



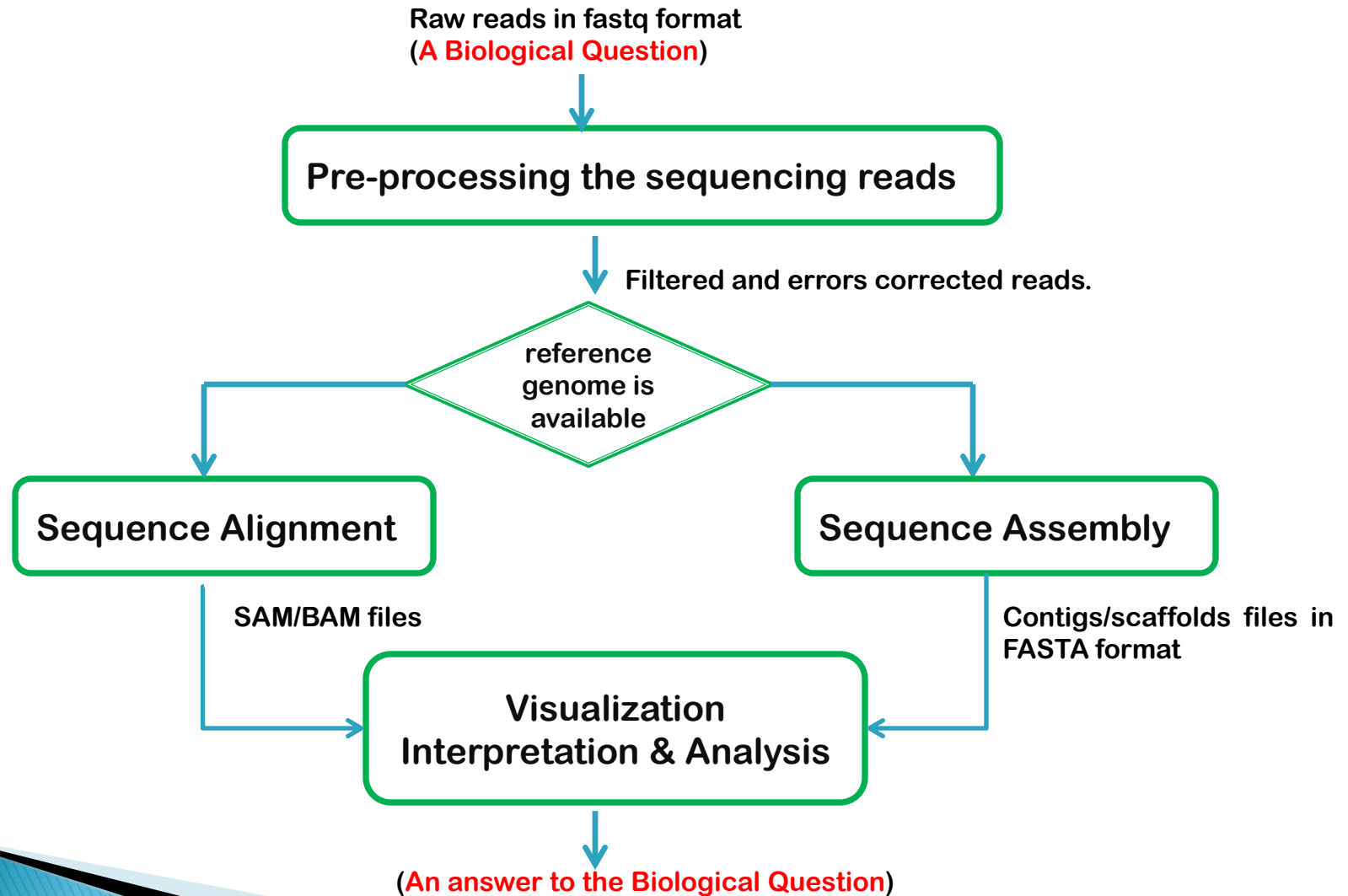
# Informatics on High-throughput Sequencing Data

(Summer Course 2020 )

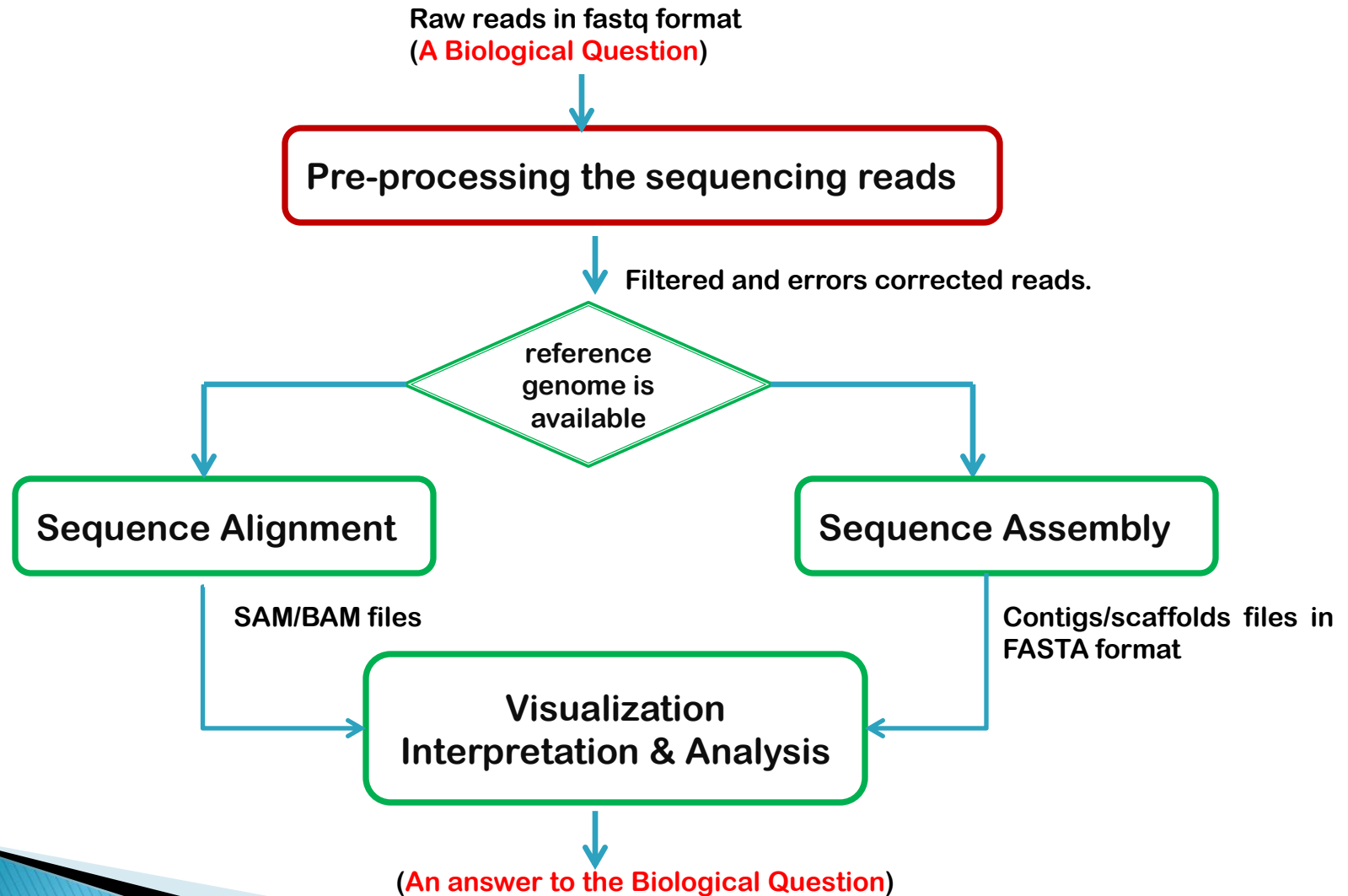
Day 13



# A typical Data Analysis pipeline



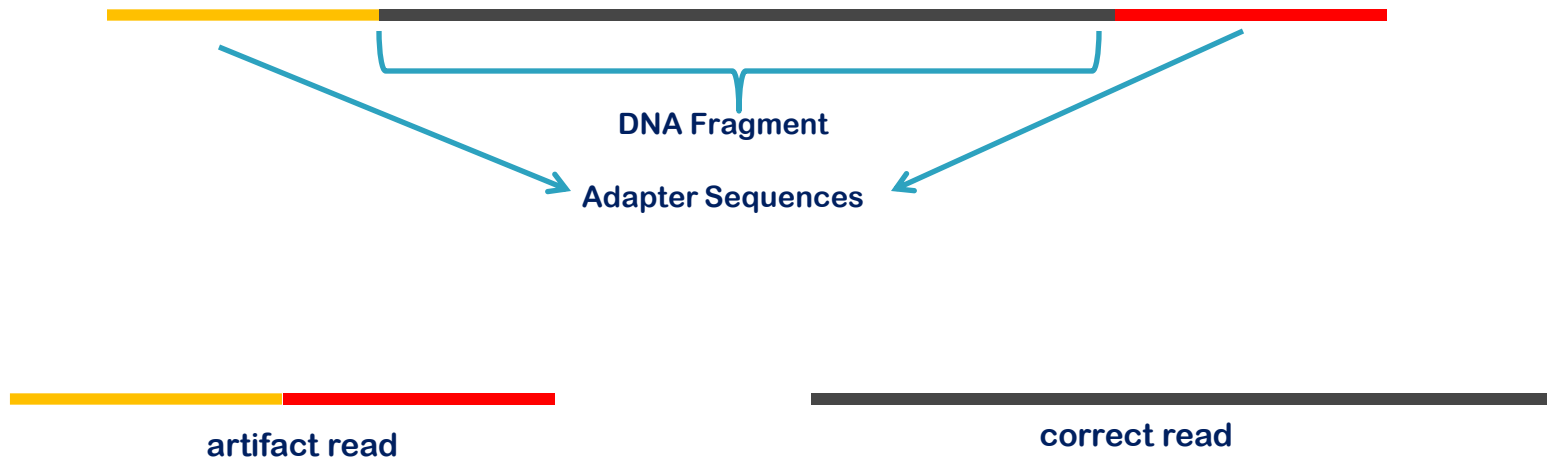
# A typical Data Analysis pipeline



# Pre-processing reads

- **Filter out garbage reads (Reads Trimming)**
  - ✓ Reads with low quality base calls.
  - ✓ Reads that are clearly artifacts with chemistry.

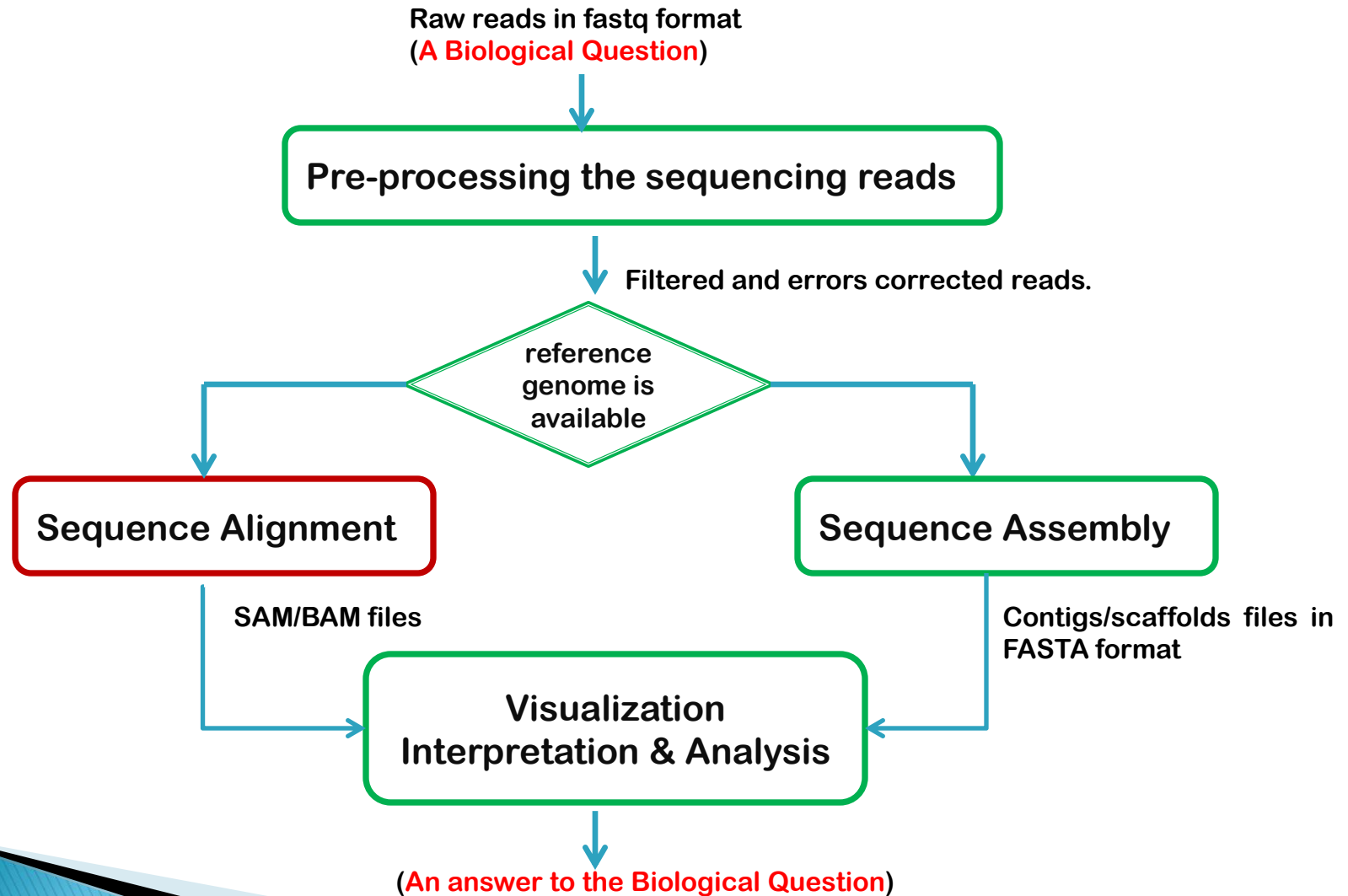
A typical read for DNA sequencing process



# Pre-processing reads

- **Filter out garbage reads (Reads Trimming)**
  - ✓ FASTX
  - ✓ AfterQC
  - ✓ Trimmomatic
- **Errors detection and correction**
  - ✓ Quake
  - ✓ Lighter
  - ✓ Musket

# A typical Data Analysis pipeline



# Sequence alignment

## Sequencing Reads

TATGTCGCAGTATCTGCGCAGTATCTG  
TATGTCGCAGTATCTT  
TATGTCGCAGTATCTG  
TATGTCGCAGTATCTG  
GTCGCAGTATCTGTCT  
CCGGACACCCCTATATATGTCGCAGTATCTT  
ACACCCTATGTCGCA  
ACACCCTATGTCGCA  
TATGTCGCAGTATCTG  
ACACCCTATGTCGCA  
CCGGACACCCCTATAT  
CCGGACACCCCTATAT  
GTCGCAGTATCTGTTN  
TGTGCGCAGTATCTGTCT

## Reference Genome

GATCACAGGTCTATCACCTATTAACCACTCACGGGAGCTCTCCATGCATTTGGTATTTT  
CGTCTGGGGGGTATGCACGCGATAGCATTGCGAGACGCTGGAGCCGGAGCACCTATGTC  
GCAGTATCTGTCTTTGATTCTGCCTCATCCTATTATTTATCGCACCTACGTTCAATATT  
ACAGGCGAACATACTTACTAAAGTGTGTTAATTAATTAATGCTTGTAGGACATAATAATA  
ACAATTGAATGTCTGCACAGCCACTTTCCACACAGACATCATAACAAAAATTTCCACCA  
AACCCCCCTCCCCGCTTCTGGCCACAGCACTTAAACACATCTCTGCCAAACCCAAAA  
ACAAAGAACCCTAACACCAGCCTAACCAGATTTCAAATTTTATCTTTTGGCGGTATGCAC  
TTTTAACAGTCACCCCCCACTAACACATTATTTCCCTCCCACTCCCATACTACTAAT  
CTCATCAATACAACCCCCGCCCATCTACCCAGCACACACACACCGCTGCTAACCCCAT  
CCCCGAACCAACCAACCCCAAGACACCCCCACAGTTTATGTAGCTTACCTCCTCAAA  
GCAATACACTGACCCGCTCAAACTCCTGGATTTTGGATCCACCCAGCGCCTTGGCCTAAA  
CTAGCCTTTCTATTAGCTCTTAGTAAGATTACACATGCAAGCATCCCCGTTCCAGTGAGT  
TCACCCTCTAAATCACCACGATCAAAAGGAACAAGCATCAAGCACGCAGCAATGCAGCTC  
AAAACGCTTAGCCTAGCCACACCCCAACGGGAAACAGCAGTGATTAACCTTTAGCAATAA  
ACGAAAGTTTAACTAAGCTATACTAACCCAGGGTTGGTCAATTTCTGTGCCAGCCACCGC  
GGTCACACGATTAACCAAGTCAATAGAAGCCGGCGTAAAGAGTGTTTTAGATCACCCCC  
TCCCCAATAAAGCTAAAACTCACCTGAGTTGTAAAAAACTCCAGTTGACACAAAATAGAC  
TACGAAAGTGGCTTTAACATATCTGAACACACAATAGCTAAGACCCAACTGGGATTAGA  
TACCCCACTATGCTTAGCCCTAAACCTCAACAGTTAAATCAACAAAACCTGCTCGCCAGAA  
CACTACGAGCCACAGCTTAAAACTCAAAGGACCTGGCGGTGCTTCATATCCCTCTAGAGG  
AGCCTGTTCTGTAATCGATAAACCCCGATCAACCTCACCACTCTTGCTCAGCCTATATA



# Sequence alignment software

<u>Aligner</u>	<u>Approach</u>	<u>Applications</u>	<u>Availability</u>
BWA-mem	Burrows-Wheeler	DNA, SE, PE, SV	open-source
Bowtie2	Burrows-Wheeler	DNA, SE, PE, SV	open-source
Novoalign	hash-based	DNA, SE, PE	free for academic use
TopHat	Burrows-Wheeler	RNA-seq	open-source
STAR	hash-based (reads)	RNA-seq	open-source
GSNAP	hash-based (reads)	RNA-seq	open-source



# Genome Indexing

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# Genome Indexing

Step1: hash/index the genome

CATGGTCATTGGTTCC

<u>Kmer/Hash</u>	<u>Genome Positions</u>
CAT	1, 7
ATG	2
TGG	3, 10
GGT	4, 11
GTC	5
TCA	6
ATT	8
TTG	9
GTT	12
TTC	13
TCC	14

# Genome Indexing

Step2: use the index to find reads locations

Toy genome    CATGGTCATTGGTTCC

Kmer/Hash

Genome Positions

CAT	1, 7
ATG	2
TGG	3, 10
GGT	4, 11
GTC	5
TCA	6
ATT	8
TTG	9
GTT	12
TTC	13
TCC	14



Read    TGGTCA

# BWA-MEM



# BWA-MEM workflow

*This takes a long time, but  
you do it once*

Create BWT of reference genome.

```
$ bwa index grch38.fa
```



*Output is in SAM format.  
Use multiple threads if you  
have a computer with  
multiple CPUs.*

Align paired-end FASTQ  
to BWT index.

```
$ bwa mem -t 16 grch38.fa 1.fq 2.fq > sample.sam
```

# BWA-MEM



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## Burrows-Wheeler Aligner

Status: **Beta** Brought to you by: [lh3lh3](#)

★★★★★ 8 Reviews

Downloads: 632 This Week

Last Update: 2017-11-0



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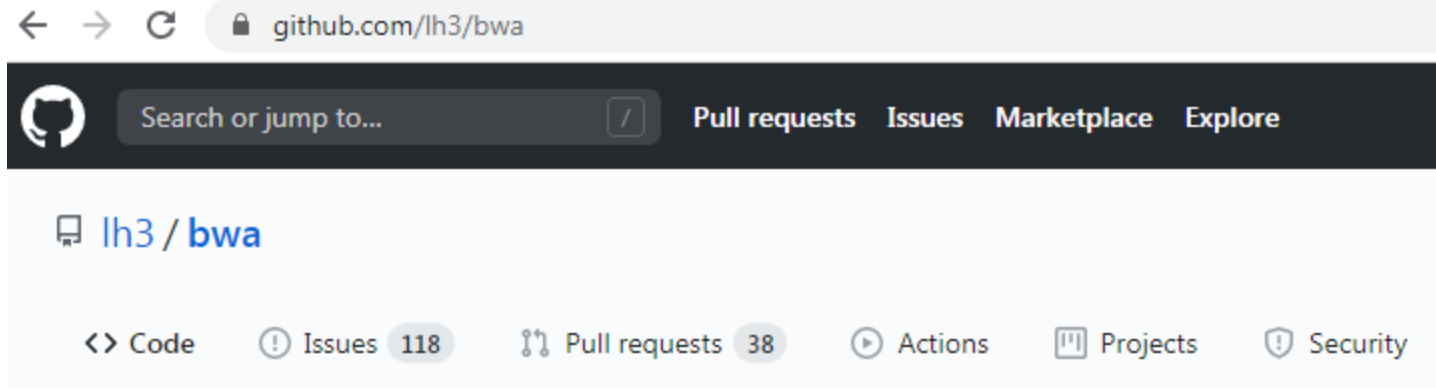
[Code](#)

BWA is a program for aligning sequencing reads against a large reference genome (e.g. human genome). It has two major components, one for read shorter than 150bp and the other for longer reads.

### Features

- Support Illumina, SOLiD, 454, Sanger reads,
- Gapped alignment and paired-end mapping
- Accurate, fast and lightweight

# BWA-MEM



## Getting started

```
git clone https://github.com/lh3/bwa.git
cd bwa; make
./bwa index ref.fa
./bwa mem ref.fa read-se.fq.gz | gzip -3 > aln-se.sam.gz
./bwa mem ref.fa read1.fq read2.fq | gzip -3 > aln-pe.sam.gz
```

## Introduction

BWA is a software package for mapping DNA sequences against a large reference genome, such as the human genome. It consists of three algorithms: BWA-backtrack, BWA-SW and BWA-MEM. The first algorithm is designed for



# BWA-MEM

- ▶ `bzip2 -d bwa-0.7.17.tar.bz2`
- ▶ `tar xvf bwa-0.7.17.tar`
- ▶ `make`
- ▶ `./bwa index wu_0.v7.fas`
  - `wu_0.v7.fas.amb`
  - `wu_0.v7.fas.ann`
  - `wu_0.v7.fas.bwt`
  - `wu_0.v7.fas.pac`
  - `wu_0.v7.fas.sa`
- ▶ `./bwa mem -t 16 wu_0.v7.fas wu_0_A_wgs.fastq > results.sam`

.amb is text file, to record appearance of N (or other non-ATGC) in the ref fasta.  
.ann is text file, to record ref sequences, name, length, etc.  
.bwt is binary, the Burrows-Wheeler transformed sequence.  
.pac is binary, packaged sequence (four base pairs encode one byte).  
.sa is binary, suffix array index.

# BWA-MEM

```
@SQ      SN:Chr1 LN:29923332
@SQ      SN:Chr2 LN:19386101
@SQ      SN:Chr3 LN:23042017
@SQ      SN:Chr4 LN:18307997
@SQ      SN:Chr5 LN:26567293
@SQ      SN:chloroplast LN:154478
@SQ      SN:mitochondria LN:366924
@PG      ID:bwa PN:bwa VN:0.7.17-r1198-dirty CL:./bwa mem -t 16 wu_0.v7.fas wu_0_A_wgs.fastq
```

# SAM

Field	Meaning
GAIIO5_0002:1:2:12086:1654	Read ID
16	Flag
Chr2	Chr
1694072	start
0	MAPQ
51M	CIGAR
*	Mate Chr
0	Mate start
0	Mate dis
CCTTGTAATAATCATTATTAATGTTTTTAAACCCCTTTTAAAAATCCTTGTA	read
CCCCCCCCCCCCCCCCCCCCBBCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	qual
NM:i:1 MD:Z:20C30 AS:i:46 XS:i:46	Tag-Type-Value

**Thanks!**

// | ?