

16SrRNA Intermediate Bioinformatics Online Course: Int\_BT\_2019

# 16S analysis pipeline QC and ASV picking using the dada2 pipeline







#### **Outline**



- Quality Control
- DADA2 background

DADA2 workflow









 Before analyzing generated sequence to draw biological conclusions, a quality control check should be performed to make sure there is no biases in the data.

 QC gives a quick impression of whether your data has any problems of which you should be aware before doing any analysis.









#### Potential problems:

- Low confidence bases (Ns)
- Sequence specific bias
- Sequence contamination
- Adapters
- •









#### Software packages for QC:

- FastQC
- MultiQC
- FastX-Toolkit
- PRINSEQ
- TagCleaner
- NGS QC Tool-Kit
- •









#### **FASTQ** format

What is a FastQ file?

**FASTQ= FASTA + Quality** 

**FastQ format** is a text-based format for storing both a biological sequence and its corresponding quality scores.











#### **FASTQ** format

- Each FastQ file contains hundreds of millions of rows.
- Each block of 4 lines, starting with "@" represents a read.

**Line 1** begins with a '@' character and is followed by a sequence identifier and an optional description (like a FASTA title line).

**Line 2** is the raw sequence letters (ATCG).

**Line 3** begins with a '+' character and is optionally followed by the same sequence identifier (and any description).

Line 4 encodes the quality values for the sequence in Line 2, and must contain the same number of symbols as letters in the sequence.









#### **FASTQ** format

A FastQ file containing a single sequence might look like this:

@read name

GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT

+ read name

!"\*((((\*\*\*+))%%%++)(%%%%).1\*\*\*-+\*"))\*\*55CCF>>>>>CCCCCC65

The character '!' represents the lowest quality while '~' is the highest.









### **Quality measurements**

Base-calling error probabilities are reported by sequencers. Usually in Phred (quality) score. Usually coded by ASCII characters

#### Phred score

$$Q = -10log_{10}P$$

If the quality of a base is 20, the probability that it is wrong is 0.01









#### **Quality measurements**

```
!"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHIJKLMNOPQRSTUVWXYZ[\]^_`abcdefghijklmnopgrstuvwxyz{|}~
33
                                               126
                                   104
0.2......41
S - Sanger
        Phred+33, raw reads typically (0, 40)
         Solexa+64, raw reads typically (-5, 40)
X - Solexa
I - Illumina 1.3+ Phred+64, raw reads typically (0, 40)
J - Illumina 1.5+ Phred+64, raw reads typically (3, 41)
  with 0=unused, 1=unused, 2=Read Segment Quality Control Indicator (bold)
  (Note: See discussion above).
L - Illumina 1.8+ Phred+33, raw reads typically (0, 41)
```









#### What is FastQC?

FastQC aims to provide a QC report which can spot problems which originate either in the sequencer or in the library material.









### **FastQC reports**











# **FastQC reports**

# Basic Statistics

Measure	Value
Filename	<pre>good_sequence_short.txt</pre>
File type	Conventional base calls
Encoding	Illumina 1.5
Total Sequences	250000
Sequences flagged as poor quality	0
Sequence length	40
%GC	45

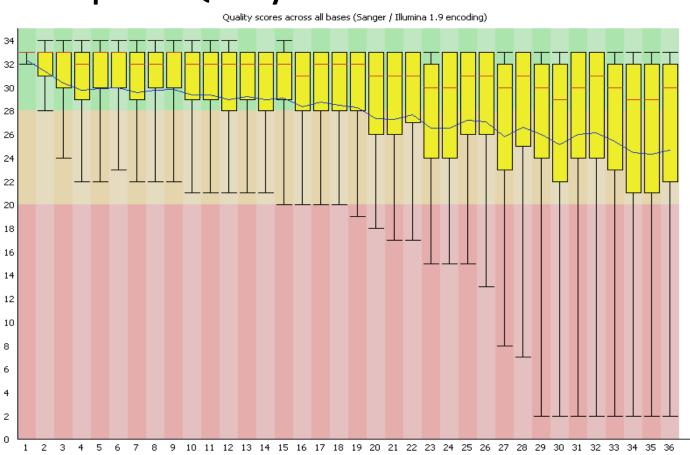








# FastQC reports Per Base Sequence Quality







Position in read (bp)

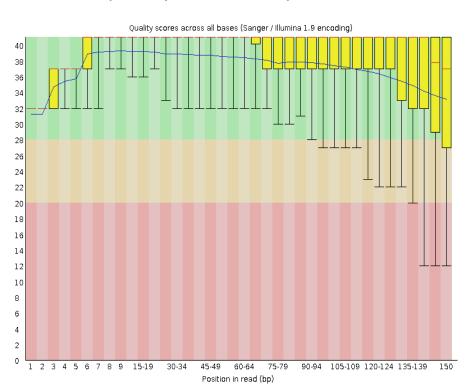




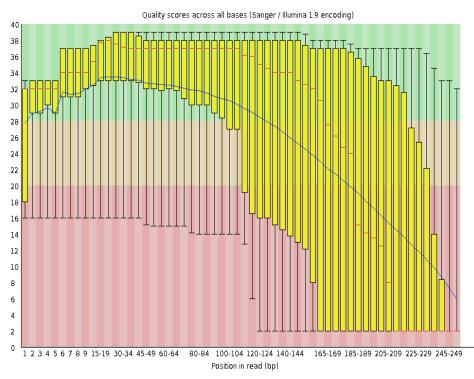
#### **FastQC reports**

#### **Per Base Sequence Quality**

Good quality FastQC report:



#### Bad quality FastQC report







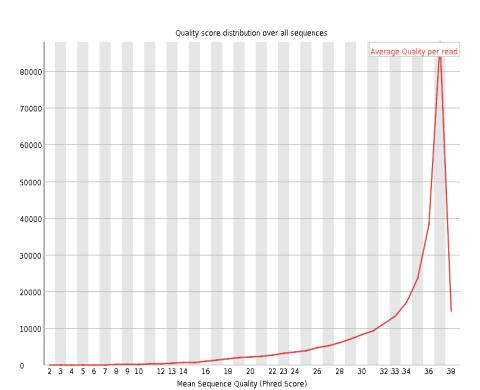




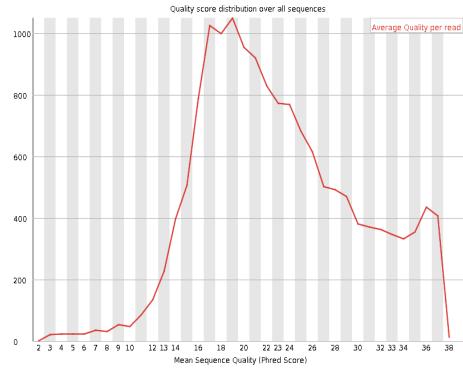
#### **FastQC reports**

#### **Per Sequence Quality Scores**

Good quality FastQC report:



#### Bad quality FastQC report







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%G

#### **FastQC reports**

#### **Per Base Sequence Content**

