

Proteomics

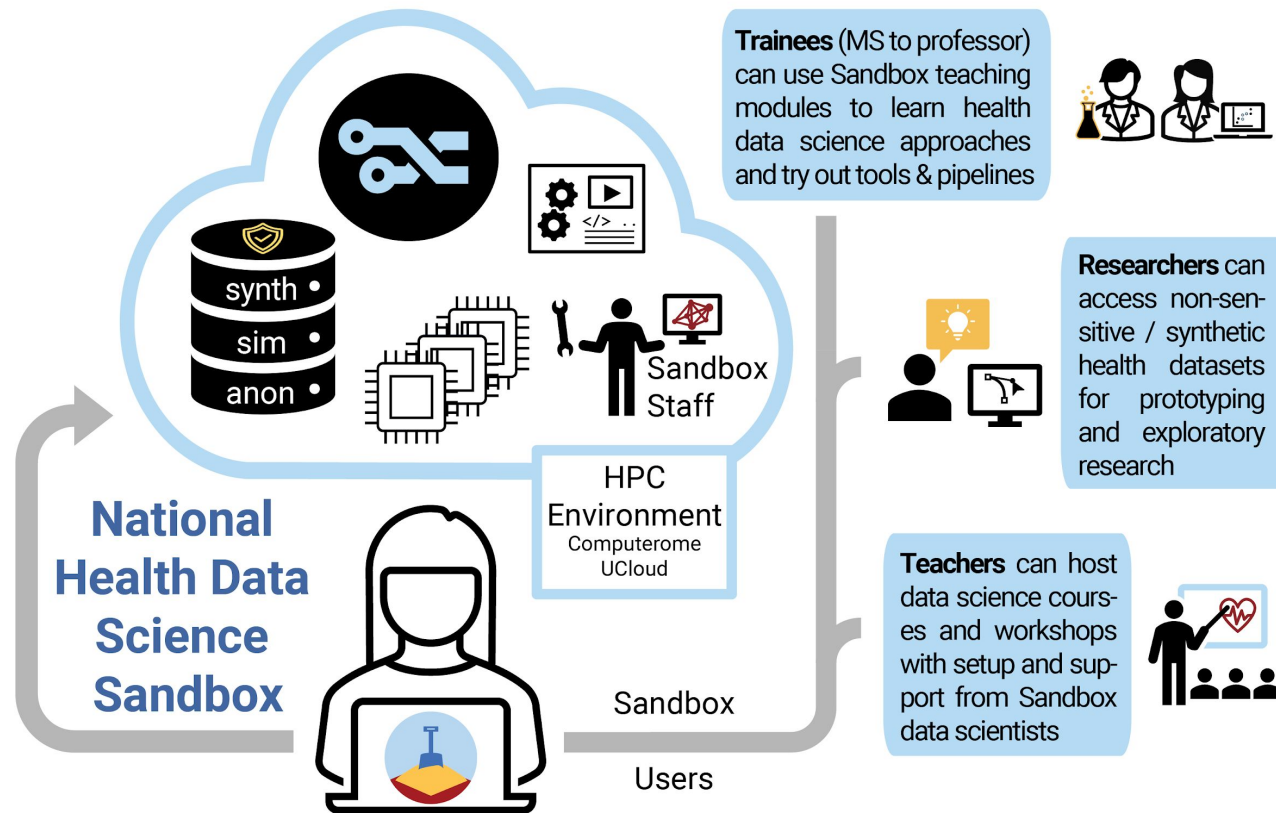
OMICS Workshop 31.august.2023



Samuele Soraggi
Health Data Science Sandbox



Health data science sandbox

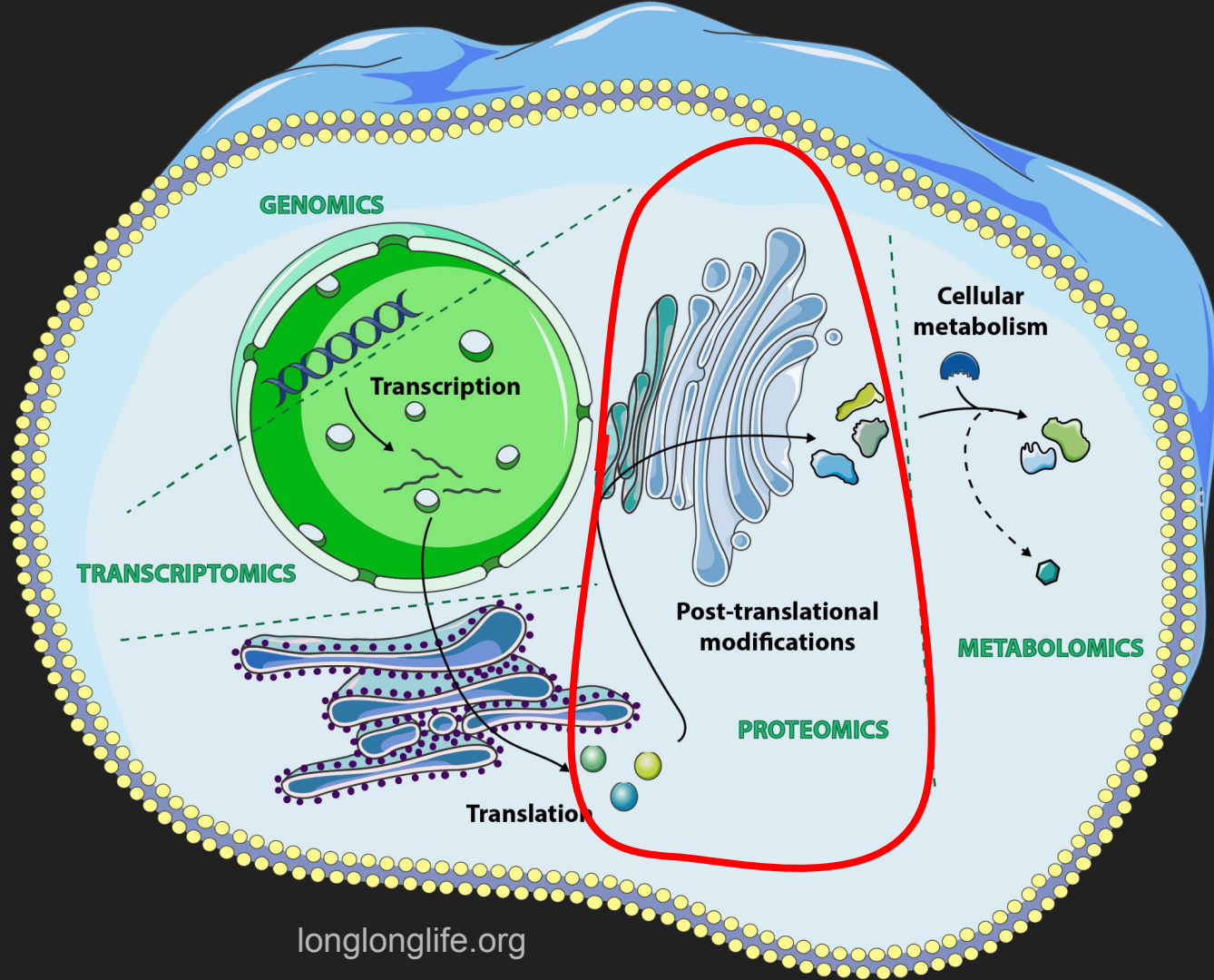


Home Page:

hds-sandbox.github.io

Contact:

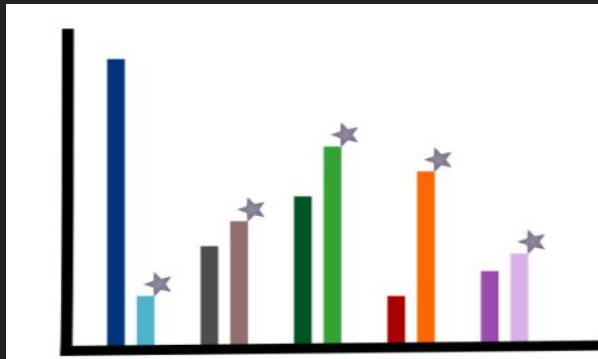
samuele@birc.au.dk



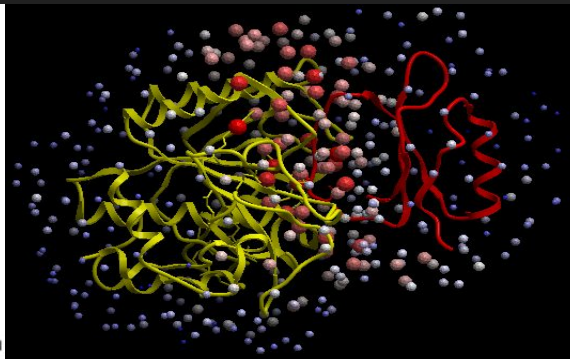
Program

9-9.30	Introduction to proteomics and tutorial format
9.30-9.45	Questions/Small break
9.45 - X	Log into uCloud, start the tutorial
X - X+15	Discussion and questions
X+15 - 13.00	Continue with the tutorial (If X small enough, we use the final time for discussion again)

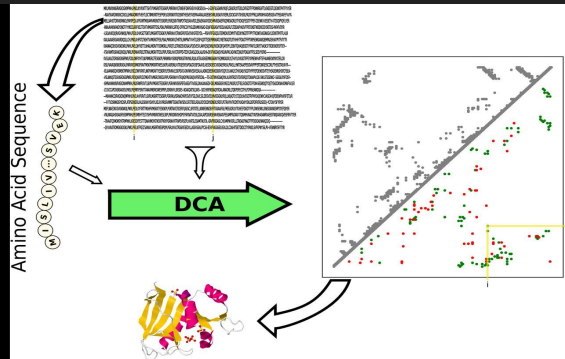
Proteomics - applications



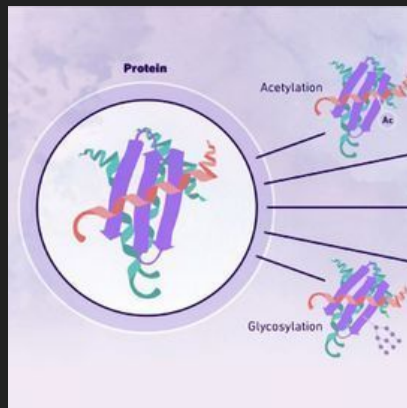
Peptide Identification and quantification



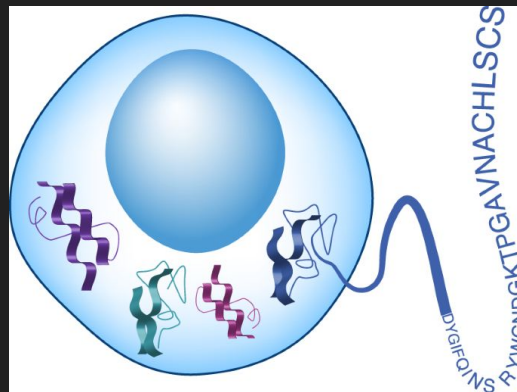
Protein-Protein Interactions



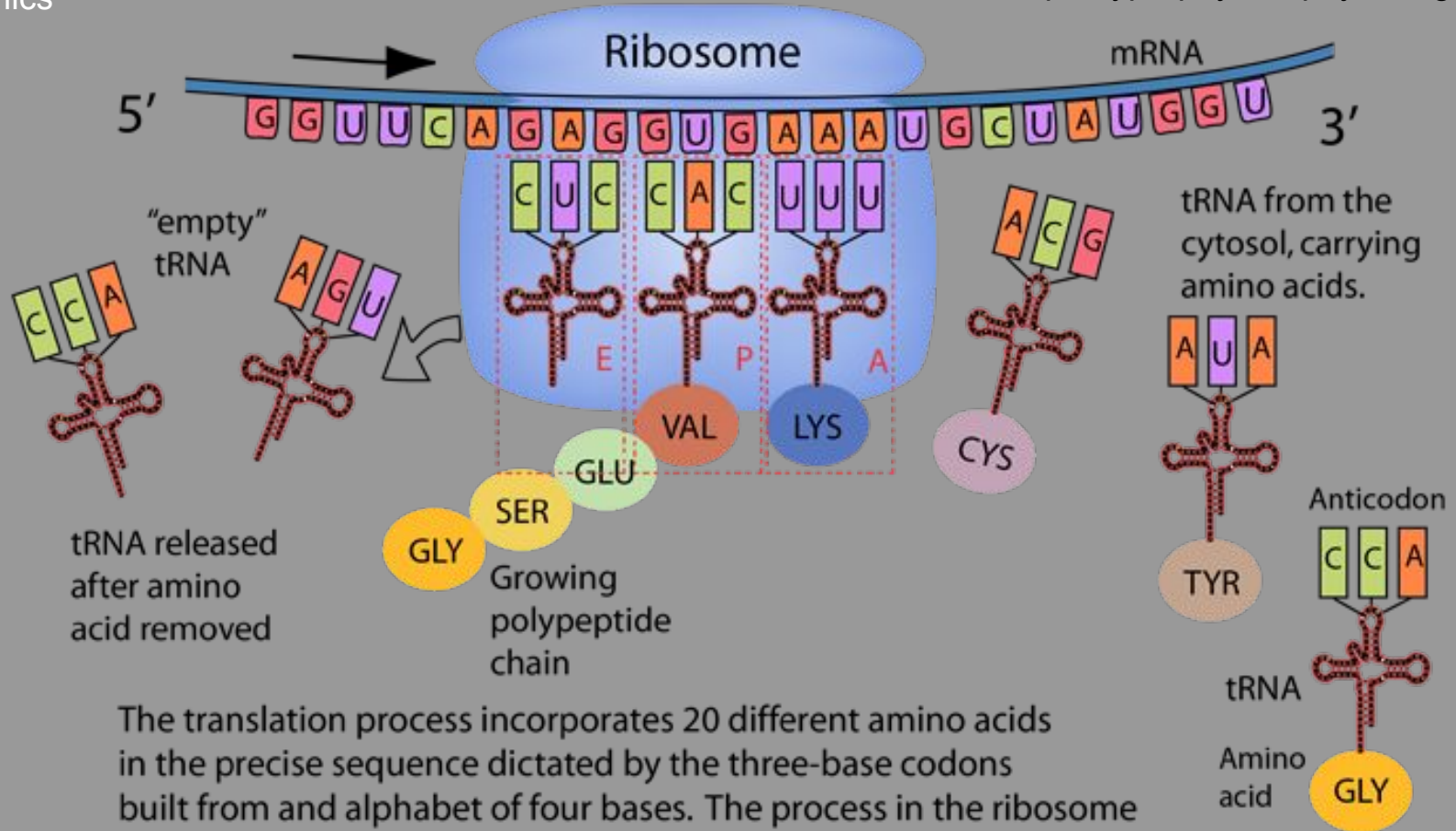
Structural prediction from aminoacids sequences



Post translational modification



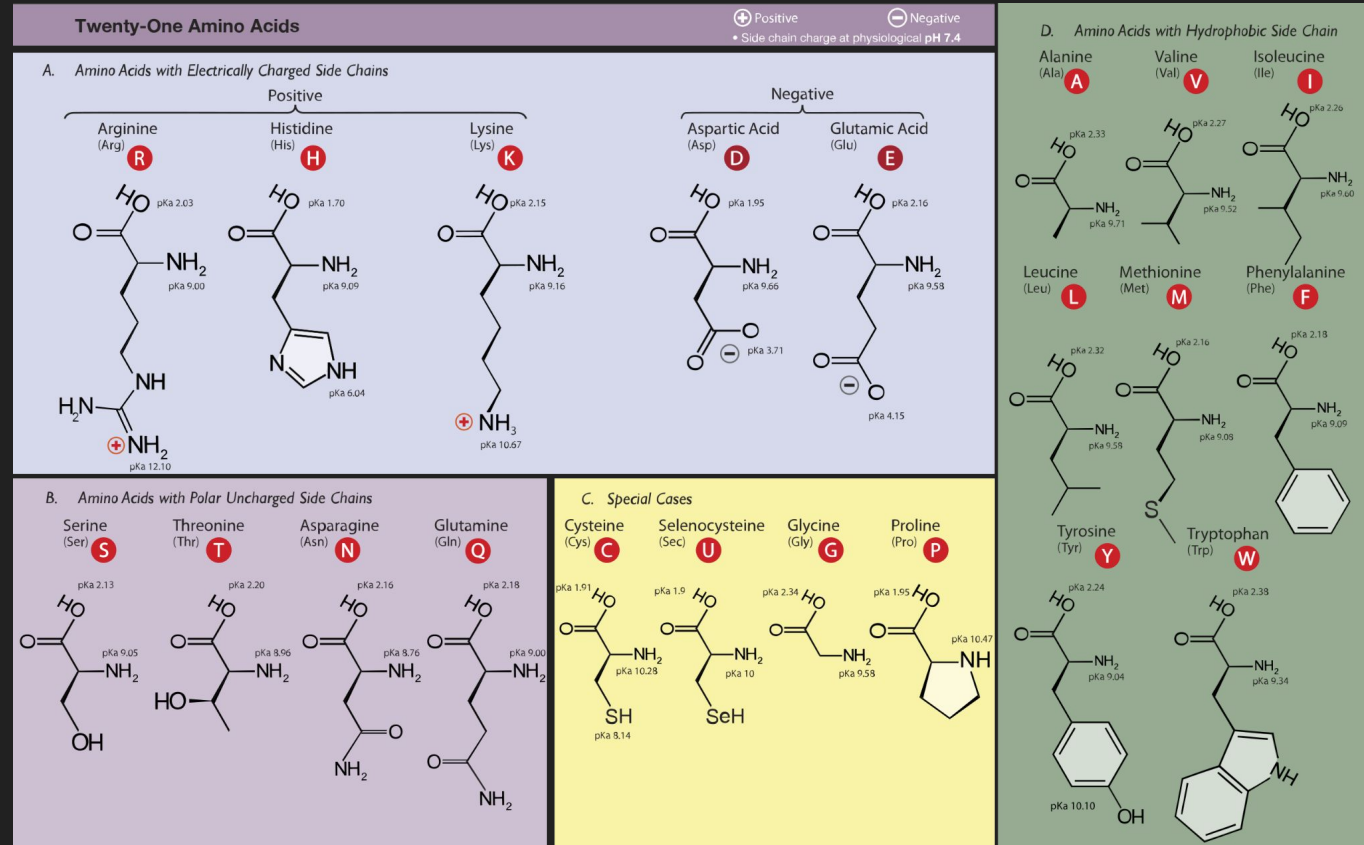
Single cell proteomics



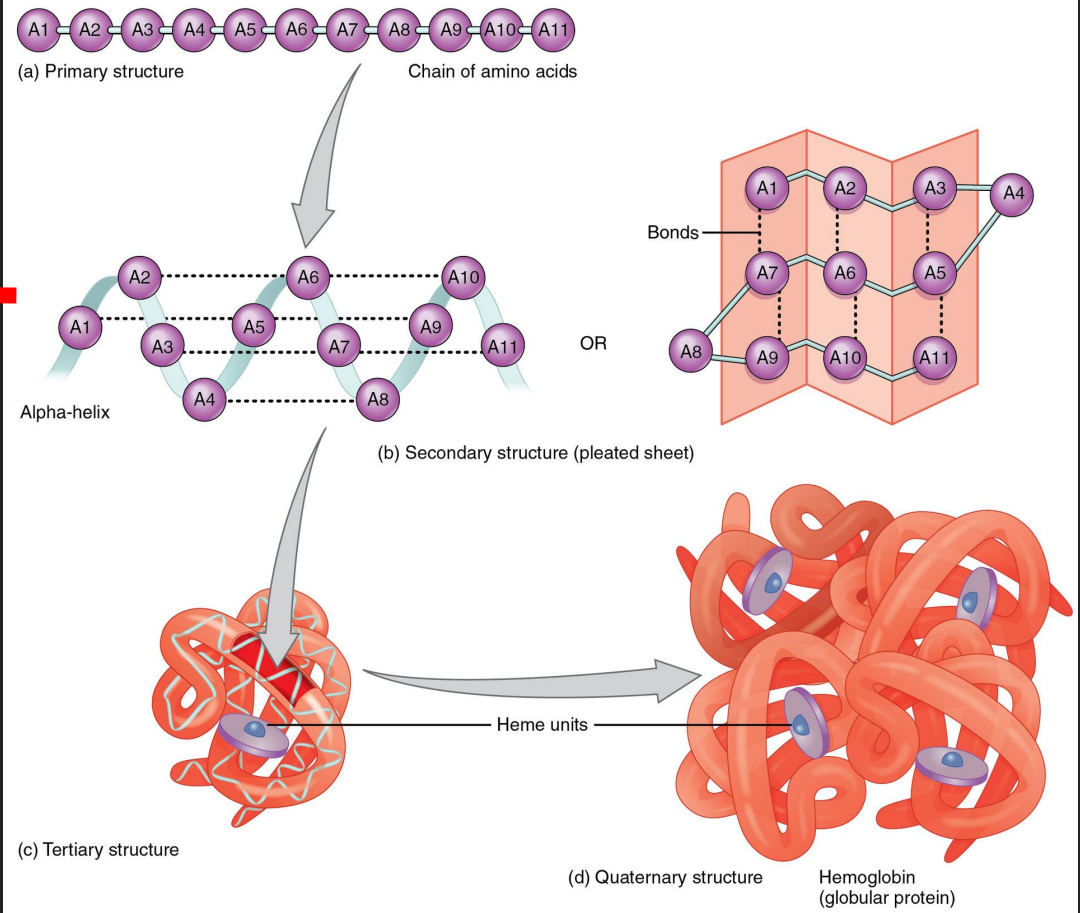
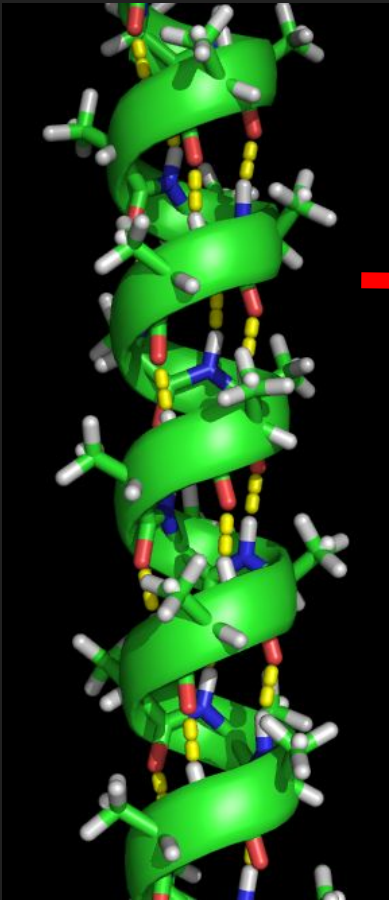
Aminoacids have specific properties such as

- charged
- uncharged
- hydrophobic

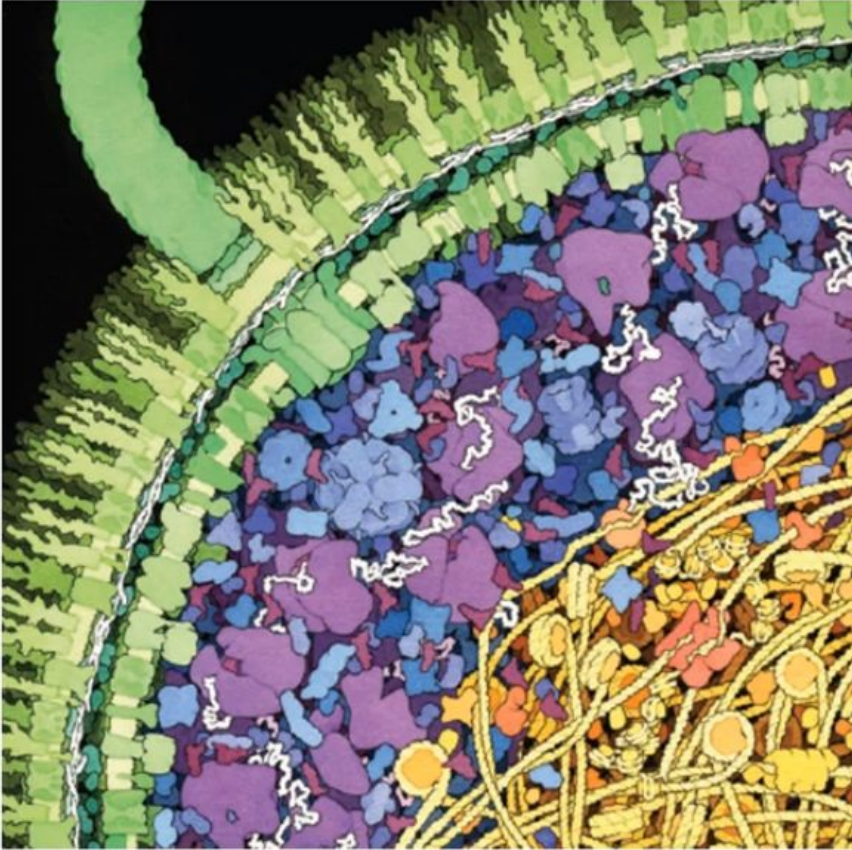
Those can create bonds between peptides



Proteomics - Protein geometry



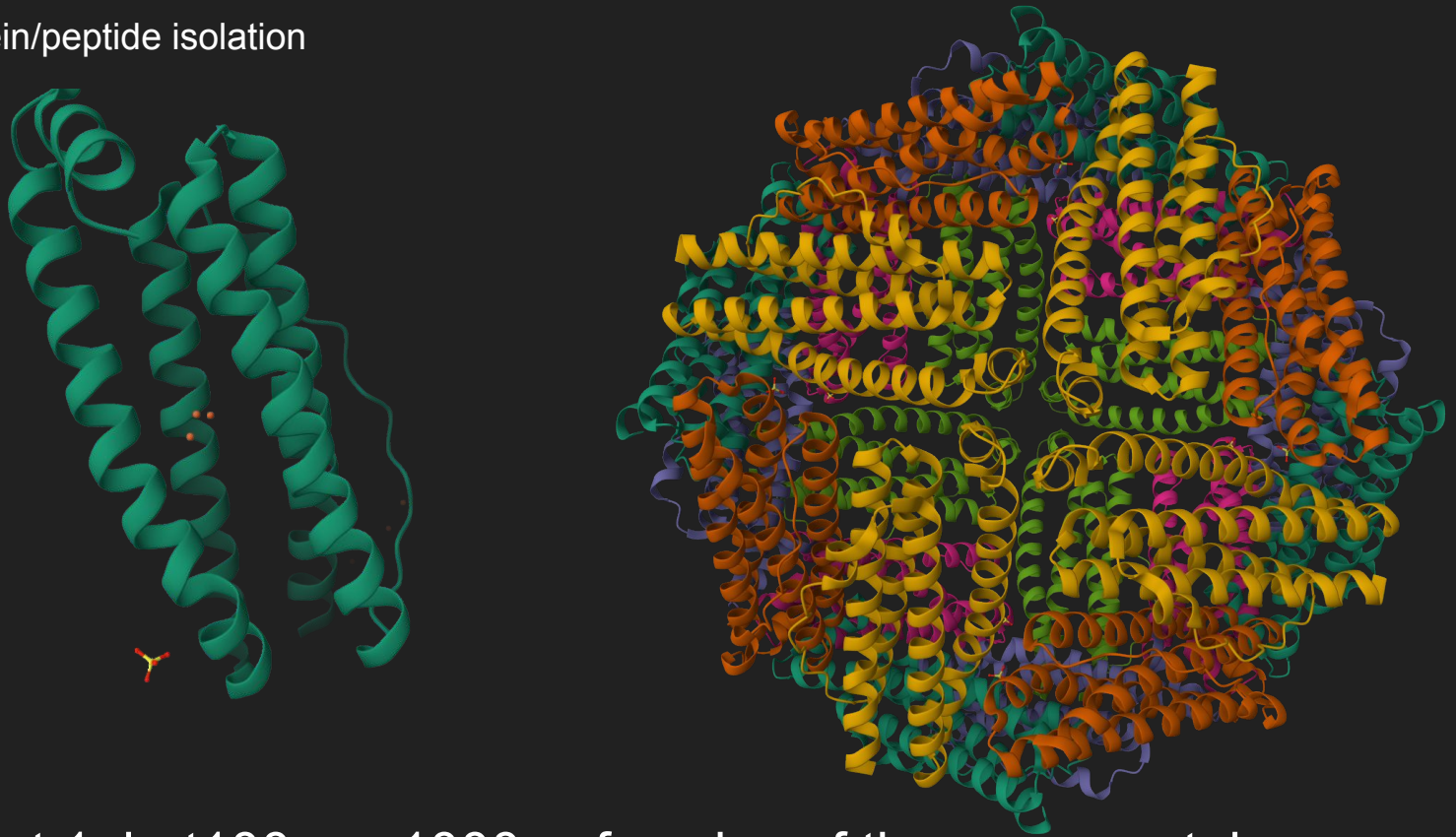
Nat Chem Biol 5:774-7 (2009)



Challenge:

isolate proteins to detect the distinct chemical structures

Note: there is no PCR here



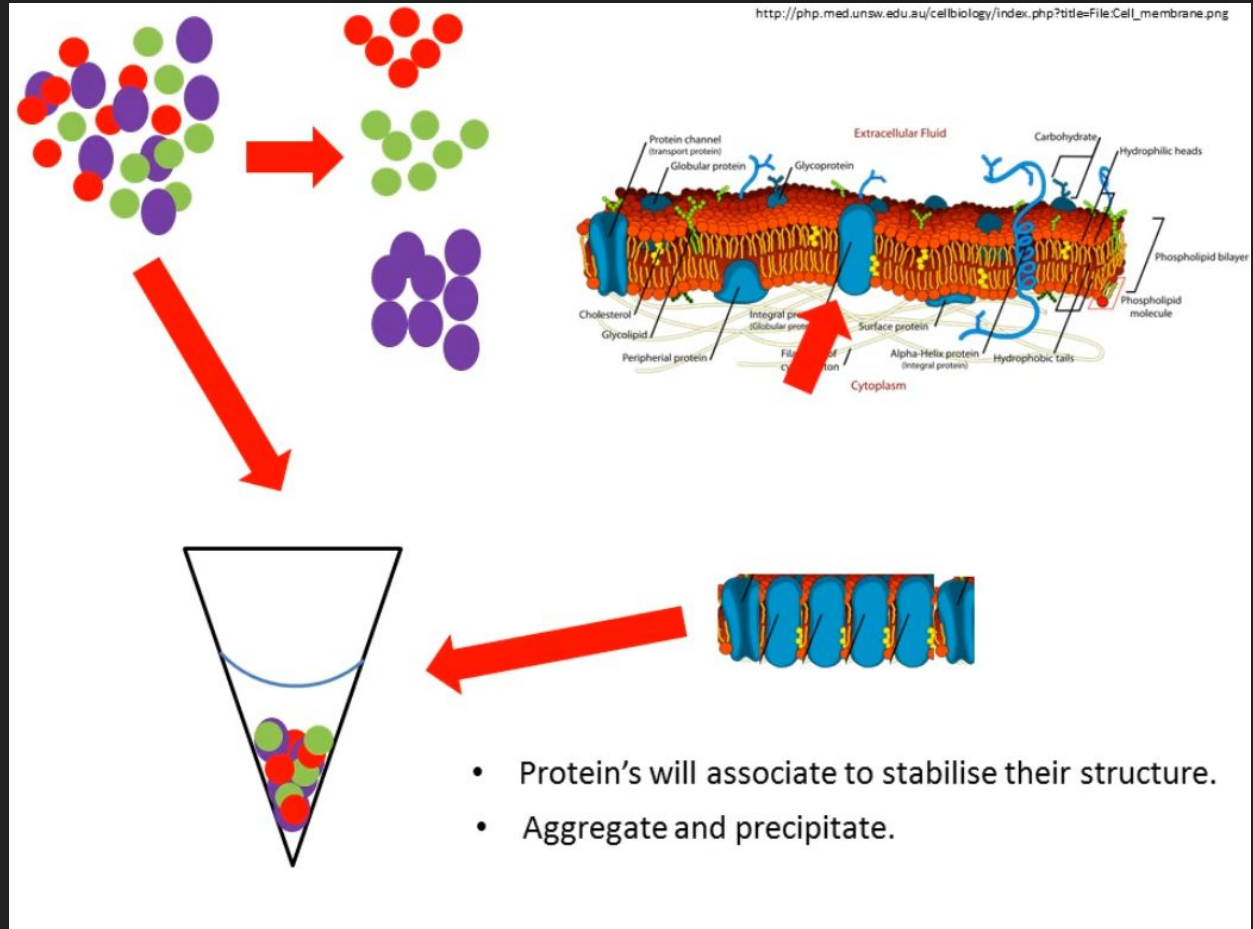
Not 1, but 100s or 1000s of copies of the same protein

A Marchetti *et al.* *Nature* **000**, 1-4 (2008)

doi:10.1038/nature07539

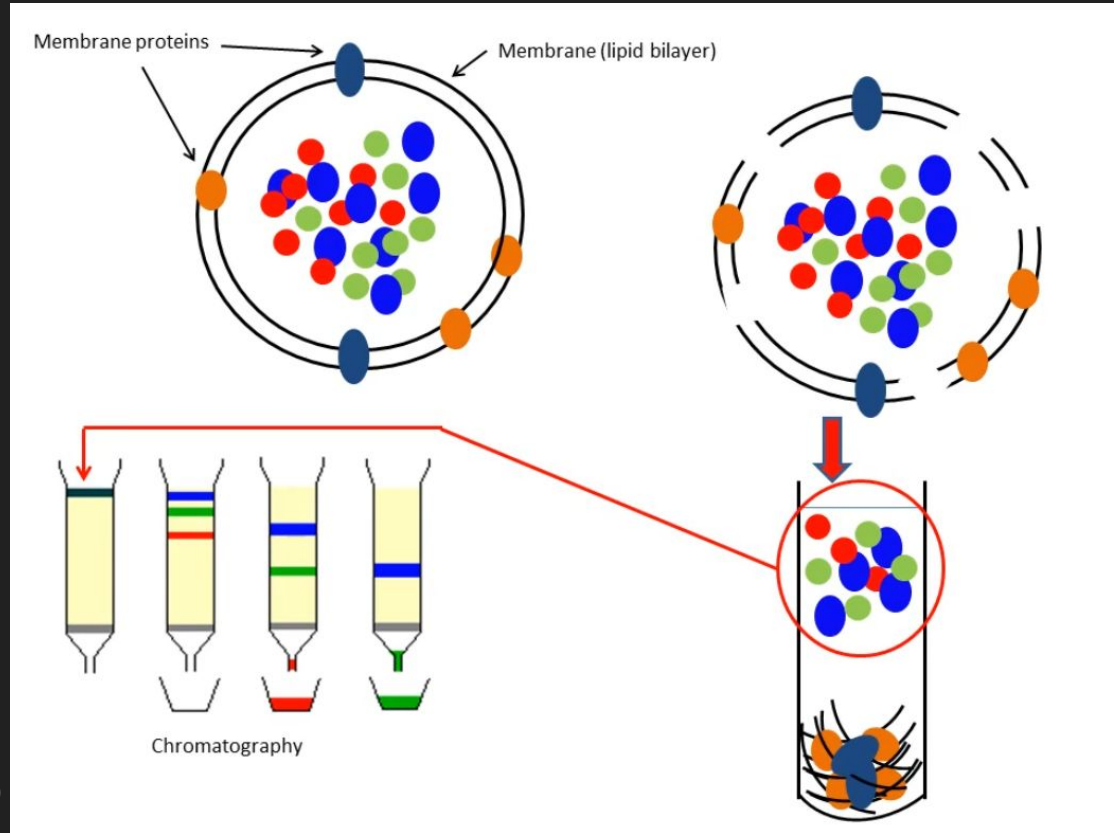
Fluidic isolation

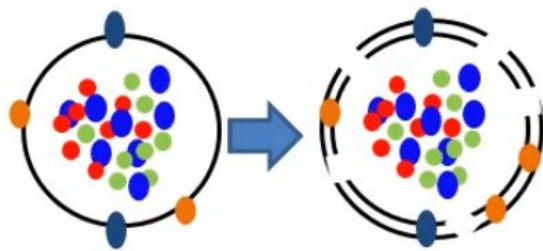
- recovers some peptides
- new associations to stabilize peptides (e.g. hydrophobic bonds)
- heavy bonds will be lost by precipitation



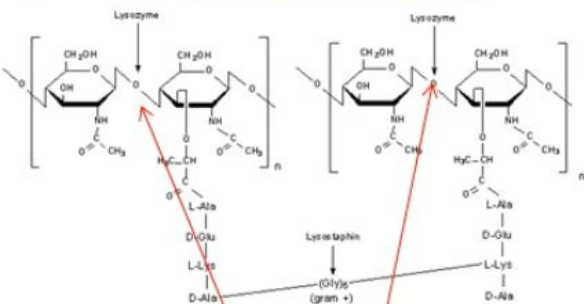
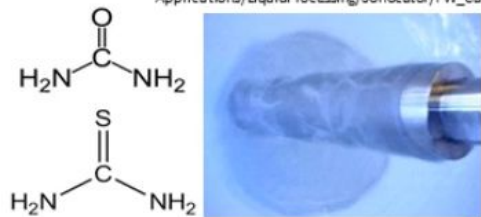
Fluidic isolation

- centrifugation recovers more peptides
- a chromatograph can separate them
- specific liquid solutions reduce the precipitation loss further by avoiding bonds (e.g. hydrophobic)

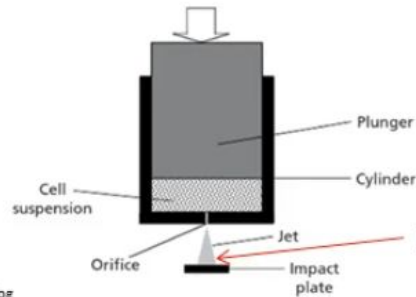




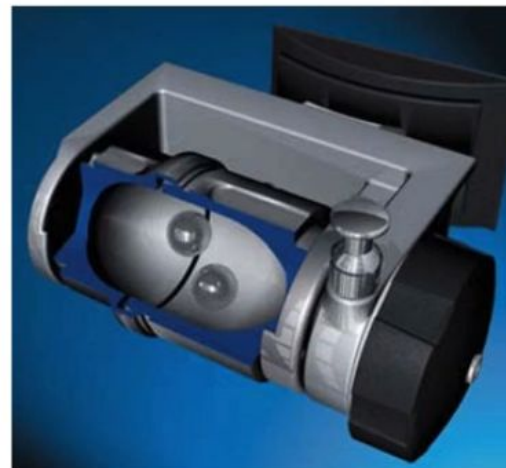
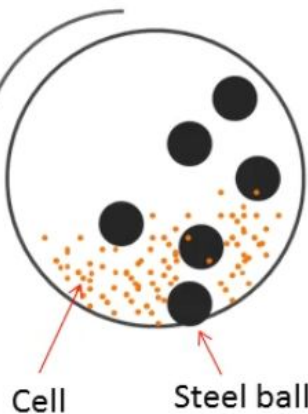
http://www.activeultrasonic.com/applications/files/images/Applications/LiquidProcessing/Sonocator/FW_Cavitation2aw.jpg



Cell wall broken here



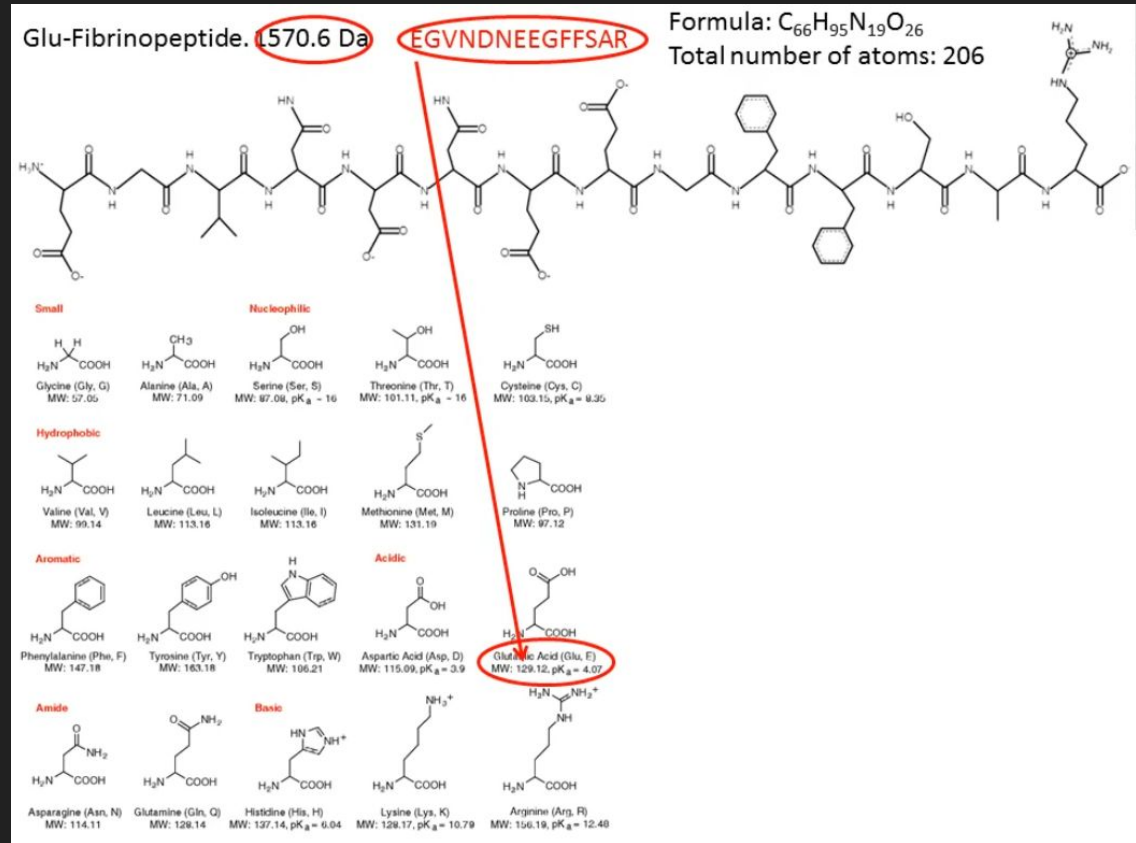
Broken cells here



Mass spectrometry

Mass is important

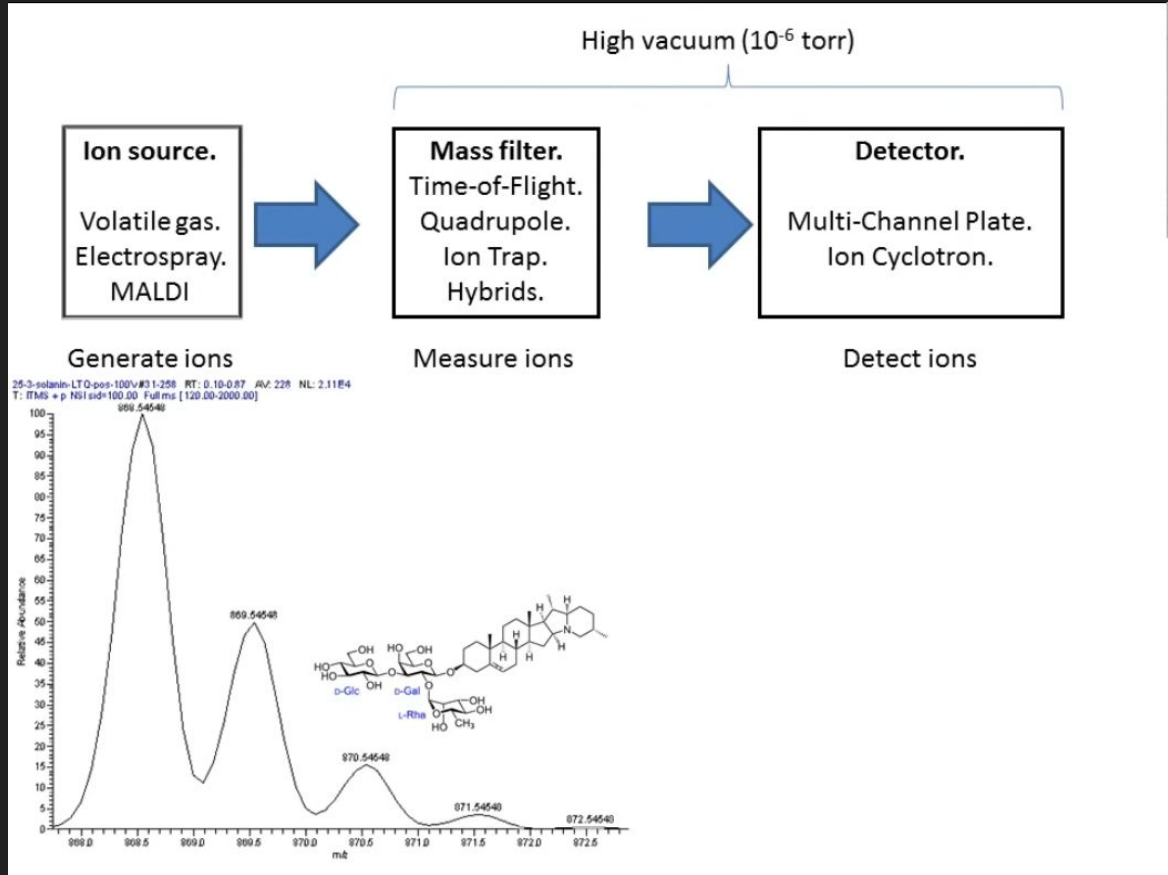
- elements in the periodic table have unique mass
- their compounds also do, including peptides
- We cannot calculate the mass of peptides when we do not know the sequence



Matt Padula

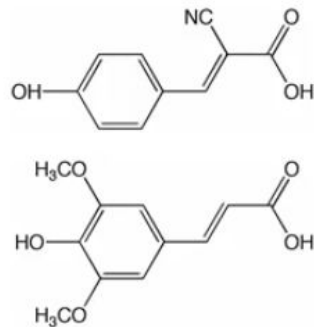
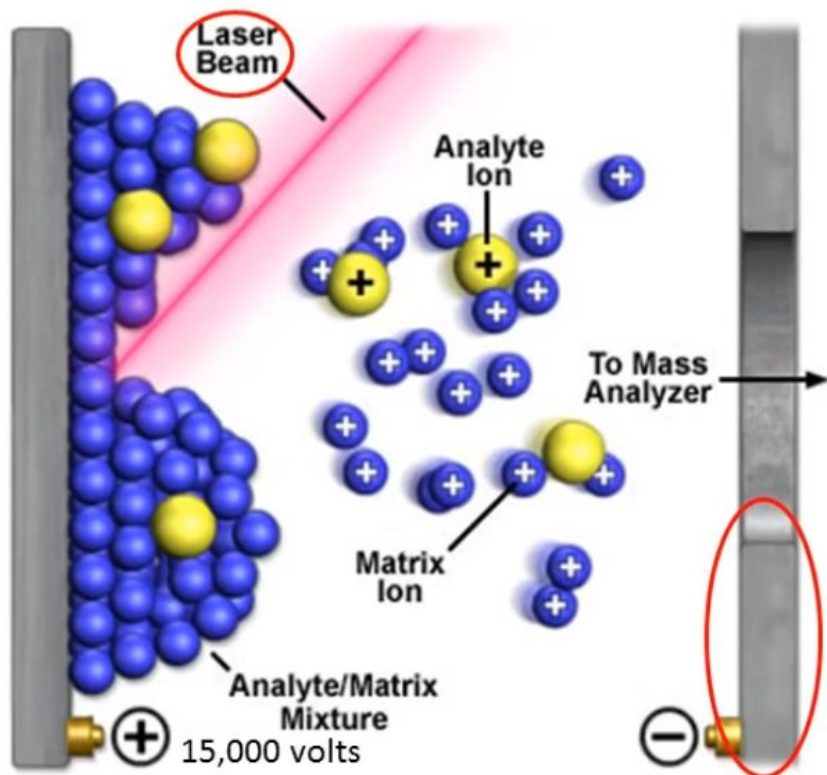
Mass spectrometry

- We charge instead our particles
- We make them go through a dedicated device (e.g. a Time Of Flight device)
- We measure their mass

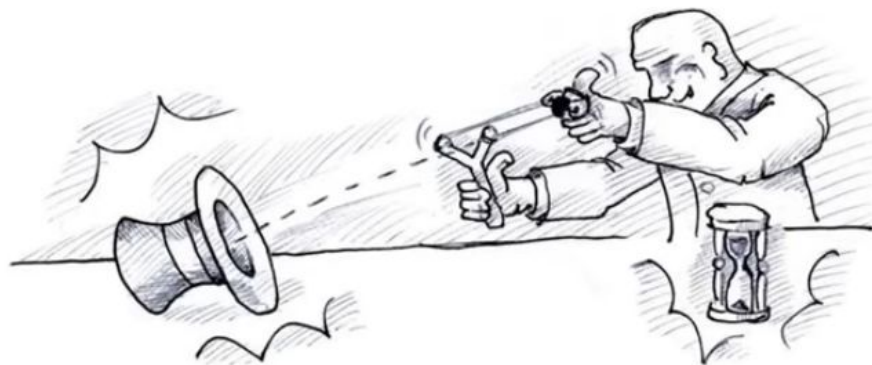
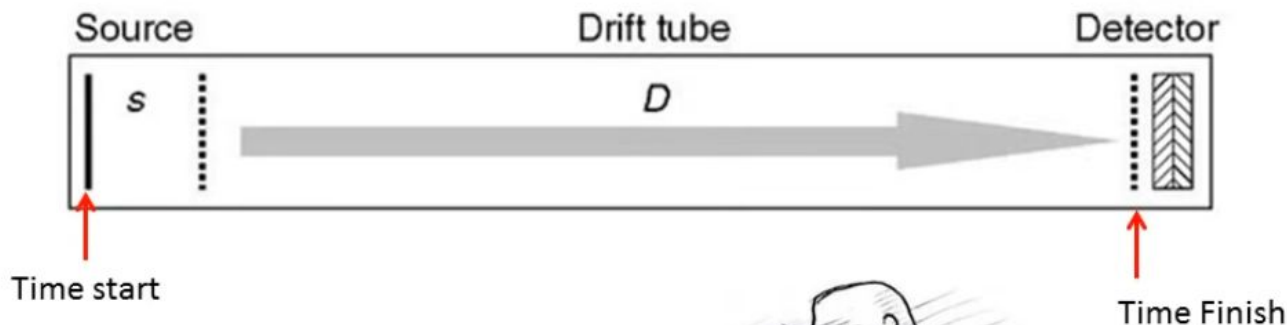


Matrix Assisted Laser Desorption/Ionisation

- Hillenkamp and Karas (1985) ionised amino acids and small peptides.



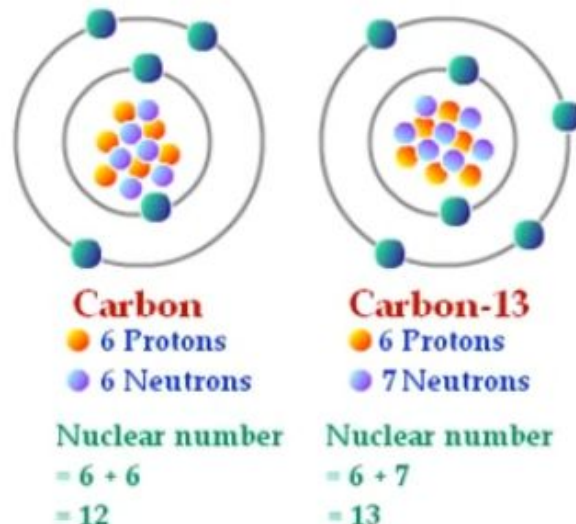
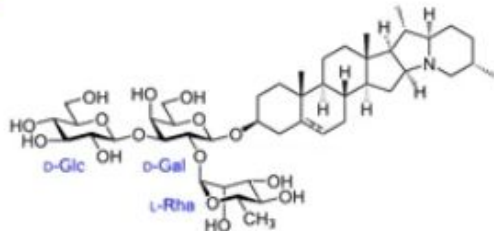
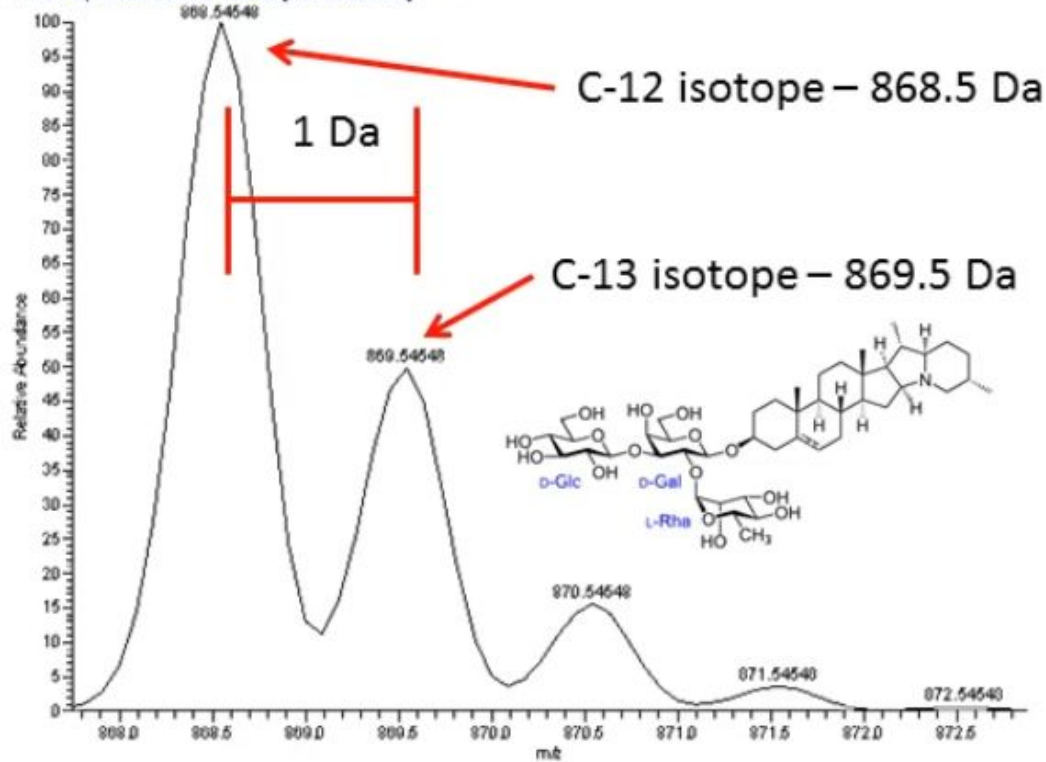
- Time of Flight: The time it takes an ion to go from ion source to detector is directly related to it's mass.
 - Heavier particles reach lower speeds.



$$velocity = \sqrt{\frac{2 \times \text{energy}}{\text{mass}}}$$

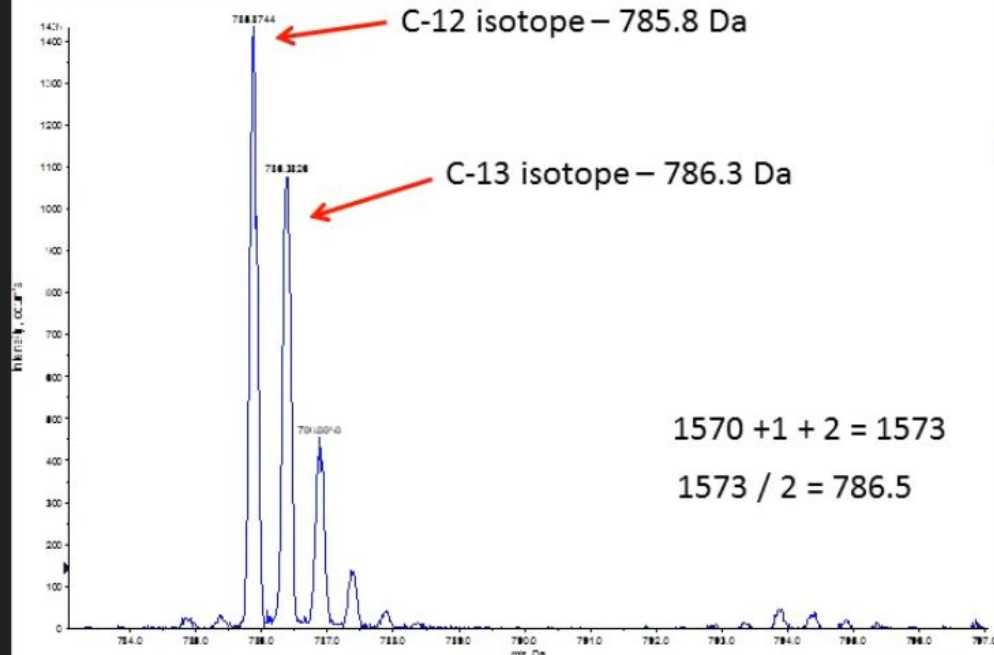
$$\text{Flight_time} = \frac{\text{drift_length}}{\text{velocity}} = \text{drift_length} \times \sqrt{\frac{\text{mass}}{2 \times \text{energy}}}$$

25-3-solanin-LTQ-pos-100v#31-258 RT: 0.10-0.87 AV: 228 NL: 2.11E4
T: ITMS + p NSI sid=100.00 Full ms [120.00-2000.00]



Mass/charge = m/z

-TOP MS: Exp 1, 18.380 min from sample 1 (GluF02).wiff
3.89424154280370900e-006, SD -3.08641800000770210e+001 M; (Nanospray)

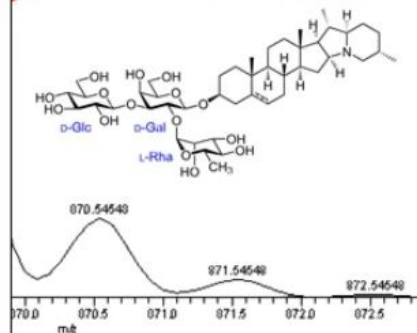


25-3-estran-17-OH-100V#3 1-258 RT: 0.10-0.97 AM: 229 NL: 2.1164
T: ITMS + p NSI sld=100.00 Full ms [120.00-2000.00]

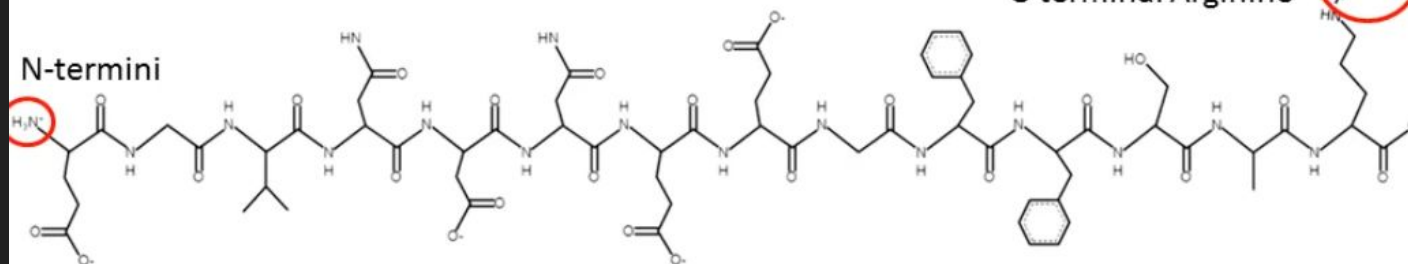
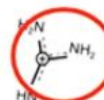


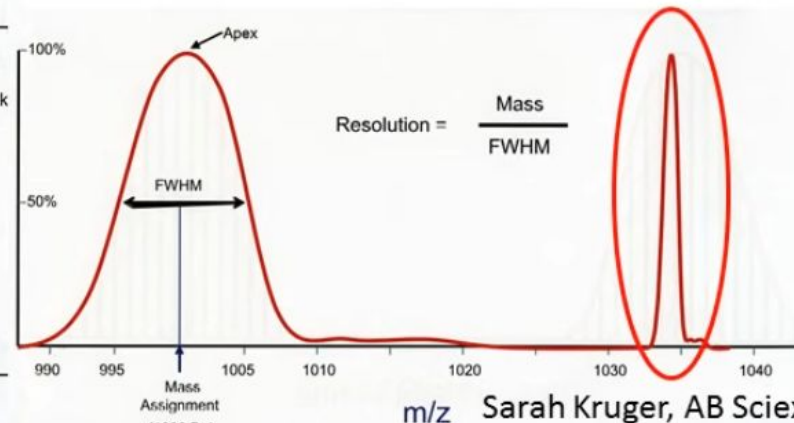
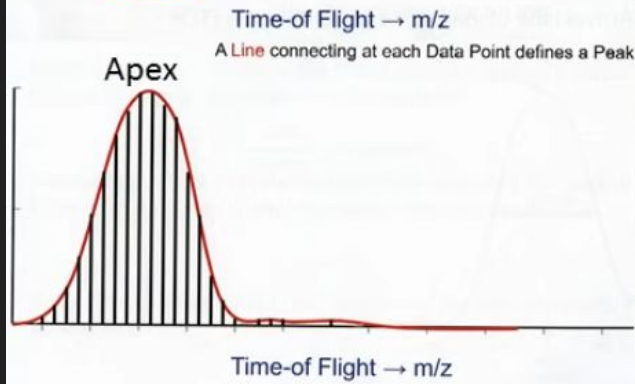
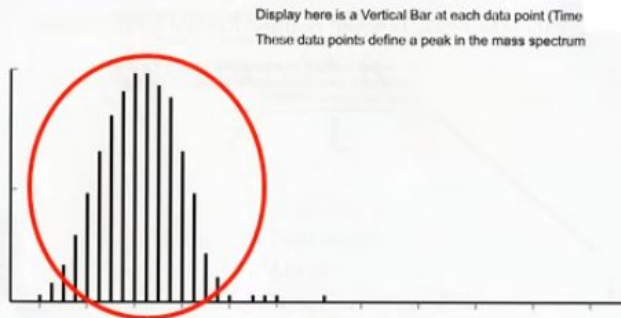
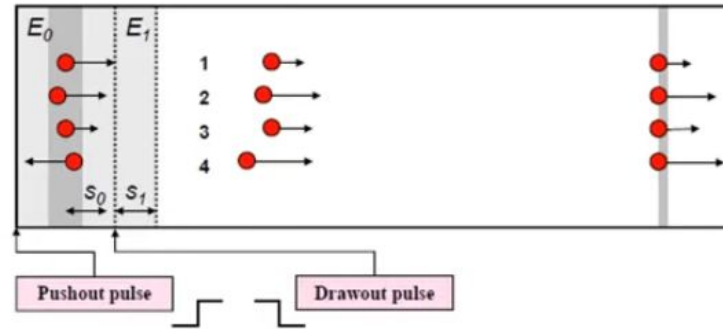
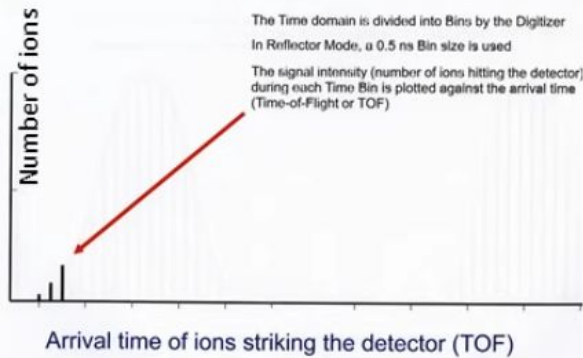
C-12 isotope – 868.5 Da

C-13 isotope – 869.5 Da

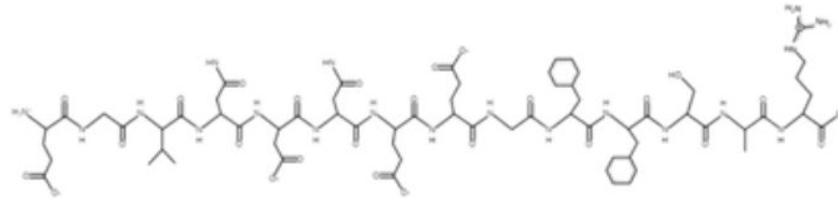


C-terminal Arginine



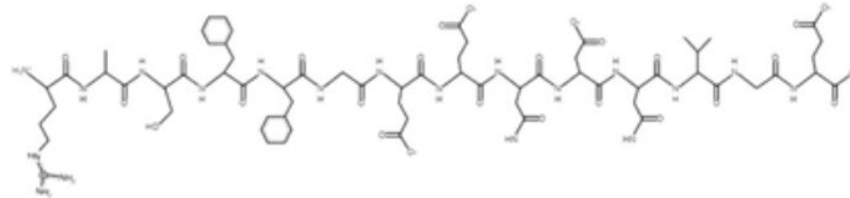


- But, in a more complex mixture of peptides from different protein isoforms, isobaric peptides exist.



EGVNDNEEGFFSAR

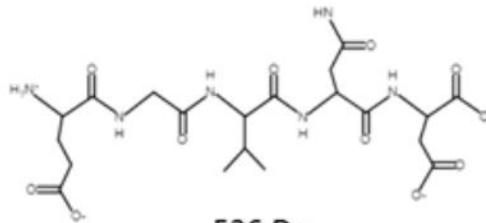
1570 Da



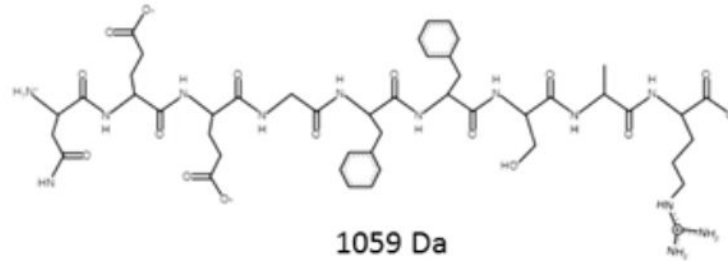
RASFFGEENDNVGE

- Same mass, but different structure.
- Measuring intact mass doesn't reveal isomers.

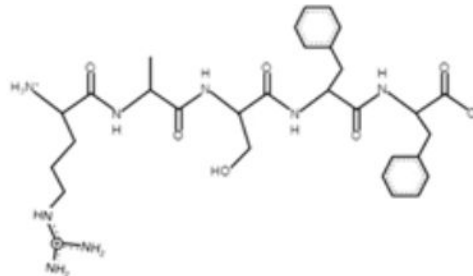
- Break molecule into smaller pieces and measure their masses.



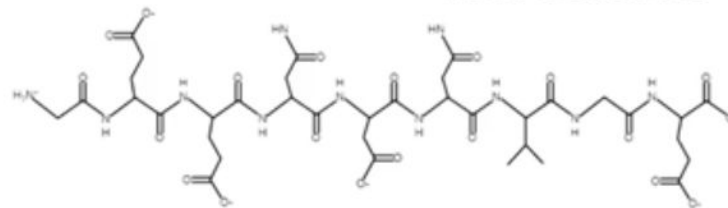
536 Da



1059 Da



626 Da

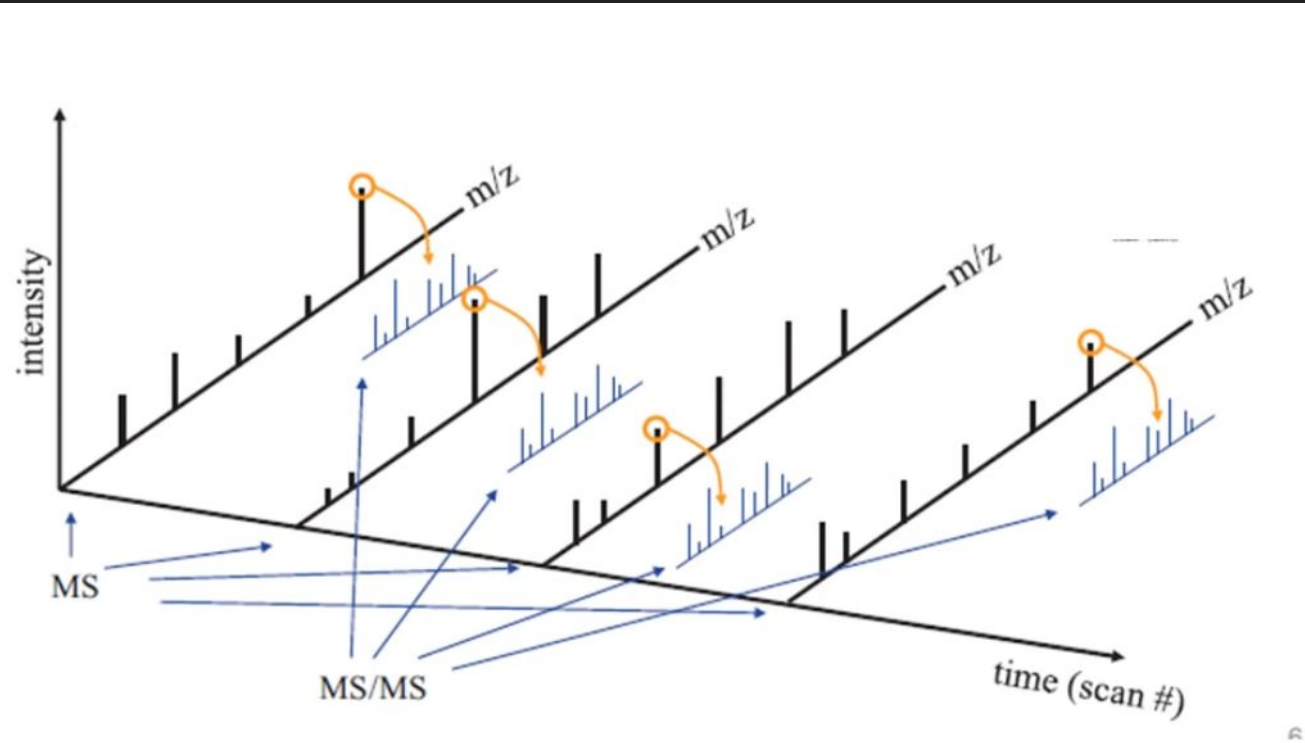


969 Da

EGVNDNEEGFFSAR

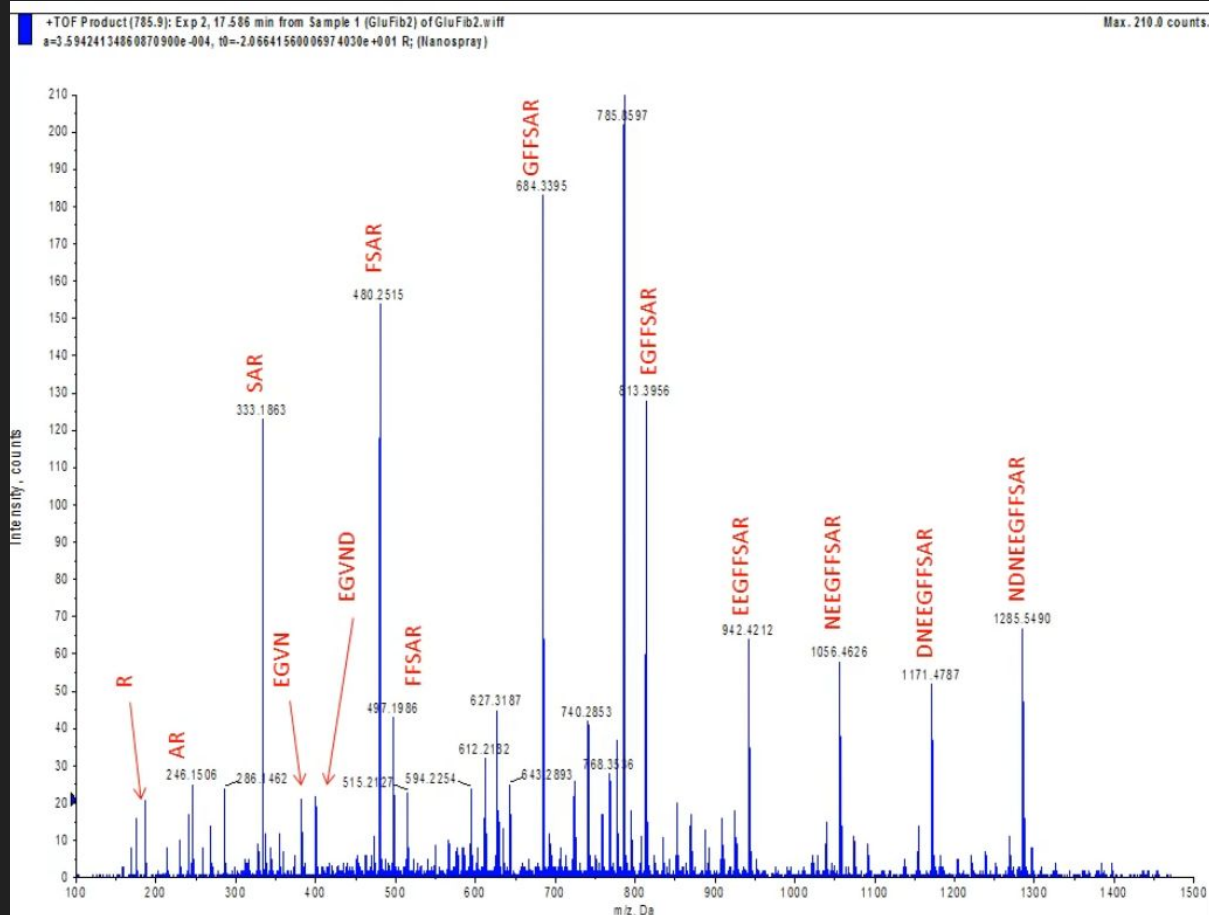
RASFFGEENDNVGE

The MS scans continuously for intact peptide ions, selecting one of those ions, fragmenting it, and measuring the mass of each fragment



Mass spectrometry

The compound sequence is not known, but can be built back by putting together peaks which difference is the mass of an amino acid.

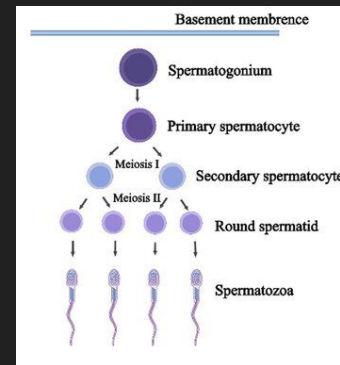
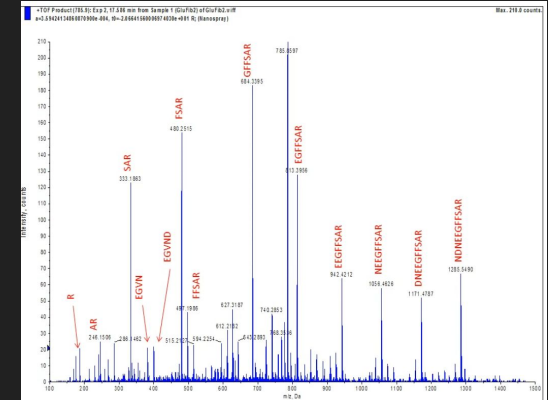


Acknowledgements:

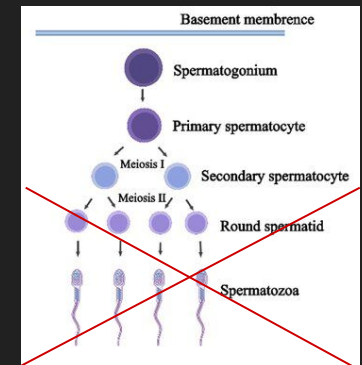
Most slides are taken from or inspired by the course in Proteomics by Matthew Padula. All his proteomics lectures are stored on the youtube channel <https://www.youtube.com/@MatthewPadula/videos> and are highly recommended for a more thorough introduction than my own.

Tutorial

- Raw data from 6 samples: 3 fertile and 3 infertile men using Liquid Chromatography Mass Spectrometry (LC-MS)
- Pipeline to identify peptides and infer protein occurrences in the spectra
- Export-Import data and find significant proteins between the conditions
- Examples of biologically relevant information from the protein/gene list



VS



- Go to <https://hds-sandbox.github.io/OMICS-workshop/>
- Follow the "**uCloud access**" instructions in the menu if this is your first access on uCloud
- Go on "**Day 3 - Proteomics**" on the webpage for instructions to work on the tutorial