

# Human Genetic Variation Influences Vitamin C Homeostasis by Altering Vitamin C Transport and Antioxidant Enzyme Function

Alexander J. Michels,<sup>1</sup> Tory M. Hagen,<sup>1,2</sup>  
and Balz Frei<sup>1,2</sup>

<sup>1</sup>Linus Pauling Institute, <sup>2</sup>Department of Biochemistry and Biophysics, Oregon State University, Corvallis, Oregon 97331; email: michelsa@onid.orst.edu, tory.hagen@oregonstate.edu, balz.frei@oregonstate.edu

Annu. Rev. Nutr. 2013. 33:45–70

First published online as a Review in Advance on  
April 29, 2013

The *Annual Review of Nutrition* is online at  
<http://nutr.annualreviews.org>

This article's doi:  
10.1146/annurev-nutr-071812-161246

Copyright © 2013 by Annual Reviews.  
All rights reserved

## Keywords

ascorbate, SVCT, haptoglobin, GST, polymorphism, SNP

## Abstract

New evidence for the regulation of vitamin C homeostasis has emerged from several studies of human genetic variation. Polymorphisms in the genes encoding sodium-dependent vitamin C transport proteins are strongly associated with plasma ascorbate levels and likely impact tissue cellular vitamin C status. Furthermore, genetic variants of proteins that suppress oxidative stress or detoxify oxidatively damaged biomolecules, i.e., haptoglobin, glutathione-S-transferases, and possibly manganese superoxide dismutase, affect ascorbate levels in the human body. There also is limited evidence for a role of glucose transport proteins. In this review, we examine the extent of the variation in these genes, their impact on vitamin C status, and their potential role in altering chronic disease risk. We conclude that future epidemiological studies should take into account genetic variation in order to successfully determine the role of vitamin C nutrition or supplementation in human vitamin C status and chronic disease risk.

## Contents

INTRODUCTION .....	46
SODIUM-DEPENDENT	
VITAMIN C TRANSPORTERS ..	47
SVCT1 ( <i>SLC23A1</i> ) .....	49
Genomic Characterization	
of <i>SLC23A1</i> .....	49
Genetic Variation in <i>SLC23A1</i>	
and Vitamin C Status .....	52
Genetic Variation in <i>SLC23A1</i>	
and Chronic Disease Risk .....	53
SVCT2 ( <i>SLC23A2</i> ) .....	54
Genomic Characterization	
of <i>SLC23A2</i> .....	54
Genetic Variation in <i>SLC23A2</i>	
and Vitamin C Status .....	55
Genetic Variation in <i>SLC23A2</i>	
and Chronic Disease Risk .....	56
HAPTOGLOBIN .....	57
Function of Haptoglobin .....	57
Characterization of Human	
Haptoglobin Genotypes .....	57
Haptoglobin Genetic Variation	
and Vitamin C Status .....	58
GLUTATHIONE	
S-TRANSFERASES .....	59
Functions of Glutathione	
S-Transferase Isoforms .....	59
Human GST Polymorphisms	
and Vitamin C Status .....	60
OTHER GENETIC	
INTERACTIONS	
WITH VITAMIN C .....	61
Manganese Superoxide Dismutase	
Polymorphisms and Vitamin C	
Status .....	61
Glucose Transporters and	
Vitamin C Transport .....	61
CONSEQUENCES OF GENETIC	
VARIATION .....	62
SUMMARY .....	63
FUTURE DIRECTIONS .....	64

## INTRODUCTION

Maintaining adequate vitamin C (ascorbic acid) levels in the body is essential for collagen, catecholamine, and carnitine biosynthesis; absorption of nonheme iron; and antioxidant protection and hence plays a critical role in normal functioning of the body and optimum health (62). Numerous factors, both endogenous and exogenous, contribute to vitamin C body status, including vitamin C transporters that regulate the vitamin's bioavailability and plasma and tissue concentrations; enzymatic reactions in which vitamin C is used up by reducing redox-active metal cofactors; and a host of environmental factors and endogenous stresses, such as oxidative stress, infection, and inflammation.

Thus, over the past decades, several proteins have been identified as critical regulators of vitamin C homeostasis. These proteins affect ascorbate levels by different mechanisms, such as vitamin C transport, transition metal regulation, enzymatic redox activity, and antioxidant protection (**Figure 1**). However, much of this work has been performed in animal models and cell culture, the results of which often are difficult to translate to humans or are confounded by artifacts in study design. Controlled studies of vitamin C status in humans are difficult to perform, challenged by design, and limited in interpretation (56).

Despite these difficulties, new evidence for the regulation of vitamin C status has emerged from studies of human genetic variations. Although a relatively new and burgeoning field, there is strong evidence that several genetic alterations, in the form of single-nucleotide polymorphisms (SNPs), gene duplications, or gene deletions, affect ascorbate levels in the human body. The phenotypes of these genomic changes are supporting previous evidence of the physiological functions of vitamin C and its role in human health.

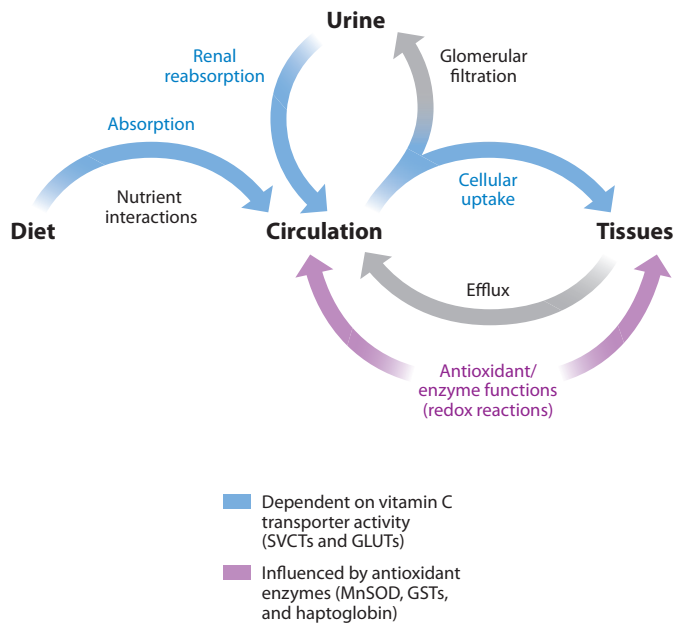
Although the list will undoubtedly grow with future research, this review focuses on variations within the genes for which there is direct evidence of alterations in vitamin C plasma levels. These genes can be subdivided

into two categories based on their interactions with ascorbic acid. First, genes involved in the direct transport and regulation of vitamin C concentrations in tissues and extracellular fluids are discussed. This discussion focuses on the known polymorphisms in the sodium-dependent vitamin C transporters (SVCTs) of the SLC23 family (*SLC23A1* and *SLC23A2*). The second category of genes is related to the antioxidant and redox activities of vitamin C and involves genetic variants affecting iron homeostasis and oxidative stress. Hence, we examine iron dysregulation through haptoglobin (*HP*) gene polymorphisms and genetic variants impacting the activity of glutathione S-transferases (*GSTs*). In addition, as candidates for future studies, we present some limited data on the influence of genetic changes in manganese superoxide dismutase (*SOD2*) and glucose transport proteins [solute carrier family 2 (*SLC2*)] on ascorbate levels in humans.

## SODIUM-DEPENDENT VITAMIN C TRANSPORTERS

The two sodium-dependent vitamin C transport proteins, SVCT1 and SVCT2, are specific for the cotransport of sodium ions and ascorbic acid across cell membranes. Although sodium-dependent vitamin C transport in cells had been characterized decades before (35), the genes encoding these members of the solute carrier family 23 (*SLC23A1* and *SLC23A2*) were not identified until 1999 (86). Initially characterized in rodents, human analogs were isolated and mapped shortly afterward (26, 73, 91).

Transcriptional regulation of the *SLC23* genes controls the tissue distribution of SVCTs and is responsible for the maintenance of vitamin C levels in nearly all cells, tissues, and extracellular fluids (Figure 2). The only cells without functional SVCT proteins are erythrocytes, which selectively lose SVCT2 expression during erythropoiesis (59). Consequently, ascorbate levels in erythrocytes are low, generally in equilibrium with the surrounding blood plasma.



**Figure 1**

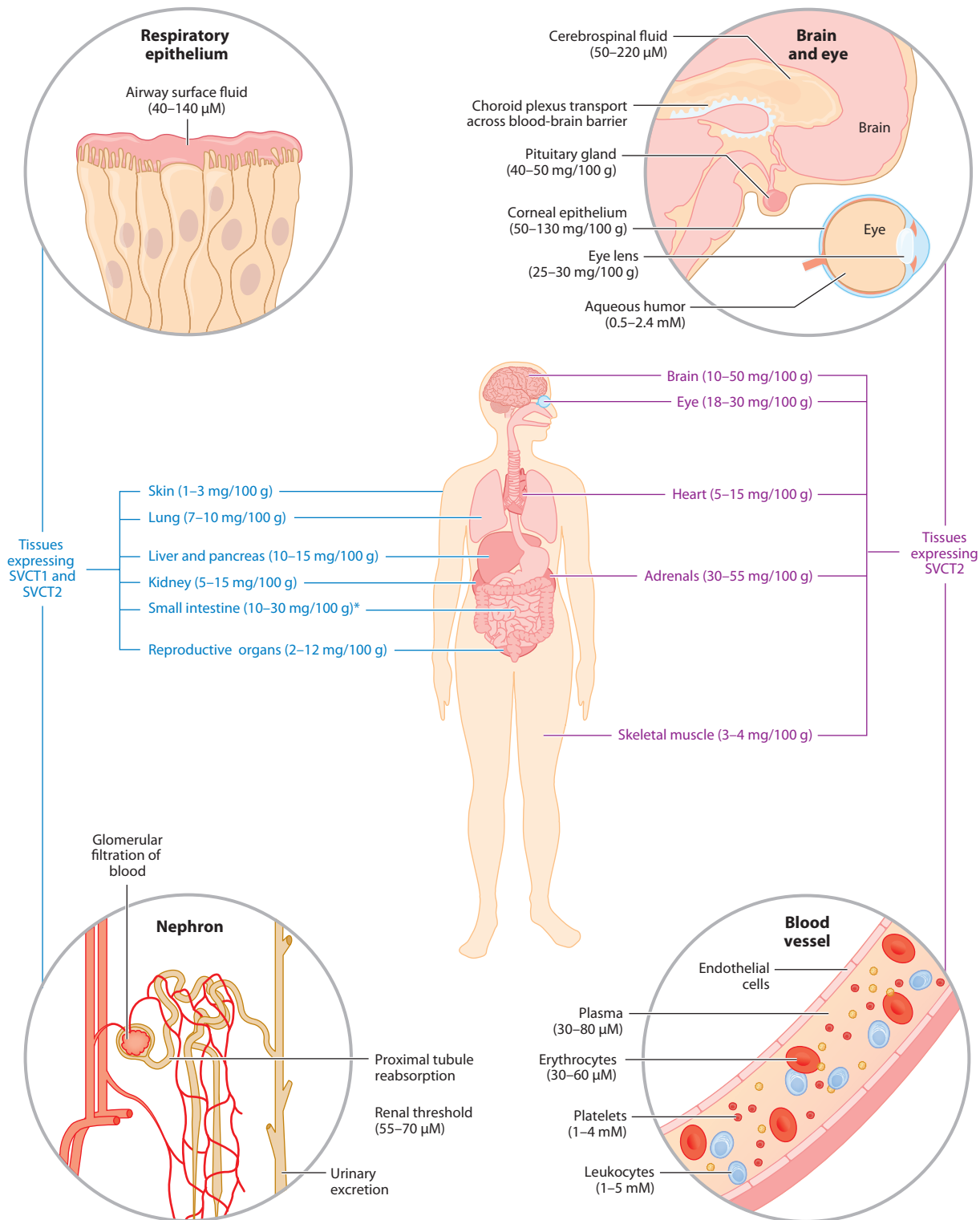
Vitamin C homeostasis. Vitamin C levels in the circulation and tissues are influenced by several regulatory mechanisms in the body. Proteins involved in vitamin C homeostasis fall under the general-categories transporters (blue) that transport vitamin C across cell membranes and those that modulate oxidative stress (purple) and would interact with ascorbate due to its antioxidant properties. Abbreviations: GLUT, glucose transporter protein; GST, glutathione S-transferase; MnSOD, manganese superoxide dismutase; SVCT, sodium-dependent vitamin C transporter.

Due to the direct interaction of SVCTs with ascorbic acid and their roles in absorption and tissue accumulation of vitamin C, genetic alterations in *SLC23A1* and *A2* appear to have the greatest effect on human vitamin C status of all the genetic factors investigated to date. Each *SLC23A1* and *SLC23A2* is thought to have emerged via gene duplication before vertebrate evolution and expansion (26). The divergent evolution of the two genes has led to distinct differences in the expression, regulation, and function of SVCT1 and SVCT2, and therefore the two genes encoding these proteins are considered separately. It should be noted that genetic variations in both of these genes is the latest area of research in this field, with these and other polymorphisms having the greatest potential for phenotypic effects that will impact vitamin C homeostasis and human health.

**Vitamin C:** includes ascorbic acid and its one- and two-electron oxidation products, ascorbyl radical and dehydroascorbic acid, respectively

**Single-nucleotide polymorphisms (SNPs):** changes in genetic structure between individuals differing by a single nucleotide

**Glutathione:** a cellular tripeptide with a reactive sulfhydryl group; it is a cellular antioxidant capable of reducing dehydroascorbic acid to ascorbic acid



## SVCT1 (*SLC23A1*)

SVCT1, encoded by the gene *SLC23A1*, is predominantly responsible for high-capacity vitamin C transport across membranes. Found on the apical side of polarized epithelial cells (8, 58), SVCT1 is highly expressed in the liver, kidney, lung, small intestine, and pancreas (86, 91). Of these tissues, it is the expression of SVCT1 in the kidney proximal tubule that is considered most important for the regulation of whole-body vitamin C status, as it is involved in the renal reabsorption of vitamin C following glomerular filtration of blood plasma (Figures 2 and 3*a*). Mice carrying a genetic knockout of *SLC23A1* excrete large amounts of ascorbic acid in the urine and have low plasma and tissue ascorbate levels (17). Because we cannot synthesize ascorbic acid in our body, loss of SVCT1 expression may cause a far more severe vitamin C deficiency in humans than in ascorbic acid–synthesizing animals. However, there are currently no reports of spontaneous loss of SVCT1 in humans.

Outside of an absence in renal reabsorption, it is unclear what other impact the loss of SVCT1 expression may have in humans. *SLC23A1* knockout mice experience increased perinatal mortality, but with no apparent physiological or cell-specific phenotype (17). Unlike SVCT2, SVCT1 appears to be expressed in specific locations within tissues. For instance, SVCT1 expression in the kidney is limited to the proximal tubule of the nephron (Figure 3*a*). It is possible that the loss of SVCT1 would only impact ascorbate levels in specific cellular environments or specific cell types within tissues. Although SVCT1 expression has been noted in the skin, bile duct, lung, pancreas, and reproductive organs (29,

45, 57, 91), it is not clear what consequences a loss of SVCT1 would have in these tissues. Another possibility is that the apparent lack of a phenotype in *SLC23A1*-null animals may be obscured, as all cell types that express SVCT1 also express SVCT2 (Figure 2). SVCT2 or glucose transporters (see below) may compensate for the lack of SVCT1, albeit with a limited capacity. For example, although SVCT1 is expressed in the intestine, *SLC23A1* genetic knockout mice are still capable of absorbing ascorbic acid from the diet (17).

## Genomic Characterization of *SLC23A1*

*SLC23A1* consists of 16 kb found on human chromosome 5 (26). The gene has 15 exons that encode for a 598 amino acid protein. A non-functional splice variant with four extra amino acids has been described in intestinal cells but has not been well characterized (91). The last intron and exon of *SLC23A1* overlap with the promoter and first exon and intron of the gene for polyadenylate-binding protein-interacting protein 2 (*PAIP2*), which encodes a transcriptional repressor protein (26). No genetic interaction between *PAIP2* and *SLC23A1* has been reported.

The promoter region of *SLC23A1* is small, with critical regulatory elements defined only within a few hundred base pairs of the transcription start site. Two independent studies have found functional binding sites for hepatocyte nuclear factor 1 (HNF1) in this region, and regulation of SVCT1 by HNF1 $\alpha$  has been demonstrated in HepG2 cells (63, 75). This would support regulation of the protein by HNF4 $\alpha$  as well. Although binding of other transcription factors has been postulated, none

**Superoxide:** reduced form of molecular oxygen (O<sub>2</sub><sup>•−</sup>); a free radical and reactive oxygen species

**Solute carrier family 2 (SLC2):** includes *SLC2A1*–*SLC2A14*, encoding glucose transporters GLUT1–GLUT14, respectively

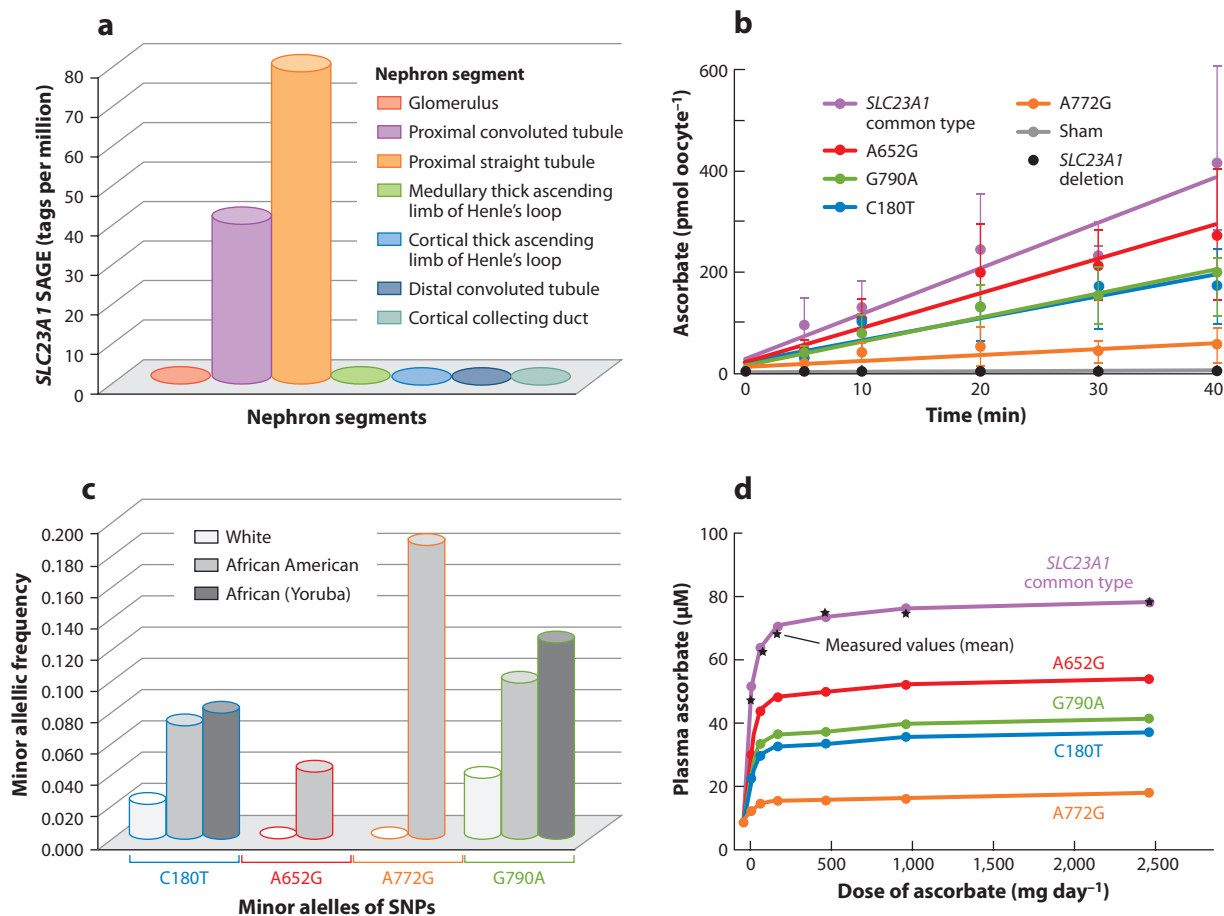
**Solute carrier family 23 (SLC23):** includes both *SLC23A1* (SVCT1) and *SLC23A2* (SVCT2). A gene identified as *SLC23A3* has an undefined function

**Apical:** in renal physiology, refers to the side of the cell facing the lumen (interior) of the proximal tubule

**Hepatocyte nuclear factor 1 $\alpha$  (HNF1 $\alpha$ ):** encoded by the gene *HNF1A*, it is a transcription factor highly expressed in the liver

## Figure 2

Tissue distribution of vitamin C and its transport proteins in the human body. Ascorbate concentrations are from limited human data and are represented as mg/100 g wet weight for tissues and molarity (mM or  $\mu$ M) for intracellular and extracellular fluids. Asterisk indicates that the tissue concentrations of ascorbate in the small intestine are heavily dependent on recent dietary intake. Sodium-dependent vitamin C transporter (SVCT) distribution is based on mRNA expression data in animals and human tissues. Redrawn with permission from Saunders/Elsevier, from A Michels and B Frei, Vitamin C, in *Biochemical, Physiological, and Molecular Aspects of Human Nutrition*, third edition, ed. MH Stipanuk, MA Caudill, pp. 626–54. Copyright 2012 (62).



**Figure 3**

*SLC23A1* activity and human genetic variation. (a) Distribution of *SLC23A1* in kidney nephron segments by Serial Analysis of Gene Expression (SAGE) libraries containing expression data from microdissected glomeruli and six different nephron segments; values are expressed as tags per million. (b) Effect of *SLC23A1* single-nucleotide polymorphisms (SNPs) on ascorbate transport. *Xenopus laevis* oocytes were microinjected with the following *SLC23A1* cRNAs: common type; sham injected; human deletion construct; and SNPs A652G (rs34521685), G790A (rs33972313), A772G (rs35817838), and C180T (rs6886922). (c) Population prevalences of *SLC23A1* polymorphisms. Shown are averaged minor allelic frequencies of *SLC23A1* genotypes in white ( $n = 1,874$ ), African American ( $n = 438$ ), and African ( $n = 48$ ) individuals, using pooled genotype data. (d) Modeled effects of *SLC23A1* polymorphisms on plasma ascorbate concentrations in humans. Values in healthy young women for common type *SLC23A1* are measured and calculated fasting steady-state plasma ascorbate concentrations. For women with SNPs, values are calculated from wild-type data comparisons to transport data in panel b. Adapted with permission of the American Society for Clinical Investigations, from Corpe CP, Tu H, Eck P, Wang J, Faulhaber-Walter R, Schnermann J, Margolis S, Padayatty S, Sun H, Wang Y, Nussbaum RL, Espey MG, Levine M. Vitamin C transporter Slc23a1 links renal reabsorption, vitamin C tissue accumulation, and perinatal survival in mice. *J. Clin. Invest.* 120(4):1069–83. Copyright 2010 (17).

have been identified. In some cells, SVCT1 expression declines with increasing exposure to ascorbate, but the regulatory factors mediating this response are unknown.

Over 150 SNPs have been identified in *SLC23A1*, although many of these have not been verified in large populations to determine global allele frequencies. Prevalence of



**Table 1** Phenotypic effects of single-nucleotide polymorphisms (SNPs) found in the human *SLC23A1* gene

Reference SNP ID	Location	Genetic variant <sup>a</sup>	References	Alterations in phenotype <sup>b</sup>
rs10063949	5' UTR	c.-97-487 A>G	(27, 28, 85, 94, 97)	Increased plasma and serum ascorbate levels in the BWHHS cohort but not in the EPIC cohort
rs6886922	Exon 3	c.180 C>T	(17)	Modeled in <i>Xenopus</i> oocytes: 40%–50% reduction in ascorbate transport
rs34521685	Exon 7	c.652 A>G	(17)	Isoleucine-to-valine substitution at position 218 of SVCT1. Modeled in <i>Xenopus</i> oocytes: 40%–50% reduction in ascorbate transport
rs35817838	Exon 8	c.772 A>G	(17, 27)	Methionine-to-valine substitution at position 258 of SVCT1. Modeled in <i>Xenopus</i> oocytes: 75% reduction in ascorbate transport
rs33972313	Exon 8	c.790 G>A	(17, 27, 85)	Valine-to-methionine substitution at position 264 of SVCT1. Modeled in <i>Xenopus</i> oocytes: 40%–50% reduction in ascorbate transport. Strongly associated with decreased plasma and serum ascorbate levels in multiple cohorts
rs11950646	Intron 9	c.1074-101 A>G	(82, 94)	Increased risk of follicular lymphoma observed in a US cohort
rs4257763	Intron 10	c.1179+109 T>C	(9, 85)	Decreased serum ascorbate levels in the BWHHS cohort
rs34063983	Intron 11	c.1310-41 A>G	(27)	None
rs6876106	Intron 13	c.1550-2088 C>T	(94)	None
rs6596473	Intron 13	c.1549+2515 G>C	(9, 27, 82, 85, 94)	Trend toward decreased plasma ascorbate levels in two studies. Increased risk of follicular lymphoma in a US cohort
rs6596471	Intron 14	c.*19+2088 T>C	(85)	None

<sup>a</sup>Variations listed relative to the coding region in the reference sequence NM\_005847.4.

<sup>b</sup>Effects of genetic polymorphism as described in the references provided.

Abbreviations: BWHHS, British Women's Heart and Health Study; EPIC, European Prospective Investigation into Cancer and Nutrition; SVCT, sodium-dependent vitamin C transport protein.

13 SNPs has been noted in Caucasian, African American, and, to a lesser degree, Asian populations (9, 26). Overall, African American populations show the greatest diversity of SNPs present in *SLC23A1*—many of which are not detectable in Caucasians (9). However, as studies into the genetic variations in *SLC23A1* continue, a different portrait of SVCT1 genetic variability may emerge.

At least four of the SNPs present in *SLC23A1* are found within the coding region

(exons 3, 7, and 8), which results in one synonymous and three nonsynonymous changes to the protein (**Table 1**). All nonsynonymous polymorphisms produce a functional SVCT1 protein, but each of these proteins displays functional declines in ascorbate transport when expressed in *Xenopus laevis* oocytes (17) (**Figure 3b**). For example, the amino acid substitution encoded by the rs35817838 SNP results in a methionine-to-valine missense mutation at position 258. Oocytes expressing this

**Nonsynonymous:** DNA change in the coding region of the protein that results in an amino acid change

particular variant demonstrate a 75% reduction in the rate of intracellular ascorbate accumulation (**Figure 3b**; A772G). This SNP is found in very low frequency in the general population, primarily in African Americans (**Figure 3c**; A772G). It is currently unclear if there are individuals homozygous for the expression of this *SLC23A1* variant, and its impact on vitamin C homeostasis in humans is unknown.

### Genetic Variation in *SLC23A1* and Vitamin C Status

SVCT1 is considered to play a central role in regulating whole-body vitamin C status and is involved in bulk vitamin C transport throughout the body (**Figure 2**). Although results from *SLC23A1* knockout mice suggest that absorption of ascorbic acid may not be impaired by the loss of SVCT1, the lack of renal reabsorption results in reduced plasma levels of ascorbate. Therefore, genetic variations in *SLC23A1* may lower the renal threshold for vitamin C and increase its urinary excretion, limiting steady-state plasma and body vitamin C levels. The reduced rate of ascorbate transport has been demonstrated in *Xenopus* oocytes as described above (**Figure 3b**). It is expected that individuals carrying these SNPs would have lower plasma ascorbate levels even with high dietary vitamin C intake (**Figure 3d**).

In 2010, two studies were published demonstrating the effects of SNPs in *SLC23A1* on circulating concentrations of vitamin C (**Table 1**). Lower serum ascorbate levels were found in Caucasian and East Asian subjects displaying the rs4257763 SNP variant (9). Similarly, this SNP was associated with lower ascorbate levels in subjects participating in the British Women's Heart and Health Study (BWHHS) (91). Strong trends toward a decline in circulating ascorbate levels were also observed in both aforementioned studies (9, 91) when a different intronic SNP (rs6596473) was examined (**Table 1**). Interestingly, this same SNP was also associated with a decreased risk of follicular lymphoma (82), as further discussed below. Another SNP (rs10063949)

was also found in the BWHHS cohort to be associated with a small but significant increase in circulating ascorbate levels, but this per allele effect was not observed in the European Prospective Investigation into Cancer and Nutrition (EPIC)-Norfolk cohort (85).

The most robust finding for the influence of *SLC23A1* genetic variation on plasma or serum ascorbate levels was found by Timpson et al. (85) in a multiple cohort evaluation of more than 15,000 individuals in the United Kingdom. One of the SNPs examined (rs33972313) was associated with consistent, reproducible declines in circulating ascorbate levels across all study cohorts. As discussed below, this SNP encodes an amino acid change in the SVCT1 protein that results in decreased vitamin C transport activity. Overall, in these human populations, this genetic feature was associated with an approximately 6  $\mu$ M lower plasma or serum ascorbate concentration per allele present. These declines were noted despite differences in vitamin C intake, smoking status, alcohol consumption, geographical location, and socioeconomic status.

The rs33972313 variant is a nonsynonymous SNP in *SLC23A1* that results in a valine-to-methionine substitution at position 264 of SVCT1 (17). The rs33972313 SNP is found in Caucasians, but it is present at a higher frequency in African American and Yoruba African populations (**Figure 3c**; G790A). This variation results in an approximate 50% decline in the rate of ascorbate accumulation in cells (**Figure 3b**). It is postulated that the overall effects of this mutation in humans would increase excretion of ascorbic acid from the body and alter the dose-response relationship between plasma ascorbate levels and dietary vitamin C intake (**Figure 3d**). However, no individuals carrying this variant or homozygous for this allele have been examined to determine the strength of this relationship with vitamin C homeostasis.

To date, the association of *SLC23A1* genetic variation and plasma levels of vitamin C is the strongest evidence for the influence of SVCT1 on human vitamin C status. Since the



regulation of plasma vitamin C homeostasis ultimately influences the availability of ascorbic acid to all tissues and other fluids in the body, it is possible that further associations between these SNPs in *SLC23A1* and physiological function or chronic disease risk will be discovered. However, caution must be taken in such analyses. Associations should not be based merely on genetic variation, but also on measured plasma or serum levels of ascorbic acid, and possibly on urinary excretion rates. Dietary records of vitamin C intake do not adequately reflect vitamin C plasma levels or body status (30, 43) and should not be used to evaluate the possible role of vitamin C in chronic disease risk. Hence, any future studies into phenotypic changes associated with *SLC23A1* genetic variation will require measuring the effects on circulating vitamin C levels. Without this data, it will be difficult to draw any conclusions in relation to vitamin C status.

### Genetic Variation in *SLC23A1* and Chronic Disease Risk

Early studies of genetic variation in *SLC23A1* had focused on associations with disease risk. Because higher dietary intake of vitamin C has been associated with lower risk of cancers of the digestive tract (13), and SVCT1 plays an important role in ascorbic acid uptake from the diet, SNPs in *SLC23A1* may play a role in gastrointestinal cancer development. However, neither incidence of gastric cancer (94) nor advanced colorectal adenoma (28) was related to *SLC23A1* genotype. These results are not surprising given the functional redundancy of vitamin C absorption, as both SVCT1 and SVCT2 are expressed in the digestive tract.

Since many of the genetic variations in *SLC23A1* are strongly associated with decreased plasma vitamin C levels, they might affect cancer development in tissues other than the gastrointestinal tract by altering the supply of ascorbic acid in the circulation. Only one study supports this idea: Skibola et al. (82) observed an increased risk of follicular lymphoma associated with two SNPs (rs6596473

and rs11950646), one of which (rs6596473) is associated with decreased plasma ascorbate levels (9, 85) (**Table 1**). Interestingly, the increased cancer risk was observed in a US but not a German cohort (82). The reason for this discrepancy is not known but may be due to the small population size, lower lymphoma rate in the European cohort, or interactions with diet and environmental exposures that were not assessed in the study.

Higher vitamin C intakes or plasma levels are consistently associated with decreased incidence and risk of death from cardiovascular diseases (30, 32, 47, 65, 90, 95). Furthermore, several studies have shown that vitamin C supplementation and higher plasma ascorbate levels are associated with decreased blood pressure, improved endothelial function, lowered circulating C-reactive protein levels, and increased serum high-density lipoprotein cholesterol levels (5, 7, 31, 44). Given these strong associations with vitamin C status, it is surprising that no studies have investigated the effects of SNPs in *SLC23A1* on cardiovascular function. There is a significant need to determine the impact of genetic variation in the *SLC23* family on the risk of cardiovascular diseases, including coronary artery disease, ischemic stroke, and hypertension.

Taken together, the current data suggest that there is only a weak association, at best, between *SLC23A1* polymorphisms and disease risk. However, it should be noted that in all of the aforementioned studies, plasma levels of ascorbate were not measured, and dietary vitamin C intake was evaluated in only two of the studies (28, 94). It is often assumed that dietary vitamin C intake accurately reflects vitamin C body status, but the presence of SNPs in *SLC23A1* may alter SVCT1-based absorption of vitamin C from the diet. Because steady-state levels of plasma ascorbate are affected by polymorphisms in *SLC23A1* (see above), the supply of ascorbate to tissues may also be affected. This suggests that in studies of chronic disease risk, the influence of *SLC23A1* SNPs must be evaluated with regard to changes in measured, circulating levels of ascorbate.

---

**Environmental exposures:** both man-made and natural toxins present in the environment (smoke, asbestos, heavy metals, etc.)

---

**microRNA:** short, noncoding RNA involved in the regulation of messenger RNA levels

Furthermore, it must be noted that the studies on *SLC23A1* variants and cancer risk were limited to Caucasian populations. The only nonsynonymous SNP in *SLC23A1* found at high frequency in Caucasian populations is the rs33972313 variant, which—as mentioned above—results in a valine-to-methionine substitution at position 264 of SVCT1 and decreased vitamin C transport activity (17). This SNP was not evaluated for cancer risk. Since other changes in the SVCT1 protein are found at greater frequency in African and African American populations (**Figure 3c**), there may be associations between risk for certain diseases and genetic variations in *SLC23A1* that have yet to be discovered.

### SVCT2 (*SLC23A2*)

Encoded by the gene *SLC23A2*, SVCT2 is a vitamin C transport protein found in nearly every tissue and cell of the body (**Figure 2**). SVCT2 is a high-affinity ascorbate transport protein responsible for the accumulation of vitamin C into tissues that have the highest concentrations of ascorbate: the brain, eyes, and adrenals (86). SVCT2 is also the only vitamin C transport protein expressed in leukocytes (70) and platelets (79); however, it is notably absent from erythrocytes (59).

Mouse models in which *SLC23A2* has been ablated survive until birth but die soon afterward from cerebral hemorrhaging and respiratory failure due to lack of lung expansion (83). Thus, it is believed that the expression of SVCT2 and adequate ascorbate levels in the brain are critical for normal fetal development. Studies in animal models and human cell lines support a role of redox signaling in the transcriptional regulation of SVCT2 (69, 70, 80). Intracellular ascorbic acid levels regulate the expression of SVCT2 in many tissues, although the brain appears to be an exception (61).

### Genomic Characterization of *SLC23A2*

*SLC23A2* is approximately 10 times larger than *SLC23A1*, exceeding 150 kbp on chromosome

20 in humans (26, 73). Corresponding to this increased gene size, larger 5' and 3' untranslated regions (UTRs) and wider intronic sequences have been noted in *SLC23A2*. Two distinct promoter regions exist in this gene, allowing for alternative sites for the initiation of transcription and different lengths of mRNA produced (76). *SLC23A2* has 17 exons, with the alternative starting exons labeled exon 1a and exon 1b. Transcription through both exons has been detected in human cells, but exon 1b predominates in mature mRNA transcripts. Transcriptional regulation of both promoter regions has been described through KLF, Sp1, YY1, and factors of the EGR family (71, 72), though the functional aspects of this regulation are unclear. Additional regulation may come in the form of microRNA binding to the 3' UTR of the full *SLC23A2* transcript. A recently published study links the expression of miR-30b and miR-30d with lowered *SLC23A2* expression (33). These microRNAs have been implicated in regulating p53 and catalase expression in response to hydrogen peroxide exposure (38, 50) and may mediate redox regulation of SVCT2 expression.

Due to the relatively large size of the gene, variants within the genetic structure are relatively frequent and widespread. Over 2,200 SNPs have been identified, and at least 250 of those are present in greater than 5% of all populations tested. Published reports have mentioned 30 different SNPs that span the entire length of the gene (26). Although some specific variants have not been detected in different groups, many SNPs are present in individuals of different ethnic backgrounds.

Overall, most of the SNPs analyzed to date fall in intronic or untranslated regions of *SLC23A2* (**Table 2**) and do not directly affect the coding of the SVCT2 protein. SNPs that do lie within the protein coding region of the gene have all been synonymous polymorphisms, suggesting that amino acid changes in SVCT2 are generally unfavorable mutations. However, SNPs not involved in protein coding are not precluded from impacting cellular SVCT2 levels. For example, one relatively rare

**Table 2** Phenotypic effects of single-nucleotide polymorphisms (SNPs) found in the human *SLC23A2* gene

Reference SNP ID	Location	Genetic variant <sup>a</sup>	References	Alterations in phenotype <sup>b</sup>
rs1279683	Intron 1	c.-375–947 C>T	(97)	Associated with decreased plasma ascorbate levels and increased risk of developing glaucoma
rs12479919	Intron 2	c.-282+1312 G>A	(2, 27, 28, 94)	Trend toward decreased risk of gastric cancer; considered part of a high-risk bladder cancer phenotype
rs2681118	Intron 2	c.-282+14050 C>A	(27, 28, 94)	None
rs6139591	Intron 3	c.-155+80 C>T	(9, 27, 28, 94)	Trend toward increased risk of preterm delivery; no association with serum ascorbate levels
rs2681116	Intron 3	c.-155+108 A>G	(9, 27, 28, 94)	None, including no association with serum ascorbate levels
rs13037458	Intron 3	c.-155+224 T>G	(28, 94)	None
rs4813725	Intron 3	c.-154–4623 G>A	(27, 28, 94)	None
rs1776948	Intron 3	c.-154–17751 C>T	(82)	None
rs1715365	Intron 4	c.109–5464 G>A	(27, 28)	None
rs6133175	Intron 5	c.207+1767 T>C	(82)	None
rs1776964	Exon 7	c.375 C>T	(27, 28, 94)	None
rs4987219	Intron 9	c.642+453 G>C	(15, 27, 28)	Decreased risk of head and neck cancer with concurrent HPV infection
rs1110277	Exon 12	c.1002 C>T	(27, 28, 94)	None
rs16990301	Exon 17	c.*2203 G>A	(33)	Limited interactions with regulating microRNA; associated with a decreased expression of SVCT2 in cells
rs35560557	Exon 17	c.*2724 C>T	(27, 28)	None

<sup>a</sup>Variations listed relative to the coding region in the reference sequence NM\_005116.6.

<sup>b</sup>Effects of genetic polymorphism as described in the references provided.

Abbreviations: HPV, human papillomavirus; SVCT, sodium-dependent vitamin C transport protein.

SNP (rs16990301) found in a Yoruba African population appears to influence regulation by the miR-30d microRNA, leading to decreased SVCT2 mRNA levels (33).

### Genetic Variation in *SLC23A2* and Vitamin C Status

Unlike variation in *SLC23A1*, genetic changes in *SLC23A2* are not expected to have a large impact on ascorbate homeostasis in the circulation, as SVCT2 regulates the tissue accumulation of the vitamin from the plasma. To date, only two studies have addressed this question. The first found no correlation between serum ascorbate levels and two common SNPs (rs6139591 and rs2681116) in intron 3

of *SLC23A2* (9). The second study observed significantly lower plasma ascorbate levels in subjects with variants of an SNP in intron 1 (rs1279683) (97). Since this SNP lies in the intronic regions of the gene, it is difficult to explain why it affects circulating ascorbate levels. Because it lies on the 5' end of the gene, it is possible that the rs1279683 polymorphism affects the transcriptional regulation of SVCT2 mRNA.

Studies are lacking on the possible effects of genetic variation in *SLC23A2* on cellular vitamin C status. No studies have looked at SVCT2 protein levels in human cell lines expressing these SNPs, and no individuals carrying these variant alleles have been evaluated

**Haplotype:** a combination of SNPs; haplotype analysis is used to determine additive effects of separate polymorphisms on an individual's phenotype

for tissue ascorbate levels. As indicated above, measuring plasma vitamin C may not be an accurate evaluation of the effect of *SLC23A2* SNPs on the concentration of vitamin C in tissues. Future studies should assess ascorbate levels of circulating leukocytes or other cells, e.g., tissue biopsies or buccal cells, as a surrogate of tissues in subjects with genetic variations in *SLC23A2*.

### Genetic Variation in *SLC23A2* and Chronic Disease Risk

Since vitamin C concentrations in the eye are among the highest in the human body (Figure 2), changes in SVCT2 function may have a strong impact on eye health. Indeed, in a Mediterranean population, a polymorphism found in intron 1 of *SLC23A2* (rs1279683) was strongly associated with an increased risk of developing glaucoma (97). Not only was this association statistically significant after adjusting for many confounding factors that could impact disease risk or alter vitamin C status, but glaucoma patients with the variant allele had the lowest plasma ascorbate levels of all subjects enrolled in the study. Severity of cataracts has also been associated with vitamin C levels in the eye lens in humans. Although some studies have observed increased dietary vitamin C intake and increased blood levels of vitamin C to be associated with decreased risk of cataracts (22, 74, 87), no studies have been performed in conjunction with genetic variation in *SLC23A2*. Hence, this may prove an interesting avenue for future research.

Vitamin C in tissues has been postulated to affect relative cancer risk, but the overall support of this association for *SLC23A2* is limited (Table 2). A variant in *SLC23A2* observed within intron 2 (rs12479919) correlated with a trend ( $p = 0.06$ ) toward a decrease in risk of gastric cancer (94). In addition, genetic variation at this site modified risk for bladder cancer, but only in association with SNPs in 20 other “high-risk” genes (2). An SNP in intron 9 (rs4987129) was associated with a decreased incidence of head and neck cancer in

individuals testing positive for human papilloma virus infection (15). Haplotypes with the rs4987129 SNP and a SNP in exon 11 (rs1110277) revealed that the combination of the two rare alleles reduced the risk of developing advanced colorectal cancer (28). Similarly, other haplotype analyses suggested the involvement of intronic SNPs in a reduced risk of developing gastric cancer and non-Hodgkin's lymphoma (82). It should be noted that in each of these studies investigating different types of cancer, plasma or tissue ascorbate levels were not assessed in association with the SNPs in *SLC23A2*. Therefore, it is not known whether these genetic variants may affect cancer risk due to changes in vitamin C plasma or tissue levels. However, if these genetic variants indeed lead to lower tissue concentrations of vitamin C, a plausible mechanism for the increased cancer risk may be increased expression of hypoxia-inducible factor 1 $\alpha$  (34).

Although not a disease state per se, the presence of *SLC23A2* SNPs in mothers was associated with an increased risk of spontaneous preterm delivery (24–37 weeks gestational age), but the overall effects of the genetic variation were not robust (27).

As mentioned above for *SLC23A1*, a majority of the studies investigating disease risk did not assess plasma ascorbate levels. Since SVCT2 controls the transport of ascorbate from the bloodstream to tissues, circulating levels of ascorbate may influence the risk of disease in tissues. In light of the strong associations between vitamin C levels and vascular function mentioned above and the fact that the heart and vascular endothelium only express SVCT2, it is interesting to note that no studies have examined the impact of genetic variation in *SLC23A2* on cardiovascular disease risk. In addition, no studies have been performed on *SLC23A2* variation and brain function. Because vitamin C is critical for normal brain development and function and is thought to play a role in the prevention of neurodegenerative diseases, behavioral or functional associations with *SLC23A2* SNPs should be addressed in future studies.

## HAPTOGLOBIN

Unlike the SVCT proteins, the interaction between haptoglobin and vitamin C is believed to be indirect, with iron dysregulation and the generation of reactive oxygen species acting as intermediaries. Both increased iron levels and oxidative stress may lead to a decline in circulating ascorbate levels due to its antioxidant activity. Despite this indirect relationship, haptoglobin is the only circulating blood factor for which there is a known, strong relationship between genetic variation and vitamin C homeostasis.

### Function of Haptoglobin

Haptoglobin is a hemoglobin-binding protein synthesized in the liver and released into the circulation. The primary role of haptoglobin is to bind free hemoglobin after it has been released from damaged or lysed erythrocytes. It is believed that this interaction allows haptoglobin to sequester the iron-porphyrin ring and prevent it from reducing molecular oxygen to superoxide radicals and other reactive oxygen species (36). Experimental animals without a functional copy of the haptoglobin gene (*HP*) suffer from extensive oxidative damage in tissues, especially the kidneys, where iron may be released from the porphyrin ring (55).

A majority of the haptoglobin-hemoglobin complex is cleared from the circulation by parenchymal tissue in the liver. To a lesser degree, circulating macrophages also remove this complex from the blood, as the CD136 receptor expressed by these cells can bind to conjugated haptoglobin. This may be an important mechanism for the clearance of haptoglobin-hemoglobin complexes trapped in capillaries of peripheral tissues. In either case, after cellular uptake the complex is subjected to lysosomal degradation and its iron atom is recycled for use in other heme proteins (36).

### Characterization of Human Haptoglobin Genotypes

Found on chromosome 16, the human haptoglobin gene is polymorphic, with two alleles

found in the population at relatively high frequency. The ancestral allele (*HP<sup>1</sup>*) consists of 5 exons, while the variant allele (*HP<sup>2</sup>*) contains 7 exons. The *HP<sup>2</sup>* allele is thought to have emerged by a duplication of exons 3 and 4. The resulting protein has a larger  $\alpha$  chain with a duplication of the cysteine residue that forms intermolecular disulfide bonds (36, 55).

Individuals homozygous for the *HP<sup>1</sup>* allele have traditionally been termed Hp1-1 phenotype. The haptoglobin protein encoded by *HP<sup>1</sup>* is an  $(\alpha\beta)_2$ -tetramer, as the cysteine in the  $\alpha$  chain binds a cysteine residue in an adjacent  $\alpha$  chain (Figure 4). On the other hand, the  $\alpha$  chain encoded by the *HP<sup>2</sup>* allele has an additional cysteine residue that binds two other  $\alpha$  chain cysteine residues. Therefore, heterozygous individuals (Hp2-1) can form haptoglobin proteins with three or more  $\alpha$  chains in a linear polymer: a core of one or more Hp2 proteins that are capped at each end by Hp1 subunits. In contrast, Hp2-2 individuals, homozygous for the *HP<sup>2</sup>* allele, form larger haptoglobin protein complexes. The protein interactions between the cysteine residues in the *HP<sup>2</sup>*  $\alpha$  chains require that these multimeric proteins are cyclical in nature (Figure 4).

The molecular interactions of the different haptoglobin proteins distinguish their genotypes. The haptoglobin protein encoded by *HP<sup>2</sup>* binds poorly to hemoglobin compared to *HP<sup>1</sup>*. Compared to Hp1-1 individuals, individuals expressing the Hp2-2 phenotype have increased circulating levels of iron, possibly free or still bound to the porphyrin ring of hemoglobin. It is believed that this increased redox-active iron can participate in the generation of superoxide and hydroxyl radicals and is associated with increased oxidative damage to lipoproteins in Hp2-2 individuals (36, 55). Thus, it would be expected that plasma vitamin C levels decline accordingly. The Hp2-2 phenotype is particularly detrimental in diabetics, where glycosylated hemoglobin reduces the ability of haptoglobin to bind to the protein. Thus, diabetic individuals carrying the *HP<sup>2</sup>* allele have a significantly increased risk

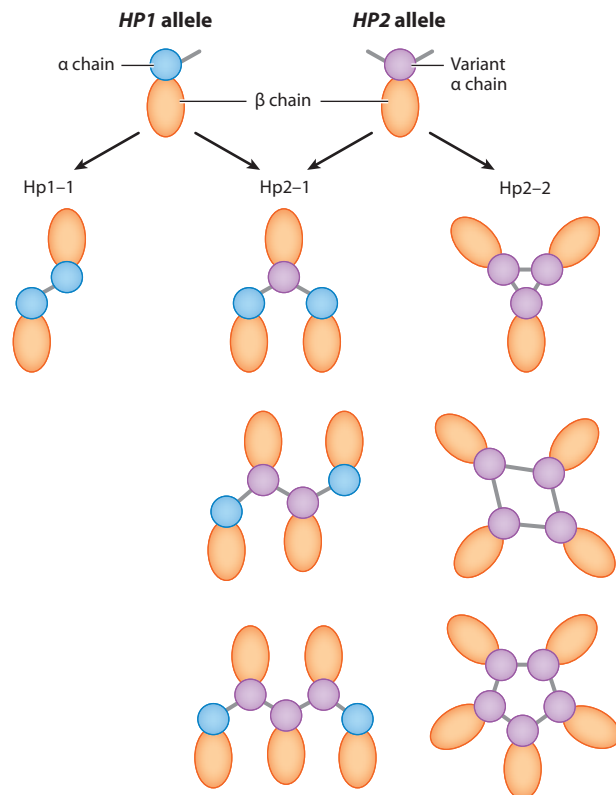
---

**Porphyrin:** a carbon- and nitrogen-based ring that can bind metals; heme (such as found in hemoglobin) is an iron-containing porphyrin

**CD136:** also known as macrophage stimulating 1 receptor, involved in response to inflammatory signals, injury, and invasive growth

---





**Figure 4**

Haptoglobin phenotypes. Haptoglobin is translated as a single polypeptide chain that undergoes posttranslational processing and cleavage to form a light chain ( $\alpha$  subunit) and a heavy chain ( $\beta$  subunit). The  $\alpha$  subunit of haptoglobin also binds to other  $\alpha$  chains through disulfide bridges (55). The haptoglobin protein encoded by *HP<sup>1</sup>* has a cysteine residue in the  $\alpha$  chain that binds a cysteine residue in an adjacent  $\alpha$  chain. The  $\alpha$  chain encoded by the *HP<sup>2</sup>* variant allele has an additional cysteine residue and hence binds two other  $\alpha$  chain cysteine residues. The resulting phenotypes are a combination of those two  $\alpha$  chains forming a tetrameric protein (Hp1-1), or different sizes of linear (Hp2-1) or cyclic (Hp2-2) haptoglobin polymers.

of cardiovascular disease and diabetes-related complications (55).

Beyond this more common allelic variation, there are reports of SNPs in the *HP<sup>2</sup>* carriers that impact circulating haptoglobin levels (18). In addition, there are also individuals who have a genetic loss of haptoglobin (*HP<sup>0</sup>* allele), but these are found in very low frequencies (67).

## Haptoglobin Genetic Variation and Vitamin C Status

As mentioned above, diabetic Hp2-2 individuals demonstrate increased oxidative damage to lipoproteins, which is likely the result of increased redox-active iron levels. In vitro studies have shown that the rate of ascorbic acid oxidation in human plasma containing free hemoglobin is decreased by the addition of haptoglobin from either Hp1-1 or Hp2-2 individuals. This is presumably due to binding of the redox-active iron in the porphyrin ring of hemoglobin by haptoglobin. However, the rate of ascorbate oxidation in the presence of haptoglobin from Hp2-2 individuals is much higher than the oxidation rate observed with the protein isolated from Hp1-1 carriers (51), in agreement with the observation that the Hp2-2 protein binds hemoglobin with weaker affinity.

Several clinical studies have found a relationship between circulating levels of ascorbic acid and haptoglobin phenotype. In two studies, serum ascorbate levels were lower in healthy Hp2-2 subjects compared to both Hp2-1 and Hp1-1 individuals (21, 66). No significant differences in plasma vitamin C concentrations were noted between subjects with the Hp2-1 and Hp1-1 phenotype. The lower serum ascorbate levels in Hp2-2 subjects were correlated with increased serum iron and an overall decrease in circulating levels of the haptoglobin protein. This phenotype-specific decline in plasma ascorbic acid was also demonstrated in individuals with peripheral artery disease (20) and HIV infection (66).

An additional study on haptoglobin phenotype showed no overall difference in plasma ascorbate levels in Hp2-2 individuals versus those carrying the *HP<sup>1</sup>* allele (10). However, Hp2-2 individuals were more likely to have plasma ascorbate levels in the deficient range ( $<11 \mu\text{M}$ ) when consuming less than the US recommended dietary allowance (RDA) of vitamin C compared to individuals with the Hp2-1 or Hp1-1 phenotype. The authors suggested that this is indicative of a significant diet-gene interaction between vitamin C and haptoglobin

### Recommended dietary allowance (RDA):

in the United States the RDA for vitamin C is currently 90 mg/day for men and 75 mg/day for women



phenotype, with Hp2-2 individuals achieving lower steady-state plasma ascorbate concentrations than Hp2-1 or Hp1-1 individuals with the same dietary intake of vitamin C. However, as mentioned above, assessment of dietary intake of vitamin C is problematic, and more carefully controlled studies are needed to establish a relationship between dietary intake of vitamin C, steady-state levels of plasma ascorbate, and haptoglobin phenotype.

It should be noted that most of the health effects of haptoglobin polymorphisms have been determined in individuals with diabetes mellitus (55). However, phenotypic changes in plasma ascorbate levels have been determined primarily in healthy Hp2-2 individuals. It is possible that, under normal conditions, ascorbate provides adequate antioxidant protection to individuals carrying the *HP*<sup>2</sup> allele against oxidative damage to lipoproteins and increased disease risk. It has been speculated that vitamin C supplementation may actually worsen iron-catalyzed oxidative damage in Hp2-2 individuals suffering from diabetes and should be avoided, but experimental evidence supporting this notion is lacking (3).

Furthermore, it is unclear whether the changes in vitamin C status associated with haptoglobin genetic variation are directly related to changes in iron regulation. For example, it is not known whether the haptoglobin phenotype is as pronounced in individuals with low dietary iron intake or chronic anemia. It is also possible that iron dysregulation exacerbates the loss of vitamin C seen with the Hp2-2 phenotype. One study of Hp2-2 individuals from Zimbabwe found that a variant allele in the *TF* gene, encoding the iron-binding protein transferrin, further reduced circulating levels of ascorbate (46). It is also possible that genetic variations in hemoglobin or the CD136 receptor may compound any effects of the haptoglobin phenotype, increasing the levels of circulating redox-active iron. Therefore, diet-gene interactions in Hp2-2 versus Hp1-1 individuals may extend beyond vitamin C regulation and should be examined with respect to iron status as well.

## GLUTATHIONE S-TRANSFERASES

Similar to haptoglobin, most of the interactions between vitamin C and GSTs are thought to be mediated by reactive oxygen species. The genetic variations in GST may also impact ascorbate levels through glutathione status, either by preventing the oxidation of ascorbate by reactive oxygen species—hence “sparing” it—or by recycling ascorbate from its oxidized form, dehydroascorbic acid (DHA).

### Functions of Glutathione S-Transferase Isoforms

GSTs are phase II enzymes involved in the detoxification of harmful electrophilic compounds of endogenous or exogenous origin. This is accomplished by the conjugation of glutathione to a target molecule recognized by the GST enzyme. Several members of the GST family exist, each with different ranges of substrate targets. Based on sequence homology, eight distinct classes of soluble human GSTs have been identified: alpha, kappa, mu, omega, pi, sigma, theta, and zeta. It is believed that the various members emerged over time as a result of a progressive series of gene duplication, recombination, and mutation (39, 60). Inactive GST pseudogenes and repeated DNA sequences lie on chromosomes near active GST members, suggesting the flexible nature of this genetic sequence. This flexibility is also seen in the protein activity, as substrate specificities overlap between isoforms of different GST families.

As part of their response to toxic environmental factors, including the detoxification of carcinogens that otherwise may cause oxidative damage, the GSTs are also induced under conditions of oxidative stress. Alpha-, pi-, mu-, and theta-class GSTs have been implicated in detoxification of numerous reactive lipid oxidation products, such as  $\alpha,\beta$ -unsaturated aldehydes (39). Therefore, it is conceivable that decreased GST activity results in lower levels of ascorbate due to its direct reaction

---

**Transferrin:** an iron-carrying protein found in extracellular fluids

---

with products of lipid peroxidation (14, 64). Furthermore, changes in GST activity may also affect glutathione levels, which in turn may alter ascorbate levels through sparing or recycling in cells and plasma, as explained above (98).

Multiple isoforms exist in the GST mu (*GSTM1*-5) and theta (*GSTT1*-2) families (39). Both *GSTM1* and *GSTT1* have deletion variants occurring in relatively high frequencies in human populations from diverse ethnic backgrounds. These are found on chromosome 1 and chromosome 22, respectively. Other common variants are a missense mutation in *GSTM1* and a deletion mutation in *GSTM3*. The GST pi class consists of a single member encoded by the gene *GSTP1*, found on chromosome 11. Several SNPs have been described in this gene, with the most commonly reported being a missense mutation at position 105, resulting in an isoleucine-to-valine (Ile105Val) amino acid substitution. This mutation reportedly affects the substrate specificity of the protein.

In general, subjects with *GSTM1* and *GSTT1* deletions are considered to be at high risk from exposure to environmental toxins. Since these two isoforms can also recognize lipid peroxidation products as substrates, a loss of activity may predispose the body to increased oxidative stress.

### Human GST Polymorphisms and Vitamin C Status

The relationship between plasma vitamin C and GST genetic variation was first explored in 2001 in a study of middle-aged men in Slovakia, which found that the *GSTM1*-null genotype was associated with increased circulating levels of ascorbate (25). This was also observed in a more recent study in a US cohort, where individuals with the highest vitamin C levels were more likely to be *GSTM1*-null than to have a functional copy of *GSTM1* (6). However, this relationship between vitamin C and the presence of *GSTM1* has not been universally observed. Two studies, one in a Chinese cohort (96) and another with a

mixed population in Canada (11), showed no differences in vitamin C status associated with the *GSTM1* genotype. In addition, a study of factory workers in Slovakia reported an overall decline in plasma vitamin C levels associated with the *GSTM1*-null genotype (42).

Although difficult to explain, these disparate results may reflect a more complex relationship of the *GSTM1* genotype with vitamin C homeostasis, involving, e.g., dietary vitamin C intake and environmental exposures. The *GSTM1*-null genotype appeared to be associated with increased plasma levels of ascorbate when the dietary intake of vitamin C also was high (6, 25). Conversely, the populations with low dietary intake of vitamin C and the *GSTM1* deletion showed lower plasma ascorbate levels than did individuals with functional *GSTM1* at the same intake level (11, 42). This suggests a heretofore uncharacterized interaction between dietary vitamin C, plasma ascorbate, and the activity of GSTs, possibly involving lipid peroxidation products or environmental toxins. There were many different types of environmental exposure in these studies, from cigarette smoking to hazardous working conditions. Together, these data would suggest that further studies on genetic variation in *GSTM1* and vitamin C levels in plasma, tissues, and diet are warranted, with special attention to known substrates for GST enzymes.

*GSTP1* variation has a poorly defined relationship with vitamin C status. One report in middle-aged men from Slovakia showed that individuals heterozygous for the Ile105Val variant had lower circulating ascorbate levels than did those homozygous for the valine substitution (25). However, this relationship with *GSTP1* was not observed in a study of other factory workers in Slovakia (42). A more detailed study of bioavailability of vitamin C in Japanese women showed that individuals heterozygous for the *GSTP1* variant had lower absorption after a single oral dose of ascorbic acid, despite having nearly identical plasma ascorbate levels before the supplementation began (41). This would suggest that *GSTP1* variations may alter vitamin C bioavailability.

An additional study on vitamin C intake in pregnant women showed that *GSTP1* genotype and dietary vitamin C appeared to interact with fetal development and exposure to environmental carcinogens. Although circulating levels of ascorbic acid were not measured in this study, *GSTP1* variation and vitamin C intake were associated with differential effects of benzo(a)pyrene exposure (24). Overall, women with the Ile/Val or Val/Val variants of *GSTP1* exposed to high levels of benzo(a)pyrene were at significantly higher risk of having children with a low birth weight when their vitamin C intakes were low. However, high vitamin C intake negated the increased susceptibility of those carrying the *GSTP1* polymorphism.

In contrast to other GSTs, a loss of *GSTT1* does not appear to impact circulating vitamin C levels in the general population. Although an early study of individuals with the *GSTT1*-null genotype showed a decline in serum ascorbate (25), this was not seen in four other study populations (6, 11, 42, 96). One study reported that in individuals who consumed less than the RDA of vitamin C, *GSTT1* gene deletion may contribute to lower circulating levels of ascorbate (11), but the same was not observed in two other studies (6, 96). On the other hand, isolated lymphocytes from individuals missing a functional *GSTT1* gene were more susceptible to hydrogen peroxide-induced DNA damage (92), suggesting that cellular ascorbate levels, rather than those in the plasma, should be measured as a functional consequence of *GSTT1* disruption.

In general, it appears that interactions between vitamin C levels and genetic variation of GSTs exist, but overall conclusions are difficult to reach. Future studies need to focus on the interactions of GSTs not only with vitamin C bioavailability, but also the interaction of these polymorphisms with each other and with exposure to environmental toxins. Other classes of GSTs should also be examined besides deletions in *GSTM1* and *GSTT1*, or variations in *GSTP1*. Genetic variation in other members of the mu and theta classes of the enzyme might provide further insight into vitamin C interac-

tions. GST omega has recently been postulated to have DHA reductase activity (98), and future work may address the effect of variation in the *GSTO* gene family on vitamin C recycling.

## OTHER GENETIC INTERACTIONS WITH VITAMIN C

### Manganese Superoxide Dismutase Polymorphisms and Vitamin C Status

Superoxide dismutase (SOD) is responsible for the removal of superoxide radicals by conversion to hydrogen peroxide and molecular oxygen and is thus considered one of the major antioxidant enzymes in the body. Three SOD isoforms exist in humans. The isoform found in mitochondria contains manganese as a cofactor (MnSOD) and is encoded by the gene *SOD2* (16). A common polymorphism exists in *SOD2* that causes a missense mutation at codon 16, causing an alanine-to-valine substitution. The valine-containing isoform of MnSOD is thought to affect the mitochondrial targeting sequence (84) and, hence, resistance of mitochondria to oxidative stress.

Although many studies have found associations between vitamin C intake, MnSOD polymorphisms, and cancer risk (1, 37), only one has measured the effect of MnSOD variants on plasma vitamin C levels (23). Interestingly, healthy individuals homozygous for the valine isoform had higher levels of serum ascorbate. Conversely, those individuals with hypercholesterolemia and at least one copy of the valine-containing MnSOD showed a significant decline in serum ascorbate levels compared to subjects homozygous for the alanine variant. Further work is needed to confirm and expand these observations before a definite conclusion on the effect of *SOD2* polymorphisms on vitamin C status can be drawn.

### Glucose Transporters and Vitamin C Transport

In addition to the SVCTs, vitamin C accumulation in cells may occur through the transport of DHA by glucose transporter

(GLUT) proteins. Specifically, GLUT1, GLUT3, and GLUT10, the gene products of *SLC2A1*, *SLC2A3*, and *SLC2A10*, respectively, have been shown to transport DHA (52, 78). Other GLUT proteins, such as GLUT2 and GLUT4, also have been shown to transport DHA, albeit at a lower rate (77, 89). In many cell types, the rate of transport of DHA through GLUTs can exceed the transport of ascorbic acid through the SVCTs (93). Once brought into the intracellular environment, DHA is rapidly reduced by a variety of nicotinamide adenine dinucleotide phosphate (NADPH)- or glutathione-dependent mechanisms to ascorbic acid. Therefore, supplementing cells with DHA can lead to increased intracellular ascorbate. However, DHA is chemically unstable, with a half-life of only a few minutes in aqueous solutions, and usually is not detectable in tissues or plasma except in times of oxidative stress. In addition, animal models unable to express SVCT1 or SVCT2 have severe phenotypes due to ascorbate deficiency (17, 83), demonstrating that GLUT transporters cannot compensate for the lack of SVCT-mediated vitamin C transport. Thus, it is unclear what, if any, contribution GLUT expression makes to cell and tissue vitamin C levels under normal physiological conditions.

Human genetic variation in glucose transport follows a broad range of gene changes in the *SLC2* family. With respect to GLUT variation and vitamin C homeostasis, the evidence is scant. Erythrocytes from individuals with GLUT1 deficiency syndrome demonstrate a decreased ability to accumulate DHA *ex vivo* (48, 49), but whether this affects the concentration of ascorbate in erythrocytes *in vivo* is doubtful. Arterial tortuosity syndrome is a genetic disorder that results in the development of elongated and twisted arteries, including the aorta, and is caused by several loss-of-function mutations in *SLC2A10* (GLUT10). It has been postulated that GLUT10 is a mitochondrial DHA transport protein since mitochondria isolated from murine models expressing one of these *SLC2A10* genetic variants exhibit decreased DHA uptake *in vitro* and increased

oxidative damage (52). However, there is no evidence to date that these genetic changes in GLUT10 result in altered circulating or tissue vitamin C levels in humans. Overall, genetic variation in the various members of the *SLC2* family may affect vitamin C status, but more studies are needed to support this notion.

## CONSEQUENCES OF GENETIC VARIATION

Pharmacokinetic studies have demonstrated that plasma ascorbate rises in a linear fashion with increasing doses of vitamin C until plasma and tissues become saturated and a maximum steady-state level is reached (53, 54) (**Figure 3d**). However, a key aspect to understanding the impact of human genetic variation on vitamin C homeostasis is that the relationship between dietary and plasma vitamin C levels may be altered. It is possible that those individuals expressing particular variant gene products might require more vitamin C in the diet to achieve the same plasma and tissue ascorbate levels as those seen in individuals without these genetic variants. Another possibility is that these individuals simply may not achieve the same maximum steady-state level of plasma ascorbate even when consuming large doses of vitamin C, as described for SVCT1 variants (**Figure 3d**).

It is likely that individuals with the lowest dietary intake of vitamin C are the most susceptible to the effects of genetic variation. The differences in plasma vitamin C levels are relatively small between those considered severely deficient (<11  $\mu\text{M}$ ) and marginally deficient (11–28  $\mu\text{M}$ ). Severe vitamin C deficiency is associated with clinical symptoms of scurvy, such as impaired wound healing and gingivitis, whereas marginally deficient plasma ascorbate levels are associated with early signs of scurvy, including malaise and fatigue, gingival inflammation, and bone abnormalities (30). In the presence of a genetic polymorphism, the number of individuals in the marginally and severely deficient vitamin C categories might be increased compared to other members of

the population without this polymorphism, as postulated for *GST* variations (11).

Numerous epidemiological studies have observed that individuals with the highest plasma ascorbate levels display a reduced risk of cardiovascular diseases and cancer compared to individuals with the lowest plasma ascorbate levels (30). For example, the EPIC-Norfolk study concluded that each 20- $\mu$ M increase in plasma ascorbate level was associated with a 20% decreased risk of all-cause mortality (47) and a 9% relative reduction in risk of heart failure (68). To achieve these benefits, it is commonly recommended that dietary consumption of vitamin C-rich fruits and vegetables should be increased. Individuals with genetic variants might require even higher dietary vitamin C intakes or vitamin C supplements to achieve maximal plasma and tissue ascorbate levels or may be unable to reach the same levels as individuals without these genetic variants, even at very high vitamin C intakes (see **Figure 3d**).

Human genetic variation is a confounding factor in epidemiological studies looking at vitamin C because it increases the variability in response to dietary intake or supplementation of vitamin C and consequently reduces the statistical power to detect significant changes with an intervention. In order to overcome these differences, a further subgrouping of the study population(s) by the polymorphisms described in this review may be necessary to detect specific effects on vitamin C homeostasis or disease risk (30, 56). This has already been observed with haptoglobin, where a beneficial effect of vitamin E supplementation is observed only in diabetics with the Hp2-2 phenotype, who are under increased oxidative stress, but not the Hp1-1 or Hp2-1 phenotypes (88). Future studies conducted without any regard to genetic variation in the population are likely to result in a poor association of vitamin C supplementation with disease prevention.

## SUMMARY

Understanding the complex network of interactions involved in the regulation of vitamin C

homeostasis in humans has proven difficult because data from animal and cell culture studies cannot be easily extrapolated to humans. However, studies into the normal genetic variation in the human population have proven to be a useful guide and, as evidenced by this review, are starting to provide meaningful information about the importance of several factors controlling vitamin C levels in the human body.

Due to the specific and direct interaction with ascorbic acid, genetic variation in the *SLC23* family provides the best information about how alterations in SVCT-mediated vitamin C transport affect vitamin C homeostasis. The SNPs present in the protein-coding region of *SLC23A1*, in particular, reveal the critical relationship of SVCT1 function with plasma ascorbate levels. The profound nature of this genetic effect cannot be overemphasized. Despite contributions from diet, environmental exposure, health status, and lifestyle, a statistically significant decline in steady-state plasma ascorbate levels has been demonstrated for one of these *SLC23A1* polymorphisms (rs33972313), and two others (rs4257763 and rs6596473) showed nonsignificant but still notable reductions in plasma vitamin C levels. It also should be noted that rs4257763 is not a rare allele found only in very specific populations but rather has been found in many study populations. Therefore, there is potential for *SLC23A1* genetic changes to have a widespread impact on human vitamin C homeostasis. In addition, studies on variation in the gene coding SVCT2 strongly suggest an important role of vitamin C in disease prevention, especially in certain cancers and glaucoma. It is likely that the effect of SNPs in *SLC23A2* will be evident in more tissues once more exacting studies are performed.

Although many of these genetic factors have been examined for their influence on vitamin C homeostasis because of their known role in lowering oxidative stress in the body, the observed interactions with vitamin C reinforce the role of ascorbic acid as a critical biological antioxidant in tissues and extracellular fluids. Genetic variation in the haptoglobin gene demonstrates



this relationship directly, as variants in the *HP* gene show direct correlation with iron dysregulation and oxidative stress. The loss of ascorbate from the plasma reflects the increased demands for antioxidant protection the variant proteins have placed on the system. Genetic variation in *SOD2*, leading to changes in MnSOD activity, likely will show similar, nonspecific interactions with vitamin C due to its nature as an antioxidant enzyme.

Similar antioxidant-related interactions between vitamin C and GSTs may occur, but, due to the pluripotent nature of glutathione, the relationship is more complex and currently incompletely understood. This uncertainty reflects the complex interactions that govern vitamin C homeostasis, as a wide range of dietary, environmental, and lifestyle factors affect the interaction between ascorbate, glutathione, and GST activity.

One topic only briefly touched on in this review is the impact that epigenetic regulation of these genes may have on vitamin C status. MicroRNA regulation and methylation-dependent transcription of *SLC23A2* were mentioned above, but other types of epigenetic regulation may also apply to this gene and other genes examined in this review. This type of genetic regulation of vitamin C status goes beyond base-pair changes in DNA sequence and may play an important role in vitamin C homeostasis in development and aging.

## FUTURE DIRECTIONS

Early studies of the human *SLC23* family investigated variations in these genes and chronic disease risk or health outcomes. However, genetic variation is only a single variable among many factors affecting vitamin C homeostasis. We now know that these studies were limited by the nature of these interactions that affect the interplay between dietary, circulating, and tissue levels of vitamin C. Without assessment of vitamin C status, either by measuring plasma, serum, or cellular ascorbic acid concentrations, such genetic studies are likely to result in a null outcome. Measuring dietary

intake of vitamin C is an inaccurate measure of vitamin C body status, due to human recall error, questionnaire validity, and variations in food sources, storage, and preparation that affect vitamin C content (4, 19, 40). Future studies must rely only on direct measurement of circulating or cellular vitamin C levels to accurately reflect vitamin C status. Only when held to this rigorous standard can we expect to find robust interactions between vitamin C-related genetic variation and disease risk.

Furthermore, the need for more mechanistic studies is apparent. For example, changes in GST isoforms appear to have an effect on vitamin C levels in some studies, but whether this is due to increased oxidative stress and, hence, depletion of ascorbate by free-radical scavenging is not clear. More mechanistic studies are needed to better understand the interaction between GST activity and ascorbate function. Although genetic changes in *SLC23A1* present in the coding region of the SVCT1 protein have been linked to a decline in transport activity, we know little about the effects of changes in the noncoding regions of these genes. This is particularly true for SNPs in *SLC23A2*, which show only synonymous codon mutations, intronic polymorphisms, and changes in the untranslated regions of the gene.

The human genetic variations covered in this review are by no means an exhaustive list of factors that can influence vitamin C homeostasis. Partially, this is due to the manner in which these genes were discovered. In the various studies reviewed, genetic variants in *HP*, *GSTs*, and *SOD2* were not associated a priori with ascorbic acid but rather were examined because of their known role in oxidative stress. Because SNPs in each of these genes were found to be related to ascorbate status, these findings not only confirm the role of vitamin C as a powerful antioxidant in the human body but also suggest that genetic variations of other antioxidant or oxidative stress-related genes may affect vitamin C status. For example, genes other than *GSTs* involved in glutathione homeostasis, such as glutathione peroxidase (*GPX*), glutathione reductase (*GR*),



glutamate-cysteine ligase (*GCLC* or *GCLM*), or the transcription factor Nrf2 (*NFE2L2*), should be examined. Interactions of polymorphisms in some of these genes with vitamin C (12, 81) have already been proposed to affect lung function. Other obvious candidates to explore for a possible role in vitamin C homeostasis include *SOD1* and *SOD3* and genes involved in iron and heme metabolism, such as transferrin (*TF*), ferritin (*FT*), and heme oxygenase (*HO*). Perhaps with a focus on changes in vitamin C homeostasis, an exploration of human genetic variation will reveal more genes and enzymes that may influence ascorbate levels in the body.

Along these lines, we have only scratched the surface of the protein-gene or RNA interactions for many of the genes discussed. We do not know if polymorphisms in the untranslated regions of the *SLC23* genes lead to a change in gene transcription due to changes in transcription factor binding, for instance. On the other hand, polymorphisms in the transcription factors themselves might lead to a change in expression. A genetic alteration in *HNF1 $\alpha$* , found in patients with mature-onset diabetes of the young (MODY) 3, would be expected to lead to a substantial decline in *SVCT1* synthesis and the renal threshold for vitamin C reabsorption (63). As *HNF4 $\alpha$*  is an upstream regulating factor of *HNF1 $\alpha$* , the former is also a potential target for *SVCT1* regulation. If indeed genetic variants of *HNF1A* and *HNF4A* are found to

play a role in vitamin C homeostasis, this would suggest that vitamin C is regulated in the body as a carbohydrate in addition to an antioxidant. This intriguing approach to vitamin C genetic regulation has never been examined.

What is the overall impact of human genetic variation in light of the myriad of factors that affect vitamin C homeostasis? The answer to this question is not immediately forthcoming. However, it is becoming increasingly clear that the optimum intake of vitamin C to support normal physiological function and promote good health differs widely between individuals and hence needs to be addressed on an individual basis. Not only are vitamin C levels influenced by age, diet, lifestyle, and various environmental exposures, but also—as evident from this review—by multiple genetic variations. These genetic variations may modify the relationships between ascorbic acid and each of the other variables, raising intriguing new questions. For example, do *GST* deletions affect cellular ascorbate levels differently in smokers and nonsmokers? Are haptoglobin polymorphisms an important determinant of vitamin C levels in the presence of iron deficiency? Do changes in *SLC23A2* result in an altered relationship between dietary intake and plasma and tissue levels of vitamin C? Only as our knowledge of vitamin C homeostasis grows can we make these more complex associations to determine their effects on human health and disease.

---

**Mature-onset diabetes of the young (MODY):** a hereditary form of diabetes mellitus. MODY3 is associated with mutations in *HNF1A*

---

## DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

## LITERATURE CITED

1. Ambrosone CB, Freudenheim JL, Thompson PA, Bowman E, Vena JE, et al. 1999. Manganese superoxide dismutase (MnSOD) genetic polymorphisms, dietary antioxidants, and risk of breast cancer. *Cancer Res.* 59:602–6
2. Andrew AS, Gui J, Sanderson AC, Mason RA, Morlock EV, et al. 2009. Bladder cancer SNP panel predicts susceptibility and survival. *Hum. Genet.* 125:527–39
3. Asleh R, Levy AP. 2010. Divergent effects of alpha-tocopherol and vitamin C on the generation of dysfunctional HDL associated with diabetes and the Hp 2-2 genotype. *Antioxid. Redox Signal.* 12:209–17

4. Birlouez-Aragon I, Saavedra G, Tessier FJ, Galinier A, Ait-Ameur L, et al. 2010. A diet based on high-heat-treated foods promotes risk factors for diabetes mellitus and cardiovascular diseases. *Am. J. Clin. Nutr.* 91:1220–26
5. Block G, Jensen CD, Dalvi TB, Norkus EP, Hudes M, et al. 2009. Vitamin C treatment reduces elevated C-reactive protein. *Free Radic. Biol. Med.* 46:70–77
6. Block G, Shaikh N, Jensen CD, Volberg V, Holland N. 2011. Serum vitamin C and other biomarkers differ by genotype of phase 2 enzyme genes *GSTM1* and *GSTT1*. *Am. J. Clin. Nutr.* 94:929–37
7. Boekholdt SM, Meuwese MC, Day NE, Luben R, Welch A, et al. 2006. Plasma concentrations of ascorbic acid and C-reactive protein, and risk of future coronary artery disease, in apparently healthy men and women: the EPIC-Norfolk prospective population study. *Br. J. Nutr.* 96:516–22
8. Boyer JC, Campbell CE, Sigurdson WJ, Kuo SM. 2005. Polarized localization of vitamin C transporters, SVCT1 and SVCT2, in epithelial cells. *Biochem. Biophys. Res. Commun.* 334:150–56
9. Cahill LE, El-Sohehy A. 2009. Vitamin C transporter gene polymorphisms, dietary vitamin C and serum ascorbic acid. *J. Nutrigenet. Nutrigenomics* 2:292–301
10. Cahill LE, El-Sohehy A. 2010. Haptoglobin genotype modifies the association between dietary vitamin C and serum ascorbic acid deficiency. *Am. J. Clin. Nutr.* 92:1494–500
11. Cahill LE, Fontaine-Bisson B, El-Sohehy A. 2009. Functional genetic variants of glutathione S-transferase protect against serum ascorbic acid deficiency. *Am. J. Clin. Nutr.* 90:1411–17
12. Canova C, Dunster C, Kelly FJ, Minelli C, Shah PL, et al. 2012. PM10-induced hospital admissions for asthma and chronic obstructive pulmonary disease: the modifying effect of individual characteristics. *Epidemiology* 23:607–15
13. Carr AC, Frei B. 1999. Toward a new recommended dietary allowance for vitamin C based on antioxidant and health effects in humans. *Am. J. Clin. Nutr.* 69:1086–107
14. Chavez J, Chung WG, Miranda CL, Singhal M, Stevens JF, Maier CS. 2010. Site-specific protein adducts of 4-hydroxy-2(E)-nonenal in human THP-1 monocytic cells: Protein carbonylation is diminished by ascorbic acid. *Chem. Res. Toxicol.* 23:37–47
15. Chen AA, Marsit CJ, Christensen BC, Houseman EA, McClean MD, et al. 2009. Genetic variation in the vitamin C transporter, *SLC23A2*, modifies the risk of HPV16-associated head and neck cancer. *Carcinogenesis* 30:977–81
16. Church SL, Grant JW, Meese EU, Trent JM. 1992. Sublocalization of the gene encoding manganese superoxide dismutase (MnSOD/SOD2) to 6q25 by fluorescence in situ hybridization and somatic cell hybrid mapping. *Genomics* 14:823–25
17. Corpe CP, Tu H, Eck P, Wang J, Faulhaber-Walter R, et al. 2010. Vitamin C transporter *Slc23a1* links renal reabsorption, vitamin C tissue accumulation, and perinatal survival in mice. *J. Clin. Invest.* 120:1069–83
18. Cox SE, Doherty C, Atkinson SH, Nweneka CV, Fulford AJ, et al. 2007. Haplotype association between haptoglobin (Hp2) and Hp promoter SNP (A-61C) may explain previous controversy of haptoglobin and malaria protection. *PLoS ONE* 2:e362
19. Dehghan M, Akhtar-Danesh N, McMillan CR, Thabane L. 2007. Is plasma vitamin C an appropriate biomarker of vitamin C intake? A systematic review and meta-analysis. *Nutr. J.* 6:41
20. Delanghe J, Langlois M, Duprez D, De Buyzere M, Clement D. 1999. Haptoglobin polymorphism and peripheral arterial occlusive disease. *Atherosclerosis* 145:287–92
21. Delanghe JR, Langlois MR, Boelaert JR, Van Acker J, Van Wanzeele F, et al. 1998. Haptoglobin polymorphism, iron metabolism and mortality in HIV infection. *AIDS* 12:1027–32
22. Dherani M, Murthy GV, Gupta SK, Young IS, Maraini G, et al. 2008. Blood levels of vitamin C, carotenoids and retinol are inversely associated with cataract in a North Indian population. *Investig. Ophthalmol. Vis. Sci.* 49:3328–35
23. Duarte MM, Moresco RN, Duarte T, Santi A, Bagatini MD, et al. 2010. Oxidative stress in hypercholesterolemia and its association with Ala16Val superoxide dismutase gene polymorphism. *Clin. Biochem.* 43:1118–23
24. Duarte-Salles T, Mendez MA, Morales E, Bustamante M, Rodriguez-Vicente A, et al. 2012. Dietary benzo(a)pyrene and fetal growth: effect modification by vitamin C intake and glutathione S-transferase P1 polymorphism. *Environ. Int.* 45:1–8

---

17. Creation of the *SLC23A1* knockout mouse, linking renal reabsorption to SVCT1 function.

---

25. Dusinska M, Ficek A, Horska A, Raslova K, Petrovska H, et al. 2001. Glutathione S-transferase polymorphisms influence the level of oxidative DNA damage and antioxidant protection in humans. *Mutat. Res.* 482:47–55
26. Eck P, Erichsen HC, Taylor JG, Yeager M, Hughes AL, et al. 2004. Comparison of the genomic structure and variation in the two human sodium-dependent vitamin C transporters, *SLC23A1* and *SLC23A2*. *Hum. Genet.* 115:285–94
27. Erichsen HC, Engel SA, Eck PK, Welch R, Yeager M, et al. 2006. Genetic variation in the sodium-dependent vitamin C transporters, *SLC23A1*, and *SLC23A2* and risk for preterm delivery. *Am. J. Epidemiol.* 163:245–54
28. Erichsen HC, Peters U, Eck P, Welch R, Schoen RE, et al. 2008. Genetic variation in sodium-dependent vitamin C transporters *SLC23A1* and *SLC23A2* and risk of advanced colorectal adenoma. *Nutr. Cancer* 60:652–59
29. Fischer H, Schwarzer C, Illek B. 2004. Vitamin C controls the cystic fibrosis transmembrane conductance regulator chloride channel. *Proc. Natl. Acad. Sci. USA* 101:3691–96
30. Frei B, Birlouez-Aragon I, Lykkesfeldt J. 2012. Authors' perspective: What is the optimum intake of vitamin C in humans? *Crit. Rev. Food Sci. Nutr.* 52:815–29
31. Frikke-Schmidt H, Lykkesfeldt J. 2009. Role of marginal vitamin C deficiency in atherogenesis: in vivo models and clinical studies. *Basic Clin. Pharmacol. Toxicol.* 104:419–33
32. Gale CR, Martyn CN, Winter PD, Cooper C. 1995. Vitamin C and risk of death from stroke and coronary heart disease in cohort of elderly people. *BMJ* 310:1563–66
33. Gamazon ER, Ziliak D, Im HK, LaCroix B, Park DS, et al. 2012. Genetic architecture of microRNA expression: implications for the transcriptome and complex traits. *Am. J. Hum. Genet.* 90:1046–63
34. Gao P, Zhang H, Dinavahi R, Li F, Xiang Y, et al. 2007. HIF-dependent antitumorigenic effect of antioxidants in vivo. *Cancer Cell* 12:230–38
35. Goldenberg H, Schweinzer E. 1994. Transport of vitamin C in animal and human cells. *J. Bioenerg. Biomembr.* 26:359–67
36. Goldenstein H, Levy NS, Levy AP. 2012. Haptoglobin genotype and its role in determining heme-iron mediated vascular disease. *Pharmacol. Res.* 66:1–6
37. Han J, Colditz GA, Hunter DJ. 2007. Manganese superoxide dismutase polymorphism and risk of skin cancer (United States). *Cancer Causes Control* 18:79–89
38. Haque R, Chun E, Howell JC, Sengupta T, Chen D, Kim H. 2012. MicroRNA-30b-mediated regulation of catalase expression in human ARPE-19 cells. *PLoS ONE* 7:e4 2542
39. Hayes JD, Flanagan JU, Jowsey IR. 2005. Glutathione transferases. *Annu. Rev. Pharmacol. Toxicol.* 45:51–88
40. Henriquez-Sanchez P, Sanchez-Villegas A, Doreste-Alonso J, Ortiz-Andrellucchi A, Pfrimer K, Serra-Majem L. 2009. Dietary assessment methods for micronutrient intake: a systematic review on vitamins. *Br. J. Nutr.* 102(Suppl. 1):S10–37
41. Higasa S, Tsujimura M, Hiraoka M, Nakayama K, Yanagisawa Y, et al. 2007. Polymorphism of glutathione S-transferase P1 gene affects human vitamin C metabolism. *Biochem. Biophys. Res. Commun.* 364:708–13
42. Horska A, Mislanova C, Bonassi S, Ceppi M, Volkovova K, Dusinska M. 2011. Vitamin C levels in blood are influenced by polymorphisms in glutathione S-transferases. *Eur. J. Nutr.* 50:437–46
43. Jenab M, Riboli E, Ferrari P, Sabate J, Slimani N, et al. 2006. Plasma and dietary vitamin C levels and risk of gastric cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST). *Carcinogenesis* 27:2250–57
44. Juraschek SP, Guallar E, Appel LJ, Miller ER 3rd. 2012. Effects of vitamin C supplementation on blood pressure: a meta-analysis of randomized controlled trials. *Am. J. Clin. Nutr.* 95:1079–88
45. Kang JS, Kim HN, Jung DJ, Kim JE, Mun GH, et al. 2007. Regulation of UVB-induced IL-8 and MCP-1 production in skin keratinocytes by increasing vitamin C uptake via the redistribution of SVCT-1 from the cytosol to the membrane. *J. Investig. Dermatol.* 127:698–706
46. Kasvosve I, Delanghe JR, Gomo ZA, Gangaidzo IT, Khumalo H, et al. 2002. Effect of transferrin polymorphism on the metabolism of vitamin C in Zimbabwean adults. *Am. J. Clin. Nutr.* 75:321–25

---

25. First association of GST polymorphisms with ascorbic acid levels in humans.

---



---

26. An in-depth genetic characterization of *SLC23A1* and *SLC23A2*, including the first mention of genetic variants.

---



---

30. Extensive review of the factors involved in vitamin C research in humans, including evaluation of current RDAs.

---

---

51. First association with the Hp2-2 phenotype and decreased levels of plasma ascorbate.

---



---

62. A broad overview of vitamin C, with a review of ascorbate chemistry, biology, nutrition, and health effects of supplementation.

---

47. Khaw KT, Bingham S, Welch A, Luben R, Wareham N, et al. 2001. Relation between plasma ascorbic acid and mortality in men and women in EPIC-Norfolk prospective study: a prospective population study. European Prospective Investigation into Cancer and Nutrition. *Lancet* 357:657–63
48. Klepper J, Vera JC, De Vivo DC. 1998. Deficient transport of dehydroascorbic acid in the glucose transporter protein syndrome. *Ann. Neurol.* 44:286–87
49. Klepper J, Wang D, Fischbarg J, Vera JC, Jarjour IT, et al. 1999. Defective glucose transport across brain tissue barriers: a newly recognized neurological syndrome. *Neurochem. Res.* 24:587–94
50. Kumar M, Lu Z, Takwi AA, Chen W, Callander NS, et al. 2011. Negative regulation of the tumor suppressor p53 gene by microRNAs. *Oncogene* 30:843–53
51. Langlois MR, Delanghe JR, De Buyzere ML, Bernard DR, Ouyang J. 1997. Effect of haptoglobin on the metabolism of vitamin C. *Am. J. Clin. Nutr.* 66:606–10
52. Lee YC, Huang HY, Chang CJ, Cheng CH, Chen YT. 2010. Mitochondrial GLUT10 facilitates dehydroascorbic acid import and protects cells against oxidative stress: mechanistic insight into arterial tortuosity syndrome. *Hum. Mol. Genet.* 19:3721–33
53. Levine M, Conry-Cantilena C, Wang Y, Welch RW, Washko PW, et al. 1996. Vitamin C pharmacokinetics in healthy volunteers: evidence for a recommended dietary allowance. *Proc. Natl. Acad. Sci. USA* 93:3704–9
54. Levine M, Wang Y, Padayatty SJ, Morrow J. 2001. A new recommended dietary allowance of vitamin C for healthy young women. *Proc. Natl. Acad. Sci. USA* 98:9842–46
55. Levy AP, Asleh R, Blum S, Levy NS, Miller-Lotan R, et al. 2010. Haptoglobin: basic and clinical aspects. *Antioxid. Redox Signal.* 12:293–304
56. Lykkesfeldt J, Poulsen HE. 2010. Is vitamin C supplementation beneficial? Lessons learned from randomised controlled trials. *Br. J. Nutr.* 103:1251–59
57. Macias RI, Hierro C, de Juan SC, Jimenez F, Gonzalez-San Martin F, Marin JJ. 2011. Hepatic expression of sodium-dependent vitamin C transporters: ontogeny, subtissular distribution and effect of chronic liver diseases. *Br. J. Nutr.* 106:1814–25
58. Maulen NP, Henriquez EA, Kempe S, Carcamo JG, Schmid-Kotsas A, et al. 2003. Up-regulation and polarized expression of the sodium-ascorbic acid transporter SVCT1 in post-confluent differentiated CaCo-2 cells. *J. Biol. Chem.* 278:9035–41
59. May JM, Qu ZC, Qiao H, Koury MJ. 2007. Maturation loss of the vitamin C transporter in erythrocytes. *Biochem. Biophys. Res. Commun.* 360:295–98
60. McIlwain CC, Townsend DM, Tew KD. 2006. Glutathione S-transferase polymorphisms: cancer incidence and therapy. *Oncogene* 25:1639–48
61. Meredith ME, Harrison FE, May JM. 2011. Differential regulation of the ascorbic acid transporter SVCT2 during development and in response to ascorbic acid depletion. *Biochem. Biophys. Res. Commun.* 414:737–42
62. Michels AJ, Frei BB. 2012. Vitamin C. In *Biochemical, Physiological, and Molecular Aspects of Human Nutrition*, ed. MH Stipanuk, MA Caudill, pp. 626–54. St. Louis, Mo.: Elsevier/Saunders
63. Michels AJ, Hagen TM. 2009. Hepatocyte nuclear factor 1 is essential for transcription of sodium-dependent vitamin C transporter protein 1. *Am. J. Physiol. Cell Physiol.* 297:C1220–27
64. Miranda CL, Reed RL, Kuiper HC, Alber S, Stevens JF. 2009. Ascorbic acid promotes detoxification and elimination of 4-hydroxy-2(E)-nonenal in human monocytic THP-1 cells. *Chem. Res. Toxicol.* 22:863–74
65. Myint PK, Luben RN, Welch AA, Bingham SA, Wareham NJ, Khaw KT. 2008. Plasma vitamin C concentrations predict risk of incident stroke over 10 y in 20 649 participants of the European Prospective Investigation into Cancer Norfolk prospective population study. *Am. J. Clin. Nutr.* 87:64–69
66. Na N, Delanghe JR, Taes YE, Torck M, Baeyens WR, Ouyang J. 2006. Serum vitamin C concentration is influenced by haptoglobin polymorphism and iron status in Chinese. *Clin. Chim. Acta* 365:319–24
67. Park KU, Song J, Kim JQ. 2004. Haptoglobin genotypic distribution (including Hp0 allele) and associated serum haptoglobin concentrations in Koreans. *J. Clin. Pathol.* 57:1094–95
68. Pfister R, Sharp SJ, Luben R, Wareham NJ, Khaw KT. 2011. Plasma vitamin C predicts incident heart failure in men and women in European Prospective Investigation into Cancer and Nutrition—Norfolk prospective study. *Am. Heart. J.* 162:246–53

69. Portugal CC, da Encarnacao TG, Socodato R, Moreira SR, Brudzewsky D, et al. 2012. Nitric oxide modulates sodium vitamin C transporter 2 (SVCT-2) protein expression via protein kinase G (PKG) and nuclear factor- $\kappa$ B (NF- $\kappa$ B). *J. Biol. Chem.* 287:3860–72
70. Qiao H, May JM. 2009. Macrophage differentiation increases expression of the ascorbate transporter (SVCT2). *Free Radic. Biol. Med.* 46:1221–32
71. Qiao H, May JM. 2011. CpG methylation at the USF-binding site mediates cell-specific transcription of human ascorbate transporter SVCT2 exon 1a. *Biochem. J.* 440:73–84
72. Qiao H, May JM. 2011. Regulation of the human ascorbate transporter SVCT2 exon 1b gene by zinc-finger transcription factors. *Free Radic. Biol. Med.* 50:1196–209
73. Rajan DP, Huang W, Dutta B, Devoe LD, Leibach FH, et al. 1999. Human placental sodium-dependent vitamin C transporter (SVCT2): molecular cloning and transport function. *Biochem. Biophys. Res. Commun.* 262:762–68
74. Ravindran RD, Vashist P, Gupta SK, Young IS, Maraini G, et al. 2011. Inverse association of vitamin C with cataract in older people in India. *Ophthalmology* 118:1958–65.e2
75. Reidling JC, Rubin SA. 2011. Promoter analysis of the human ascorbic acid transporters SVCT1 and 2: mechanisms of adaptive regulation in liver epithelial cells. *J. Nutr. Biochem.* 22:344–50
76. Rubin SA, Dey S, Reidling JC. 2005. Functional analysis of two regulatory regions of the human Na<sup>+</sup>-dependent vitamin C transporter 2, SLC23A2, in human vascular smooth muscle cells. *Biochim. Biophys. Acta* 1732:76–81
77. Rumsey SC, Daruwala R, Al-Hasani H, Zarnowski MJ, Simpson IA, Levine M. 2000. Dehydroascorbic acid transport by GLUT4 in *Xenopus* oocytes and isolated rat adipocytes. *J. Biol. Chem.* 275:28246–53
78. Rumsey SC, Kwon O, Xu GW, Burant CF, Simpson I, Levine M. 1997. Glucose transporter isoforms GLUT1 and GLUT3 transport dehydroascorbic acid. *J. Biol. Chem.* 272:18982–89
79. Savini I, Catani MV, Arnone R, Rossi A, Frega G, et al. 2007. Translational control of the ascorbic acid transporter SVCT2 in human platelets. *Free Radic. Biol. Med.* 42:608–16
80. Savini I, Rossi A, Catani MV, Ceci R, Avigliano L. 2007. Redox regulation of vitamin C transporter SVCT2 in C2C12 myotubes. *Biochem. Biophys. Res. Commun.* 361:385–90
81. Siedlinski M, Postma DS, van Diemen CC, Blokstra A, Smit HA, Boezen HM. 2008. Lung function loss, smoking, vitamin C intake, and polymorphisms of the glutamate-cysteine ligase genes. *Am. J. Respir. Crit. Care Med.* 178:13–19
82. Skibola CF, Bracci PM, Halperin E, Nieters A, Hubbard A, et al. 2008. Polymorphisms in the estrogen receptor 1 and vitamin C and matrix metalloproteinase gene families are associated with susceptibility to lymphoma. *PLoS ONE* 3:e2816
83. Sotiriou S, Gispert S, Cheng J, Wang Y, Chen A, et al. 2002. Ascorbic-acid transporter Slc23a1 is essential for vitamin C transport into the brain and for perinatal survival. *Nat. Med.* 8:514–17
84. Sutton A, Imbert A, Igoudjil A, Descatoire V, Cazanave S, et al. 2005. The manganese superoxide dismutase Ala16Val dimorphism modulates both mitochondrial import and mRNA stability. *Pharmacogenet. Genomics* 15:311–19
85. Timpson NJ, Forouhi NG, Brion MJ, Harbord RM, Cook DG, et al. 2010. Genetic variation at the *SLC23A1* locus is associated with circulating concentrations of L-ascorbic acid (vitamin C): evidence from 5 independent studies with >15,000 participants. *Am. J. Clin. Nutr.* 92:375–82
86. Tsukaguchi H, Tokui T, Mackenzie B, Berger UV, Chen XZ, et al. 1999. A family of mammalian Na<sup>+</sup>-dependent L-ascorbic acid transporters. *Nature* 399:70–75
87. Valero MP, Fletcher AE, De Stavola BL, Vioque J, Alepuz VC. 2002. Vitamin C is associated with reduced risk of cataract in a Mediterranean population. *J. Nutr.* 132:1299–306
88. Vardi M, Levy AP. 2012. Is it time to screen for the haptoglobin genotype to assess the cardiovascular risk profile and vitamin E therapy responsiveness in patients with diabetes? *Curr. Diab. Rep.* 12:274–79
89. Vera JC, Rivas CI, Fischbarg J, Golde DW. 1993. Mammalian facilitative hexose transporters mediate the transport of dehydroascorbic acid. *Nature* 364:79–82

---

**83. Development of the SVCT2 knockout mouse, originally named *SLC23A1*, linking perinatal survival with ascorbate levels.**

---



---

**85. First association of *SLC23A1* polymorphisms with plasma or serum ascorbate levels in human volunteers.**

---



---

**86. First characterization of the genes *SLC23A1* and *SLC23A2* and their function as vitamin C transporters.**

---



90. Voko Z, Hollander M, Hofman A, Koudstaal PJ, Breteler MM. 2003. Dietary antioxidants and the risk of ischemic stroke: the Rotterdam Study. *Neurology* 61:1273–75
91. Wang H, Dutta B, Huang W, Devoe LD, Leibach FH, et al. 1999. Human Na(+)-dependent vitamin C transporter 1 (hSVCT1): primary structure, functional characteristics and evidence for a non-functional splice variant. *Biochim. Biophys. Acta* 1461:1–9
92. Wilms LC, Claughton TA, de Kok TM, Kleinjans JC. 2007. GSTM1 and GSTT1 polymorphism influences protection against induced oxidative DNA damage by quercetin and ascorbic acid in human lymphocytes in vitro. *Food Chem. Toxicol.* 45:2592–96
93. Wilson JX. 2005. Regulation of vitamin C transport. *Annu. Rev. Nutr.* 25:105–25
94. Wright ME, Andreotti G, Lissowska J, Yeager M, Zatonski W, et al. 2009. Genetic variation in sodium-dependent ascorbic acid transporters and risk of gastric cancer in Poland. *Eur. J. Cancer* 45:1824–30
95. Ye Z, Song H. 2008. Antioxidant vitamins intake and the risk of coronary heart disease: meta-analysis of cohort studies. *Eur. J. Cardiovasc. Prev. Rehabil.* 15:26–34
96. Yuan LH, Meng LP, Ma WW, Li S, Feng JF, et al. 2012. The role of glutathione S-transferase M1 and T1 gene polymorphisms and fruit and vegetable consumption in antioxidant parameters in healthy subjects. *Br. J. Nutr.* 107:928–33
97. Zanon-Moreno V, Ciancotti-Olivares L, Asencio J, Sanz P, Ortega-Azorin C, et al. 2011. Association between a *SLC23A2* gene variation, plasma vitamin C levels, and risk of glaucoma in a Mediterranean population. *Mol. Vis.* 17:2997–3004
98. Zhou H, Brock J, Liu D, Board PG, Oakley AJ. 2012. Structural insights into the dehydroascorbate reductase activity of human omega-class glutathione transferases. *J. Mol. Biol.* 420:190–203





# Contents

Functional Organization of Neuronal and Humoral Signals Regulating Feeding Behavior <i>Gary J. Schwartz and Lori M. Zeltser</i> .....	1
Extrarenal Vitamin D Activation and Interactions Between Vitamin D <sub>2</sub> , Vitamin D <sub>3</sub> , and Vitamin D Analogs <i>Glenville Jones</i> .....	23
Human Genetic Variation Influences Vitamin C Homeostasis by Altering Vitamin C Transport and Antioxidant Enzyme Function <i>Alexander J. Michels, Tory M. Hagen, and Balz Frei</i> .....	45
Vitamin D Biology Revealed Through the Study of Knockout and Transgenic Mouse Models <i>Sylvia Christakos, Tanya Seth, Jennifer Hirsch, Angela Porta, Anargyros Moulas, and Puneet Dhawan</i> .....	71
Vitamin E Trafficking in Neurologic Health and Disease <i>Lynn Ulatowski and Danny Manor</i> .....	87
Cocoa and Human Health <i>Samantha Ellam and Gary Williamson</i> .....	105
Novel Aspects of Dietary Nitrate and Human Health <i>Eddie Weitzberg and Jon O. Lundberg</i> .....	129
Prevention of Chronic Diseases by Tea: Possible Mechanisms and Human Relevance <i>Chung S. Yang and Jungil Hong</i> .....	161
Nutrient Deficiencies After Gastric Bypass Surgery <i>Edward Saltzman and J. Philip Karl</i> .....	183
O-GlcNAc Cycling: A Link Between Metabolism and Chronic Disease <i>Michelle R. Bond and John A. Hanover</i> .....	205
Omega-3 Fatty Acids, Hepatic Lipid Metabolism, and Nonalcoholic Fatty Liver Disease <i>E. Scorletti and C.D. Byrne</i> .....	231
Fatty Acid-Regulated Transcription Factors in the Liver <i>Donald B. Jump, Sasmita Tripathy, and Christopher M. Depner</i> .....	249

Iron Nutrition and Premenopausal Women: Effects of Poor Iron Status on Physical and Neuropsychological Performance <i>James P. McClung and Laura E. Murray-Kolb</i> .....	271
Evolutionary Perspectives on the Obesity Epidemic: Adaptive, Maladaptive, and Neutral Viewpoints <i>John R. Speakman</i> .....	289
Dietary Fat in Breast Cancer Survival <i>Nour Makarem, Urmila Chandran, Elisa V. Bandera, and Niyati Parekh</i> .....	319
Quantifying Diet for Nutrigenomic Studies <i>Katherine L. Tucker, Caren E. Smith, Chao-Qiang Lai, and Jose M. Ordovas</i> .....	349
The Role of Cost-Effectiveness Analysis in Developing Nutrition Policy <i>Linda J. Cobiac, Lennert Veerman, and Theo Vos</i> .....	373
African Lessons on Climate Change Risks for Agriculture <i>Christoph Müller</i> .....	395
Making, Baking, and Breaking: The Synthesis, Storage, and Hydrolysis of Neutral Lipids <i>Kelly V. Ruggles, Aaron Turkish, and Stephen L. Sturley</i> .....	413

## Indexes

Cumulative Index of Contributing Authors, Volumes 29–33 .....	453
Cumulative Index of Article Titles, Volumes 29–33 .....	456

## Errata

An online log of corrections to *Annual Review of Nutrition* articles may be found at  
<http://nutr.annualreviews.org/errata.shtml>