

A Mathematical model for Thelaziasis

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Abstract

In the present manuscript we present a mathematical model for thelaziasis. We consider outbreak studies and estimate the severity of the disease. We base our study on a multiple hosts ...

Keywords: Thelaziasis, Mathematical Model, Parameter Estimation, Basic Reproductive number

1. Introduction

The disease. Hosts. What are the effects on the infected individuals. Where has been found. What is the vector in each case. Case studies. The use of a mathematical model to study a particular place.

Thelaziasis is a vector neglected disease that affects mainly mammals, including humans and in a minor scales, birds. The transmission takes place due to the presence of a vector which is the face fly *Musca autumnalis*. Depending on the region this vector might vary as well as the host species.

The transmission depends upon the presence of vectors and therefore thelaziasis has a seasonal occurrence [1]

The transmitted by the face fly [2]. The disease has spread in animals but in humans it has been reported but in a very isolated cases [3], [4], [5].

In [6] it was found the presence of *Thelazia gulosa* and *Thelazia lacrymalis* in cattle where the main responsible vector is the face fly (*Musca autumnalis*) in which of larvae of *Thelazia spp* were found. Data from slaughtered cattle was collected from April to October 1978. In [7] the authors present a survey for different diseases in equids in Kentucky USA. In their study, they found the presence *Thelazia Lacrymalis* in which it is presumed that the face fly (*Musca autumnalis*) is the vector responsible for transmission. Otranto et. al. [2] made a survey in different regions in Italy to observe

the current status on dogs, cats and foxes. In their work they present the proportion of infected animals (by *Thelazia Callipaeda*) in each of the regions they studied. In [8] data about the proportions of mule deer from Wyoming and Utah by *T. californiensis* was reported. Asrat [1] study the prevalence of Thelaziasis in Ethiopia whereas Beitel [9] studied the prevalence of eyeworms in the columbian black tailed deer in Oregon, USA by *Thelazia californiensis*. Khedri et. al. [10] present a one year data about infected bovine in Southeast Iran (puede ser útil).

In [11], the authors present a study about the prevalence and intensity of *Thelazia spp* in a flies population in Alberta, Canada.

In [9] studied the prevalence of eyeworms in the Columbian Black-Tailed Deer in Oregon.

A special work was done in [6] were it was estimated the proportion of infected animals as well as the proportion of infected vectors.

1.1. Some questions to explore.

An important issue in this disease is that the propagation coincides with the presence of flies that carry the disease. If the life expectancy of the fly is reduced, then the complete cycle of the thelazia within the vector does not complete and therefore, the disease no longer can be transmitted. Therefore, it might be expected that as soon as the temperature of a place of study is reduced, then the levels of the infected individuals with thelazia, must reach a final steady level.

In the mathematical side, analyse the model about stability, persistance, what would happen if stochasticity gets implemented? how?

1.2. Model parameters

Flies have a life expectancy of about 28 days, but it might live up to two months ([12]). The first larval stage (L1) of the worm is ingested by the fly when it feeds from lachrymal secretions, where in the internal organs, the worm develops into its second (L2) and third (L3) larval stages within 21 days post infection [2]. Other studies [13], show that flies infected with *Thelazia lacrymalis* can reach the infective stage in 12-15 days, while this takes 28-32 days for flies infected with *T. gulosa* [13]. Once in the infective stage, the fly releases L3 larvae into the definite host. Finally, once in the definite host, the L3 larvae matures within 3 to 6 weeks, where the new worm deposits new eggs into the definite host becoming infective [13]. Foxes lifespan is 2 years [14].

58 We will use the model to fit two data sets. One referring to a multi-host case
 59 given by dogs and foxes and the second in a one host study, particularly the
 60 case of cattle.

61 1.2.1. Cattle only.

62 The problem can be seen as a simple host or multi-host when considering
 63 beef and milk cattle. Some considerations about the life expectancy of the
 64 individuals. A common technique to detect thelazias in farming animals is
 65 done by sacrificing the animal. In this case, the infected individual is no
 66 longer part of the infection cycle and basically out of the dynamics. In this
 67 work we consider that the sample used to observe the proportion of infected
 68 individuals is of little to neglected significance respect to the total population.
 69 The life expectancy of beef cattle is approximately 16 to 24 months (and can
 70 be up to 30 months [15]), whereas for dairy cattle is 5 to 6 years. The natural
 71 cattle life expectancy is 18 to 22 years.

72 2. Mathematical Model

73 Our model is based on the interaction of flies and cattle. Following the
 74 formulation in Esteva [16] we obtain the following SI vector host model for
 75 cattle and flies.

$$\begin{aligned}
 \dot{S}_f &= \Lambda_f - \frac{\beta_f}{N_c} I_c S_f - \mu_f S_f \\
 \dot{L}_f &= \frac{\beta_f}{N_c} I_c S_f - \kappa_f L_f - \mu_f L_f \\
 \dot{I}_f &= \kappa_f L_f - \mu_f I_f \\
 \dot{S}_c &= \Lambda_c - \frac{\beta_c}{N_c} I_f S_c - \mu_c S_c \\
 \dot{L}_c &= \frac{\beta_c}{N_c} I_f S_c - \kappa_c L_c - \mu_c L_c \\
 \dot{I}_c &= \kappa_c L_c - \mu_c I_c
 \end{aligned} \tag{1}$$

76 where $N_c = S_c + L_c + I_c$. For this model, the basic reproductive number is
 77 given by

$$R_0 = \left(\left(\frac{k_f}{\mu_f + k_f} \right) \left(\frac{\beta_c}{\mu_f} \right) \left(\frac{k_c}{k_c + \mu_c} \right) \left(\frac{F_c \beta_f}{\mu_c} \right) \right)^{1/4} \tag{2}$$

78 where $F_c = \frac{N_f^\infty}{N_c^\infty}$, $N_f^\infty = \frac{\Lambda_f}{\mu_f}$ and $N_c^\infty = \frac{\Lambda_c}{\mu_c}$. Table 2 show the meaning and
 79 the values of the parameters considered in this study.

Parameter	Meaning	Interval	Reference
N_c	Total number of individuals at time t	1000	This study
Λ_f	Fly recruitment rate		This study
Λ_c	Cattle recruitment rate		This study
β_c	Number of successful contacts of a fly that infects a cattle host		This study
β_f	Number of successful contacts in which a fly gets infected by a cattle host		This study
k_v^{-1}	average latency time for vectors	14-21 days 12-15 days (<i>T. Lacrymalis</i>) 28-32 days (<i>T. Gulosa</i>)	[17] [13] [13]
k_i^{-1}	average latency time for hosts $i = 1, 2$	≈ 35 days 21-42 days	[17] [13]
μ_v^{-1}	vector average lifespan	30-60 months	[12]
μ_c^{-1}	cows average lifespan	1080 days	[18]

Table 1: Parameter meaning and values.

80 3. Local and global stability analysis

81 System 1 has two equilibrium points. The disease free equilibrium $S_1 =$
82 $(S_{f1}^*, L_{f1}^*, I_{f1}^*, S_{c1}^*, L_{c1}^*, I_{c1}^*) = (\frac{\Lambda_f}{\mu_f}, 0, 0, \frac{\Lambda_c}{\mu_c}, 0, 0)$ and the endemic equilibrium
83 $S_2 = (S_{f2}^*, L_{f2}^*, I_{f2}^*, S_{c2}^*, L_{c2}^*, I_{c2}^*) = .$

Theorem. The disease free equilibrium point S_1 is globally asymptotically stable if $R_0 < 1$. Consider the Lyapunov function

$$V(L_f, I_f, L_c, I_c) = a_1 L_f + a_2 I_f + a_3 L_c + a_4 I_c$$

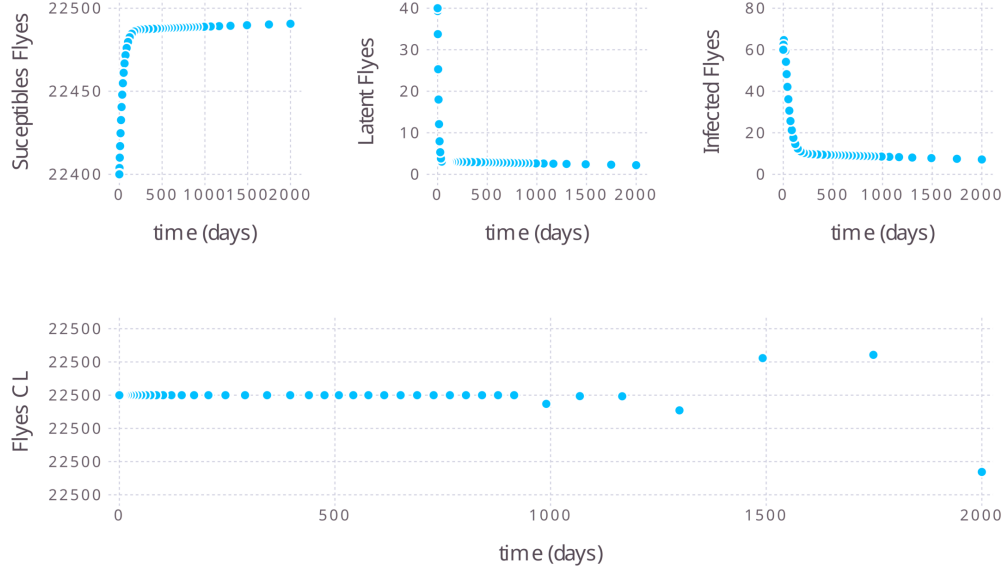


Figure 1: Solution with parameters according to $R_0 < 1$.

$$\begin{aligned}
\dot{V}(t) &= a_1 \left[\frac{\beta_f}{N_c} S_c I_c - (\mu_f + k_f) L_f \right] + a_2 [k_f L_f - \mu_f I_f] \\
&\quad + a_3 \left[\beta_c \frac{S_c I_f}{N_c} - (k_c + \mu_c) L_c \right] + a_4 [k_c L_c - \mu_c I_c] \\
&\leq \left[\frac{a_1 \beta_f N_f}{N_c} - a_4 \mu_c \right] I_c + [a_2 k_f - a_1 (\mu_f + k_f)] L_f + \\
&\quad [a_3 \beta_c - a_2 \mu_f] I_f + [a_4 k_c - a_3 (k_c + \mu_c)] L_c
\end{aligned}$$

Then, by taking $a_1 = \frac{1}{\mu_c} \frac{k_c}{k_c + \mu_c} \frac{\beta_c}{\mu_f} \frac{k_f}{k_f + \mu_f}$, $a_2 = \frac{1}{\mu_c} \frac{k_c}{k_c + \mu_c} \frac{\beta_c}{\mu_f}$, $a_3 = \frac{1}{\mu_c} \frac{k_c}{k_c + \mu_c}$ and $a_4 = \frac{1}{\mu_c}$, we arrive to

$$\dot{V}(t) \leq R_0 - 1,$$

84 which completes the proof.

85 *3.1. Persistence*

86 **4. Discussion**

87 **5. Numerical Results**

88 **Bibliography**

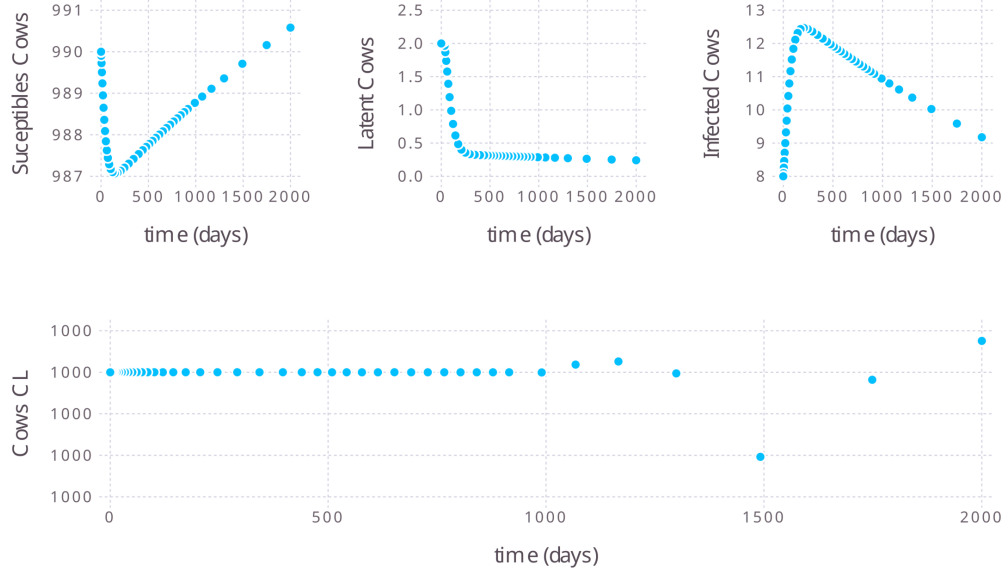


Figure 2: Solution with parameters according to $R_0 < 1$

- 89 [1] M. Asrat, Prevalence and risk factors for bovine thelaziasis at mersa
90 town of south wollo zone, amhara regional state, ethiopia, J. Ecosys.
91 Ecograph. 6 (2016) 1000212 (1–4).
- 92 [2] D. Otranto, E. Ferroglio, R. P. Lia, D. Traversa, L. Rossi, Current
93 status and epidemiological observation of *Thelazia callipaeda* (spirurida,
94 thelaziidae) in dogs, cats and foxes in italy: a “coincidence” or a parasitic
95 disease of the old continent?, Vet. Parasit. 116 (2003) 315–325.
- 96 [3] X.-L. Wang, J.-L. Guo, X.-L. Wang, X.-L. Ma, Y. Wang, C.-L. An, Two
97 cases of human thelaziasis as confirmed by mitochondrial *cox1* sequenc-
98 ing in china, Path. and Global Health 108 (2014) 298–301.
- 99 [4] D. Otranto, M. Dutto, Human thelaziasis, europe, Emerg. Inf. diseases.
100 14 (2008) 647–649.
- 101 [5] J. Shen, R. B. Gasser, D. Chu, Z. Wang, X. Yuan, C. Cantacessi, O. D.,
102 Human thelaziosis—a neglected parasitic disease of the eye, J. Parasitol.
103 92 (2006) 872–876.

- 104 [6] W. J. Moolenbeek, S. G. A., Southern ontario survey of eyeworms,
105 *Thelazia gulosa* and *Thelazia lacrymalis* in cattle and larvae of *Thelazia*
106 *spp.* in the face fly, *Musca autumnalis*, Can. Vet. J. 21 (1980) 50–52.
- 107 [7] E. T. Lyons, T. W. Swerczek, S. C. Tolliver, H. D. Bair, J. H. Drudge,
108 L. E. Ennis, Prevalence of selected species of internal parasites in equids
109 at necropsy in central kentucky (1995–1999), Vet Parasit. 92 (2000) 51–
110 62.
- 111 [8] S. A. Dubay, E. S. Williams, K. Mills, A. M. Boerger-Filedts, Bacteria
112 and nematodes in the conjunctiva of mule deer from wyoming and utah,
113 J. Wildlife disease 36 (2000) 783–787.
- 114 [9] S. E. Beitel, R. J. an Knapp, P. A. Vohs, Jr., Prevalence of eyeworm in
115 three populations of columbian black-tailed deer in northwestern oregon,
116 The J. of Parasitology 60 (1974) 972–975.
- 117 [10] J. Khedri, M. H. Radfar, H. Borji, M. Azizzadeh, Epidemiological survey
118 of bovine thelaziosis in southeastern of iran, Iran J. Parasitol. 11 (2016)
119 221–225.
- 120 [11] J. E. O’hara, J. K. Murray, Prevalence and intensity of thelazia spp.
121 (nematoda: Thelazioidea) in a musca autumnalis (diptera:muscidae)
122 population from central alberta, J. Parasit. 75 (1989) 803–806.
- 123 [12] H. Sanchez-Arroyo, J. L. Capinera, House fly, *Musca domestica* linnaeus
124 (insecta: Diptera: Muscidae), UF/IFAS Extension (1998) 1–8.
- 125 [13] M. Chanie, B. Bogale, Thelaziasis: Biology, species affected and pathol-
126 ogy (conjunctivitis): A review, Acta Parasitologica Globalis 5 (2014)
127 65–68.
- 128 [14] E. S. Devenish-Nelson, S. A. Richards, S. Harris, C. Soulsbury, S. P. A.,
129 Demonstrating frequency-dependent transmission of sarcoptic mange in
130 red foxes, Biol. Lett. 10 (2014) 1–5.
- 131 [15] K. Stanley, K. Jones, Cattle and sheep farms as reservoirs of campy-
132 lobacter, J. Appl. Microbiol. 94 (2003) 104S–113S.
- 133 [16] L. Esteva, C. Vargas, Analysis of a dengue disease transmission model,
134 Math. Biosci. 150 (1998) 131–151.

- 135 [17] D. Otranto, F. Dantas-Torres, Thelaziosis, in: C. Brisola-Marcondes
136 (Ed.), Arthropod borne diseases, Springer, Cham, Switzerland, 2017,
137 pp. 457–464.
- 138 [18] FAO, Guidelines for slaughtering, meat cutting and further processing,
139 <http://www.fao.org/3/T0279E/T0279E00.htm>, 1991.