

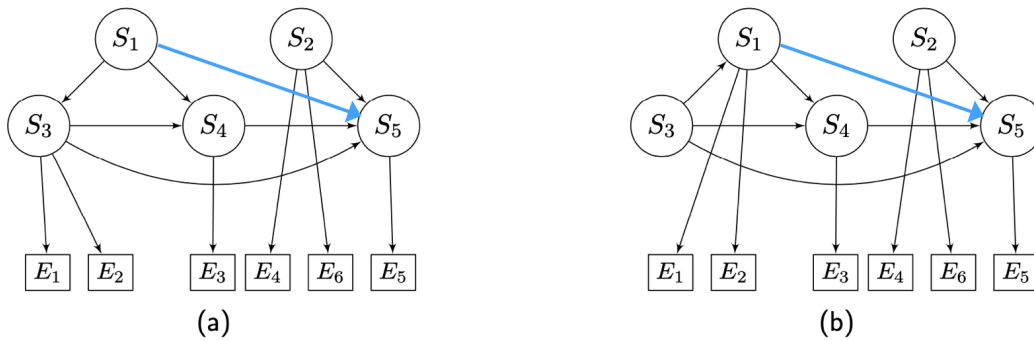
Project 8

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Problem 20: Classical NEMs

1. For each model, construct the transitive closure (by adding edges) and define the corresponding adjacency matrices Φ and Θ , which represent the signalling pathways and the E-gene attachments. Determine the corresponding expected effect patterns (F).



Construct phi

```
phi1 = t(array(c(c(1,0,1,1,1),
                 c(0,1,0,0,1),
                 c(0,0,1,1,1),
                 c(0,0,0,1,1),
                 c(0,0,0,0,1)),
               dim = c(5, 5), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                               c("S1", "S2", "S3", "S4", "S5"))))

phi2 = t(array(c(c(1,0,0,1,1),
                 c(0,1,0,0,1),
                 c(1,0,1,1,1),
                 c(0,0,0,1,1),
                 c(0,0,0,0,1)),
               dim = c(5, 5), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                               c("S1", "S2", "S3", "S4", "S5"))))
```

phi1

a) Φ

```
##      S1 S2 S3 S4 S5
## S1   1  0  1  1  1
## S2   0  1  0  0  1
## S3   0  0  1  1  1
## S4   0  0  0  1  1
## S5   0  0  0  0  1
```

phi2

b) Φ

```
##      S1 S2 S3 S4 S5
## S1   1  0  0  1  1
## S2   0  1  0  0  1
## S3   1  0  1  1  1
## S4   0  0  0  1  1
## S5   0  0  0  0  1
```

```
theta1 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                              c("E1", "E2", "E3", "E4", "E5", "E6")))

theta1["S1",] = c(0,0,0,0,0,0)
theta1["S2",] = c(0,0,0,1,0,1)
theta1["S3",] = c(1,1,0,0,0,0)
theta1["S4",] = c(0,0,1,0,0,0)
theta1["S5",] = c(0,0,0,0,1,0)

theta2 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"),
                                              c("E1", "E2", "E3", "E4", "E5", "E6")))

theta2["S1",] = c(1,1,0,0,0,0)
theta2["S2",] = c(0,0,0,1,0,1)
theta2["S3",] = c(0,0,0,0,0,0)
theta2["S4",] = c(0,0,1,0,0,0)
theta2["S5",] = c(0,0,0,0,1,0)
```

Construct theta

theta1

a) Θ

```
##      E1 E2 E3 E4 E5 E6
## S1   0  0  0  0  0  0
## S2   0  0  0  1  0  1
## S3   1  1  0  0  0  0
## S4   0  0  1  0  0  0
## S5   0  0  0  0  1  0
```

```
theta2
```

b) Θ

```
##      E1 E2 E3 E4 E5 E6
## S1   1  1  0  0  0  0
## S2   0  0  0  1  0  1
## S3   0  0  0  0  0  0
## S4   0  0  1  0  0  0
## S5   0  0  0  0  1  0
```

```
F1 = phi1*%theta1
F2 = phi2*%theta2
```

Calculate $F = \Phi\Theta$

```
F1
```

a) F

```
##      E1 E2 E3 E4 E5 E6
## S1   1  1  1  0  1  0
## S2   0  0  0  1  1  1
## S3   1  1  1  0  1  0
## S4   0  0  1  0  1  0
## S5   0  0  0  0  1  0
```

```
F2
```

b) F

```
##      E1 E2 E3 E4 E5 E6
## S1   1  1  1  0  1  0
## S2   0  0  0  1  1  1
## S3   1  1  1  0  1  0
## S4   0  0  1  0  1  0
## S5   0  0  0  0  1  0
```

2. Assuming no noise, determine the discrete data D1 and D2 from both models. Given only the data, can you tell apart the two models?

```
D1 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                           c("S1", "S2", "S3", "S4", "S5")))

D1["E1",] = c(1,0,1,0,0)
D1["E2",] = c(1,0,1,0,0)
D1["E3",] = c(1,0,1,1,0)
D1["E4",] = c(0,1,0,0,0)
D1["E5",] = c(1,1,1,1,0)
D1["E6",] = c(0,1,0,0,1)

D2 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                           c("S1", "S2", "S3", "S4", "S5")))

D2["E1",] = c(1,0,1,0,0)
D2["E2",] = c(1,0,1,0,0)
D2["E3",] = c(1,0,1,1,0)
D2["E4",] = c(0,1,0,0,0)
D2["E5",] = c(1,1,1,1,0)
D2["E6",] = c(0,1,0,0,1)
```

D1

a) *D1*

```
##      S1 S2 S3 S4 S5
## E1   1  0  1  0  0
## E2   1  0  1  0  0
## E3   1  0  1  1  0
## E4   0  1  0  0  0
## E5   1  1  1  1  0
## E6   0  1  0  0  1
```

D2

2) *D2*

```
##      S1 S2 S3 S4 S5
## E1   1  0  1  0  0
## E2   1  0  1  0  0
## E3   1  0  1  1  0
## E4   0  1  0  0  0
## E5   1  1  1  1  0
## E6   0  1  0  0  1
```

Since the Data matrices D1 and D2 are identical, we cannot tell the two models apart.

3. Use the `mnem1` package for this question: Take `D1` and `D2` from the previous question. For each model, calculate the marginal log-likelihood ratio (network score) given the data by setting the false positive rate to be 5 and the false negative rate to be 1.

```
library(mnem)

## Registered S3 methods overwritten by 'RcppEigen':
##   method      from
##   predict.fastLm  RcppArmadillo
##   print.fastLm    RcppArmadillo
##   summary.fastLm  RcppArmadillo
##   print.summary.fastLm RcppArmadillo

network_score_1 = scoreAdj(D1,adj = phi1,method="disc",marginal=TRUE,fpfn=c(0.05,0.01))$score
network_score_2 = scoreAdj(D2,adj = phi2,method="disc",marginal=TRUE,fpfn=c(0.05,0.01))$score
```

```
network_score_1
```

Network score of (a)

```
## [1] 52.42384
```

```
network_score_2
```

Network score of (b)

```
## [1] 52.42384
```

Problem 21: Hidden Markov NEMs

1. Using the definitions for HM-NEMs from the lecture, compute the transition probabilities from $G_t = u$ to $G_{t+1} \in v1, v2$ for different smoothness parameter $\lambda \in 0.1, \dots 0.9$.

```
u = t(array(c(c(1,1,1,0),
              c(0,1,1,1),
              c(0,0,1,1),
              c(0,0,0,1)),
            dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                             c("S1", "S2", "S3", "S4"))))

v1 = t(array(c(c(1,1,1,0),
               c(0,1,1,1),
               c(0,0,1,0),
               c(0,0,0,1)),
```

```

        dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                         c("S1", "S2", "S3", "S4")))
v2 = t(array(c(c(1,0,0,0),
               c(1,1,1,0),
               c(1,0,1,0),
               c(1,0,0,1)),
            dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                         c("S1", "S2", "S3", "S4"))))

lambda = seq(0.1, 0.9, by=0.1)

s_uv1 = sum(u!=v1)
s_uv2 = sum(u!=v2)
w = mnem:::enumerate.models(c("S1","S2","S3","S4"),trans.close = FALSE)

```

```
## Generated 4096 unique models ( out of 4096 )
```

```

s_uw = array(dim = c(length(w), 1))
for(i in 1:length(w)){
  s_uw[i] = sum(u!=w[[i]])
}

power <- function(x, y) sign(x) * abs(x)^y

T = array(dim = c(9,2), dimnames = list(lambda,c("v1", "v2")))
C = array(dim = c(9,1), dimnames = list(lambda,c("C")))
for(i in lambda){
  C[as.character(i),] = sum(power((1-i),s_uw)*i)
  T[as.character(i),"v1"] = (1/C[as.character(i),])*((1-i)^s_uv1)*i
  T[as.character(i),"v2"] = (1/C[as.character(i),])*((1-i)^s_uv2)*i
}

```

T

Transition probabilities

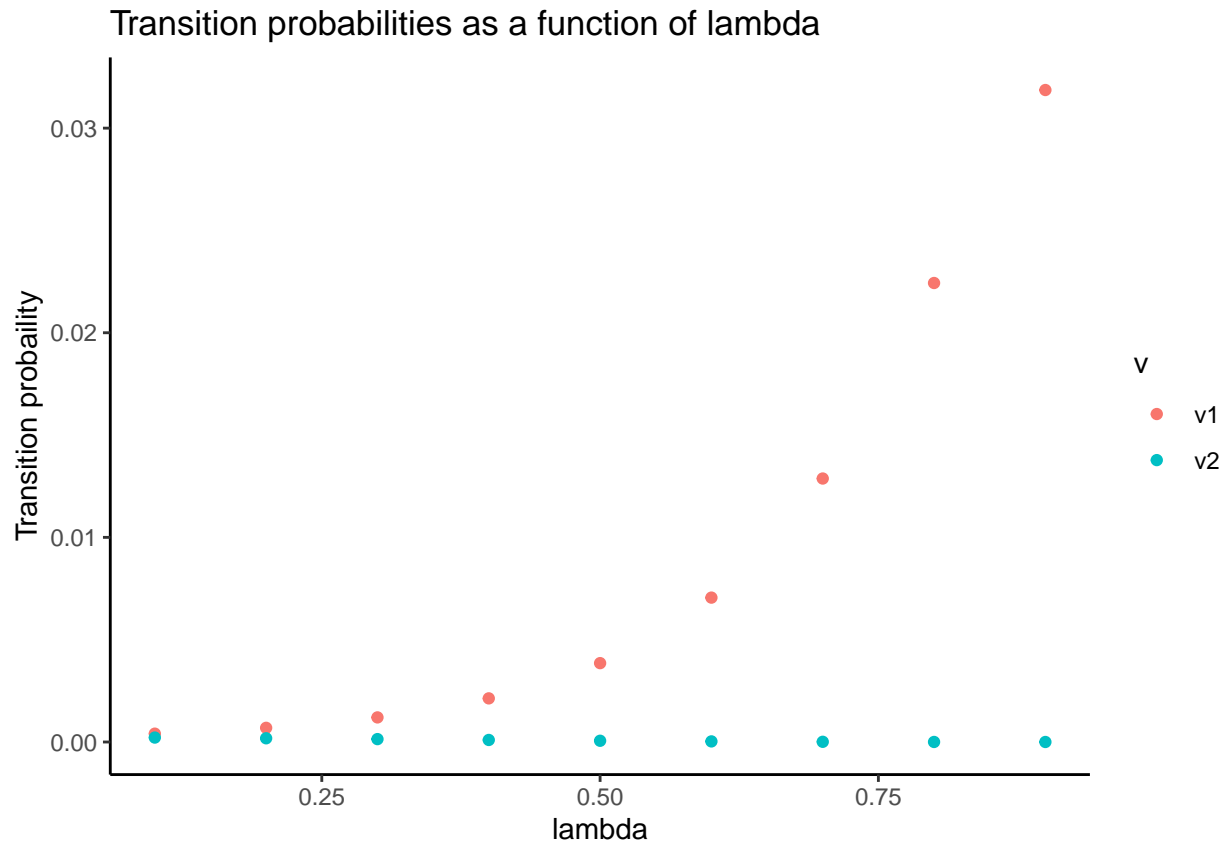
```

##           v1           v2
## 0.1 0.0004066299 2.160998e-04
## 0.2 0.0006915442 1.812842e-04
## 0.3 0.0012014646 1.413511e-04
## 0.4 0.0021316282 9.945325e-05
## 0.5 0.0038536733 6.021365e-05
## 0.6 0.0070554312 2.889905e-05
## 0.7 0.0128765947 9.387038e-06
## 0.8 0.0224313310 1.435605e-06
## 0.9 0.0318630818 3.186308e-08

```

2. Plot the transition probabilities for v_1 and v_2 as a function of λ . Describe the transition probabilities as a function of λ .

```
library(reshape2)
library(ggplot2)
library(RColorBrewer)
data = data.frame(melt(T))
colnames(data) <- c("lambda", "v", "T")
plot <- ggplot(data, aes(x=lambda, y=T, color=v)) +
  geom_point() +
  theme_classic() +
  ylab("Transition probability") +
  labs(title="Transition probabilities as a function of lambda")
plot
```



As λ increases the similarity between networks becomes more relevant the transition probabilities, that is, dissimilar networks get more highly penalized with a lower transition probability as λ increases. Since v_1 is more similar to u than v_2 , the probability to transition into v_1 increases as we increase λ . Conversely as we bring λ close to zero, network similarity becomes less relevant and we see the transition probabilities of v_1 and v_2 converge.

Problem 22: Mixture NEMs

1. Determine the cellular perturbation map ρ , where $\rho_{ic} = 1$ if cell c is perturbed by a knock-down of S-gene i .

```
rho = array(dim = c(2,4), dimnames = list(c("S1","S2"),
                                           c("C1", "C2", "C3", "C4")))
rho["S1",] = c(1,0,1,0)
rho["S2",] = c(0,1,1,1)
rho
```

```
##      C1 C2 C3 C4
## S1   1  0  1  0
## S2   0  1  1  1
```

2. Assume that C_1, C_2 are generated from F_1 and C_3, C_4 are generated from F_2 , compute the noiseless log odds matrix R , where $R_{jc} > 0$ means that the perturbation on cell c has an effect on E-gene j :

```
phi_F1 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("S1", "S2")))
phi_F1["S1",] = c(1,1)
phi_F1["S2",] = c(0,1)

phi_F2 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("S1", "S2")))
phi_F2["S1",] = c(1,0)
phi_F2["S2",] = c(1,1)

theta_F1 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("E1", "E2")))
theta_F1["S1",] = c(1,0)
theta_F1["S2",] = c(0,1)

theta_F2 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                              c("E1", "E2")))
theta_F2["S1",] = c(0,1)
theta_F2["S2",] = c(1,0)

EEP_F1 = t(t(rho)%*%phi_F1)%*%theta_F1
EEP_F1[EEP_F1>1] = 1
EEP_F2 = t(t(rho)%*%phi_F2)%*%theta_F2
EEP_F2[EEP_F2>1] = 1
```

(a) For each component k , compute the expected effect pattern $(\rho^T \phi^k \theta^k)^T$. Replace all non-zeros by 1.


```
EEP_F1
```

Expected effect pattern of F1

```
##      C1 C2 C3 C4
## E1   1  0  1  0
## E2   1  1  1  1
```

```
EEP_F2
```

Expected effect pattern of F2

```
##      C1 C2 C3 C4
## E1   0  1  1  1
## E2   1  1  1  1
```

```
R= cbind(EEP_F1[,1:2],EEP_F2[,3:4])
R[R==0] = -1
```

```
R
```

(b) Based on the component assignment for each cell, extract the corresponding column from the expected effect patterns computed above and put it into R. Replace all zeros by -1 .

```
##      C1 C2 C3 C4
## E1   1 -1  1  1
## E2   1  1  1  1
```

3. Take R from the previous question. Given the vector of mixture weights $\pi = (0.44, 0.56)$, calculate the responsibilities Γ . Then, update the mixture weights.

```
L1 = t(EEP_F1)%*%R
L2 = t(EEP_F2)%*%R
print("L1")
```

```
## [1] "L1"
```

```
L1
```

```
##      C1 C2 C3 C4
## C1   2  0  2  2
## C2   1  1  1  1
## C3   2  0  2  2
## C4   1  1  1  1
```

```
print("L2")
```

```
## [1] "L2"
```

```
L2
```

```
##      C1 C2 C3 C4
## C1   1  1  1  1
## C2   2  0  2  2
## C3   2  0  2  2
## C4   2  0  2  2
```

```
pi = c(0.44,0.56)
```

```
gamma = array(dim = c(2,4), dimnames = list(c("F1","F2"),
                                              c("C1", "C2", "C3", "C4")))
gamma["F1",] = pi[1]*exp(diag(L1))/(pi[1]*exp(diag(L1))+pi[2]*exp(diag(L2)))
gamma["F2",] = pi[2]*exp(diag(L2))/(pi[2]*exp(diag(L2))+pi[1]*exp(diag(L1)))
```

Responsibilities gamma

```
pi[1] = sum(gamma["F1",])/(sum(gamma["F1",])+sum(gamma["F2",]))
pi[2] = sum(gamma["F2",])/(sum(gamma["F1",])+sum(gamma["F2",]))
```

```
pi
```

Updated mixture weights

```
## [1] 0.5066091 0.4933909
```