

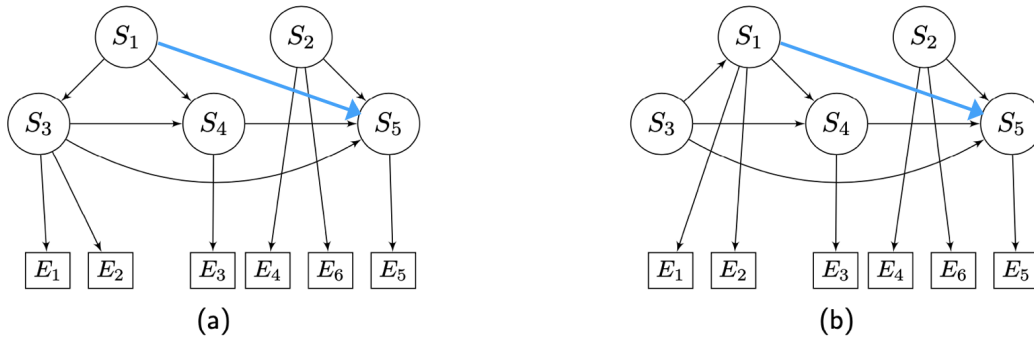
# Project 8

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## Problem 20: Classical NEMs

1. For each model, construct the transitive closure (by adding edges) and define the corresponding adjacency matrices  $\Phi$  and  $\Theta$ , which represent the signalling pathways and the E-gene attachments. Determine the corresponding expected effect patterns (F).



#### Construct phi

```
phi1 = t(array(c(c(1,0,1,1,1),
                 c(0,1,0,0,1),
                 c(0,0,1,1,1),
                 c(0,0,0,1,1),
                 c(0,0,0,0,1)),
               dim = c(5, 5), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                               c("S1", "S2", "S3", "S4", "S5")))))

phi2 = t(array(c(c(1,0,0,1,1),
                 c(0,1,0,0,1),
                 c(1,0,1,1,1),
                 c(0,0,0,1,1),
                 c(0,0,0,0,1)),
               dim = c(5, 5), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                               c("S1", "S2", "S3", "S4", "S5")))))
```

phi1

a)  $\Phi$

```
##      S1 S2 S3 S4 S5
## S1  1  0  1  1  1
## S2  0  1  0  0  1
## S3  0  0  1  1  1
## S4  0  0  0  1  1
## S5  0  0  0  0  1
```

phi2

b)  $\Phi$

```
##      S1 S2 S3 S4 S5
## S1  1  0  0  1  1
## S2  0  1  0  0  1
## S3  1  0  1  1  1
## S4  0  0  0  1  1
## S5  0  0  0  0  1
```

```
theta1 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                              c("E1", "E2", "E3", "E4", "E5", "E6")))

theta1["S1",] = c(0,0,0,0,0,0)
theta1["S2",] = c(0,0,0,1,0,1)
theta1["S3",] = c(1,1,0,0,0,0)
theta1["S4",] = c(0,0,1,0,0,0)
theta1["S5",] = c(0,0,0,0,1,0)

theta2 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                              c("E1", "E2", "E3", "E4", "E5", "E6")))

theta2["S1",] = c(1,1,0,0,0,0)
theta2["S2",] = c(0,0,0,1,0,1)
theta2["S3",] = c(0,0,0,0,0,0)
theta2["S4",] = c(0,0,1,0,0,0)
theta2["S5",] = c(0,0,0,0,1,0)
```

Construct theta

theta1

a)  $\Theta$

```
##      E1 E2 E3 E4 E5 E6
## S1  0  0  0  0  0  0
## S2  0  0  0  1  0  1
## S3  1  1  0  0  0  0
## S4  0  0  1  0  0  0
## S5  0  0  0  0  1  0
```

theta2

b)  $\Theta$

```
##      E1 E2 E3 E4 E5 E6
## S1  1  1  0  0  0  0
```

```
## S2  0  0  0  1  0  1
## S3  0  0  0  0  0  0
## S4  0  0  1  0  0  0
## S5  0  0  0  0  1  0
```

```
F1 = phi1%*%theta1
F2 = phi2%*%theta2
```

Calculate  $F = \Phi\Theta$

F1

a)  $F$

```
##      E1 E2 E3 E4 E5 E6
## S1  1  1  1  0  1  0
## S2  0  0  0  1  1  1
## S3  1  1  1  0  1  0
## S4  0  0  1  0  1  0
## S5  0  0  0  0  1  0
```

F2

b)  $F$

```
##      E1 E2 E3 E4 E5 E6
## S1  1  1  1  0  1  0
## S2  0  0  0  1  1  1
## S3  1  1  1  0  1  0
## S4  0  0  1  0  1  0
## S5  0  0  0  0  1  0
```

2. Assuming no noise, determine the discrete data D1 and D2 from both models. Given only the data, can you tell apart the two models?

```
D1 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                             c("S1", "S2", "S3", "S4", "S5")))
```

```
D1["E1",] = c(1,0,1,0,0)
D1["E2",] = c(1,0,1,0,0)
D1["E3",] = c(1,0,1,1,0)
D1["E4",] = c(0,1,0,0,0)
D1["E5",] = c(1,1,1,1,0)
D1["E6",] = c(0,1,0,0,1)
```

```
D2 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                             c("S1", "S2", "S3", "S4", "S5")))
```

```
D2["E1",] = c(1,0,1,0,0)
D2["E2",] = c(1,0,1,0,0)
D2["E3",] = c(1,0,1,1,0)
D2["E4",] = c(0,1,0,0,0)
```

```
D2["E5",] = c(1,1,1,1,0)
D2["E6",] = c(0,1,0,0,1)
```

D1

a)  $D1$

```
##      S1 S2 S3 S4 S5
## E1  1  0  1  0  0
## E2  1  0  1  0  0
## E3  1  0  1  1  0
## E4  0  1  0  0  0
## E5  1  1  1  1  0
## E6  0  1  0  0  1
```

D2

2)  $D2$

```
##      S1 S2 S3 S4 S5
## E1  1  0  1  0  0
## E2  1  0  1  0  0
## E3  1  0  1  1  0
## E4  0  1  0  0  0
## E5  1  1  1  1  0
## E6  0  1  0  0  1
```

Since the Data matrices D1 and D2 are identical, we cannot tell the two models apart.

**3. Use the `mnem1` package for this question: Take D1 and D2 from the previous question. For each model, calculate the marginal log-likelihood ratio (network score) given the data by setting the false positive rate to be 5 and the false negative rate to be 1.**

```
library(mnem)
```

```
## Registered S3 methods overwritten by 'RcppEigen':
##      method      from
## predict.fastLm    RcppArmadillo
## print.fastLm      RcppArmadillo
## summary.fastLm    RcppArmadillo
## print.summary.fastLm RcppArmadillo
```

```
network_score_1 = scoreAdj(D1,adj = phi1,method="disc",fpfn=c(0.05,0.01))$score
network_score_2 = scoreAdj(D2,adj = phi2,method="disc",fpfn=c(0.05,0.01))$score
```

```
network_score_1
```

Network score of (a)

```
## [1] 51.68914
```

```
network_score_2
```

Network score of (b)

```
## [1] 51.68914
```

## Problem 21: Hidden Markov NEMs

1. Using the definitions for HM-NEMs from the lecture, compute the transition probabilities from  $G_t = u$  to  $G_{t+1} \in v1, v2$  for different smoothness parameter  $\lambda \in 0.1, \dots 0.9$ .

```
u = t(array(c(c(1,1,1,0),
              c(0,1,1,1),
              c(0,0,1,1),
              c(0,0,0,1)),
           dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                           c("S1", "S2", "S3", "S4"))))

v1 = t(array(c(c(1,1,1,0),
              c(0,1,1,1),
              c(0,0,1,0),
              c(0,0,0,1)),
           dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                           c("S1", "S2", "S3", "S4"))))

v2 = t(array(c(c(1,0,0,0),
              c(1,1,1,0),
              c(1,0,1,0),
              c(1,0,0,1)),
           dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                           c("S1", "S2", "S3", "S4"))))

lambda = seq(0.1, 0.9, by=0.1)

s_uv1 = sum(u!=v1)
s_uv2 = sum(u!=v2)
w = mnem:::enumerate.models(c("S1","S2","S3","S4"),trans.close = FALSE)

## Generated 4096 unique models ( out of 4096 )

s_uw = array(dim = c(length(w), 1))
for(i in 1:length(w)){
  s_uw[i] = sum(u!=w[[i]])
}

power <- function(x, y) sign(x) * abs(x)^y

T = array(dim = c(9,2), dimnames = list(lambda,c("v1", "v2")))
C = array(dim = c(9,1), dimnames = list(lambda,c("C")))
for(i in lambda){
  C[as.character(i),] = sum(power((1-i),s_uw)*i)
  T[as.character(i),"v1"] = (1/C[as.character(i),])*((1-i)^s_uv1)*i
  T[as.character(i),"v2"] = (1/C[as.character(i),])*((1-i)^s_uv2)*i
}
```

T

Transition probabilities

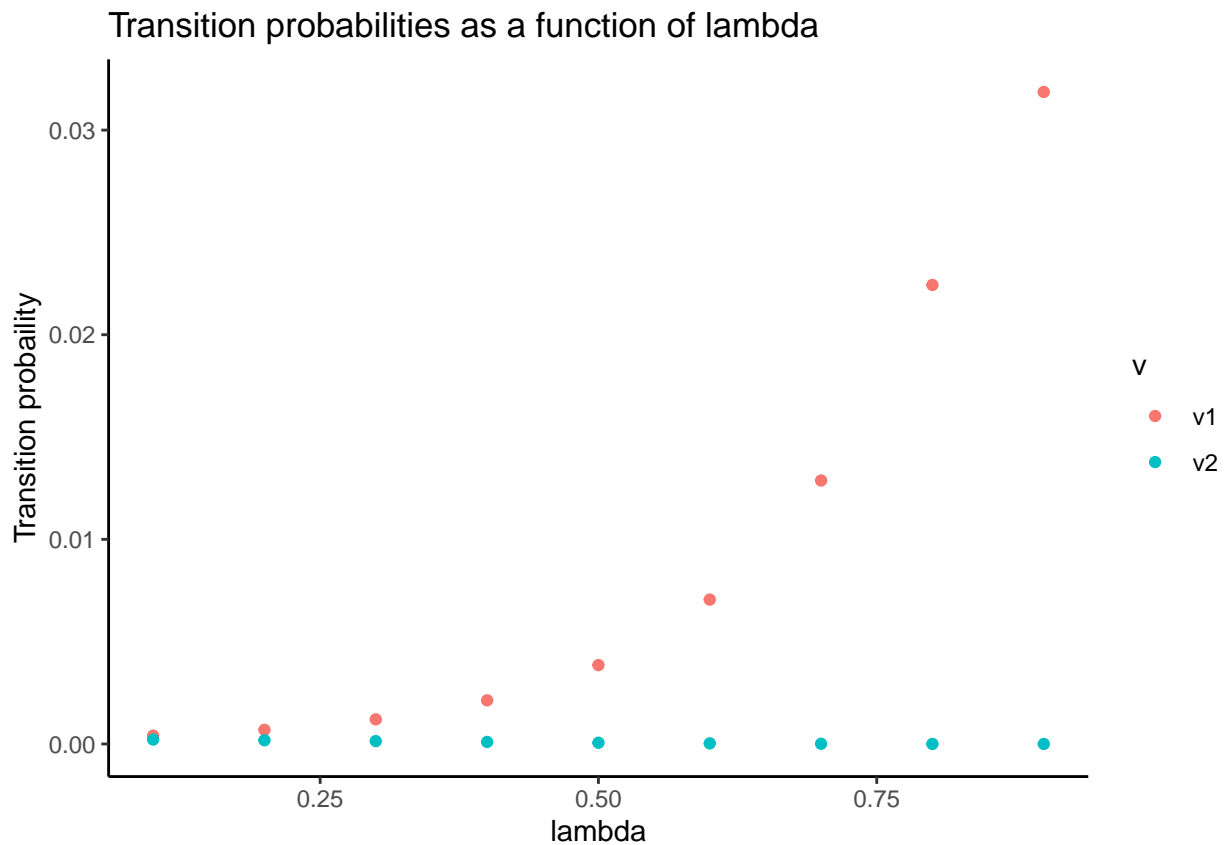
```
##           v1           v2
## 0.1 0.0004066299 2.160998e-04
## 0.2 0.0006915442 1.812842e-04
## 0.3 0.0012014646 1.413511e-04
## 0.4 0.0021316282 9.945325e-05
## 0.5 0.0038536733 6.021365e-05
## 0.6 0.0070554312 2.889905e-05
## 0.7 0.0128765947 9.387038e-06
## 0.8 0.0224313310 1.435605e-06
## 0.9 0.0318630818 3.186308e-08
```

2. Plot the transition probabilities for  $v_1$  and  $v_2$  as a function of  $\lambda$ . Describe the transition probabilities as a function of  $\lambda$ .

```
library(reshape2)
library(ggplot2)
library(RColorBrewer)
```

```
## Warning: package 'RColorBrewer' was built under R version 4.0.5
```

```
data = data.frame(melt(T))
colnames(data)<-c("lambda","v","T")
plot<-ggplot(data,aes(x=lambda,y=T,color=v))+
  geom_point()+
  theme_classic()+
  ylab("Transition probaility")+
  labs(title="Transition probabilities as a function of lambda")
plot
```



As  $\lambda$  increases the similarity between networks becomes more relevant for the transition probabilities, that is, dissimilar networks get more highly penalized with a lower transition probability as  $\lambda$  increases. Since  $v_1$  is more similar to  $u$  than  $v_2$ , the probability to transition into  $v_1$  increases as we increase  $\lambda$ . Conversely as we bring  $\lambda$  close to zero, network similarity becomes less relevant and we see the transition probabilities of  $v_1$  and  $v_2$  converge.

## Problem 22: Mixture NEMs

1. Determine the the cellular perturbation map  $\rho$ , where  $\rho_{ic} = 1$  if cell  $c$  is perturbed by a knock-down of S-gene  $i$ .

```
rho = array(dim = c(2,4), dimnames = list(c("S1","S2"),
                                           c("C1", "C2", "C3", "C4")))
rho["S1",] = c(1,0,1,0)
rho["S2",] = c(0,1,1,1)
rho

##      C1 C2 C3 C4
## S1   1  0  1  0
## S2   0  1  1  1
```

2. Assume that  $C_1, C_2$  are generated from  $F_1$  and  $C_3, C_4$  are generated from  $F_2$ , compute the noiseless log odds matrix  $R$ , where  $R_{jc} > 0$  means that the perturbation on cell  $c$  has an effect on E-gene  $j$ :

```
phi_F1 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("S1", "S2")))
phi_F1["S1",] = c(1,1)
phi_F1["S2",] = c(0,1)

phi_F2 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("S1", "S2")))
phi_F2["S1",] = c(1,0)
phi_F2["S2",] = c(1,1)

theta_F1 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("E1", "E2")))
theta_F1["S1",] = c(1,0)
theta_F1["S2",] = c(0,1)

theta_F2 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("E1", "E2")))
theta_F2["S1",] = c(0,1)
theta_F2["S2",] = c(1,0)

EEP_F1 = t(t(rho))%*%phi_F1%*%theta_F1
EEP_F1[EEP_F1>1] = 1
EEP_F2 = t(t(rho))%*%phi_F2%*%theta_F2
EEP_F2[EEP_F2>1] = 1
```

(a) For each component  $k$ , compute the expected effect pattern  $(\rho^T \phi^k \theta^k)^T$ . Replace all non-zeros by 1.

```
EEP_F1
```

Expected effect pattern of F1

```
##      C1 C2 C3 C4
## E1   1  0  1  0
## E2   1  1  1  1
```

```
EEP_F2
```

Expected effect pattern of F2

```
##      C1 C2 C3 C4
## E1   0  1  1  1
## E2   1  1  1  1
```

```
R= cbind(EEP_F1[,1:2],EEP_F2[,3:4])
R[R==0] = -1
```

```
R
```

(b) Based on the component assignment for each cell, extract the corresponding column from the expected effect patterns computed above and put it into R. Replace all zeros by  $-1$ .

```
##      C1 C2 C3 C4
## E1   1 -1  1  1
## E2   1  1  1  1
```

3. Take R from the previous question. Given the vector of mixture weights  $\pi = (0.44, 0.56)$ , calculate the responsibilities  $\Gamma$ . Then, update the mixture weights.

```
L1 = t(EEP_F1)%*%R
L2 = t(EEP_F2)%*%R
print("L1")
```

```
## [1] "L1"
```

```
L1
```

```
##      C1 C2 C3 C4
## C1   2  0  2  2
## C2   1  1  1  1
## C3   2  0  2  2
## C4   1  1  1  1
```

```
print("L2")
```

```
## [1] "L2"
```

```
L2
```

```
##      C1 C2 C3 C4
## C1   1  1  1  1
## C2   2  0  2  2
## C3   2  0  2  2
## C4   2  0  2  2
```



```

pi = c(0.44,0.56)

gamma = array(dim = c(2,4), dimnames = list(c("F1","F2"),
                                              c("C1", "C2", "C3", "C4")))
gamma["F1",] = pi[1]*exp(diag(L1))/(pi[1]*exp(diag(L1))+pi[2]*exp(diag(L2)))
gamma["F2",] = pi[2]*exp(diag(L2))/(pi[2]*exp(diag(L2))+pi[1]*exp(diag(L1)))

```

**Responsibilities**    gamma

```

pi[1] = sum(gamma["F1",])/(sum(gamma["F1",])+sum(gamma["F2",]))
pi[2] = sum(gamma["F2",])/(sum(gamma["F1",])+sum(gamma["F2",]))

```

```

pi

```

**Updated mixture weights**

```
## [1] 0.5066091 0.4933909
```