# Project 8

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#### Problem 20: Classical NEMs

1. For each model, construct the transitive closure (by adding edges) and define the corresponding adjacency matrices  $\Phi$  and  $\Theta$ , which represent the signalling pathways and the E-gene attachments. Determine the corresponding expected effect patterns (F).

## Construct phi

```
theta1 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"), c("E1", "E2", "E3", "E4", "E5", "E6"))) theta1["S1",] = c(0,0,0,0,0,0) theta1["S2",] = c(0,0,0,1,0,1) theta1["S3",] = c(1,1,0,0,0,0) theta1["S4",] = c(0,0,1,0,0,0) theta1["S5",] = c(0,0,0,0,1,0) theta2 = array(dim = c(5,6), dimnames = list(c("S1", "S2", "S3", "S4", "S5"), c("E1", "E2", "E3", "E4", "E5", "E6"))) theta2["S1",] = c(1,1,0,0,0,0) theta2["S2",] = c(0,0,0,1,0,1)
```

```
theta2["S3",] = c(0,0,0,0,0,0)
theta2["S4",] = c(0,0,1,0,0,0)
theta2["S5",] = c(0,0,0,0,1,0)
theta1
Construct thetha
##
    E1 E2 E3 E4 E5 E6
## S1 0 0 0 0 0 0
## S2 0 0 0 1 0 1
## S3 1 1 0 0 0 0
## S4 0 0 1 0 0 0
## S5 0 0 0 0 1 0
theta2
## E1 E2 E3 E4 E5 E6
## S1 1 1 0 0 0 0
## S2 0 0 0 1 0 1
## S3 0 0 0 0 0 0
## S4 0 0 1 0 0 0
## S5 0 0 0 0 1 0
F1 = phi1%*%theta1
F2 = phi2%*%theta2
print("F1")
Calculate F = \Phi\Theta
## [1] "F1"
F1
## E1 E2 E3 E4 E5 E6
## S1 1 1 1 0 1 0
## S2 0 0 0 1 1 1
## S3 1 1 1 0 1 0
## S4 0 0 1 0 1 0
## S5 0 0 0 0 1 0
print("F2")
```

## [1] "F2"

F2

```
##
      E1 E2 E3 E4 E5 E6
## S1
      1
         1
            1 0 1
      0
          0
            0
## S3
                0
      1
          1
            1
                  1
## S4
      0
          0
            1
                0
                   1
                      0
## S5
      0
          0
            0
               0
                  1
```

2. Assuming no noise, determine the discrete data D1 and D2 from both models. Given only the data, can you tell apart the two models?

```
D1 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                           c("S1", "S2", "S3", "S4", "S5")))
D1["E1",] = c(1,0,1,0,0)
D1["E2",] = c(1,0,1,0,0)
D1["E3",] = c(1,0,1,1,0)
D1["E4",] = c(0,1,0,0,0)
D1["E5",] = c(1,1,1,1,0)
D1["E6",] = c(0,1,0,0,1)
D2 = array(dim = c(6, 5), dimnames = list(c("E1", "E2", "E3", "E4", "E5", "E6"),
                                           c("S1", "S2", "S3", "S4", "S5")))
D2["E1",] = c(1,0,1,0,0)
D2["E2",] = c(1,0,1,0,0)
D2["E3",] = c(1,0,1,1,0)
D2["E4",] = c(0,1,0,0,0)
D2["E5",] = c(1,1,1,1,0)
D2["E6",] = c(0,1,0,0,1)
D1
      S1 S2 S3 S4 S5
##
## E1
       1
          0
             1
## E2
                0
                   0
       1
          0
             1
## E3
       1
          0
             1
## E4
       0
             0
                0
                   0
          1
## E5
       1
          1
             1
                1
                   0
## E6
       0
             0
          1
D2
##
      S1 S2 S3 S4 S5
## E1
       1
          0
            1
                0
                   0
       1
          0
             1
                0
                   0
## E3
       1
          0
             1
                1
                   0
## E4
       0
          1
             0
                0
                   0
## E5
       1
          1
                   0
       0
             0
                0
## E6
          1
```

Since the Data matrices D1 and D2 are identical, we cannot tell the two models apart. ### 3. Use the mnem1 package for this question: Take D1 and D2 from the previous question. For each model, calculate

the marginal log-likelihood ratio (network score) given the data by setting the false positive rate to be 5 and the false negative rate to be 1.

```
library(mnem)
## Registered S3 methods overwritten by 'RcppEigen':
     method
##
                           from
##
     predict.fastLm
                          RcppArmadillo
##
                          RcppArmadillo
    print.fastLm
##
    summary.fastLm
                          RcppArmadillo
##
    print.summary.fastLm RcppArmadillo
\#nem1 = nem(D1, marqinal = TRUE, fpfn = c(0.05, 0.01))
#nem1$score
\#nem2 = nem(D2, marginal = TRUE, fpfn = c(0.05, 0.01))
#nem2$score
###Not sure which one it is
scoreAdj(D1,adj = phi1,method="disc",marginal=TRUE,fpfn=c(0.05,0.01))$score
## [1] 52.42384
scoreAdj(D2,adj = phi2,method="disc",marginal=TRUE,fpfn=c(0.05,0.01))$score
## [1] 52.42384
```

#### Problem 21: Hidden Markov NEMs

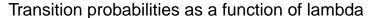
1. Using the definitions for HM-NEMs from the lecture, compute the transition probabilities from  $G_t = u$  to  $G_{t+1} \in v1, v2$  for different smoothness parameter  $\lambda \in 0.1, \dots 0.9$ .

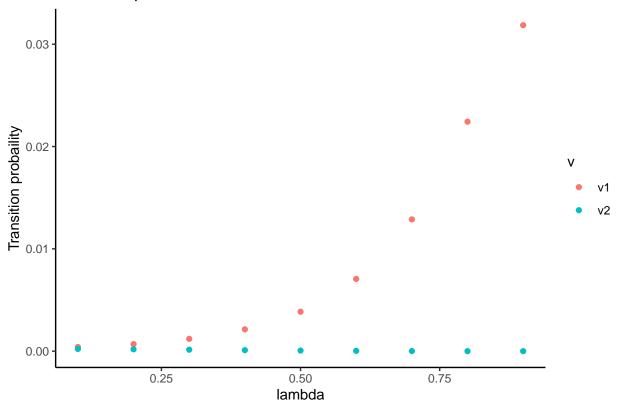
```
## not sure if the enumerate has to be used and how?
u = t(array(c(c(1,1,1,0)),
              c(0,1,1,1),
              c(0,0,1,1),
              c(0,0,0,1)),
              \dim = c(4, 4), \dim = list(c("S1", "S2", "S3", "S4"),
                                             c("S1", "S2", "S3", "S4"))))
v1 = t(array(c(c(1,1,1,0),
              c(0,1,1,1),
              c(0,0,1,0),
              c(0,0,0,1)),
              dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                              c("S1", "S2", "S3", "S4"))))
v2 = t(array(c(c(1,0,0,0)),
              c(1,1,1,0),
              c(1,0,1,0),
              c(1,0,0,1)),
              dim = c(4, 4), dimnames = list(c("S1", "S2", "S3", "S4"),
                                              c("S1", "S2", "S3", "S4"))))
```

```
lambda = seq(0.1, 0.9, by=0.1)
s_uv1 = sum(u!=v1)
s_uv2 = sum(u!=v2)
w = mnem:::enumerate.models(c("S1", "S2", "S3", "S4"), trans.close = FALSE)
## Generated 4096 unique models ( out of 4096 )
s_uw = array(dim = c(length(w), 1))
for(i in 1:length(w)){
  s_uw[i] = sum(u!=w[[i]])
power <- function(x, y) sign(x) * abs(x)^y</pre>
T = array(dim = c(9,2), dimnames = list(lambda,c("v1", "v2")))
C = array(dim = c(9,1), dimnames = list(lambda,c("C")))
for(i in lambda){
  C[as.character(i),] = sum(power((1-i),s_uw)*i)
  T[as.character(i),"v1"] = (1/C[as.character(i),])*((1-i)^s_uv1)*i
  T[as.character(i),"v2"] = (1/C[as.character(i),])*((1-i)^s_uv2)*i
Τ
##
                 v1
                               v2
## 0.1 0.0004066299 2.160998e-04
## 0.2 0.0006915442 1.812842e-04
## 0.3 0.0012014646 1.413511e-04
## 0.4 0.0021316282 9.945325e-05
## 0.5 0.0038536733 6.021365e-05
## 0.6 0.0070554312 2.889905e-05
## 0.7 0.0128765947 9.387038e-06
## 0.8 0.0224313310 1.435605e-06
## 0.9 0.0318630818 3.186308e-08
```

2. Plot the transition probabilities for  $v_1$  and  $v_2$  as a function of  $\lambda$ . Describe the transition probabilities as a function of  $\lambda$ .

```
library(reshape2)
library(ggplot2)
library(RColorBrewer)
data = data.frame(melt(T))
colnames(data)<-c("lambda","v","T")
plot<-ggplot(data,aes(x=lambda,y=T,color=v))+
    geom_point()+
    theme_classic()+
    ylab("Transition probability")+
    labs(title="Transition probabilities as a function of lambda")
plot</pre>
```





## Problem 22: Mixture NEMs

## S2 0 1 1 1

1. Determine the the cellular perturbation map  $\rho$ , where  $\rho_{ic} = 1$  if cell c is perturbed by a knock-down of S-gene i.

2. Assume that  $C_1, C_2$  are generated from  $F_1$  and  $C_3, C_4$  are generated from  $F_2$ , compute the noiseless log odds matrix R, where  $R_{jc} > 0$  means that the perturbation on cell c has an effect on E-gene j:

```
phi_F1["S1",] = c(1,1)
phi_F1["S2",] = c(0,1)
phi_F2 = array(dim = c(2,2), dimnames = list(c("S1", "S2"),
                                              c("S1", "S2")))
phi_F2["S1",] = c(1,0)
phi_F2["S2",] = c(1,1)
theta_F1 = array(\dim = c(2,2), \dim = list(c("S1","S2"),
                                                c("E1", "E2")))
theta_F1["S1",] = c(1,0)
theta_F1["S2",] = c(0,1)
theta_F2 = array(dim = c(2,2), dimnames = list(c("S1","S2"),
                                                c("E1","E2")))
theta_F2["S1",] = c(0,1)
theta_F2["S2",] = c(1,0)
EEP_F1 = t(t(rho)%*%phi_F1%*%theta_F1)
EEP_F1[EEP_F1>1] = 1
EEP_F2 = t(t(rho)%*\%phi_F2%*\%theta_F2)
EEP_F2[EEP_F2>1] = 1
print("Expected effect pattern of F1")
(a) For each component k, compute the expected effect pattern (\rho^T \phi^k \theta^k)^T. Replace all non-
zeros by 1.
## [1] "Expected effect pattern of F1"
EEP_F1
```

```
## C1 C2 C3 C4
## E1 1 0 1 0
## E2 1 1 1 1

print("Expected effect pattern of F2")

## [1] "Expected effect pattern of F2"
```

```
EEP_F2
```

```
## C1 C2 C3 C4
## E1 0 1 1 1
## E2 1 1 1 1
```

```
R= cbind(EEP_F1[,1:2],EEP_F2[,3:4])
R[R==0] = -1
R
```

(b) Based on the component assignment for each cell, extract the corresponding column from the expected effect patterns computed above and put it into R. Replace all zeros by -1.

```
## C1 C2 C3 C4
## E1 1 -1 1 1
## E2 1 1 1 1
```

3. Take R from the previous question. Given the vector of mixture weights  $\pi = (0.44, 0.56)$ , calculate the responsibilities  $\Gamma$ . Then, update the mixture weights.

```
library("expm")
## Loading required package: Matrix
##
## Attaching package: 'expm'
## The following object is masked from 'package:Matrix':
##
##
      expm
L1 = t(EEP_F1)%*%R
L2 = t(EEP_F2)\%*\%R
print("L1")
## [1] "L1"
L1
##
     C1 C2 C3 C4
## C1 2 0 2 2
## C2 1 1 1 1
## C3 2 0 2 2
## C4 1 1 1 1
print("L2")
## [1] "L2"
L2
     C1 C2 C3 C4
## C1
     1
         1
           1 1
## C2
      2 0 2 2
## C3 2 0 2 2
## C4 2 0 2 2
```

```
pi = c(0.44, 0.56)
gamma = array(dim = c(2,4), dimnames = list(c("F1","F2"),
                                            c("C1", "C2", "C3", "C4")))
gamma["F1",] = pi[1]*exp(diag(L1))/(pi[1]*exp(diag(L1))+pi[2]*exp(diag(L2)))
gamma["F2",] = pi[2]*exp(diag(L2))/(pi[2]*exp(diag(L2))+pi[1]*exp(diag(L1)))
##Responsibilities should be in [0,1]??
print("Responsibilities")
## [1] "Responsibilities"
gamma
             C1
                       C2
                            СЗ
##
                                      C4
## F1 0.6811014 0.6811014 0.44 0.2242338
## F2 0.3188986 0.3188986 0.56 0.7757662
pi[1] = sum(gamma["F1",])/(sum(gamma["F1",])+sum(gamma["F2",]))
pi[2] = sum(gamma["F2",])/(sum(gamma["F1",])+sum(gamma["F2",]))
print("Updated mixture weights")
## [1] "Updated mixture weights"
рi
```

## [1] 0.5066091 0.4933909