

Rosetta Basics: IO and Navigating Rosetta

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Goals for this Talk



1. General Rosetta Concepts:
 - How do I run basic Rosetta applications?
 - Input/Output: file types, options, etc.

2. Learn where things are in Rosetta
 - Your working directory is independent of these Rosetta directories (AKA your data is stored outside of Rosetta)

QUESTIONS ARE ENCOURAGED!

Files for this talk are in

~/rosetta_workshop/tutorials/short_talks/RosettaIO/



Notice these files are located outside of Rosetta
you do NOT want to store your input/output files in
the Rosetta directories

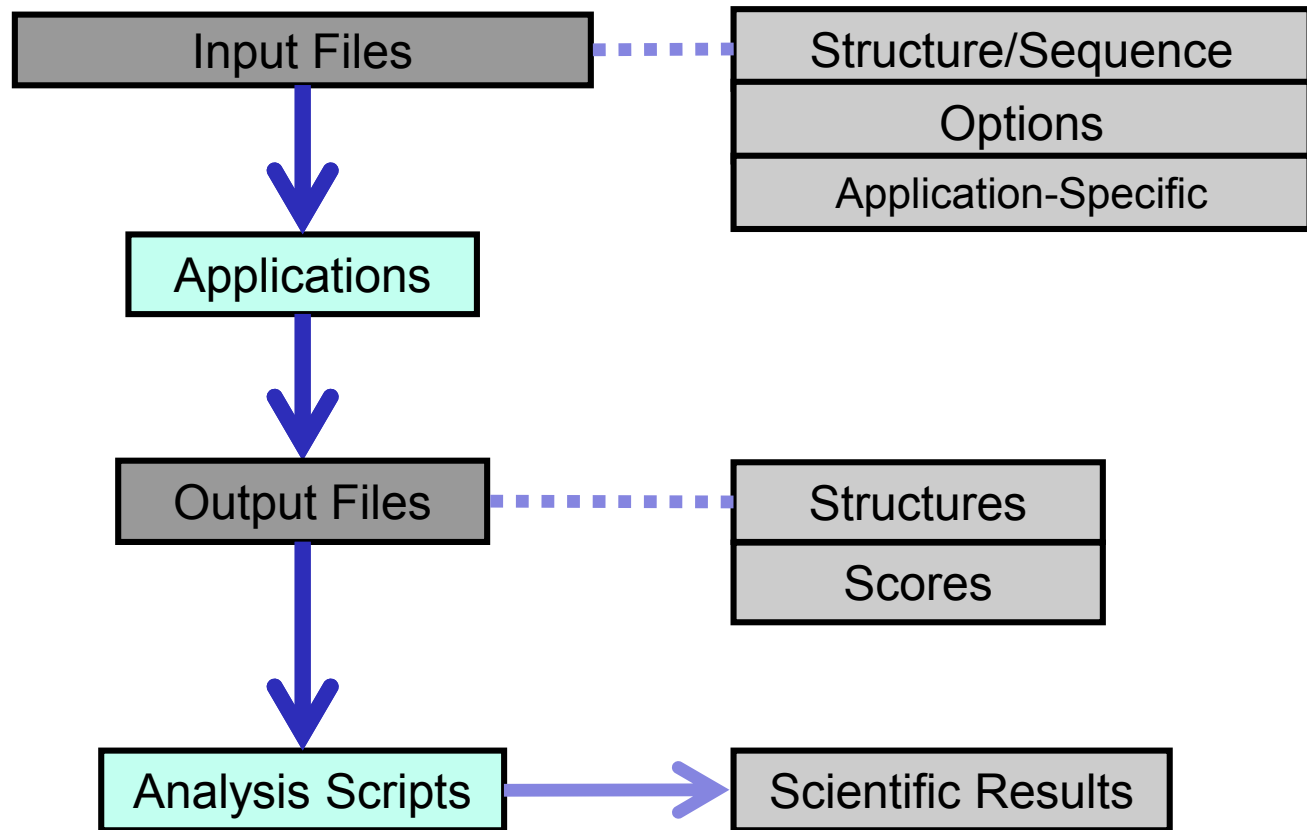
Tip: Open each file in your favorite text editor (gedit, vi,
emacs, etc.) as we introduce them through the talk

How do I get Rosetta?



- <https://www.rosettacommons.org/software/license-and-download>
- Weekly Releases: (e.g. “2019.07”)
 - Latest version of the code, released roughly every week
 - Every revision passes scientific performance tests
- Numbered Releases (e.g. “3.11”)
 - A weekly release that’s relabeled, released roughly every 6 months
- All tutorials use version 3.11
- Links to documentation, forum and demos:
 - <https://www.rosettacommons.org/docs/latest/Home>
 - <https://www.rosettacommons.org/demos/latest/Home>

General Workflow



Reading structures into Rosetta



PDB files

- International standard
- Readable by PyMol, MOE, Chimera, etc
- One line per atom
- Useful for small number of structures

www.wwpdb.org/documentation/file-format

Silent files

- Specific to Rosetta
- Compact
- One line per residue
- Useful for archiving many structures
- Binary silent files: more compact, but not human-readable

https://www.rosettacommons.org/docs/latest/rosetta_basics/file_types/silent-file



How do I run a Rosetta command?

Every command has the same basic layout:

Path to the application

`<path_to_rosetta>/main/source/bin/<app_name>.default.linuxgccrelease`

Arguments

`–arg1 –arg2`

Example for you first Rosetta command:

```
<path_to_rosetta>/main/source/bin/score_jd2.default.linuxgccrelease –in:file:s \
input_files/test1.pdb –out:file:scorefile output_files/test1.sc –out:pdb \
output_files/test1_out.pdb
```

****Note:** the “\” represents a continuous line



Common command line arguments

- Input options:
 - `-parser:protocol example.xml ##` RosettaScripts XML file
 - `-in:file:s example_in.pdb ##` input a PDB structure file
 - `-in:file:fasta example.fasta ##` input a FASTA sequence file
 - `-in:file:silent example_in.silent ##` input a Rosetta silent file
 - `-in:file:extra_res_fa example.params ##` input a parameters file for uncommon compounds
 - `-nstruct 42 ##` produce 42 outputs
- Output options:
 - `-out:file:silent example_out.silent ##` output structures to silent file
 - `-out:file:scorefile example_out.sc ##` output scorefile for run

Scores can be found in output scorefiles and PDBs



output_files/test1.sc

```
SEQUENCE:|
SCORE:      score      fa_atr      fa_rep      fa_sol      fa_intra_rep      fa_elec      ...      omega      fa_dun      p_aa_pp      ref      description
SCORE: -1217.209 -2778.696 266.309 1545.149 5.900 -301.320 ... 63.032 684.989 -109.110 -32.534 3gbm_HA_3gbn_Ab_full_0011
SCORE: -1217.028 -2792.422 263.906 1549.738 5.867 -295.799 ... 66.036 682.694 -108.402 -32.534 3gbm_HA_3gbn_Ab_full_0012
SCORE: -1204.280 -2760.354 259.175 1534.072 5.913 -293.050 ... 65.391 674.840 -108.393 -32.534 3gbm_HA_3gbn_Ab_full_0013
SCORE: -1207.127 -2768.191 260.443 1541.857 5.881 -301.847 ... 67.951 686.381 -110.919 -32.534 3gbm_HA_3gbn_Ab_full_0014
SCORE: -1208.390 -2769.872 262.398 1539.668 5.879 -297.571 ... 64.073 681.731 -109.633 -32.534 3gbm_HA_3gbn_Ab_full_0015
-----
```

Bottom of output_files/test1.pdb

```
ATOM 3378 HB THR L 227 -36.166 22.580 28.848 1.00 0.00 H
ATOM 3379 HG1 THR L 227 -34.994 19.987 29.136 1.00 0.00 H
ATOM 3380 1HG2 THR L 227 -34.138 22.579 30.246 1.00 0.00 H
ATOM 3381 2HG2 THR L 227 -35.593 22.831 31.238 1.00 0.00 H
ATOM 3382 3HG2 THR L 227 -34.799 21.240 31.213 1.00 0.00 H
TER
# All scores below are weighted scores, not raw scores.
#BEGIN_POSE_ENERGIES_TABLE 3gbn_Ab_0005.pdb
label fa_atr fa_rep fa_sol fa_intra_rep fa_elec pro_close hbond_sr_bb hbond_lr_bb hbond_bb_sc hbond_sc dslf_fa13 rama omega fa_dun p_aa_pp yhh_planarity ref total
weights 1 0.55 0.9375 0.005 0.875 1.25 1.17 1.17 1.17 1.1 1.25 0.25 0.625 0.7 0.4 0.625 1 NA
pose -994.338 137.719 561.027 2.15688 -112.612 19.7634 -12.2069 -79.5391 -24.1449 -23.4441 -1.15166 -7.47192 71.8572 276.633 -29.8673 0.09431 13.9828 -201.541
GLU:NtermProteinFull_1 -1.58225 0.53996 1.45283 0.00353 0.06909 0 0 0 0 0 0 0.01109 6.53174 0 0 -1.96094 5.06505
VAL_2 -3.34255 0.45648 1.46378 0.01322 -0.08167 0 0 0 0 0 -0.16095 0.87346 0.30715 0.39992 0 0.97964 0.90848
GLN_3 -2.79445 0.10936 1.74929 0.00451 -0.52743 0 0 0 -0.35772 0 0 -0.09682 0.35321 2.59775 0.02034 0 -1.51717 -0.45911
LEU_4 -5.13483 0.73792 1.6574 0.00685 -0.16379 0 0 0 0 0 0.06265 0.2281 2.29891 -0.1217 0 0.76113 0.33264
VAL_5 -2.72905 0.12167 1.72074 0.00789 -0.45069 0 0 0 0 0 -0.27382 0.01969 0.02557 -0.49649 0 0.97964 -1.07485
```

How do I run a Rosetta command?



Toward high-resolution prediction and design of transmembrane helical protein structures

P. Barth, J. Schonbrun*, and D. Baker†

Department of Biochemistry and Howard Hughes Medical Institute, University of Washington, Seattle, WA 98195

```
$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease -database
../../rosetta_database -in:file:fasta ./input_files/1elwA.fasta -
in:file:native ./input_files/1elw.pdb -in:file:frag3
./input_files/aalelwA03_05.200_v1_3 -in:file:frag9
./input_files/aalelwA09_05.200_v1_3 -abinitio:relax -relax:fast -
abinitio::increase_cycles 10 -abinitio::rg_reweight 0.5 -
abinitio::rsd_wt_helix 0.5 -abinitio::rsd_wt_loop 0.5 -use_filters
true -psipred_ss2 ./input_files/1elwA.psipred_ss2 -kill_hairpins -
out:file:silent 1elwA_silent.out
-nstruct 10
```

OR

```
$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt
```

Use an options file for your runs



\$ROSETTA/main/source/bin/AbinitioRelax.linuxgccrelease @options.txt

```
-in:file
  -fasta ./input_files/1elwA.fasta
  -native ./input_files/1elw.pdb
  -frag3 ./input_files/aa1elwA03_05.200_v1_3
  -frag9 ./input_files/aa1elwA09_05.200_v1_3
-psipred_ss2 ./input_files/1elwA.psipred_ss2
-abinitio:relax
-relax:fast
-abinitio::increase_cycles 10
-abinitio::rg_reweight 0.5
...
-out:file:silent ./output_files/1elwA_silent.out
-nstruct 10
```

Other files: application-specific



- Span File: Defines which residues are in the membrane
- Loops File: Identifies the loop residues for loop closure
- Params File: Custom parameters for small molecules or non-canonical amino acids
- Constraint File: Experimentally derived restraints
- Fragment File: Short protein segments used for comparative modeling and de novo folding
- Res Files: Indicates which residue positions should be designed

Tracer output (log files)



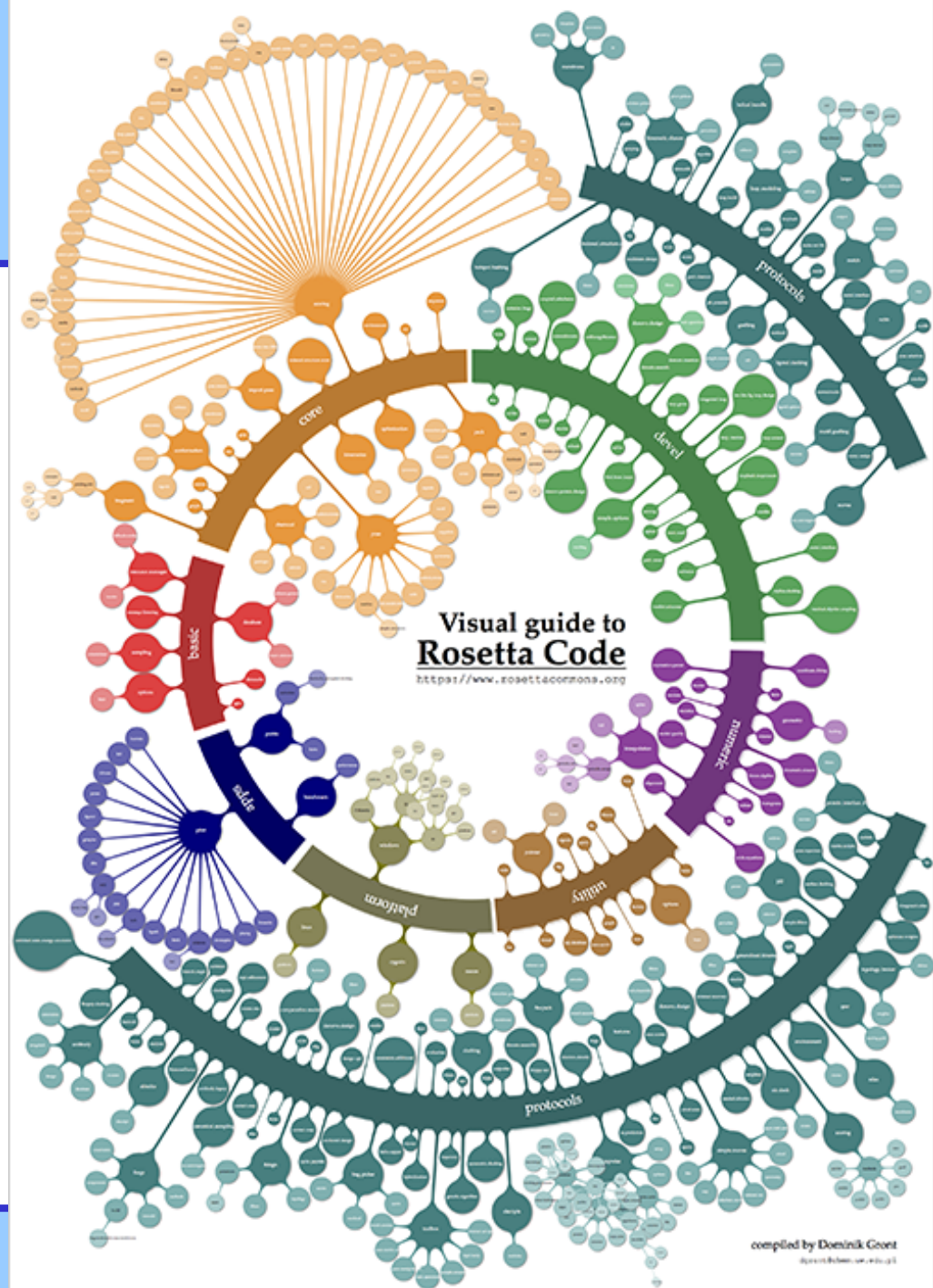
```
core.init: Rosetta version exported from http://www.rosettacommons.org
core.init: command: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/bin/rosetta_scripts.default.linuxgccrelease @docking.options -parser:protocol docking
core.init: 'RNG device' seed mode, using '/dev/urandom', seed=1059677151 seed_offset=0 real_seed=1059677151
core.init.random: RandomGenerator:init: Normal mode, seed=1059677151 RG_type=mt19937
core.init: Resolved executable path: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/source/build/src/release/linux/2.6/64/x86/gcc/5.2/default/rosetta_scripts.d
core.init: Looking for database based on location of executable: /dors/meilerlab/apps/rosetta/rosetta_2016.08.58479/main/database/
protocols.jd2.PDBJobInputter: Instantiate PDBJobInputter
protocols.jd2.PDBJobInputter: PDBJobInputter::fill_jobs
protocols.jd2.PDBJobInputter: pushed 3gbm_HA_3gbn_Ab.pdb nstruct indices 1 - 50
protocols.evaluation.ChiWellRmsdEvaluatorCreator: Evaluation Creator active ...
protocols.jd2.JobDistributor: Parser is present. Input mover will be overwritten with whatever the parser creates.
protocols.jd2.PDBJobInputter: PDBJobInputter::pose_from_job
protocols.jd2.PDBJobInputter: filling pose from PDB 3gbm_HA_3gbn_Ab.pdb
core.chemical.ResidueTypeSet: Finished initializing fa_standard residue type set. Created 384 residue types
core.chemical.ResidueTypeSet: Total time to initialize 0.43 seconds.
core.conformation.Conformation: Found disulfide between residues 7 461
...
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====End report for =====
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====Begin report for =====
protocols.rosetta_scripts.ParsedProtocol.REPORT: =====End report for =====
protocols.jd2.JobDistributor: 3gbm_HA_3gbn_Ab_full_0050 reported success in 381 seconds
protocols.jd2.JobDistributor: no more batches to process...
protocols.jd2.JobDistributor: 50 jobs considered, 50 jobs attempted in 16297 seconds
~
~
~
```

Options to control tracer output:

- Silence certain tracers:
 - mute core.chemical.ResidueTypeSet
- Change verbosity level (Error/Warning/Info/Debug/Trace)
 - out:levels all:Warning core.init:Info



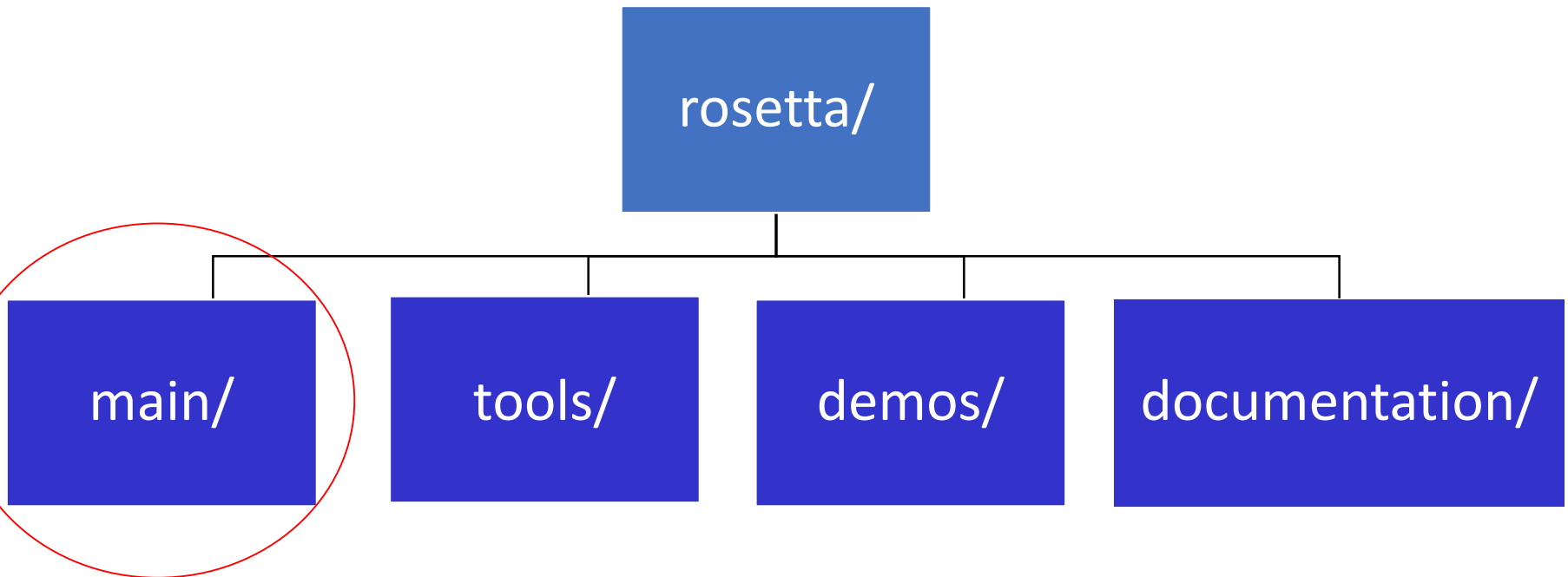
Any Questions?



Basic Rosetta Structure



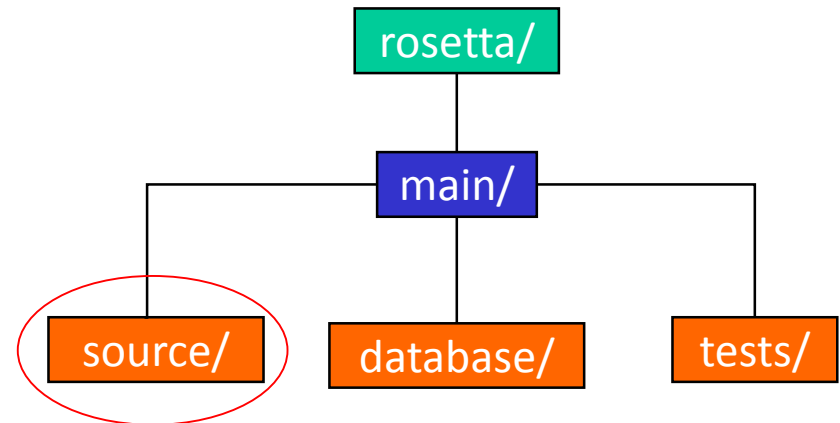
`cd ~/rosetta_workshop/rosetta/`



~/rosetta_workshop/rosetta/main/



- [Rosetta/main/source/bin/](#)
 - Most applications you will run are
 - calling programs within the bin directory
 - rosetta_scripts.default.linuxgccrelease
 - score_jd2.default.linuxgccrelease
 - relax.default.linuxgccrelease
- [Rosetta/main/source/scons.py](#)
 - Used for compiling
- [Rosetta/main/source/src/](#)
 - This is where all of the code lives
- [Rosetta/main/source/scripts/](#)
 - Some useful scripts live here
 - (e.g. params file generation)





~/rosetta_workshop/rosetta/main/database/

Contains pre-defined information that an application needs
— users generally don't change things here.

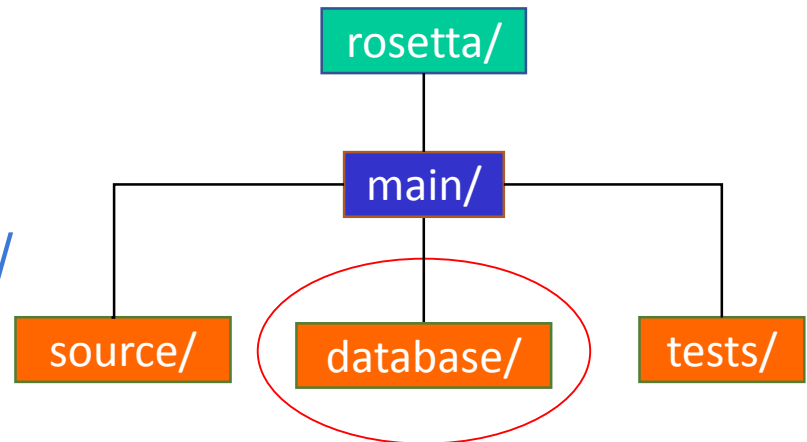
Note: Most of the time, applications know where the database is without having to specify it.

- [Rosetta/main/database/scoring/](#)

- Default weights files
- Rotamer libraries

- [Rosetta/main/database/chemical/](#)

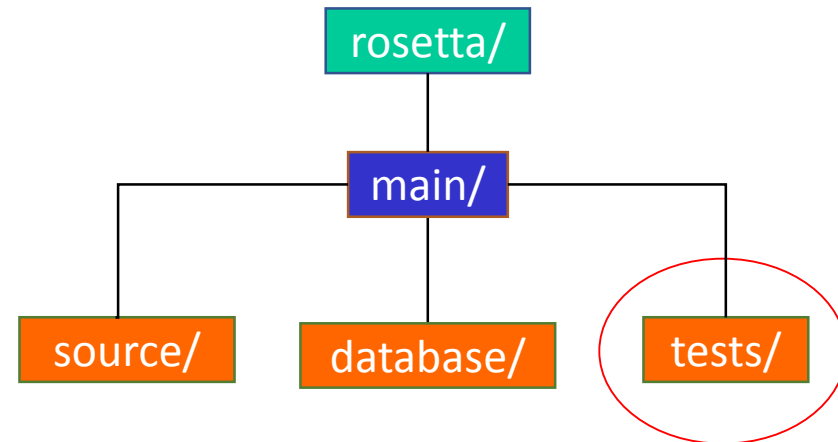
- Residue information--params files
- Atom sets





~/rosetta_workshop/rosetta/main/tests/

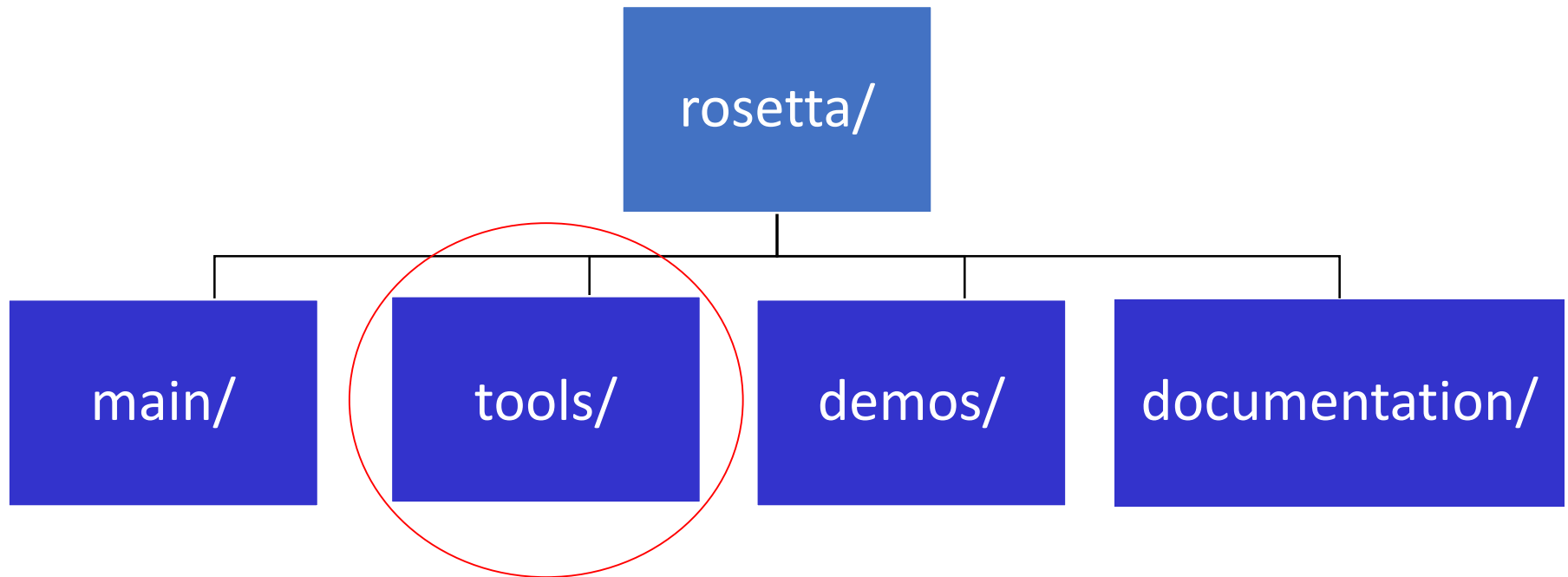
- Rosetta/main/tests/
 - Tests for Rosetta code (useful for developers only)



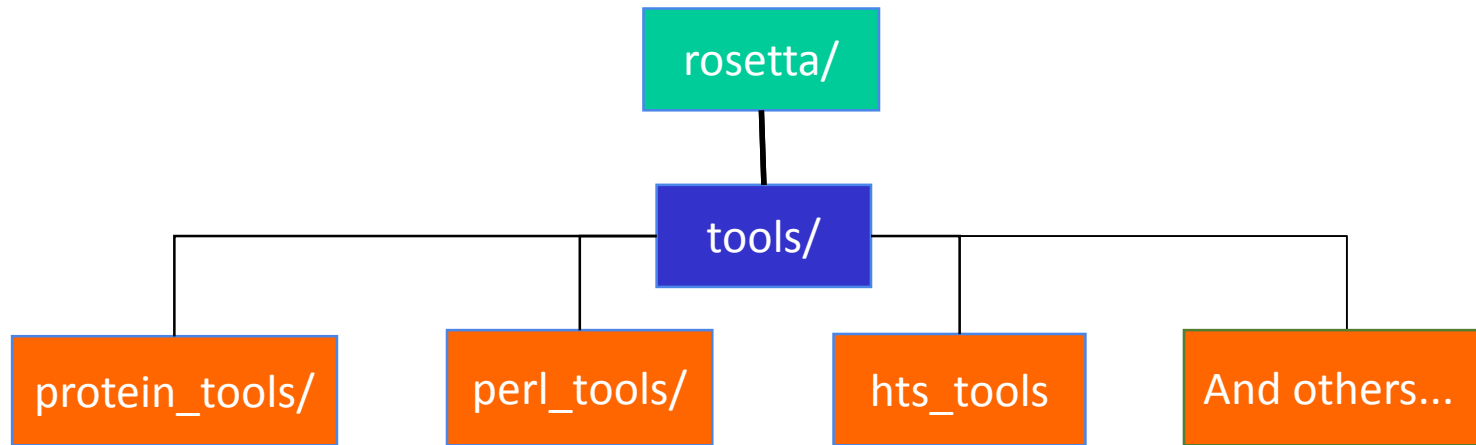
Basic Rosetta Structure



cd ~/rosetta_workshop/rosetta/



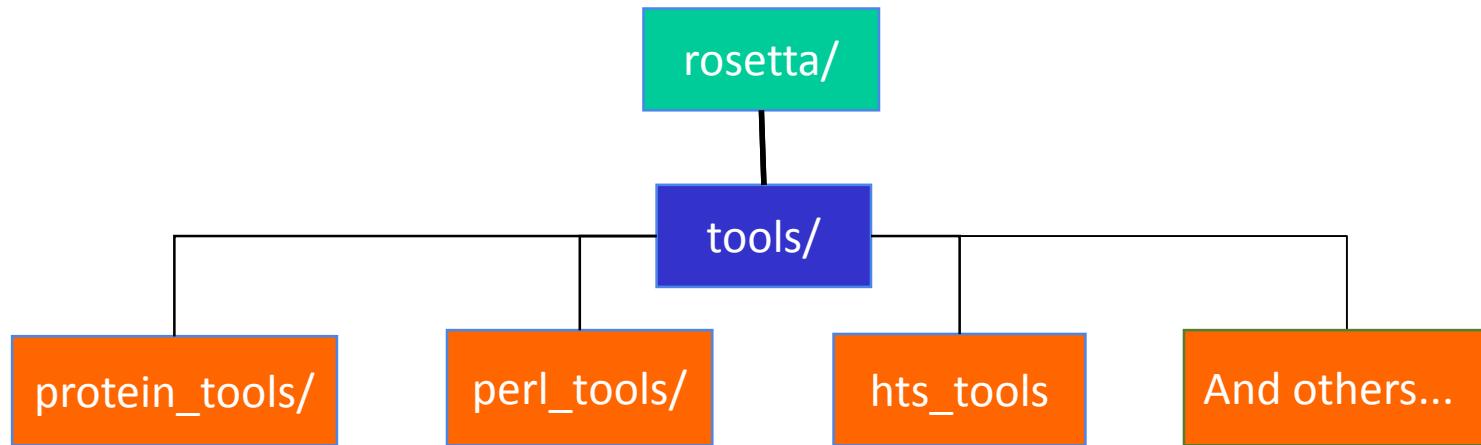
~/rosetta_workshop/rosetta/tools/



- These scripts are incredibly help for smaller, more basic tasks
- Used mainly to setup or analyze runs



~/rosetta_workshop/rosetta/tools/protein_tools/scripts/



Some useful scripts to be aware of:

clean_pdb.py*

- Makes a PDB “Rosetta-proof” and used at the beginning of almost any protocol

pdb_renumber.py

- Renumbering your PDB starting from 1

score_vs_rmsd.py

- Setup for score vs. RMSD plots

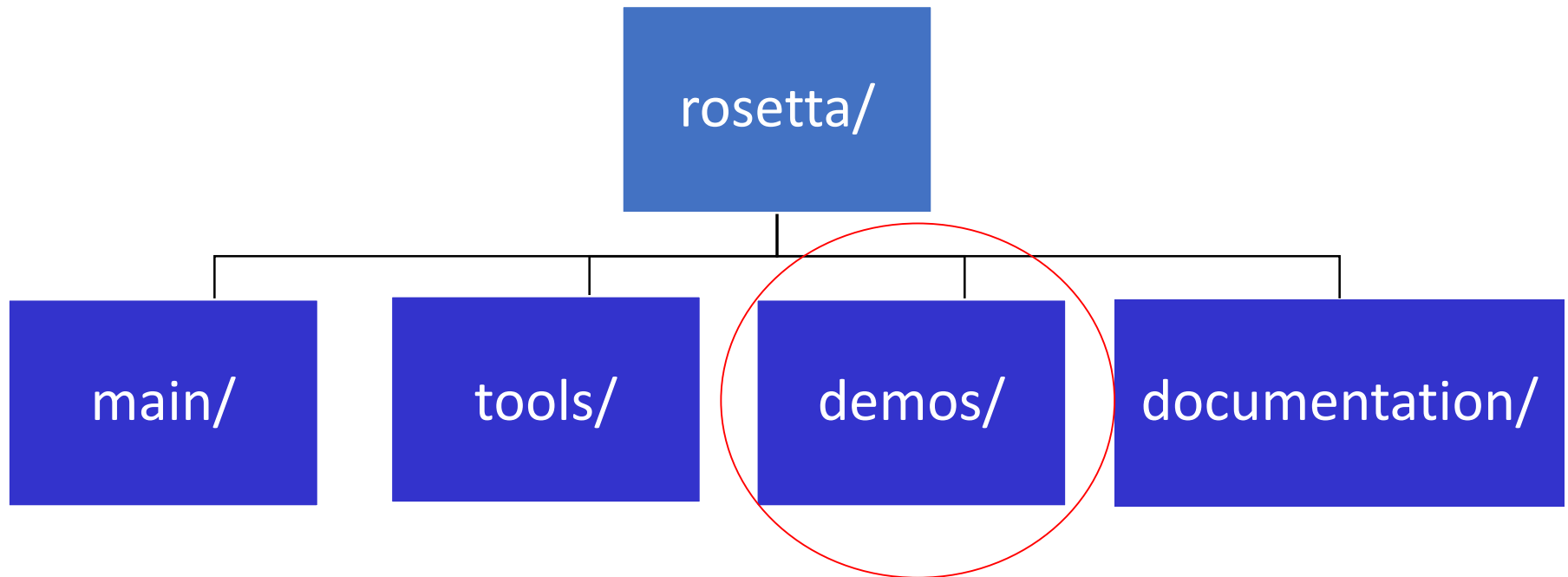
top_n_percent.py

- Extracts tags (protein names) for top given percent of models based on score term

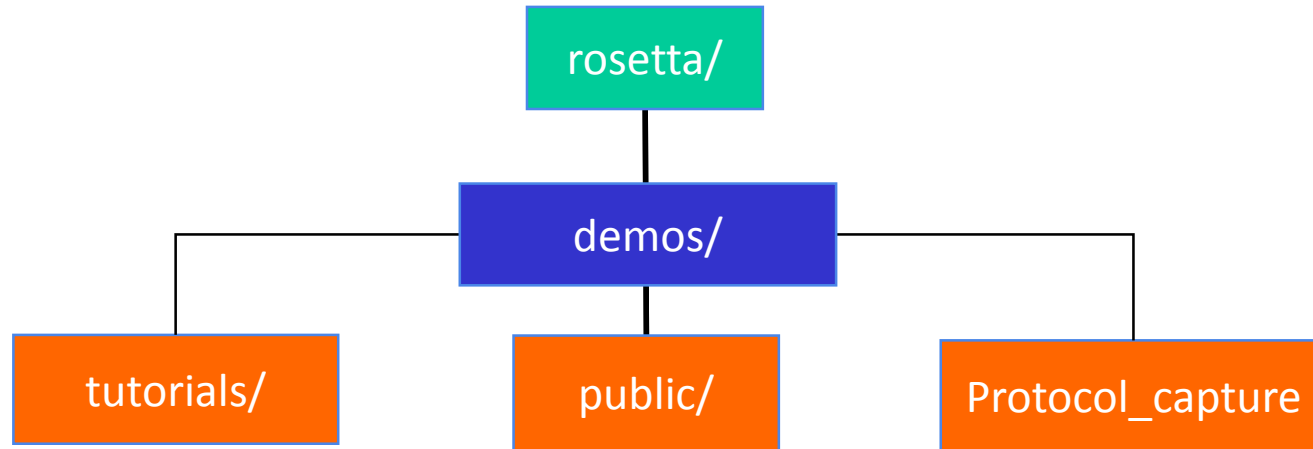
Basic Rosetta Structure



`cd ~/rosetta_workshop/rosetta/`



~/rosetta_workshop/rosetta/demos



[tutorials/](#)

- Past tutorials

[public/](#)

- Protocol examples

[protocol_capture/](#)

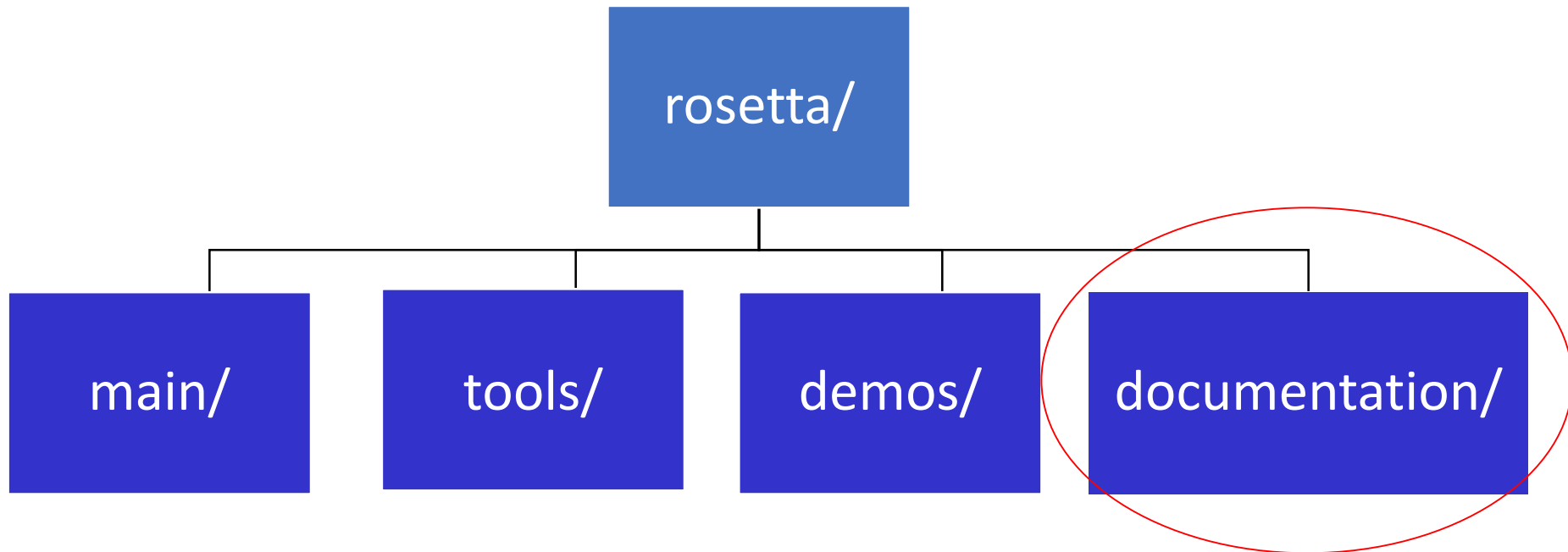
- protocols associated with a publication

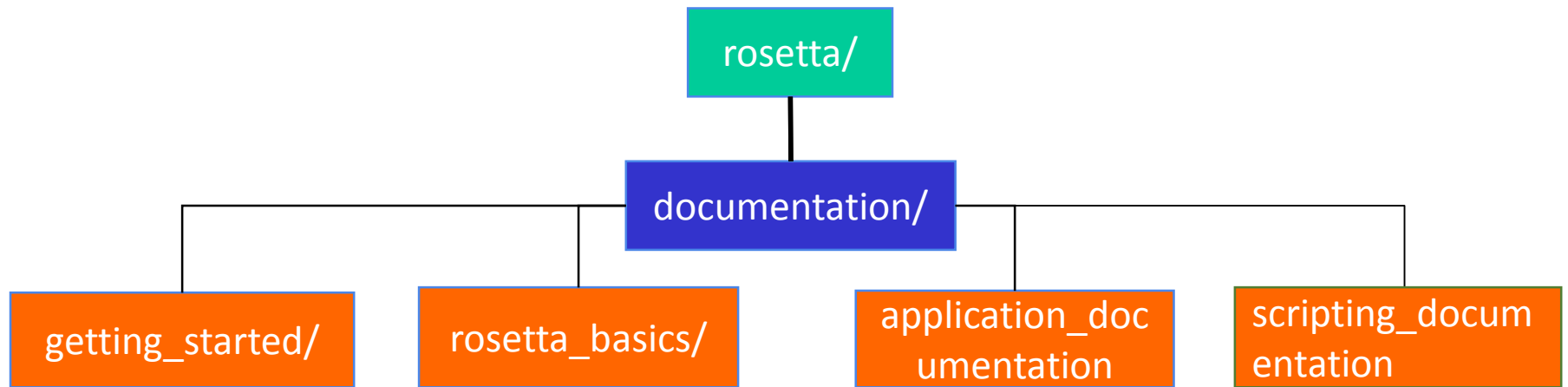
DISCLAIMER: May be out of date, always check Wiki/RosettaCommons/forum for latest information!

Basic Rosetta Structure



`cd ~/rosetta_workshop/rosetta/`





Very useful to go through when you're just getting started in
Rosetta or any structural biology software

Understanding general Rosetta concepts

- Where to find FAQs (How long does this run take?)
- Options list, file types

Protocols you can use

- General structural biology FAQs
 - How do I do X?

<https://www.rosettacommons.org/docs/latest/Home>



Other Rosetta Resources:

- <https://www.rosettacommons.org>
 - Documentation
 - User guides
 - Forum
 - Software Download