameters and targets is automatically generated by the testing suite. The suite then examines the results of the various API calls to ensure that each returned set of data and targets, include or exclude observations as expected. Tests also ensure that the API behaves appropriately for incorrect user input, including incorrect or completely unknown parameters and targets.

Chapter 5

Results

This chapter examines some of the main results encountered from application of **Frontier** and **scikit-learn** to the task of replicating classification results from the current **auto_qc** system. Following this chapter, Part II will begin exploring the effects lanelet quality carries to downstream variant calling analysis.

Note that the figures of constructed decision tree in this chapter are presented for illustration of size and shape only and full figures may be found in the project repository[35].

5.1   Introduction

5.1.1   Objectives

The investigation considers the following main objectives:

- Outline some parameter sets for initial examination

- Investigate other potential data handling methods

- Use parameter selection for more rigorous modelling

- Attempt to minimise generated trees

5.1.2   Decision Trees

Our primary motivation for using a decision tree is its interpretability; trees create human-readable rule sets and are very easy to explain and understand. The current **auto_qc** system performs logical tests of simple rules in a similar fashion and thus it seemed sensible to compare results from both systems by using a decision tree as output. A "black box" machine learning classifier such as a neural network would not produce any tangible rules that could be explained to another human. Whilst the first goal is to briefly investigate whether

auto_qc's rules actually can be recovered, future work may wish to make improved rules that could still be interpreted by lab technicians, which would be impossible with a "black box" algorithm.

Decision trees are robust to noise and missing data (within reason) and can be realistically deployed in a variety of applications including medicine and finance. Decision trees also scale well to large inputs, which is useful given the size of the data set (approximately 1.15 million observation-parameter combinations)[36].

However, trees are prone to a process called **overfitting**, where decisions have too much basis on the data seen so far and don't generalise well to future observations. **Cross-validation** can help overcome this barrier by splitting the available data and targets into training and testing subsets. The tree will be constructed using the training set and the accuracy of the tree is then measured on the unseen testing data (*i.e.* did classifications made by the tree match the known targets for those observations?)[37].

5.2   Naive Parameter Sets

5.2.1   Introduction

For initial training and testing purposes, the following arbitrary parameter sets were created as a starting point:

- **ALL**

  A parameter set consisting of all the summary numbers in a "BAMcheckr'd" file

- **AQC**

  A set consisting of all available parameters used by **auto_qc**.

- **AQCN**

  Replaces groups of the **AQC** set with aggregated parameters.

- **ERROR**

  An experimental toy set consisting of only the "error-rate" parameter.

- **NO_ERROR**

  Another toy set designed to test results gained by excluding the "error-rate" parameter.

- **BASELINE**

  Include any parameter which includes "baseline" as a substring.

- **NOBASELINE**

  Exclude any parameter which includes "baseline" as a substring.

- **MARP**

  Include any parameter which includes any of the following as substrings: mean, average, rate or percent.

- **NO_MARP**

  Exclude any parameter which includes any of the following as substrings: mean, average, rate or percent.

Set	Nº	CV ± SD	SCV ± SD	Depth	Most Important Feature
ALL	86	90 ± 4	97 ± 1	38	T-percent-max-baseline-deviation (27%)
AQC	27	87 ± 4	95 ± 1	36	T-percent-max-baseline-deviation (31%)
AQCN	21	86 ± 4	95 ± 1	39	max-max-baseline-deviation (31%)
ERROR	1	60 ± 6	61 ± 2	53	error-rate (100%)
NO_ERROR	85	90 ± 4	97 ± 1	38	T-percent-max-above-baseline(27%)
BASELINE	34	82 ± 5	89 ± 1	46	T-percent-max-above-baseline(28%)
NOBASELINE	52	72 ± 10	91 ± 1	31	error-rate (24%)
MARP	47	87 ± 4	95 ± 1	39	T-percent-max-above-baseline (27%)
NO_MARP	39	75 ± 7	87 ± 1	38	max-max-baseline-deviation (34%)

Table 5.1 **Parameter Set Cross Validation Scores**: Results of classifying testing data into one of three classes; pass, fail or warn. Columns left to right; parameter set name, number of parameters included, average cross-validation score (max 100) ± std. deviation, average stratified cross-validation score (max 100) ± std. deviation, average depth of the generated tree and the most important parameter by Gini importance (max 100). Tree depth and parameter importance was estimated on experiments using the stratified data.

Of particular note is the **AQC** set, constructed from the parameters available from the "BAMcheckr'd" data that are taken into account by the current **auto_qc** system. However not all these parameters were initially available in the input files and **auto_qc** relies upon data made available from **vr-pipe** where the rules of **auto_qc** are actually applied; Appendix D.2 discusses contributions I made to **bamcheckr** for recovering these parameters for use in our analysis. Ultimately, **bamcheckr** performed too slowly and it was possible to use **Frontier** itself to extract the additional features needed to better represent the decisions made by the current system.

5.2.2 Experiment Results

For each parameter set, data and targets were extracted via **Frontier**'s API before being subset by **scikit-learn**'s **StratifiedKFold**[38] function. All available data (13,455 "BAMcheckr'd" lanelets) were used as input and the target variable could be one of three classes; pass, fail or warn. The subsets are referred to as "folds" (of which we used 10) and for each fold a **DecisionTreeClassifier**[39] was trained and tested. Table 5.1 outlines the average cross validation scores across each such experiment.

Whilst these parameter sets are certainly crude – a majority of them having been constructed without close examination just by matching or excluding substrings using **Frontier**'s parameter inspection functions – we can learn quite a lot about the available data.

Firstly it should be of no surprise that the **ALL** model scores highly in both cross-validation and stratified cross-validation testing. The feature group is a superset over the same parameters used by **auto_qc** itself. Similarly the **NO_ERROR** set which uses all but one parameter (**error-rate**) scores highly.

It's interesting to compare the scores between the **AQC** and **AQCN** parameter sets against **ALL**. Both of the former score highly despite having significantly fewer parameters. This appears to indicate that many of the parameters in the **ALL** model are redundant for use in classification. This should of course be expected; you'd hope to see that a set that includes parameters known to be used by the current **auto_qc** system is capable of replicating results made by that system.

In general the other models perform reasonably well, potentially reflecting the simple linear nature of the decisions made by the **auto_qc** system.

Stratified cross-validation scores are consistently better and less variable (smaller standard deviations) than their non-stratified counterparts. This should be due to the more "balanced" training and testing sets gained by assigning observations to folds to match their proportions in the whole data set. The differences between the pairs of validation scores is predictable – averaging 8.66 percentage points – but the **NOBASELINE** set presents a difference of almost 20pp and exhibits more variable behaviour. This suggests that excluding the baseline related parameters is problematic for one (or two) of the three class labels (as the only difference expected between the experiments will be the proportions of the classes).

The average maximum tree depth – ranging between 31 and 53 splits – may be highlighting a possible shortcoming of using a decision tree classifier; complexity could be indicative that a tree may have been overfitted on the training data. Later sections will apply best practice in an attempt to minimise this effect.

Whilst **T-percent-max-baseline-deviation** is a parameter used by **auto_qc**, it was not believed to be a parameter of particularly major importance and yet it is ranked as the *Most Important Feature* by **scikit-learn**'s feature importance algorithm[1] for all parameter sets it was a member of. Further investigation will yield a more authoritative conclusion as to whether the influence of this parameter should be reassessed.

5.2.3   Summary of Trees

Figures 5.1 and 5.2 are two of the ten trees constructed (one per fold) during the testing of the **ALL** parameter set. The breadth and depth of the trees are somewhat reasonable and stand up to scrutiny by cross validation (stratified or not). The depth of the trees appear to be a result of a particular decision path consisting of a large number of observations passing through nodes and splitting off a small subset of observations each time, akin to a busy train dropping small groups of passengers off at small stations. This behaviour matches the **auto_qc** process where all observations pass through the same tests and failed observations are left behind.

Figures 5.3 and 5.4 are two of the ten trees generated via training and testing of the significantly smaller **AQC** parameter set. Whilst the size of these trees has grown (though the average depth for the **AQC** set is in fact lower than **ALL**), they also exhibit the linear **auto_qc** behaviour.

It is interesting to note the apparent difference in size between these two trees, although they were generated using parameters from the same set, the results of training the trees are more variable. This could hint that there are parameters we have excluded (that are present in **ALL**) that may be useful in maintaining tree stability.

Figures 5.5 and 5.6 are examples of poor decision trees. Splitting decisions are overly complicated, creating an uninterpretable sprawl of nodes and leaves. Rules will be incredible difficult to extract, if not impossible and will be very unlikely to offer any real meaning. Figure 5.6 in particular is a highly overfit tree, constructed on just one parameter (**error-rate**); every node in the tree is an arbitrary decision, classifying the trained observations exactly.

---

[1]**scikit-learn** measures this using **Total Gini Importance**[40]
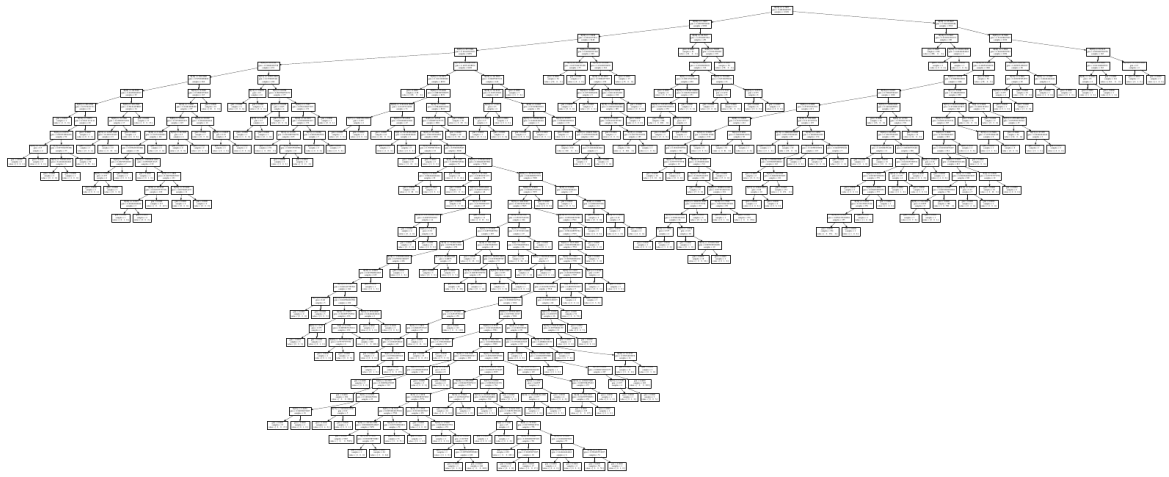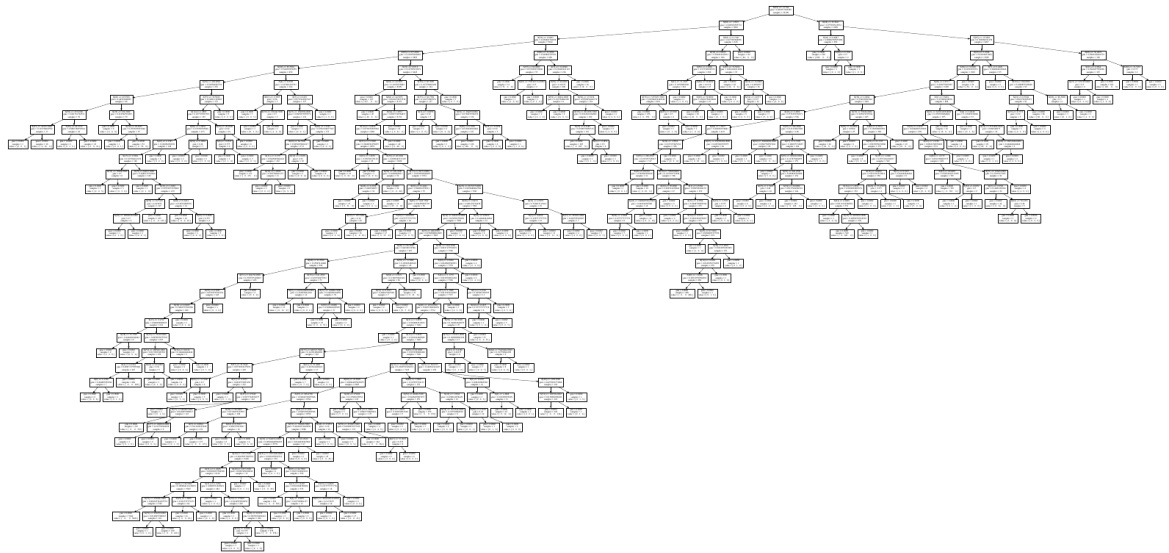
Fig. 5.1 **ALL Set Decision Tree A**
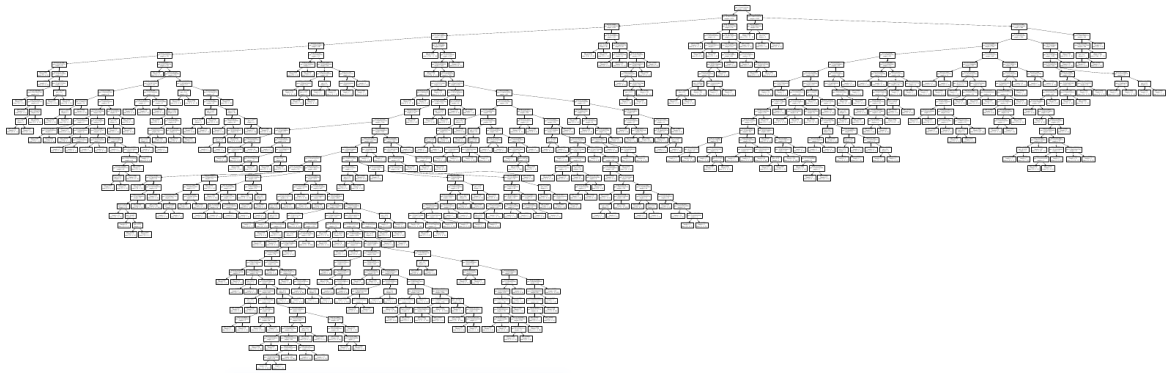

Fig. 5.2 **ALL Set Decision Tree B**

Fig. 5.3 **AQC Set Decision Tree (a)**
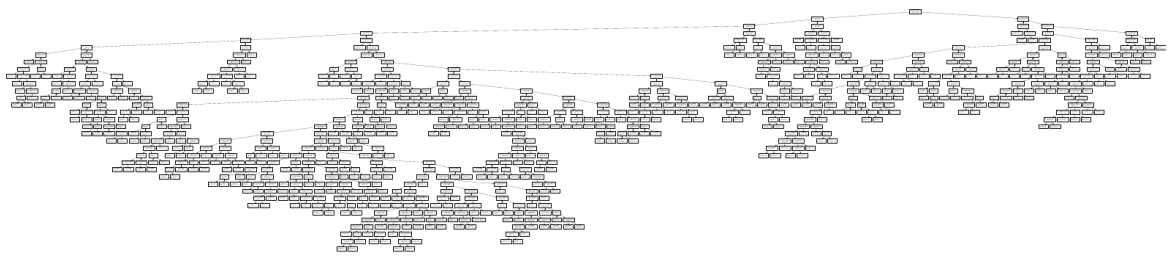
Fig. 5.4 **AQC Set Decision Tree (b)**

28
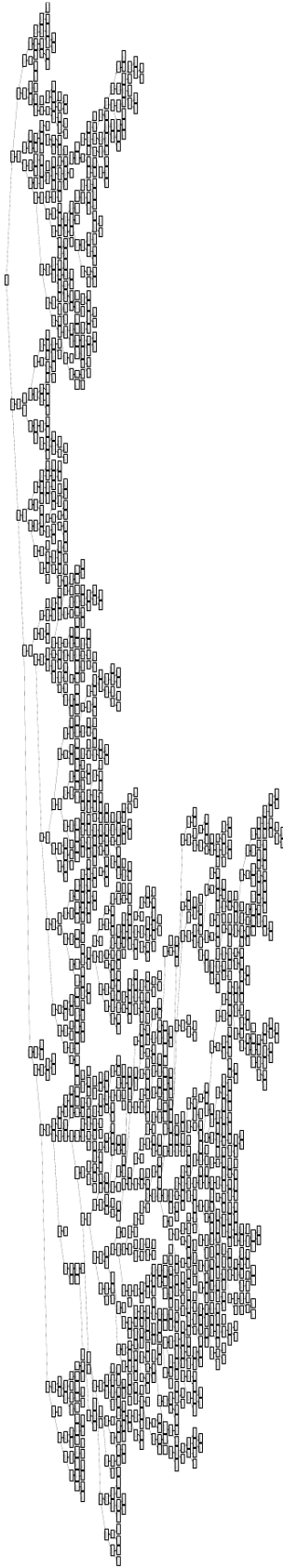
Fig. 5.5 **BASELINE Set Decision Tree**: A decision tree trained using the **BASELINE** set.

Fig. 5.6 **ERROR Set Decision Tree**: A highly overfit decision tree trained on the single parameter **ERROR** set.

The **ERROR** set in Table 5.1 however achieves a cross-validation score of 60%, highlighting the necessity of examining created trees and not just blindly accepting the validation scores. Despite its somewhat acceptable performance (60% accuracy in a three class problem), no useful rules can be extracted. In addition, you would require a 42m wide wall to hang it on if you wanted a handy classification guide in a lab.

5.3 Alternative Handling of Data Sets

The presentation of data to **scikit-learn** will greatly influence the predictions made by the resulting classifier. For example, one may choose to determine criteria to identify outlier observations that will be excluded from a study to prevent the creating rules from anomalous results.

The following subsections introduce alternative ways to present the data and targets to **scikit-learn**, in particular these methods dictate handling of the warnings class label. It is important to note that when using these suggestions the "warnings" class no longer exists and the learning task becomes the problem of predicting passes and failures.

5.3.1 Ignoring Warnings

The aim of the chapter is to firstly mimic the **auto_qc** system by training and testing on data sets with known targets as classified by **auto_qc**. However, attempting to learn what **auto_qc** doesn't know might be counter-intuitive. Lanelets are classed as warnings because the current system doesn't have the rules to mark it otherwise, such classifications could be considered as **auto_qc** reporting "I don't know".

After manual inspection a lanelet flagged as a warning will be categorised as a "pass" or "fail" by a human operator. So for all **Frontier** and **scikit-learn** know, a sample with an **auto_qc** "warning" could be either a pass or a fail.

In other words, the set of warnings is actually a union between some unknown subset of the passes and failure. Training on this now seems impractical and inherently noisy. Users may now specify `-dIGNWARN` to discard all warnings from the data set.

5.3.2 Treating Warnings as Passes

auto_qc's rule set is typically able to catch the worst quality lanelets the system will overcautiously flag other lanelets for manual inspection. Many of these flagged lanelets are approved after human intervention, to the degree where by default, lanelets classified with a warning are allowed to proceed downstream for further analysis – as if they were a pass.

If **auto_qc** is technically treating warnings as passes, then why shouldn't we? Thus, using **Frontier**'s *recode* functionality, a user may specify `-dWARNPASS` to recode all warnings as passes.

5.3.3   Classifier Performance

PSet	DSet	Nº	CV ± SD	SCV ± SD	Depth	Most Important Feature
ALL	IGNWARN	86	96 ± 3	99 ± 0	22	error-rate (43%)
ALL	WARNPASS	86	95 ± 3	99 ± 0	33	quality-dropoff-rev-mean-runmed -decline-low-value (32%)
AQC	IGNWARN	27	94 ± 4	98 ± 1	26	error-rate (44%)
AQC	WARNPASS	27	92 ± 4	98 ± 1	33	quality-dropoff-rev-mean-runmed -decline-low-value (33%)

Table 5.2 **Parameter Set Cross Validation Scores using Alternative Warning Handling**: Results of classifying testing data. Columns left to right; parameter set name, data set name, number of parameters included, average cross-validation score (max 100) ± std. deviation, average stratified cross-validation score (max 100) ± std. deviation, average depth of the generated tree and the most important parameter by Gini importance (max 100). Tree depth and parameter importance was estimated on experiments using the stratified data. *N.B.* **IGNWARN** and **WARNPASS** data sets perform classifications on two classes (pass and fail) rather than three.

Table 5.2 details the cross-validation and depth data for classifiers built using the naive **ALL** and **AQC** parameter sets in conjunction with the aforementioned data set augmentations. We can instantly see a major improvement in scores and a reduction in average maximum depth when compared to the results in Table 5.1.

At first glance it would certainly appear that the method in which warnings are handled has great influence over the performance of the decision tree classifier.

5.4   Informed Parameter Selection

5.4.1   Introduction

Whilst the crude parameter sets of the previous section were useful to gain understanding of the data that we have, it was necessary to obtain more deliberate decisions for parameters to utilise in the model. Using **backward elimination** – a method borrowed from statistical model design – the trees were repeatedly refitted after removing the least important feature (as calculated by **scikit-learn** using total Gini information).

The smallest parameter set that could be used for training and testing before cross-validation fell by more than 1% below the running average was selected as the "winner". For our purposes the winner will be referred to as **TOP9**. All data was provided, rather than using the **IGNWARN** or **WARNPASS** augmentations for this process. In future it would be interesting to repeat the selection experiment with augmentations activated.

A-percent-mean-below-baseline

duplicate-mapped-ratio

fwd-percent-insertions-above-baseline

insert-size-average

max-max-baseline-deviation

quality-dropoff-fwd-mean-runmed-decline-low-value

quality-dropoff-rev-mean-runmed-decline-low-value

rev-percent-insertions-above-baseline

rev-percent-insertions-below-baseline

Table 5.3 **TOP9 Parameter Set**: Features selected by a backward elimination experiment providing all observations to an iterative decision tree classifier and removing the least important feature until cross-validation fell below a threshold.

5.4.2   Experiment Results

PSet	DSet	Nº	CV ± SD	SCV ± SD	Depth	Most Important Feature
RTOP9	ALL	9	87 ± 6	95 ± 1	32	max-max-baseline-deviation (32%)
RTOP9	IGNWARN	9	98 ± 1	99 ± 0	20	rev-pct-insertions-above-baseline (38%)
RTOP9	WARNPASS	9	95 ± 3	99 ± 1	24	quality-dropoff-rev-mean-runmed -decline-low-value (34%)

Table 5.4 **Backward Elimination Parameter Set Cross Validation Scores**: Results of classifying testing data. Columns left to right; parameter set name, data set name, number of parameters included, average cross-validation score (max 100) ± std. deviation, average stratified cross-validation score (max 100) ± std. deviation, average depth of the generated tree and the most important parameter by Gini importance (max 100). Tree depth and parameter importance was estimated on experiments using the stratified data. *N.B.* **IGNWARN** and **WARNPASS** data sets perform classifications on two classes (pass and fail) rather than three.

Table 5.4 lists some quite impressive cross-validation results (particularly for classifiers which augment the warnings class data) considering only nine parameters were used in the training and testing. The average maximum tree depths range between 20 and 32, the upper bound now briefly overlapping the lower bound of the depth range of the naive sets found in Table 5.1. Clearly we are able to gain substantial accuracy on class prediction without requiring to use each and every parameter at our disposal.

5.4.3   Summary of Trees

Figure 5.7 displays a tree generated with the backward elimination **TOP9** parameter set against all available data. Note how the tree is more complex and than Figure 5.1 that uses all data and all parameters. Perhaps there are some parameters we could ideally add back in to the set. Further experimentation with the stopping criteria for backward elimination could be considered (perhaps looking at a weighted combination of the number of parameters, cross-validation score and maximum depth).
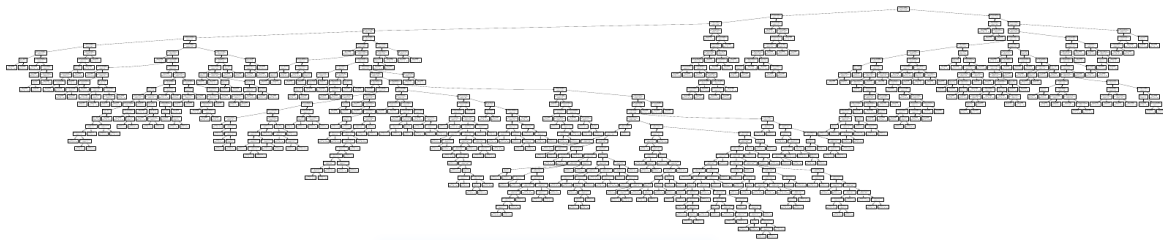
Fig. 5.7 **TOP9 Set Decision Tree with ALL Data**

A drastic change is however apparent in Figures 5.8 and 5.9. The graphs are far more simple and linear than previous figures; both width and depth are of manageable size. It would certainly appear that augmenting the treatment of the warnings class assists in reducing the amount of noise and thus inhibiting overfitting – there are no longer hundreds of uncommon combinations of parameter values that each need a node of their own to be plotted.

Given the results in Tables 5.2 and 5.4, it is arguable that treatment of warnings has a greater influence over cross validation score than the parameter set does. But it should be noted that decreasing the number of parameters used does appear to cause a difference in the average maximum tree depth between the experiments.

5.5 Conclusion

In conclusion we have investigated the cross-validation accuracy of decision tree classifiers for a series of parameter sets, examined the effects of handling the warnings label class differently and conducted parameter selection using backward elimination to reduce the number of parameters required.

Initial results seem to indicate that ignoring warnings or treating them as passes significantly improves validation scores by removing the noise of **auto_qc**'s indecisiveness. Later trees more closely resemble the simple linear rules of the current system; it's certainly possible that **auto_qc**'s rule set can be recovered using machine learning techniques.

An important part of building decision trees that hasn't been covered in this chapter is **pruning**; **scikit-learn** doesn't provide any pruning functionality and instead recommends use of ensemble methods such as **random forests**. For interpretability I continued using the single tree classifier and regardless, implementing my own pruning algorithms would haven fallen outside the scope of this project. Pruning would certainly have assisted with some of the earlier trees but our warning-augmented, minimal-parameter trees were well sized even without post-pruning.

From here, Part II will undertake an investigation in to lanelet quality, identifying lanelets that improve or detriment downstream analysis. Searching for relationships or trends in the parameters of these particular lanelets will fuel further investigation of some of the lines in inquiry introduced in this chapter.

Fig. 5.8 **TOP9 Set Decision Tree with IGNWARN Data (a)**
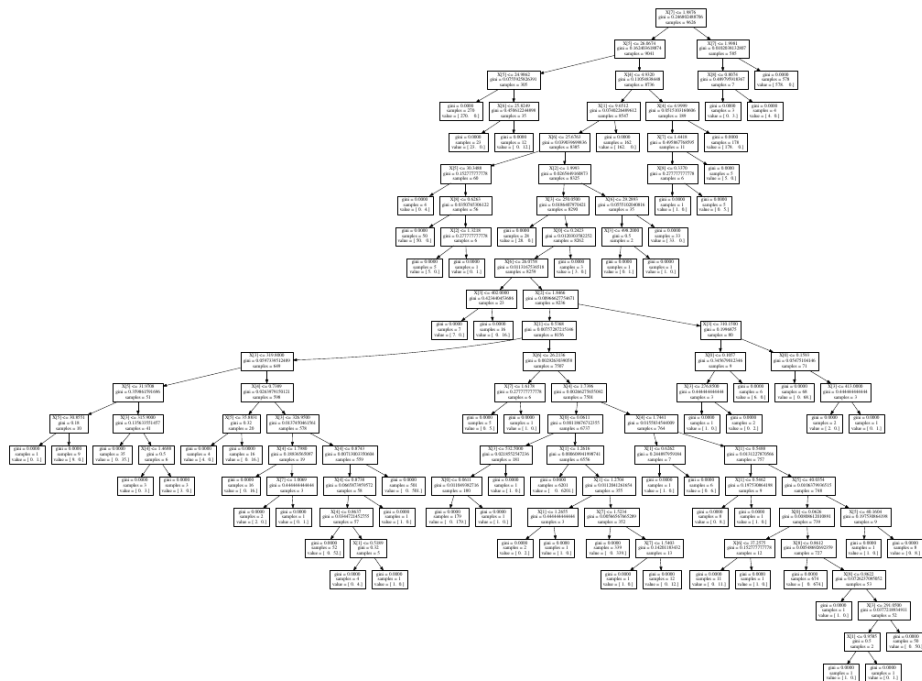
Fig. 5.9 **TOP9 Set Decision Tree with IGNWARN Data (b)**

Part II

Identification of Qualitative Sample Properties

Chapter 6

Introduction and Motivation

6.1 Introduction

The second part of the project can be outlined as follows:

- Collation of variant locations from Sanger data sets

- Construction of script to select a 'representative' genomic region

- Extraction of regions from whole-genome sequence data

- Establishment of leave-one-out analysis pipeline

- Comparison of concordance between pipeline results and SNP chip

6.2 Background

6.2.1 GWAS and iCHIP Data Sets

As briefly explained in Section 1.1.2 this part of the project focuses on measuring the similarity between called variants in pairs of samples which have been sequenced in different ways. The Sanger Institute has granted access to a set of human samples which have been sequenced twice, once with next-generation sequencing (NGS) and the other with SNP genotyping. The resultant data sets will hereafter be referred to as:

- **GWAS**[1] *NGS*
  Every base in the genome of the sample has been sequenced with some level of accuracy.

- **iCHIP** *SNP Genotyping*
  Only particular bases of interest in the genome of the sample has been sequenced with high accuracy.

---

[1]Genome-Wide Association Study

Each sample will thus have a corresponding result from each of the two studies. A result in this context refers to the output from a variant calling pipeline at the Sanger Institute – a **VCF** file (introduced in Chapter 7.1) detailing variant sites that were located during analysis of the sequenced sample.

6.2.2   The 'Goldilocks' Region

For the next step of the project we are looking to document what effects lanelet quality has on analysis that occurs downstream from whole-genome sequencing. To investigate this I'll be performing a leave-one-out analysis on the **GWAS** data set: consisting of leaving a particular lanelet out, re-performing variant calling and comparing the resulting variants called for each sample to their corresponding **iCHIP** variants.

The basic idea is to answer the question of what constitutes "good" or "bad" lanelet quality. Does leaving out a lanelet from the full sequence data lead to variant calls that better match the SNP chip data, or cause the correspondence between the two sets to decrease? In which case, having identified such lanelets, can we look back to the quality parameters we've been analysing so far and find whether they have something in common in those cases?

If we can, these parameters can be used to identify "good" or "bad" lanelets straight out of the machine. We know that lanelets that exhibit these quality variables will go on to improve or detriment analysis.

However, variant calling is both computationally and time intensive. Whilst the Sanger Institute has significant computing resources available, the project time scale is too small to conduct the analysis on thousands of full sequence samples and thus we must focus on a smaller region of the human genome.

It is for this reason we are looking for what Sanger described as a "representative autosomal region". The region must not have too many variants, or too few: a "Goldilocks genome".

6.3   Concepts and Terminology

In the search for the aforementioned "Goldilocks genome", we define the terminology found in Appendix A.3.

Chapter 7

Materials and Methods

7.1 Input Data and Format

7.1.1 Variant Query File

Named such for the command that creates it, the **Variant Query File** (or *Query File*) is an input format for populating the candidate searching script with variants. The file is formed of line-delimited, tab-separated entries where the first field consists of a colon-delimited pair representing a chromosome and position respectively (*e.g.* 1:5000 is the base at position 5000 on the first chromosome). Other fields may be provided but are currently ignored.

The **Query File** is generated by the **bcftools query**[41] command (demonstrated in Listing 5) which is a member of the **bcftools** collection – a tool set designed specifically for handling **VCF** – Variant Call Format[42][43] – files and their binary counterparts. **VCF** files are typically compressed text files developed for the 1000 Genomes Project, the files contain a meta-information header followed by records detailing the chromosome and position of variants and the base found at that site for the samples contained within.

```
bcftools query -f '%CHROM:%POS\t%REF\t%ALT\n' cd-seq.vcf.gz > cd-seq.vcf.gz.q
```

Listing 5 : Extracting variant positions from a **VCF** file with **bcftools**

7.1.2 Paths File

The **Paths File** is a trivial and esoteric file type used to handle grouping of **Variant Query Files**. The format is simple, using lines beginning with a '#' to delimit desired file groups. Entries following that line (until the next group definition) are considered to be a member of that group. Entries are line-seperated, tab-delimited and consist of only two fields: a unique slug that serves only for simple user identification of a file, followed by the the actual file path (relative or absolute).

It is an error to provide a file entry before the first group definition. An exception will also be raised if a user defines a group more than once or uses a duplicate slug due to the ambiguity this introduces.

An example can be found in Appendix B.3.

7.2 Development Environment

Many of the features of the development environment for Part I are just as applicable to Part II. Differences and additions to the environment described in Chapter 3.2 are outlined below.

7.2.1 Language

Once again, Python was the language of choice for many of the reasons previously discussed in Chapter 3.2.1. Additionally, Python is useful for rapid prototyping, thanks partly to its emphasis on readability; concise syntax, its dynamic type system and extensive standard library; making it a suitable choice to author a working script in a short time span.

Although performance was more of a concern, with the use of **NumPy** (whose mathematical functions are typically wrappers around C or Fortran implementations) reasonable efficiency was expected to be obtainable.

7.2.2 Testing

Given the more traditional development methodology, testing need not follow a continuous integration style roadmap. A test suite would need to be written to ensure the components of the script work as intended, simulating input variants and ensuring the correct candidate region densities are found.

Chapter 8

Goldilocks

This chapter introduces **Goldilocks**; the first programming hurdle for this part of the project. **Goldilocks** is a Python script designed specifically to read in and store locations of user-selected variants on the human genome and to then find regions of a given size on the genome where variants are of a desired density.

8.1 Design

8.1.1 Purpose

Expanding on the introduction, **Goldilocks** is responsible for parsing input files that contain chromosome and base position entries (in the format described in Section 7.1.1). These files can be grouped to pool the set of unique variants between each file group to treat them as one search space.

For each group, a list of positions for each chromosome is stored in memory. Once all variants have been loaded **Goldilocks** is designed to iterate over each chromosome[1] and load all of the variants from the lists in to an array and essentially count the number of variants present (in each group) between a given start and end position, determined by the arguments specified by the user as described in Section A.3.2.

This combination of a start and end position and the counts of variants present inbetween forms the structure of a **candidate**. Having performed this search over all the autosomes, candidates can then be ranked by user-specified criteria related to the density of the counted variants.

8.1.2 Format

Goldilocks is a modestly sized but elegantly complex Python script which can also serve as a stand-alone module to be imported in to other programs. Whilst the use case is certainly more esoteric than **Frontier**, I

---

[1]Strictly speaking we are only handling the 22 *autosomes* and ignoring the remaining 2 sex chromosomes (*allosomes*)

made the choice to modularise the functionality, in the hope that if an end user were to find some function of the script particularly useful they may call it as part of their own program.

8.1.3 Method

Goldilocks had to be constructed quickly but carefully. Although avoiding a quick and dirty solution, the early codebase was arguably cluttered with blocks of repeated code to perform similar actions for different hard coded groups. Initial testing was not automated and utilised eyeballing of resulting data and occasional checking of input files to ensure results were as expected.

Development was akin to a more traditional methodology; requirements were simple and available up front and formal testing was scheduled to be completed following the completion of a working prototype.

8.1.4 Pitfalls

Python lists and other sequence based structures such as **NumPy** arrays are 0-indexed, however base positions in a genome are commonly 1-indexed. These differing indices must be normalised in some fashion; for example subtracting 1 from all positions or simply ignoring the presence of element 0 in the structure. The method that is used does not matter so much as consistency; it is trivially easy to make 1-base-out mistakes when handling genomic data.

Of note also; after a discussion with the Sanger Institute it has been decided to exclude candidates located on chromosome 6 from being utilised as a "Goldilocks region". Chromosome 6 contains a system of genes called the Human Leukocyte Antigen (HLA)[2] which encode for Major Histocompatibility Complex (MHC)[46]: a critical component of the body's immune system and of particular importance to anyone requiring an organ transplant. The HLA can cause difficulties during the variant calling process due to the high level of variability expressed between samples in these areas and could thus give less accurate results if we were to measure concordance using candidates that include these genes.

8.2 Implementation

This section considers the implementation of the major components of **Goldilocks**:

- Handling user defined Paths File

- Loading variants from specified Variant Query Files

- Searching for candidates with user specified length and stride

- Ranking suitable candidates based on variant density

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[2]A much younger version of me was introduced to this concept via the partwork *How Your Body Works*[44], which also distributed English dubs of a 1987 French television show titled *Il était une fois... la vie (Once upon a time... Life)[45]* a relevant episode can be viewed here: http://www.youtube.com/watch?v=yvhU2UyRv-Q

8.2.1   Parsing Paths File

Goldilocks requires the user provide their own valid **Paths File**, meeting the specification introduced in Section 7.1.2 (an actual example of such a file can be found in Appendix B.3).

As previously described the file merely documents the locations of the **Variant Query Files** to be loaded, along with the desired grouping of those files. Any variant location that appears in at least one file of a group will be added to the pooled set of variants for that group.

Goldilocks provides the simple **load_variant_files** to parse the user's **Paths File**, where for each group, the following structures are initialized and added to a dictionaries encapsulated by **Goldilocks** itself, using the group name as a key:

- **groups** *Dictionary*
  Keys refer to a particular autosome number, mapped to a list structure which will later be populated with the positions of all variants that appear on that chromosome in at least one **Query File** of the group. It is from this structure that chromosome **NumPy** arrays will be loaded with data.

- **group_buckets** *Dictionary*
  Keys refer to a specific number of variants counted (considering only variants from the group), mapped to a list of all i'th candidate regions which contained that density. This structure eases locating groups which match desired density criteria once summary statistics have been calculated.

- **group_counts** *List*
  The number of variants which appear in each candidate, for this variant group will be appended to this list. This structure simplifies the gathering of summary data (such as mean, median and variance) of all the candidate regions of the group once the search has completed.

A **Paths File** which violates the specification will raise an exception during execution of this function, informing the end user of their error. This was decided to ensure ambiguous input did not cause confusion later, for example should defining a file group twice merge or override the first definition?

8.2.2   Loading Variant Query Files

load_variants_from_file is responsible for loading a given **Variant Query File** and adding the position of each record contained within to the appropriate group-chromosome list pair in the **groups** structure.

It is important to note that this function is not responsible for ensuring the positions added to the list are unique to avoid performance penalties in checking whether a variant is already a member of the list (which increase proportionally for the size of the list). However as examined in Appendix A.2.4, appending a significant number of items to a list will likely induce costly memory operations as the list is resized. Thus use of a set might have been more appropriate (especially considering the container forbids duplicate entries) but a set must be constructed from an existing iterable such as a list anyway!

Indeed the container does specify an **update** function but this could in itself be a source of overhead as the structure ensures that the no duplicate condition has not been violated on every call. Given more time it would

be interesting to compare performance differences between container implementations.

load_variants_from_file has another responsibility in keeping track of the "furthest along" variant position for each chromosome across all groups, this is used in the search step later, firstly to ensure that candidates are not constructed beyond where the last variant on a chromosome lies (as by definition they would be empty and searching over them would waste resources) and secondly to normalize the candidate regions across all groups to allow for easy comparison between them.

8.2.3 Searching for Candidate Regions

search_regions is responsible for performing the main task of the script, walking over the genome and creating objects who represent candidate regions. The function is possibly a misnomer as it conducts a candidate census rather than a search, indiscriminately adding all valid candidates to the **regions** dictionary.

search_regions will iterate over each chromosome and load the listed variants in each group-chromosome pair from the **groups** structure (by calling **load_chromosome**) into a **NumPy** array whose length is set to the last position from which a full region can be extracted based on the maximum position seen by **load_variants_from_file** for this chromosome, plus one to account for the 1-indexed nature of genome positions. The function will then take slices of these arrays of the user defined *length* and count the number of variants contained within by summing across the slice (elements with variants contain a 1, otherwise 0). The next region is sliced by moving the first base of the current region forward by the user defined *stride*.

Each valid region will be entered in to the **regions** dictionary using the current region counter (**region_i**) as the key, mapping to an arbitrary object (a dictionary) as defined in Listing 6 below:

```
regions[region_i] = {
    "ichr": i,                # The i'th region on the current chromosome
    "group_counts": {},       # A dictionary mapping variant group names to
                              # the number of variants counted for that group
    "chr": chrno,             # The autosome number
    "pos_start": region_s,    # The first base of this region (inclusive)
    "pos_end": region_e       # The last base on this region (inclusive)
}
```

Listing 6 : Candidate Region Data Structure

Regions will also have their respective group densities appended to the appropriate **group_counts** list and its **region_i** added to the relevant **group_buckets** structure as established in Section 8.2.1.

8.2.4 Filtering and Ranking Candidates for Selection

For the purpose of this project we're looking to filter candidates by their absolute distance from the median density of variants located in the **GWAS** variant group (the median density will give the most representative

43

regions) and then rank those by their corresponding **iCHIP** variant group densities, descending (enriching the number of comparisons that can be made between the **GWAS** data and **iCHIP** data).

Two functions in the **Goldilocks** API provide a process to rank candidate regions by a desired group's density:

- **initial_filter**

  Uses the relevant filtering group's **group_counts** list (in this case, **GWAS**) to calculate a window around the median density. The function will then return all candidates inside the group's **group_buckets** dictionary whose density fall inside the upper and lower bounds of the window around the median.

- **enrich**

  Sorts the output of **initial_filter** by the desired ranking variant group (in our case, **iCHIP**). The function will prioritise candidates with the closest absolute value to the filter group's median and ensure that the density of the ranking group is higher than the filtering group. Returns a ranked list of candidates in the format displayed in Listing 6 for manual inspection.

Currently ranking and filtering criteria are still hard coded to meet the needs of our analysis. In future it would be useful to generalise the **Goldilocks** API to allow more operations such as filtering by a given percentile rather than only the median and to provide more 'enrichment' options such as ranking by minimum.

It would be sensible to consider re-naming the two functions to **filter** and **rank** respectively and allow users to chain together these methods to compound filtering and ranking operations as necessary.

Whilst in this state the script proposes little re-use value, we at least have a basis for repeating results in future with different data sets or additional variant groups.

8.3 Testing

The testing of **Goldilocks** presented a challenge; we'd not only like to simply ensure that the correct query files are located and loaded in to memory but also that the expected candidates are found, filtered and selected. It is thus not practical to conduct testing on a large data set such as the two we have for analysis, we need a smaller, well defined set with known results.

To achieve this the testing suite is designed to generate valid **Variant Query Files** from a **TEST_DATA** dictionary, which defines test groups and variant locations on test chromosomes and can easily be altered for future testing. Test files are re-generated from this dictionary each time the test suite is instantiated using the **setUpClass** classmethod provided by the **unittest** library.

Checking both the **Paths File** and generated **Variant Query Files** were loaded successfully is trivial and the suite goes on to test whether the input files and all generated variants within are mapped to the specified chromosome of the correct group, both in memory and also when loaded in to the **NumPy** array structure. Once regions have been searched (or more accurately, once a a candidate census has populated the **regions** dictionary via **search_regions**), additional tests ensure that all candidate regions are of the correct *length* and begin on a valid *stride*.

```
GROUP0
CHR 1                                                           Expected
          4      10 13      21  25                                Size   i
        1 |*==*=====*==*=======*===*=======================*| 50
            |          |                                            3    0
              |            |                                        2    1
                |            |                                      1    2
                  |            |                                    2    3
                    |            |                                  2    4
                      |            |                                0    5
                        |            |                              0    6
                          |            |                            0    7
                            |            |                          1    8
                              |            |                        x


GROUP0
CHR 2                                                           Expected
          5                          91                           Size   i
        1 |====*================\/...\/==========**********| 100
            |          |                                            1    9
              |            |                                        0   10
                      ...                                           .    .
                          |          |                             5   26
                            |          |                          10   27
                              |          |                         x
```

Listing 7 **Testing Expected Candidates with Goldilocks**: The test suite defines the locations of variants across many files in two groups, the figure represents the expected regions for *GROUP0*. The horizontal line represents a particular chromosome on which a * signals the location of a variant (= otherwise). The expected size column tabulates the number of variants that should appear as a member of region i (start-end inclusive).

As discussed, testing the discovery of correct regions is more complex. Listing 7 details the regions expected for the variants specified in **TEST_DATA**. The test suite checks that the number of regions located and their content is correct for each chromosome in each variant group. Tests also ensure that the previously mentioned **group_counts** and **group_buckets** structures are populated appropriately.

Finally the **initial_filter** and **enrich** functions can be tested (as we are able to manually calculate the expected candidates given the small data set) and ensure those candidates are returned.

Admittedly the suite could still be described as modest and with more time it would be appropriate to further improve coverage, possibly generating larger (or even random) data sets. Results however, have also undergone manual inspection both by myself and the Sanger Institute and the regions described in the following chapter are considered legitimate.

Chapter 9

Results

Table 9.1 lists the top 25 candidate regions selected by **Goldilocks** for inspection. Regions were constructed using a *length* of 1Mnt[1](mega nucleotides) and a *stride* of 0.5Mnt[2]. As detailed in Section 8.2.4, candidates for this study were filtered to include regions with a **GWAS** density within ±12.5% of the median **GWAS** density (*i.e.* in the middle 25%) before being ranked firstly by absolute difference from the median **GWAS**, then by **iCHIP** variant density, descending.

Empty regions were excluded from a variant group's analysis, including the calculation of any percentiles. This was to ensure that we located the most "representative" density from the regions and avoid potential skewing from 0 density candidates. However repeating the experiment including empty regions leads to highly similar results and the candidate that was actually selected as the Goldilocks region appears in both sets of results.

Candidate 1074 (marked by *****) was selected as the region with the closest absolute difference between the median **GWAS** density of 297, whilst also offering the highest **iCHIP** density of the top 25 returned candidates.

The implementation of the filtering and ranking functions in the previous chapter noted that during final ranking, the **enrich** function will discard candidates if the number of variants in the ranking group is less than the filtering group – in this case, if the **iCHIP** density is lower than the **GWAS**. This was a crude rule implemented to prevent candidates with a significantly low **iCHIP** density ($\leq$100) from being put forward following 'enrichment'; these regions would give us little to compare when it comes to measuring the concordance between variants called in our leave-one-out study and the original **iCHIP** data set.

---

[1]1,000,000 bases
[2]500,000 bases

i	GWAS	iCHIP	Chr	Start	End
0234	297	470	1	117000001	118000000
1074*	294	1540	3	46000001	47000000
5222	294	336	21	16500001	17500000
3125	298	310	10	60000001	61000000
0880	293	344	2	191500001	192500000
3560	299	772	12	9000001	10000000
4407	299	512	15	78500001	79500000
1036	292	300	3	27000001	28000000
2734	300	515	9	5000001	6000000
3426	300	486	11	76000001	77000000
0015	291	1029	1	7500001	8500000
0365	301	487	1	182500001	183500000
3415	301	419	11	70500001	71500000
1581	290	802	4	102500001	103500000
3554	290	403	12	6000001	7000000
3184	302	449	10	89500001	90500000
1580	289	603	4	102000001	103000000
1948	288	1297	5	96000001	97000000
2215	288	622	7	49500001	50500000
0414	288	346	1	207000001	208000000
2055	304	1377	5	149500001	150500000
0384	287	827	1	192000001	193000000
0959	306	406	2	231000001	232000000
4214	286	393	14	88500001	89500000
0320	307	620	1	160000001	161000000

Table 9.1 **Goldilocks Results**: "Top 25" Candidate Regions using a *length* of 1Mnt and a *stride* of 0.5Mnt. Candidates are filtered by median **GWAS** density and ranked by maximum **iCHIP** density.

Curious about the wide range and presence of so many low resolution **iCHIP** candidates, I authored an R script (making use of **ggplot2**[24] for graphing) to plot figures 9.1 and 9.2 as a means of visual inspection to confirm such results should be anticipated.

Figure 9.1 shows a large range in the densities of variants in the **iCHIP** regions, between 0 and almost 3000, with a significant baseline below 100. Figure 9.2 plots the same data set but also facetting by chromosome, again it can be seen that the **iCHIP** variant group counts are well spread when compared to the rather constant variance of their **GWAS** counterparts.

This isn't a surprise when you consider that the **GWAS** data set represents the variants sequenced from whole-genome analysis of our samples, whereas the **iCHIP** set consists of the variants output from SNP genotyping, a process that focuses on particular genomic locations of interest.

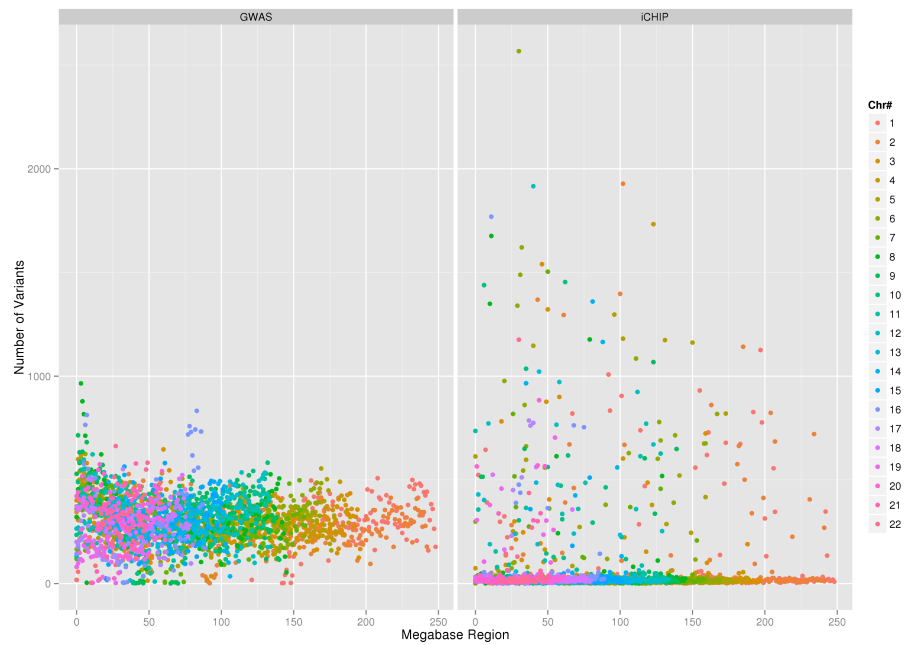Fig. 9.1 **Goldilocks Candidate Plot A**: Candidate regions of one megabase length are arranged along the x axis (coloured by chromosome) against the number of variants contained within that region.
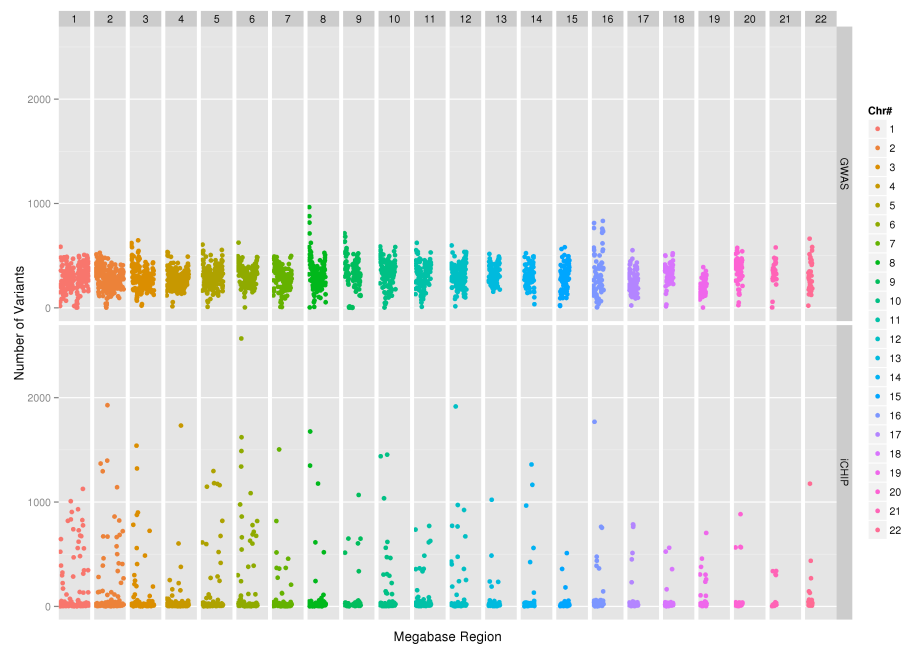
Fig. 9.2 **Goldilocks Candidate Plot B**: Candidates of one megabase length are arranged along the x axis (coloured and facetted by autosome) and plotted against the number of variants contained within that region.

Chapter 10

Analysis Pipeline

Having selected the Goldilocks region (Chromosome 3: 46,000,001—47,000,000) via **Goldilocks**, we must now begin constructing the pipeline that will conduct the leave-one-out analysis over all the **GWAS** lanelets and compare the called variants to the appropriate **iCHIP** sample.

This chapter introduces the various components of the pipeline, their purpose and any difficulties encountered.

10.1 Overview

The pipeline can be divided up in to the following analysis steps:

- **Extraction**
  For every whole-genome lanelet from the **GWAS** study, extract the Goldilocks region.

- **Indexing**
  Create indexes for the extracted Goldilocks regions, improving efficiency for tools which require random access to variants inside the region.

- **Merge**
  Merge the data from each extracted region in to one file to reduce file handling overhead in later analysis.

- **Pileup**
  Calculate genotype likelihoods based on the reads seen across all the extracted Goldilocks regions.

- **Call**
  Use the genotype likelihood scores to actually call the variants for each position of interest in each of the Goldilocks regions.

- **Compare**
  For each pair of **GWAS** and **iCHIP** samples, measure the concordance of called variants.

10.2   Region Extraction and Indexing

10.2.1   Extraction

Now the Goldilocks region has been found, it must be extracted for each **GWAS** sample in our study. The Sanger Institute currently stores the electronic samples in the **BAM**[47] format (a binary format for the storage of sequence data) at one of their in-house data centres, storage and retrieval is managed by **iRODS**[48]: the *Integrated Rule-Oriented Data System*.

iRODS is a BSD licensed, C++ based data management application, describing itself as "middleware for your critical data"[49]. The system was deployed at the institute in 2011 to cope with the data deluge brought on by constantly improving next-generation sequencing technologies. "[T]raditional data management methods [became] saturated" and other more advanced file systems did not provide the desired support for customised metadata such as project information or sequencing specific fields like sample and lanelet ID to be automatically added to files[50] which **iRODS** does support, along with improved data replication and access control policy management.

Once I had been added to the relevant access group, it was possible to extract the regions from **iRODS** via **samtools_irods**[51] which provides wrappers around both **samtools** and **iRODS** functionality to allow **samtools** commands to directly support interaction with samples stored in **iRODS**.

```
samtools_irods view -bh irods:\${file} 3:46000001-47000000
    > \$(basename \${file}).goldilocks.bam
```

Listing 8 **BAM Extraction**: Retrieve Goldilocks region for a particular sample ($file) from **iRODS**.

10.2.2   Indexing

An index file provides a mapping between genomic positions and the compressed blocks of a **BAM** file that contain them, affording programs such as members of the **samtools** collection efficient access to the relevant blocks of a **BAM** file without conducting costly searches.

Whilst indexing of **BAM** files is a somewhat optional step, many tools will emit warnings or errors if one is not available. The process itself requires little resources in terms of time and memory and can significantly improve the efficiency of later commands which are capable of reading and using the index.

Both the commands in Listing 8 and 9 can be executed in parallel for extracting and indexing many samples.

```
samtools index \$(basename \${file}).goldilocks.bam
```

Listing 9 **BAM Indexing**: Creation of a **BAM Index** (BAI) for a sample ($file). with **samtools index**[52]

10.3 Pileup

Following extraction and indexing of the Goldilocks region for each whole-genome sample from the **GWAS** study, the next step is to "pileup" the samples, a process which stacks the bases seen at each variant location from all the indexed samples up in a structure before calculating the statistical likelihood of those bases actually appearing in each sample given the data overall.

```
bsub -o ~/goldilocks/joblog/samtools_mpileup.%J.o
    -e ~/goldilocks/joblog/samtools_mpileup.%J.e
    -G hgi -J "samtools_mpileup"
    -M1000 -R "select[mem>1000] rusage[mem=1000]"
    bash -c 'samtools mpileup -b ../goldilocks-3:46000001-47000000.fofn -g -I -f
        /lustre/scratch113/resources/ref/Homo_sapiens/1000Genomes_hs37d5/hs37d5.fa
            > ~/goldilocks/all.withref.bcf'
```

Listing 10 **Pileup**: Submission of **samtools mpileup**[53] task for execution on the **Farm**. Note the request to increase the memory allocation with the `-R` flag and how the pileup command is passed as a string to `bash -c`.

However, performing a pileup is a rather intensive process, with initial test runs requiring 6.5 hours of CPU time, with an average memory usage of 905.85MB (920MB max), outputting a 13GB **BCF** file. In a scenario such as ours where there are thousands of input files, one for each of the extracted regions, we place considerable strain on the cluster's file system and introduce incredibly inefficiency given the overhead of file handling. As this pipeline will need to be executed once for each leave-one-out experiment, it would be required to reduce the strain on the file server before the job could be submitted. Usefully another member of the **samtools** collection provides functionality to merge a list of given input files such as those extracted Goldilocks regions.

10.4 Merge

samtools merge[52] is a utility for merging two or more sorted **BAM** files in to one sorted output. After viewing the source of the command I found an undocumented feature that allowed an end user to provide a file of filenames to merge rather than being required to list all inputs on the command line.

Various issues with use of **samtools merge** at scale were encountered during the assembly of the analysis pipeline. Ultimately I ran out of time attempting to overcome these problems before being able to put the pipeline to use. The difficulties experienced with **samtools merge** specifically (including tracking and patching of memory leaks) are detailed on the **Frontier** blog[54].

10.5   Calling

Variant calling was briefly introduced as a concept in Part I (See Section 1.1.2). Calling for variants is the process of finding differences between a reference sequence[1] and a given sample[2] or set of different samples.

The genotype likelihoods calculated by **samtools mpileup** in the previous step of the pipeline represent a probability distribution of the bases seen across all piled up samples at a particular location (site). This allows variant calling algorithms to make an informed decision as to whether particular sites actually contain a SNP or if a differing base is statistically unlikely to be a real variant[3] with the current data.

bcftools call[58][59] is the successor to the 'consensus' variant caller included with the **samtools** package and utilises **htslib**: a *high-throughput sequencing library* for improved file handling efficiency with common genomic data formats.

Unfortunately during the initial testing of this step, using a pileup of all the Goldilocks regions, the output contained only the standard header information generated by **bcftools** and not a single record detailing a called variant. I discovered this was due to the **samtools mpileup** job being completed without including a reference sequence, causing the *REF* (reference) column of the resulting **BCF** file to be populated with 'N': an ambiguity code which translates to 'any base'[60][61].

bcftools call does provide the `-M` flag for this scenario, where the reference is described as a "masked reference". However attempting to repeat calling with the flag set merely caused the command to produce a segmentation fault instead of an empty output file. This appeared to be due to a compatibility issue between the versions of **htslib** and **bcftools** I was using on the **Farm** – Sanger's high performance computing cluster.

Finally this initial test of the calling component of our pipeline was able to be completed in 66 minutes with an average memory usage of 11.97MB, resulting in a 114MB compressed VCF file.

A similar compressed VCF file will be output from each iteration of the leave-one-out analysis, generating thousands of files. Whilst the Sanger Institute manages petabytes of storage, it is sensible to minimise memory and storage resource use if at all possible. In future runs, output size could be reduced by generating a list of variants that appear in both the **GWAS** and **iCHIP** Goldilocks regions (as these are the only variants we are interested in anyway), **bcftools call** will then discard records pertaining to variants that do not appear in the targets list. This "targets" file actually uses the same format as the **Variant Query Files** described in Section 7.1.1 and should be provided to **bcftools call**'s `-T` (`--targets-file`) option.

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[1]Our analysis maps to the **hs37d5** reference sequence, used by the *1000 Genomes Project Phase II*[55]. **hs37d5** extends the **b37** genomic reference with "decoy sequences" that are designed to reduce false positives (incorrectly mapping to poorly referenced areas around centromeres for example[56]), optimising it for variant calling[57].

[2]Technically to be confident you have an actual SNP as opposed to a variant (or incorrect read!), you'll need to see whether such variation occurs through a population of different samples rather than just one.

[3]Although it could also be a very low frequency variant...

10.6   Measuring Concordance

The terminal step of the Goldilocks pipeline consists of analysing the differences between variants called from the left-one-out **GWAS** data and the **iCHIP** study. The two data sets will form matched pairs where an individual's DNA has been sequenced and processed by our pipeline (possibly with one of that sample's lanelets ignored) and alternatively via SNP genotyping. The pipeline will measure the concordance between the variants called for each individual, this measure will then be used to determine the effect of the lanelet that was ignored on that run.

For this step I retrofitted a Python script I had authored earlier to sanity check the bases seen at the known variant sites were sensible (*i.e.* they agreed on the same genotype – the same sets of bases appeared). Due to the difficulties encountered in assembling the pipeline (particularly with **samtools merge**), it was not possible to adequately test the pipeline and thus the concordance script with the time that we had. Considering this and for the sake of brevity, discussion on the design and implementation is considered outside the scope of this write-up.

Another option for this component was **SnpSift Concordance**[62], a Java based package which can also calculate concordances between two **VCF** files. However I was concerned with the possible performance of using Java to process thousands of concordances.

Part III

Discussions and Conclusions

Chapter 11

Critical Evaluation

Unfortunately time constraints cut the project short; the time required to assemble the pipeline outlined in Chapter 10 was massively inflated due to difficulties encountered in the tools used, which was unexpected both by myself and the Sanger Institute. Whilst this prevented completion of the Part II objectives outlined in Section 1.1.2, I was still able to collate variant data from the samples, identify the "**Goldilocks Region**" and make contributions ranging from documentation fixes to the patching of memory leaks in highly popular open-source bioinformatics tools.

In retrospect, despite the complications, Part II was probably overly ambitious for the amount of time I had. Given the project again I think I would more seriously weigh up the benefits of extending the analysis downstream, although it should be remembered that it is this element of the project that makes the research novel.

Looking at Part I, I think the requirements were more correctly balanced and **Frontier** assists greatly in achieving the objectives outlined in Chapter 2. I am confident the correct development choices were made in the construction of **Frontier** and the scripts used to actually conduct analyses. One of the most influential decisions on this part of the project was that of the machine learning framework: **scikit-learn**, whose integration with Python's scientific computing packages like **NumPy** was incredibly helpful, unlike other available packages which may have required time-consuming wrapper functions to be created to mangle data in to an obscure format. The framework also included subpackages that were useful for cross-validation testing and parameter selection saving time and simplifying interaction between various components of the scripts.

The machine learning analysis of Part I was quick and easy to set-up and document thanks to the functionality provided by **Frontier** for the storage and retrieval of data and targets. **Frontier** is still in its infancy (though that is not to say it is lacking in core function) and I plan to continue its development after the project ends.

The analysis presented in Chapter 5 introduces many interesting avenues of investigation, too many to be written up as only one part of this project! In brief I've demonstrated that the decision trees generated exhibit behaviours similar to the current **auto_qc** system and presented cross-validation results for a range of classifiers built on various parameter sets which improve as we obtain a better idea of the observations and their features.

It should be noted that the results presented consist an initial not exhaustive study. Decision trees cannot promise optimality and given more time I'd have investigated use of other machine learning classifiers, starting with random forests – collections of decision trees – to test whether sets of similar trees are created. Although **scikit-learn**'s does not provide methods to prune, future trees should consider best practice provided by the framework's developers[63]. It would also be interesting to also experiment with other algorithms (even black box methods such as neural networks) and frameworks.

Although **Frontier** documented experiments in a useful format, I do regret the lack of a more advanced testing platform. My search for a suitable application (see Section 3.2.4) for managing results of machine learning classifiers unfortunately turned up nothing. With time I'd like to take the opportunity to fill this gap and provide a front-end to **Frontier** for users to generate graphs of classification performance over time and keep track of their various parameter and data sets.

Overall I am pleased with the outcomes of this project; I feel both **Frontier** and **Goldilocks** are fit for purpose and more than meet their original design requirements. The **Pipeline**, although incomplete, has been defined and work can be continued outside of this project in conjunction with the Sanger Institute.

Chapter 12

Conclusions

This project introduces **Frontier**, a Python package which provides users with interfaces for the reading, storage and retrieval of large machine learning data sets. **Frontier** provides users with interfaces allowing for simple definition of readers, enabling any file (no matter how esoteric) to be used as input. **Frontier** also presents an API for manipulation and extraction of stored data-target pairs based on lists of parameters or desired target classes. The package also effectively supports any machine learning task by providing users the ability to specify their own class labels and codes.

The project went on to utilise **Frontier** and **scikit-learn** in a script to investigate the accuracy of a range of decision tree classifiers for a series of quality control parameter sets. I also examined the effects of augmenting the handling of observations classified by **auto_qc** as a warning and experimented with backward elimination to generate minimal yet accurate models, creating trees which closely resemble the behaviour of the simple linear rules of **auto_qc**

This part of the project left off on an excellent starting point for continuing analysis with the identification of lanelets which cause changes in the results of downstream experiments. The hope was to examine the parameters of those lanelets to motivate further lines of inquiry, picking up from the end of Part I.

Part II introduced **Goldilocks**, a Python script used to find a representative genomic region that expressed a desired density of variants across different studies, nicknamed the "Goldilocks genome". Alongside this, a **pipeline** was outlined for analysis of thousands of samples of the Goldilocks region. Whilst the **pipeline** is still in the process of being developed, I've already made several contributions to **samtools**; updating documentation, patching minor memory leaks and conducting an investigation on poor time performance.

In addition to this, as part of the project I also submitted bug fixes and minor contributions to Sanger's **bamcheckr** tool which are now in place at the institute. **Frontier** even turned out to be more efficient than **bamcheckr** for the recovery of additional parameters.

This project has produced a potentially wide reaching Python package in the form of **Frontier**, applicable to a variety of statistical or machine learning problems. Although **Goldilocks** may not appear to have much of

an audience a 2003 paper[64][1] analysed the spread of SNPs across regions of the human genome to assert whether variants could be modelled with a statistical distribution, a clear use case for **Goldilocks**.

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[1]Titled *An Exponential Dispersion Model for the Distribution of Human Single Nucleotide Polymorphisms*[64]

References

[1] T. Strachan and A. Read, *Human Molecular Genetics*, 4th ed. Garland Science, 2011, pp. 214–254.
> A concise introduction to the processes involved in massively parallel DNA sequencing.

[2] M. Kircher, U. Stenzel and J. Kelso, "Improved base calling for the Illumina Genome Analyzer using machine learning strategies," *Genome Biology*, vol. 10, no. 8, p. R83, 2009.
> Useful introduction to relevant Illumina hardware and the errors that can occur during sequencing.

[3] auto_qc: Additional high-throughput sequencing autoQC steps [Github]. [Online]. Available: https://github.com/wtsi-hgi/seq_autoqc
> Github repository for the Sanger Institute's auto_qc system.

[4] vr-pipe: A generic pipeline system [Github]. [Online]. Available: https://github.com/wtsi-hgi/vr-pipe/
> Github repository for the Sanger Institute's vr-pipe probject.

[5] M. Niemi, "Internal QC Summary Report," 09 2012, unpublished.
> Internal Sanger report outlining the decisions applied by the QC team and what circumstances may lead to failure of lanelets.

[6] Triage: A todo manager for the disaster that is your day [Github]. [Online]. Available: https://github.com/SamStudio8/triage
> Github repository for the Triage task management application.

[7] sam/Dissertation | Triage. [Online]. Available: https://triage.ironowl.io/sam/dissertation/
> Public facing todo list outlining tasks for this project.

[8] H. Li, B. Handsaker, A. Wysoker, T. Fennell, J. Ruan, N. Homer, G. Marth, G. Abecasis, R. Durbin, and G. Subgroup, "The Sequence Alignment/Map format and SAMtools." *Bioinformatics*, vol. 25, p. 2078, 2009.
> Paper outling the SAM/BAM file formats and introducing the samtools project.

[9] M. Hall, E. Frank, G. Holmes, B. Pfahringer, P. Reutemann, and I. H. Witten, "The WEKA Data Mining Software: An Update," *SIGKDD Explorations Newsletter*, vol. 11, no. 1, pp. 10–18, Nov. 2009. [Online]. Available: http://doi.acm.org/10.1145/1656274.1656278
> Article introducing the WEKA machine learning framework.

[10] B. Ripley, *tree: Classification and regression trees*, 2014, r package version 1.0-35. [Online]. Available: http://CRAN.R-project.org/package=tree
> R manual for the tree package.

[11] T. Therneau, B. Atkinson, and B. Ripley, *rpart: Recursive Partitioning*, 2013, r package version 4.1-3. [Online]. Available: http://CRAN.R-project.org/package=rpart
> R manual for the rpart package.

[12] R Core Team, *R: A Language and Environment for Statistical Computing*, R Foundation for Statistical Computing, Vienna, Austria, 2013. [Online]. Available: http://www.R-project.org/
> The R manual.

[13] T. Smith. (2012–2013) aRrgh: a newcomer's (angry) guide to R. [Online]. Available: http://tim-smith.us/arrgh/
> A useful alternate manual for the R programming language, highlighting elements of the language's syntax that may be potential pitfalls to new users.

[14] D. E. King, "Dlib-ml: A Machine Learning Toolkit," *Journal of Machine Learning Research*, vol. 10, pp. 1755–1758, 2009.
> A machine learning framework for C++

[15] C. Igel, V. Heidrich-Meisner, and T. Glasmachers, "Shark," *Journal of Machine Learning Research*, vol. 9, pp. 993–996, 2008.
> A machine learning framework for C++

[16] Python Wiki. (2014) Python for Artificial Intelligence.

[17] F. Pedregosa, G. Varoquaux, A. Gramfort, V. Michel, B. Thirion, O. Grisel, M. Blondel, P. Prettenhofer, R. Weiss, V. Dubourg, J. Vanderplas, A. Passos, D. Cournapeau, M. Brucher, M. Perrot, and E. Duchesnay, "Scikit-learn: Machine learning in Python," *Journal of Machine Learning Research*, vol. 12, pp. 2825–2830, 2011.
> A machine learning framework for Python.

[18] J. Demšar, T. Curk, A. Erjavec, Črt Gorup, T. Hočevar, M. Milutinovič, M. Možina, M. Polajnar, M. Toplak, A. Starič, M. Štajdohar, L. Umek, L. Žagar, J. Žbontar, M. Žitnik, and B. Zupan, "Orange: Data Mining Toolbox in Python," *Journal of Machine Learning Research*, vol. 14, pp. 2349–2353, 2013. [Online]. Available: http://jmlr.org/papers/v14/demsar13a.html
> Paper introducing the Orange machine learning framework.

[19] scikit-learn. About Us. [Online]. Available: http: //scikit-learn.org/stable/about.html
    About the scikit-learn project.

[20] T. E. Oliphant, "Python for Scientific Computing," *Computing in Science & Engineering*, vol. 9, no. 3, pp. 10–20, 2007.
    Paper introducing the SciPy stack, including NumPy and SciPy packages.

[21] E. Jones, T. Oliphant, P. Peterson, *et al.*, "SciPy: Open source scientific tools for Python," 2001–. [Online]. Available: http://www.scipy.org/
    The SciPy project website.

[22] SciPy.org. Scipy license. [Online]. Available: http://scipy.org/scipylib/license.html
    Licensing statement for SciPy

[23] NumPy.org. Numpy license. [Online]. Available: http://www.numpy.org/license.html
    Licensing statement for NumPy

[24] H. Wickham, *ggplot2: elegant graphics for data analysis*. Springer New York, 2009. [Online]. Available: http://had.co.nz/ggplot2/book 978-0-387-98140-6.
    A book introducing the ggplot2 graphing package for R. Of particular use for new users to the syntax of the package.

[25] S. Bethard. argparse 1.2.1: Python command-line parsing library. [Online]. Available: https: //pypi.python.org/pypi/argparse/1.2.1
    Python Package Index record for the argparse library.

[26] M. Fowler, "Continuous Integration," 2006. [Online]. Available: http://www.martinfowler.com/ articles/continuousIntegration.html
    Article detailing experiences with continuous integration and the practices required to make it work for a team.

[27] Kohsuke Kawaguchi. Meet jenkins. [Online]. Available: https://wiki.jenkins-ci.org/display/ JENKINS/Meet+Jenkins
    About the Jenkins CI platform.

[28] Jenkins Usage Statistics. [Online]. Available: http://stats.jenkins-ci.org/jenkins-stats/svg/svgs.html
    Recent Jenkins usage statistics

[29] Getting Started: Travis CI Overview. [Online]. Available: http://docs.travis-ci.com/user/ getting-started/
    An introduction and overview of the Travis CI platform.

[30] Introduction to wercker. [Online]. Available: http://devcenter.wercker.com/articles/introduction/
    An introduction and overview of the Wercker CI platform.

[31] D. E. Knuth, *Art of Computer Programming, Volume II: Seminumerical Algorithms*, 3rd ed., 1998, vol. 2, p. 232.
    An efficient algorithm for the computation of running means and variances.

[32] W. B. Van Rossum, Guido and N. Coghlan, "PEP 8 – Style Guide for Python Code," *Python Enhancement Proposals*, 2001. [Online]. Available: http://legacy.python.org/dev/peps/pep-0008/
    Considered as the Python style guide.

[33] Kirk Strauser (http://stackoverflow.com/users/ 32538/kirk-strauser), "Does python have 'private' variables in classes?" [Online]. Available: http://stackoverflow.com/questions/1641219/ does-python-have-private-variables-in-classes
    StackOverflow discussion on the concept of private variables in Python.

[34] Python Software Foundation. unittest — Unit testing framework. [Online]. Available: https: //docs.python.org/2/library/unittest.html
    Python documentation for the unittest library, providing an introduction and helpful examples.

[35] Frontier [Github]. [Online]. Available: https: //github.com/SamStudio8/Frontier
    Github repository for the Frontier and Goldilocks packages.

[36] S. Russell and P. Norvig, *Artificial Intelligence: A Modern Approach*. Prentice Hall, 2010. [Online]. Available: http://books.google.co.uk/books? id=8jZBksh-bUMC 9780136042594.
    Recommended reading for last semester's Machine Learning module, providing an in-depth explanation of various machine learning algorithms that are available.

[37] scikit-learn developers. (2010–2013) Decision Trees. [Online]. Available: http://scikit-learn.org/stable/ modules/tree.html
    scikit-learn's introduction to trees including benefits and disadvantages.

[38] ——. (2010–2013) sklearn.cross_validation.StratifiedKFold. [Online]. Available: http://scikit-learn.org/stable/modules/ generated/sklearn.cross_validation.StratifiedKFold. html
    scikit-learn manual entry for usage of the StratifiedKFold class for creating proportionally weighted training and testing folds.

[39] ——. (2010–2013) sklearn.tree.DecisionTreeClassifier. [Online]. Available: http://scikit-learn.org/stable/modules/ generated/sklearn.tree.DecisionTreeClassifier.html
    scikit-learn manual entry for usage of its DecisionTreeClassifier, including examples and tips.

[40] L. Breiman, and A. Cutler. Random Forests: Gini importance. [Online]. Available: http://www.stat. berkeley.edu/~breiman/RandomForests/cc_home.htm

Explanation of the Gini importance method for calculating feature importances.

[41] *The bcftools/htslib VCF commands: bcftools query*, VCFTools Project. [Online]. Available: http://vcftools.sourceforge.net/htslib.html#query
Manual reference page for the bcftools query command.

[42] The Variant Call Format (VCF) Version 4.2 Specification. SAM tools Project. [Online]. Available: http://samtools.github.io/hts-specs/VCFv4.2.pdf
The VCF file specification, an essential reference when creating software designed to interact with either the VCF or BCF formats.

[43] Danecek, Petr and Auton, Adam and Abecasis, Goncalo and Albers, Cornelis A. and Banks, Eric and DePristo, Mark A. and Handsaker, Robert E. and Lunter, Gerton and Marth, Gabor T. and Sherry, Stephen T. and McVean, Gilean and Durbin, Richard and 1000 Genomes Project Analysis Group, "The Variant Call Format and VCFtools," *Bioinformatics*, vol. 27, no. 15, pp. 2156–2158, 2011. [Online]. Available: http://bioinformatics.oxfordjournals.org/content/27/15/2156.abstract
Paper introducing the VCF file format and VCFTools software collection.

[44] Orbis Play and Learn, Albert Barillé, "How My Body Works," 1992–1994.
A series of books designed to introduce children to the biology of the human body.

[45] Procidis and Albert Barillé, "Once upon a time... Life," 1987, [Television broadcast].
A 1980s television show to introduce children to the biology of the human body.

[46] U.S. National Library of Medicine. (2009) HLA Gene Family. [Online]. Available: http://ghr.nlm.nih.gov/geneFamily/hla
Information on HLA and its relationship with chromosome six.

[47] The SAM/BAM Format Specification Working Group. (2014) Sequence Alignment/Map Format Specification. [Online]. Available: http://samtools.github.io/hts-specs/SAMv1.pdf
The SAM/BAM file format specification, a useful reference when implementing algorithms that must interact with either format.

[48] iRODS Consortium. iRODS: About. [Online]. Available: http://irods.org/about/
About the iRODS data management platform.

[49] irods: Middleware for your critical data [Github].

[50] G.-T. Chiang, P. Clapham, G. Qi, K. Sale, and G. Coates, "Implementing a genomic data management system using iRODS in the Wellcome Trust Sanger Institute," *BMC Bioinformatics*, vol. 12, no. 1, p. 361, 2011. [Online]. Available: http://www.biomedcentral.com/1471-2105/12/361
Paper introducing Sanger's implementation of the iRODS data management platform.

[51] K. Lewis. samtools_irods [Github]. WTSI. [Online]. Available: https://github.com/wtsi-npg/samtools_irods
Github repository for the samtools_irods tool.

[52] *Manual Reference Pages - samtools (1): Samtools Commands And Options*, SAM tools Project. [Online]. Available: http://samtools.sourceforge.net/samtools.shtml#3
Manual reference page for the samtools tool collection.

[53] Calling SNPs/INDELs with SAMtools/BCFtools. SAM tools Project. [Online]. Available: http://samtools.sourceforge.net/mpileup.shtml
Manual reference page for the samtools mpileup command, including examples and suggestions on usage.

[54] Sam Nicholls. (2014) Frontier: Quality Control for Next Generation Sequencing. [Online]. Available: http://frontier.samnicholls.net/
Self-authored blog providing updates on the project status.

[55] 1000 Genomes. 1000 Genomes FAQ: Which reference assembly do you use? [Online]. Available: http://www.1000genomes.org/faq/which-reference-assembly-do-you-use
Discussion of the reference genomes used by the 1000 Genomes Project.

[56] zam.iqbal.genome (https://www.biostars.org/u/5152/). (2013) Decoy In Reference Assembly. [Online]. Available: https://www.biostars.org/p/73100/#73120
A BioStar question related to the usage of 'decoy' sequences in the reference sequence used by the second phase of the 1000 Genomes Project.

[57] Heng Li. (2011) The missing human sequences (version 5). [Online]. Available: ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/technical/reference/phase2_reference_assembly_sequence/hs37d5.slides.pdf
An introduction in to the reference sequence used by the second phase of the 1000 Genomes Project.

[58] *The bcftools/htslib VCF commands: bcftools call*, VCFTools Project. [Online]. Available: http://vcftools.sourceforge.net/htslib.html#call
Manual reference page for the bcftools call command.

[59] *bcftools call*, Genome Research Ltd. [Online]. Available: http://samtools.github.io/bcftools/bcftools.html#call

Additional manual and reference material on the bcftools call command.

[60] A. Cornish-Bowden, "Nomenclature for Incompletely Specified Bases in Nucleic Acid Sequences: Recommendations 1984." *Nucleic Acids Research*, vol. 13, no. 9, p. 3021, 1985.
Specification for the nomenclature of nucleotide bases that are potentially ambiguous or missing.

[61] C. Liébecq, *Biochemical Nomenclature and Related Documents*. Portland Press, 1992.
Specification for the nomenclature of nucleotide bases.

[62] Pablo Cingolani, *SnpSift Concordance*, 2013. [Online]. Available: http://snpeff.sourceforge.net/SnpSift.html#concordance
SnpSift Concordance software for measuring concordance between two VCF files.

[63] scikit-learn developers. (2010–2013) Decision Trees – Tips on practical use. [Online]. Available: http://scikit-learn.org/stable/modules/tree.html#tips-on-practical-use
Helpful tips on the practical use of the scikit-learn DecisionTreeClassifier class.

[64] W. S. Kendal, "An Exponential Dispersion Model for the Distribution of Human Single Nucleotide Polymorphisms," *Molecular Biology and Evolution*, vol. 20, no. 4, pp. 579–590, 2003.
A paper investigation the density of SNPs across regions of the human genome. Presented as a use case for Goldilocks.

[65] T. Strachan and A. Read, *Human Molecular Genetics*, 4th ed. Garland Science, 2011, pp. 2–5.
An excellent introduction to DNA and RNA.

[66] ——, *Human Molecular Genetics*, 4th ed. Garland Science, 2011, p. 30.
Continuing from [65], introduces the concept of Chromosomes.

[67] Kris Wetterstrand. DNA Sequencing Costs: Data from the NHGRI Genome Sequencing Program (GSP). [Online]. Available: http://www.genome.gov/sequencingcosts/

[68] Maggie Bartlett. (2011) Illumina HiSeq Flow Cell. National Human Genome Research Institute. Image. [Online]. Available: http://www.genome.gov/dmd/img.cfm?node=Photos/Technology/Genome%20analysis%20technology&id=80102
Public domain image of a flowcell.

[69] Python Software Foundation. More on Lists. [Online]. Available: https://docs.python.org/2/tutorial/datastructures.html#more-on-lists
Python documentation for the list data structure.

[70] Python Wiki. (2012) Time Complexity. [Online]. Available: https://wiki.python.org/moin/TimeComplexity
An introduction to Python data structures and their computational complexity.

[71] Python Software Foundation. Dictionaries. [Online]. Available: https://docs.python.org/2/tutorial/datastructures.html#dictionaries
Python documentation for the dictionary data structure.

[72] ——. Sets. [Online]. Available: https://docs.python.org/2/tutorial/datastructures.html#sets
Python documentation for the set data structure.

[73] Hadley Wickham and Winston Chang, *devtools: Tools to make developing R code easier*, 2013, r package version 1.4.1. [Online]. Available: http://CRAN.R-project.org/package=devtools
R manual for the devtools package.

[74] Morandat, Floréal and Hill, Brandon and Osvald, Leo and Vitek, Jan, "Evaluating the design of the R language," in *ECOOP 2012–Object-Oriented Programming*. Springer, 2012, pp. 104–131.
A thorough investigation in to the performance of the R programming language as compared to C and Python.

Appendix A

Concept Glossary

A.1 Introduction

A.1.1 DNA, Bases and Sequencing

Practically every cell in a living organism contains deoxyribonucleic acid (**DNA**). DNA provides a coded template essential for the construction of proteins vital for ongoing development and function of a living organism. DNA is a polymer (long molecule) made up of a sequence of just four nucleic acids called nucleotides or **bases**[65]:

- **G** guanine,

- **C** cytosine,

- **T** thymine and

- **A** adenine.

DNA is arranged in to linear structures called **chromosomes**, of which humans have 23 pairs; consisting of 22 hereditary trait carrying **autosomes** (numbered from 1–22) and a pair of sex-determining **allosomes** (a combination of X and Y)[66]. The human genome is approximately three billion bases long.

A.1.2 NGS

Sequencing is the process of recovering or extracting the sequence of these nucleotide bases that appear in a particular organism's DNA.

Next Generation Sequencing (**NGS**) is a somewhat generic term used to refer to improving sequencing hardware and methods introduced from 2008 onwards, bringing a massive decrease in both the time and cost involved in the sequencing of an entire human genome[67].

A.1.3   Samples, Lanes and Lanelets

A **sample** is a distinct DNA specimen extracted from a particular person. For the purpose of sequencing, samples are pipetted in to a *flowcell* such as the one in Figure A.1 – a glass slide containing a series of very thin tubules known as **lanes**. It is throughout these lanes that the chemical reactions involved in sequencing take place.
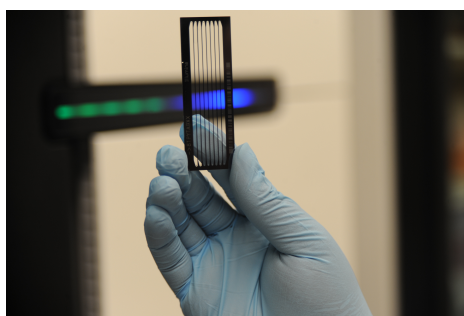
Fig. A.1 An Illumina HiSeq Flowcell[68]

Once inserted, samples are amplified *in situ*, in the flowcell itself. A process in which the genetic material of each sample is caused to multiply in magnitude to form a dense cluster of the sample around the original. Millions of clusters will be created throughout each lane of the flowcell.

Note that a lane can contain more than one sample and a sample can appear in more than one lane; this is known as *sample multiplexing* and helps to ensure that the failure of a particular lane does not hinder analysis of a sample (as it will still be sequenced as part of another lane).

The more abstract of the definitions, a **lanelet** is the aggregate read of all clusters of a particular sample in a single lane.

A.2   Part I

A.2.1   Observations and Parameters

An **observation** refers to a distinct object or "thing" from which we will attempt to understand the relationship between its known properties and classification to be able to label future observations with unknown class based on those properties. In the context of this project, this would be a lanelet. Other documentation in the field may use the term *sample* but to avoid confusion with the definition of sample introduced in Chapter A.1.3 we will use the term observation.

The aforementioned *properties* or *attributes* of an observation will be referred to as the **parameters** of that observation. Traditionally these may be described as *features* but it was felt that this wording may have connotations with discrete data. Early versions of Frontier referred to parameters as *regressors*, using terminology from statistical modelling. This wording was dropped to remove any confusion between regression and classification machine learning problems.

A.2.2   Data and Targets

Data will be used somewhat generically to refer to a matrix in which observations act as rows and their parameters act as columns. It is expected that all observations will have the same set of parameters.

For problems of a classification nature such as ours, an observation's **target** is the known classification of that particular observation. Depending on the context of the learning problem targets may be discovered manually (*e.g.* counting leaves from images of plants, one will need to manually count all the leaves in the image before being able to use it for learning) or as the output from another system such as **auto_qc**.

In the context of this project where the objective is to begin learning the rules of **auto_qc** the targets refer to whether a particular lanelet observation was labelled as a pass, fail or warn by the current system.

A.2.3   Class Labels and Codes

An observation's target may also be known as its **label**. A labelled observation is said to be a member of that label's **class**. For example a lanelet that has failed **auto_qc** is said to be labelled as a fail and is a member of the class of failures.

String based class labels can pose difficulties when handling data later on, not only taking more memory to store as opposed to a type such as an integer but also introducing accidental subsetting or discarding of data whose labels differ unexpectedly. For example are "fail", ,"fial", "FAIL" and "Failed" supposed to be members of the same class?

Often such labels are **encoded** in to simpler types such as integers.

A.2.4   Python Data Structures

The Python standard library offers many different data structures each with their own properties. I introduce three such structures below to assist the reader in understanding some of the implementation decisions made in the following section:

List

A **list**[69] essentially represents an ordered sequence of elements. *Lists* are incredibly versatile, allowing elements of any data type, including *lists* and other container structures or arbitrary objects. Elements in a *list* are accessed by their index (location) in the sequence. *Lists* permit duplicate values.

Lists are represented in memory as a contiguous block and so great expense is incurred if the *list* grows beyond its bounds and must be relocated elsewhere in memory to resize to a larger contiguous block. Whilst items can be appended to or popped from anywhere in the container, performing those actions on a location that is not at the end of the *list* will require the contents of the structure to be moved in memory to maintain the contiguous layout.

Checking whether an item is in a *list* requires iterating over all *n* elements in the container until either a match is found or there are no elements left to check[70].

Dictionary

A **dict**[71] or *dictionary* is a structure containing key-value pairs in which string or numeric keys[1] are mapped to an arbitrary value or object through a hashing function. Keys in a *dictionary* are unique; attempting to add a key-value pair where the key already exists will overwrite the original entry.

Python *dictionaries* offer constant lookup access and more importantly, unlike a list, querying the structure for whether it contains a particular key can be performed in constant time[70]. However it is important to note that a *dictionary* does not maintain order, keys are not sorted alphanumerically nor are they sorted by insertion time. Ordering is determined arbitrarily by the **dict**'s underlying hash table[2].

Set

Sets[72] represent a collection of elements and are similar to a **dict** in that they are allowed to contain any immutable (thus hashable) elements but entries are not mapped to a value or object. *Sets* are forbidden from containing duplicate objects and do not maintain order.

Like **dict**, checking for membership of an item in a *set* completes in constant time, making the structure particularly useful for membership testing and filtering duplicates out of other structures.

A.3   Part II: Goldilocks

A.3.1   Group

Rather than considering the location of all variants in our samples as a whole, we may wish to consider how variants are distributed across the two different data sets we have, namely the **GWAS** and **iCHIP** sets. Thus variants will be considered as members of a **group**. The group may be arbitrarily defined but for the purpose of our own analysis the set in which a variant appears will be considered as its group.

A particular variant location may appear in more than one group.

A.3.2   Length and Stride

The **length** of a region represents the number of bases included in the region and the **stride** refers to the number of bases to be used as an offset between the first base of one region and the beginning of the next.

Sensible choices for these parameters will need to be selected after considering both time and memory.

---

[1]Any type can actually be used as a dictionary key providing it is immutable, including tuples[71]

[2]This is probably the question I see most often from users new to Python on StackOverflow

A.3.3   Density

The **density** of a region represents the number of variants located within that region. **Density** will be recorded for each *group* of variants.

A.3.4   Candidate Regions

A **candidate** is a region of *length* bases whose first base falls on a *stride*. A **candidate** region will contain some count (described as the *density*) of variants for each group. In the context of this project we consider the number of variant locations that fall within this region for both the **GWAS** and **iCHIP** data sets.

Candidates must meet all (any any additional) criteria to be considered. For example, regions whose first position falls on a base in such a way that the chromosome ends before a region of *length* can be constructed will violate the size criteria.

Appendix B

Input Examples

B.1   BAMcheckR'd Example Output

```
# Summary Numbers. Use 'grep ^SN | cut -f 2-' to extract this part.
SN      raw total sequences:          41400090
SN      filtered sequences:        0
SN      sequences:         41400090
SN      is paired:       1
SN      is sorted:       1
SN      1st fragments:         20700045
SN      last fragments:         20700045
SN      reads mapped:       41291484
SN      reads unmapped:        108606
SN      reads unpaired:        60000
SN      reads paired:       41231484
SN      reads duplicated:        5756822
SN      reads MQ0:      1038644
SN      reads QC failed:       0
SN      non-primary alignments:         0
SN      total length:        3105006750
SN      bases mapped:       3096861300
SN      bases mapped (cigar):         3090885143
SN      bases trimmed:        0
SN      bases duplicated:         431761650
SN      mismatches:        9107833
SN      error rate:        0.002946675
SN      average length:        75
SN      maximum length:        75
SN      average quality:         36
SN      insert size average:         178.7
SN      insert size standard deviation:        44.1
SN      inward oriented pairs:         20577242
SN      outward oriented pairs:        3140
SN      pairs with other orientation:        3711
SN      pairs on different chromosomes:        31535
SN      fwd percent insertions above baseline:        1.43135383851191
SN      fwd percent insertions below baseline:        0.686265539012562
SN      fwd percent deletions above baseline:        1.38326380878871
SN      fwd percent deletions below baseline:        0.44923551909251
SN      rev percent insertions above baseline:        1.08264446659241
SN      rev percent insertions below baseline:        0.457290262062496
```

```
SN        rev percent deletions above baseline:        1.15931214598243
SN        rev percent deletions below baseline:        0.413119424753248
SN        contiguous cycle dropoff count:        36
SN        fwd.percent.insertions.above.baseline:        1.43135383851191
SN        fwd.percent.insertions.below.baseline:        0.686265539012562
SN        fwd.percent.deletions.above.baseline:        1.38326380878871
SN        fwd.percent.deletions.below.baseline:        0.44923551909251
SN        rev.percent.insertions.above.baseline:        1.08264446659241
SN        rev.percent.insertions.below.baseline:        0.457290262062496
SN        rev.percent.deletions.above.baseline:        1.15931214598243
SN        rev.percent.deletions.below.baseline:        0.413119424753248
SN        quality.dropoff.fwd.high.iqr.start.read.cycle:        0
SN        quality.dropoff.fwd.high.iqr.end.read.cycle:        0
SN        quality.dropoff.fwd.high.iqr.max.contiguous.read.cycles:        0
SN        quality.dropoff.fwd.mean.runmed.decline.start.read.cycle:        20
SN        quality.dropoff.fwd.mean.runmed.decline.end.read.cycle:        51
SN        quality.dropoff.fwd.mean.runmed.decline.max.contiguous.read.cycles:        32
SN        quality.dropoff.fwd.mean.runmed.decline.high.value:        36.9775883578997
SN        quality.dropoff.fwd.mean.runmed.decline.low.value:        36.301749247405
SN        quality.dropoff.rev.high.iqr.start.read.cycle:        0
SN        quality.dropoff.rev.high.iqr.end.read.cycle:        0
SN        quality.dropoff.rev.high.iqr.max.contiguous.read.cycles:        0
SN        quality.dropoff.rev.mean.runmed.decline.start.read.cycle:        18
SN        quality.dropoff.rev.mean.runmed.decline.end.read.cycle:        56
SN        quality.dropoff.rev.mean.runmed.decline.max.contiguous.read.cycles:        39
SN        quality.dropoff.rev.mean.runmed.decline.high.value:        36.1517621338504
SN        quality.dropoff.rev.mean.runmed.decline.low.value:        35.3152133727245
SN        quality.dropoff.high.iqr.threshold:        10
SN        quality.dropoff.runmed.k:        25
SN        quality.dropoff.ignore.edge.cycles:        3
SN        A.percent.mean.above.baseline:        0.0991164444444441
SN        C.percent.mean.above.baseline:        0.127379555555556
SN        G.percent.mean.above.baseline:        0.0603679999999997
SN        T.percent.mean.above.baseline:        0.0868000000000005
SN        A.percent.mean.below.baseline:        0.0991164444444451
SN        C.percent.mean.below.baseline:        0.127379555555555
SN        G.percent.mean.below.baseline:        0.0603680000000002
SN        T.percent.mean.below.baseline:        0.0867999999999993
SN        A.percent.max.above.baseline:        0.601733333333332
SN        C.percent.max.above.baseline:        0.394266666666667
SN        G.percent.max.above.baseline:        0.2956
SN        T.percent.max.above.baseline:        0.768000000000001
SN        A.percent.max.below.baseline:        0.318266666666666
SN        C.percent.max.below.baseline:        0.825733333333332
SN        G.percent.max.below.baseline:        0.554400000000001
SN        T.percent.max.below.baseline:        0.251999999999999
SN        A.percent.max.baseline.deviation:        0.601733333333332
SN        C.percent.max.baseline.deviation:        0.825733333333332
SN        G.percent.max.baseline.deviation:        0.554400000000001
SN        T.percent.max.baseline.deviation:        0.768000000000001
SN        A.percent.total.mean.baseline.deviation:        0.198232888888889
SN        C.percent.total.mean.baseline.deviation:        0.254759111111111
SN        G.percent.total.mean.baseline.deviation:        0.120736
SN        T.percent.total.mean.baseline.deviation:        0.1736
# First Fragment Qualitites. Use 'grep ^FFQ | cut -f 2-' to extract this part.
# Columns correspond to qualities and rows to cycles. First column is the cycle number.
FFQ        1        8968        3619        9863        747        5094        0        6642        1609        4673        ...
FFQ        2        21676        0        0        0        0        0        0        43        1885        ...
FFQ        3        7        0        177        0        0        0        0        0        0        ...
[...]
FFQ        74        3697        39        0        919        4933        0        0        56866        1524        ...
FFQ        75        4542        0        0        0        0        0        4634        77822        0        ...
FFQ        76        0        0        0        0        0        0        0        0        ...
```

```
# Last Fragment Qualitites. Use 'grep ^LFQ | cut -f 2-' to extract this part.
# Columns correspond to qualities and rows to cycles. First column is the cycle number.
LFQ      1       8869      0        0        0        0        0       63        0        0        1156        ...
LFQ      2       3300      0        0        0        0        0        0        0        0        0          ...
LFQ      3       6816      0        0        0        573      0       83        0        7011                ...
[...]
LFQ      74      5980      3        91       0        0        0       1340     9696     72939               ...
LFQ      75      4314      0        0        168      0        848     8591        0     70358               ...
LFQ      76      0         0        0        0        0        0        0        0        0        0          ...
# Mismatches per cycle and quality. Use 'grep ^MPC | cut -f 2-' to extract this part.
# Columns correspond to qualities, rows to cycles. First column is the cycle number, second
# is the number of N's and the rest is the number of mismatches
MPC      1       14078     0        2594     6777     416      1919     0       2222                ...
MPC      2       21407     0        0        0        0        0        0        0        5          ...
MPC      3       3205      0        0        37       0        43       0       12        0          ...
[...]
MPC      74      779       0        3        0        131      440      0       93       7485       ...
MPC      75      136       0        0        0        3        0        47      704      9302       ...
MPC      76      0         0        0        0        0        0        0        0        0          ...
# GC Content of first fragments. Use 'grep ^GCF | cut -f 2-' to extract this part.
GCF      0.5        56
GCF      1.76       60
GCF      3.02       126
GCF      4.27       212
GCF      5.78       347
[...]
GCF      93.72      378
GCF      95.23      186
GCF      96.48      87
GCF      97.74      55
GCF      99.25      17
# GC Content of last fragments. Use 'grep ^GCL | cut -f 2-' to extract this part.
GCL      0.5        118
GCL      1.76       175
GCL      3.02       230
GCL      4.27       354
GCL      5.78       525
[...]
GCL      93.72      613
GCL      95.23      430
GCL      96.48      274
GCL      97.74      185
GCL      99.25      110
# ACGT content per cycle. Use 'grep ^GCC | cut -f 2-' to extract this part. The columns are: cycle, and A,C,G,T counts [%]
GCC      1       26.93      23.09      22.77      27.2
GCC      2       26.78      23.24      22.97      27.02
GCC      3       26.46      23.59      23.3       26.66
GCC      4       26.29      23.79      23.45      26.46
GCC      5       26.47      23.61      23.3       26.62
[...]
GCC      70      26.09      24.26      23.45      26.2
GCC      71      26.07      24.25      23.46      26.22
GCC      72      26.04      24.27      23.49      26.2
GCC      73      26.07      24.25      23.47      26.22
GCC      74      26.08      24.24      23.45      26.23
GCC      75      26.01      24.31      23.51      26.18
# Insert sizes. Use 'grep ^IS | cut -f 2-' to extract this part.
# The columns are: pairs total, inward oriented pairs, outward oriented pairs, other pairs
IS       0       10         0          1          9
IS       1       3          0          3          0
IS       2       4          0          4          0
IS       3       5          0          5          0
IS       4       2          0          2          0
```

```
IS        5        3        0        3        0
[...]
IS        110      33952         33952        0        0
IS        111      38433         38433        0        0
IS        112      43373         43370        0        3
IS        113      48160         48159        0        1
IS        114      53175         53171        0        4
IS        115      59504         59502        0        2
IS        116      64668         64668        0        0
IS        117      71107         71105        0        2
IS        118      77157         77156        0        1
IS        119      84044         84044        0        0
IS        120      90116         90110        3        3
[...]
IS        327      6546          6546         0        0
IS        328      6483          6483         0        0
IS        329      6201          6201         0        0
IS        330      6228          6228         0        0
IS        331      5852          5852         0        0
# Read lengths. Use 'grep ^RL | cut -f 2-' to extract this part. The columns are: read length, count
RL        75       41400090
# Indel distribution. Use 'grep ^ID | cut -f 2-' to extract this part.
# The columns are: length, number of insertions, number of deletions
ID        1        128650        183418
ID        2        26409         39770
ID        3        10213         16046
ID        4        7756          11444
ID        5        1746          3455
[...]
ID        35       0        8
ID        36       0        1
ID        37       0        1
ID        38       0        1
ID        40       0        2
# Indels per cycle. Use 'grep ^IC | cut -f 2-' to extract this part.
# The columns are: cycle, number of insertions (fwd), .. (rev) , number of deletions (fwd), .. (rev)
IC        1        0        0        105      97
IC        2        24       15       150      179
IC        3        129      138      441      509
IC        4        253      310      623      829
IC        5        557      724      786      1164
[...]
IC        70       571      710      638      761
IC        71       350      428      309      434
IC        72       154      150      38       45
IC        73       60       61       15       23
IC        74       20       19       11       12
# Coverage distribution. Use 'grep ^COV | cut -f 2-' to extract this part.
COV       [1-1]         1        332980694
COV       [2-2]         2        105004580
COV       [3-3]         3        29112182
COV       [4-4]         4        13415014
COV       [5-5]         5        6716815
[...]
COV       [996-996]         996       2
COV       [997-997]         997       2
COV       [998-998]         998       2
COV       [1000-1000]        1000       4
COV       [1000<]        1000       116
# GC-depth. Use 'grep ^GCD | cut -f 2-' to extract this part.
# The columns are: GC%, unique sequence percentiles, 10th, 25th, 50th, 75th and 90th depth percentile
GCD       0        0.001      0        0        0        0        0
GCD       0.4      0.002      0.101      0.101      0.101      0.101      0.101
```

```
GCD      19      0.003     0.049      0.049     0.049      0.049      0.049
GCD      20      0.004     0.06       0.06      0.06       0.06       0.06
GCD      21      0.004     0.045      0.045     0.045      0.045      0.045
[...]
GCD      66      99.99     0.244      2.693     6.746      11.794     15.885
GCD      67      99.994    1.279      1.279     4.305      9.667      11.483
GCD      68      99.997    4.148      4.148     4.463      5.741      7.354
GCD      69      99.999    0.499      0.499     0.499      1.935      1.935
GCD      72      100       0.476      0.476     0.476      1.219      1.219
```

B.2 auto_qc Decision Matrix

```
lanelet        sample        study       npg       aqc        ...
9999_9#1       AQCTest000000       AQCTest     pass      passed       ...
9999_9#3       AQCTest000002       AQCTest     fail      failed       ...
9999_9#2       AQCTest000001       AQCTest     pass      pass       ...
9999_9#5       AQCTest000004       AQCTest     warn      warn       ...
9999_9#4       AQCTest000003       AQCTest     warn      warning        ...
9999_9#7       AQCTest000006       AQCTest     fail      fail       ...
9999_9#6       AQCTest000005       AQCTest     pass      passed       ...
9999_9#9       AQCTest000008       AQCTest     warn      warning        ...
9999_9#8       AQCTest000007       AQCTest     pass      passed       ...
```

B.3 Goldilocks Paths File

```
# gwas
cd-gwas        /pools/encrypted/sanger/vcf/cd-seq.vcf.gz.q
uc-gwas        /pools/encrypted/sanger/vcf/uc-seq.vcf.gz.q
# ichip
cd-ichip        /pools/encrypted/sanger/vcf/cd.ichip.vcf.gz.q
uc-ichip        /pools/encrypted/sanger/vcf/uc.ichip.vcf.gz.q
```

Appendix C

Output Examples

C.1   Frontier Log Example

```
Frontier
********
Data Dir        /pools/encrypted/sanger/frontier/data/bamcheck_2013dec25_ratios_out/
AQC File        /pools/encrypted/sanger/frontier/data/crohns-uc-table-a.2013dec25.manual_qc_update.txt


Class Def         fail        pass        warn
Class Read        1542        9154        2759
Class Used        1542        9154        2759


Total Read        13455
Total Used        13455


Param Set         ALL
Param Count         86
Param List:
                1st-fragments
                A-percent-max-above-baseline
                A-percent-max-baseline-deviation
                A-percent-max-below-baseline
                A-percent-mean-above-baseline
                A-percent-mean-below-baseline
                A-percent-total-mean-baseline-deviation
                C-percent-max-above-baseline
                C-percent-max-baseline-deviation
                C-percent-max-below-baseline
                C-percent-mean-above-baseline
                C-percent-mean-below-baseline
                C-percent-total-mean-baseline-deviation
                G-percent-max-above-baseline
                G-percent-max-baseline-deviation
                G-percent-max-below-baseline
                G-percent-mean-above-baseline
                G-percent-mean-below-baseline
                G-percent-total-mean-baseline-deviation
                T-percent-max-above-baseline
                T-percent-max-baseline-deviation
                T-percent-max-below-baseline
                T-percent-mean-above-baseline
```

```
T-percent-mean-below-baseline
T-percent-total-mean-baseline-deviation
average-length
average-quality
bases-duplicated
bases-mapped
bases-mapped-(cigar)
bases-trimmed
duplicate-mapped-ratio
error-rate
filtered-sequences
fwd-percent-deletions-above-baseline
fwd-percent-deletions-below-baseline
fwd-percent-insertions-above-baseline
fwd-percent-insertions-below-baseline
indel-ratio
insert-size-average
insert-size-standard-deviation
inward-oriented-pairs
is-paired
is-sorted
last-fragments
max-max-baseline-deviation
max-total-mean-baseline-deviation
maximum-length
mismatches
non-primary-alignments
outward-oriented-pairs
pairs-on-different-chromosomes
pairs-with-other-orientation
quality-dropoff-fwd-high-iqr-end-read-cycle
quality-dropoff-fwd-high-iqr-max-contiguous-read-cycles
quality-dropoff-fwd-high-iqr-start-read-cycle
quality-dropoff-fwd-mean-runmed-decline-end-read-cycle
quality-dropoff-fwd-mean-runmed-decline-high-value
quality-dropoff-fwd-mean-runmed-decline-low-value
quality-dropoff-fwd-mean-runmed-decline-max-contiguous-read-cycles
quality-dropoff-fwd-mean-runmed-decline-start-read-cycle
quality-dropoff-high-iqr-threshold
quality-dropoff-ignore-edge-cycles
quality-dropoff-rev-high-iqr-end-read-cycle
quality-dropoff-rev-high-iqr-max-contiguous-read-cycles
quality-dropoff-rev-high-iqr-start-read-cycle
quality-dropoff-rev-mean-runmed-decline-end-read-cycle
quality-dropoff-rev-mean-runmed-decline-high-value
quality-dropoff-rev-mean-runmed-decline-low-value
quality-dropoff-rev-mean-runmed-decline-max-contiguous-read-cycles
quality-dropoff-rev-mean-runmed-decline-start-read-cycle
quality-dropoff-runmed-k
raw-total-sequences
reads-MQ0
reads-QC-failed
reads-duplicated
reads-mapped
reads-paired
reads-unmapped
reads-unpaired
rev-percent-deletions-above-baseline
rev-percent-deletions-below-baseline
rev-percent-insertions-above-baseline
rev-percent-insertions-below-baseline
sequences
total-length
```

```
Feature Importances:
            0.27      T-percent-max-above-baseline
            0.12      rev-percent-insertions-above-baseline
            0.09      fwd-percent-insertions-above-baseline
            0.07      quality-dropoff-rev-mean-runmed-decline-low-value
            0.07      insert-size-average
            0.05      quality-dropoff-fwd-mean-runmed-decline-low-value
            0.04      max-max-baseline-deviation
            0.03      insert-size-standard-deviation
            0.02      reads-duplicated
            0.02      fwd-percent-deletions-above-baseline
            0.02      duplicate-mapped-ratio
            0.02      rev-percent-insertions-below-baseline
            0.02      max-total-mean-baseline-deviation
            0.02      bases-duplicated
            0.02      T-percent-max-baseline-deviation
            0.01      average-quality
            0.01      A-percent-max-above-baseline


CV Score (Fld)        0.97 +/- 0.01 (10)


Decision PDF          pdf/2014-05-05_2031__ALL_ALL_/
********
```

Appendix D

Pre-Frontier Implementation

D.1 Classification Correlation

An important consideration for statistical analysis is the relation between observations. The "bamcheckr'd" input data described in Chapter 3.1.1 is available per lanelet, however as shown in Chapter A.1.3 a lane may contain more than one lanelet. Herein lies the trouble: if during a sequencing run the flowcell is somehow subjected to abnormal conditions (*e.g.* a temperature increase due to an air conditioning failure) or the device is depleted of reagents then every lane (and thus all lanelets within) will be of considerably poor quality.
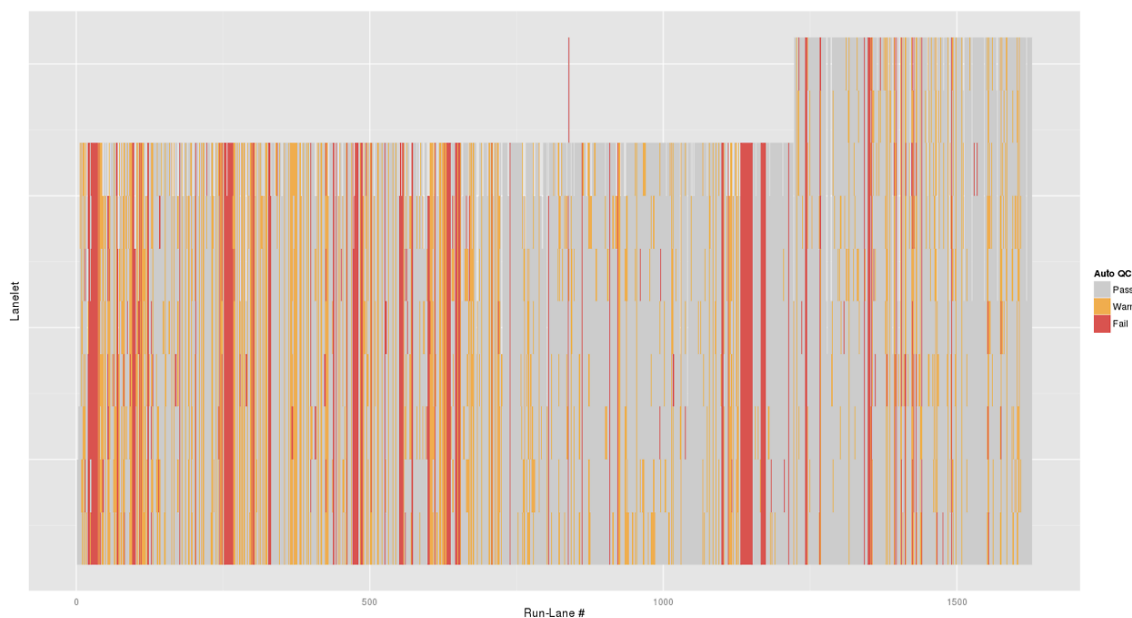

Fig. D.1 **Heatmap of lanelet QC status by lane**: Lanes are vertical bars with each lanelet cell coloured red to represent a failure, yellow for a warning and grey for a pass.

In such a case there would appear to exist a relationship between the respective qualities of each lanelet in

a lane as well as each lane in a sequencing run. To examine this further, an R script utilising **ggplot2** was authored to visually inspect whether correlation existed and if so to what degree that data is affected.

Figure D.1 displays a plot of **auto_qc** classification for each lanelet in a lane. The plot itself is a dense heatmap where each lane stands as a vertical bar, broken in to horizontal cells, each of which represents a lanelet that was sequenced in that particular lane. These lanelets are colour coded using; red for failures, yellow for warnings and grey for passes (to allow the other two classes to be more easily seen).

Therefore an unbroken vertical red line indicates that all lanelets that comprise of that line failed to pass some aspect of the current **auto_qc** thresholds. In reality there are few conditions under which a lanelet would fail irrespective of the status of the rest of the lanelets in the same lane which typically involve an error during the preparation of the sample (an easy to spot result as it will cause poor quality across all lanelets using that sample).

Overall there are a series of instances appearing to support correlation for whole-lane failures and warnings but despite this there do appear to be occasions where a lanelet has failed where the remainder of the lane has not. Having discussed this with the project supervisor and contacts at the Sanger Institute we decided to continue to the implementation stage, agreeing that whilst some evidence of correlation between lanelets in the same lane has emerged, we will still be able to recover parameters that will be useful to quality control and statistical testing may be required following this analysis to describe how powerful such parameters are taking this possible correlation in to account.

It should be noted that the proportion of failures and warnings is considerably smaller than passes and so care will need to be taken to find a balance; for example it would not be feasible to merely discard lanelets from lanes that have failed entirely as there'd hardly be any data on which to train a classifier. Indeed other solutions may be possible, perhaps weighting observations which exhibit similar behaviour to other lanelets in their lane so as to give their parameter values less priority during the construction of the classifier itself.

It is worth noting that although the plot does not make a particular distinction between lanes in the same flow cell, lanes are sequentially identified so the red bars of thicker-width arguably display some failures across entire flow cells.

As a final note it should be stressed that this plot should be regarded as a diagnostic rather than an experiment with a direct conclusion. Given more time it would be useful to investigate the nature of these possible correlations, given a lane that has failed across all lanelets: do those lanelets actually express similar quality metrics?

D.2 Recovering Ratios

An initial scan of the summary numbers available in the "bamcheckr'd" files introduced in Chapter 3.1.1 revealed 82 different parameters. However, when comparing this parameter set to the threshold rules of **auto_qc**, it appeared that some parameters were "missing".

In the pursuit of replicating the decisions of the existing system, it is ideal to have all the parameters used

in the making of those decisions at hand. In particular, the missing parameters were of a normalised nature, typically in the form of a ratio or percentage which should make them more valuable than a parameter that just represents a raw count of some property.

It was found that these missing parameters are calculated in a step preceding **auto_qc** as part of the **vr-pipe** pipeline[1]. Whilst I could have attempted to set-up my own instance of **vr-pipe** to recover this data, speaking with contacts at the Sanger Institute, it was decided that this would prove troublesome work; requiring an involved deployment to a cloud based facility such as Amazon's Elastic Compute Cloud and installation of various Perl dependencies as well as requiring access to many controlled databases within the institute.

Most of the functions that calculated these parameters turned out to be straightforward and could easily be ported to another application. At first it was intended to add these functions to the program authored for this part of the project, but the Sanger Institute suggested it would be more useful to implement such functionality in **bamcheckr** directly, removing some of **auto_qc**'s dependence on its position in **vr-pipe**.

```r
# Install and include the 'devtools' package
install.packages("devtools")
library(devtools)


# Install package directly from Github repository
install_github("samstudio8/seq_autoqc", subdir="bamcheckr")


# Install package from local directory
install("/home/sam/Projects/seq_autoqc/bamcheckr")
```

Listing 11 : Installing an in-development R package with **devtools**

With the help of **devtools**[73] (see Listing 11) it was simple to develop and test contributions to **bamcheckr** locally without needing to re-publish the package after changes. An additional script was added to **bamcheckr**'s NAMESPACE to recover the missing ratio and percentage based parameters.

Unfortunately, the performance of the new script was poor, taking 5.5s on average and increasing to 16.1s when enabling a complex function containing many vector operations – potentially inefficient due to my lack of R experience. It was simply impractical to use **bamcheckr** to update thousands of files and thus I instead used **Frontier** to load the "BAMcheckr'd" files and used its API to calculate the missing parameters and write them back to the files. An impressive feat for **Frontier** considering how simple it was to use it for something it was not actually intended for.

An evaluation of the R language[74] compares the language's performance to both Python and C. With such an intriguing difference in performance between **bamcheckr** and **Frontier**, with significantly more time it would be interesting to explore the idiosyncrasies of each implementation.

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[1]https://github.com/wtsi-hgi/vr-pipe/blob/hgi-release/modules/VRPipe/Steps/vrtrack_auto_qc_hgi_3.pm

Appendix E

Third Party Libraries