

Sawall\_lab\_Chla\_Chlc2\_coral\_tissue\_analysis\_Carbonne

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## Protocols Adapted for Sawall Lab at ASU

### Quantifying Chlorophyll-*a* and Chlorophyll-*c2* Concentrations

*Adapted from Dr. Hollie Putman's protocol by Dr. Chloe Carbonne*

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#### Materials Checklist

- ☐ 100% acetone
  - ☐ Flammable-safe fridge (4 °C)
  - ☐ Quartz 96-well plate
  - ☐ Glass plate lid (plastic lids will melt)
  - ☐ Centrifuge
  - ☐ 10 mL glass syringe
  - ☐ 2 mL microcentrifuge tubes
  - ☐ Q-tips
  - ☐ Labeling tape
  - ☐ Microplate reader ( 630, 663, 750 nm)
  - ☐ Homogenizer (Ultra-Turrax type)
  - ☐ Parafilm (optional)
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## Protocol

### A. Sample Preparation

1. Prepare a **500  $\mu$ L aliquot** of adult airbrush homogenate (coral slurry) in a **2 mL micro-centrifuge tube**.
  2. Add **1 mL of 100% acetone** to the slurry using a **10 mL glass syringe**.
  3. Vortex tubes for **15 seconds**.
    - If the pellet does not dissolve or tissue chunks remain:
      - Sonicate **2  $\times$  5 seconds, 100% amplitude**
      - Keep tubes **on ice**
      - Avoid evaporation as much as possible
  4. Place tubes **in the dark** at **4 °C** for **24 hours**.
  5. After incubation, sonicate again:
    - **2  $\times$  5 seconds, 100% amplitude**, on ice
  6. Centrifuge tubes at **2,000 rpm for 3 minutes** to pellet debris.
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### B. Plate Setup

1. Use the plate template to **map sample positions**.
  2. Pipette:
    - **200  $\mu$ L** of each sample into **triplicate wells**
    - **200  $\mu$ L acetone blanks** into:
      - First 3 wells
      - Last 3 wells
  3. As you fill the plate:
    - Cover every ~3 columns with the **glass lid or parafilm**
    - Use another plate (or object of equal height) to support the lid and maintain balance
  4. Once the plate is complete, **fully cover with the glass lid**.
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## C. Measuring Absorbance

1. Measure absorbance at:
    - **630 nm**
    - **663 nm**
    - **750 nm**
  2. Using Roger Lab's plate reader:
    - Open **Gen5 software**
    - The **Task Manager** will open automatically
    - Select protocol: "**Chloe\_Chla**"
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## 3. Calculating Chlorophyll Concentration

1. Subtract **A750** from all absorbance measurements.
  2. Calculate chlorophyll *a* and *c2* concentrations using equations from **Jeffrey & Humphrey (1975)**.
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## Path Length Correction

Because well-plate path length differs from a 1 cm cuvette, apply a correction following **Warren (2007)**.

- Path length correction factor used here: **0.6 cm**
- This value may vary by instrument

**Corrected Absorbance Calculations**    "`r datac630.corr < -(datac630 - datac750)/0.6datac663.corr`  
`<- (datac663 - datac750) / 0.6` Units of concentration:  $\mu\text{g mL}^{-1}$

To calculate a plate-specific path length, follow the procedure described in Warren (2007).

References Jeffrey, S. W. & Humphrey, G. F. (1975) New spectrophotometric equations for determining chlorophylls a, b, c1 and c2

Warren, C. R. (2007) Standards for chlorophyll measurement

Synergy HTX Operating Manual

Gen5 Software Manual