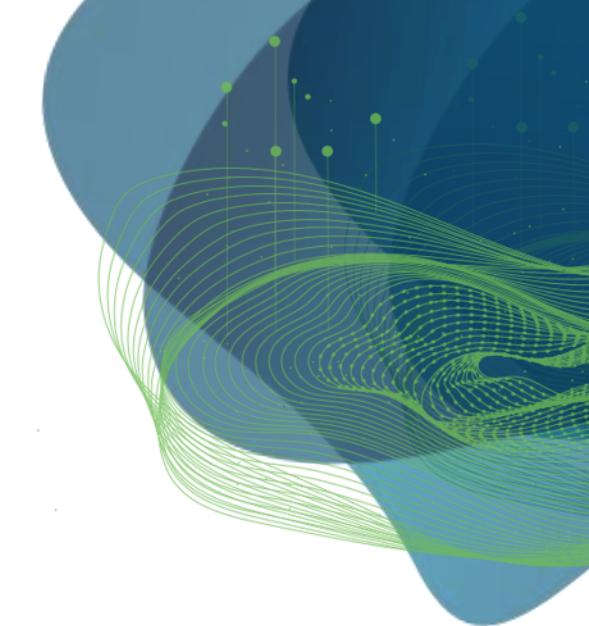




DRESDEN LEIPZIG

CENTER FOR SCALABLE DATA ANALYTICS  
AND ARTIFICIAL INTELLIGENCE



# Image segmentation

Robert Haase

Using materials from Ryan Savill George (PoL, TU Dresden)

GEFÖRDERT VOM



Bundesministerium  
für Bildung  
und Forschung

Diese Maßnahme wird gefördert durch die Bundesregierung  
aufgrund eines Beschlusses des Deutschen Bundestages.  
Diese Maßnahme wird mitfinanziert durch Steuermittel auf  
der Grundlage des von den Abgeordneten des Sächsischen  
Landtags beschlossenen Haushaltes.

# Quiz (recap)

- Which of the following is a band-pass filter?

Gaussian



Median



Top-hat



Difference  
of Gaussian



# Quiz (recap)

- Which of the following is a denoising filter?

Gaussian



Median



Top-hat

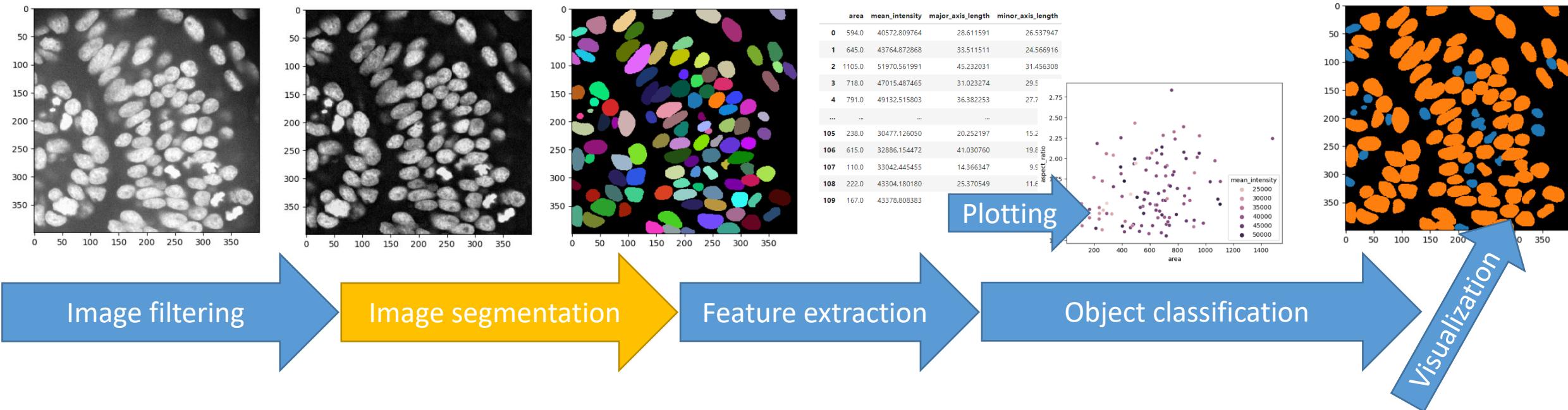


Difference  
of Gaussian



# Lecture overview: Bio-image Analysis

- Image Data Analysis workflows
- Goal: **Quantify observations, substantiate conclusions with numbers**



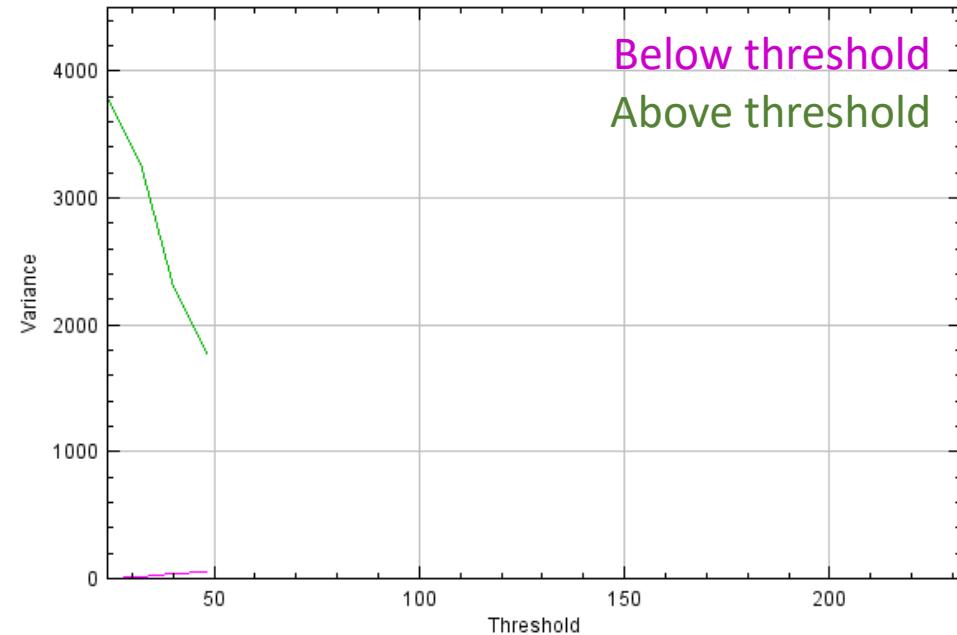
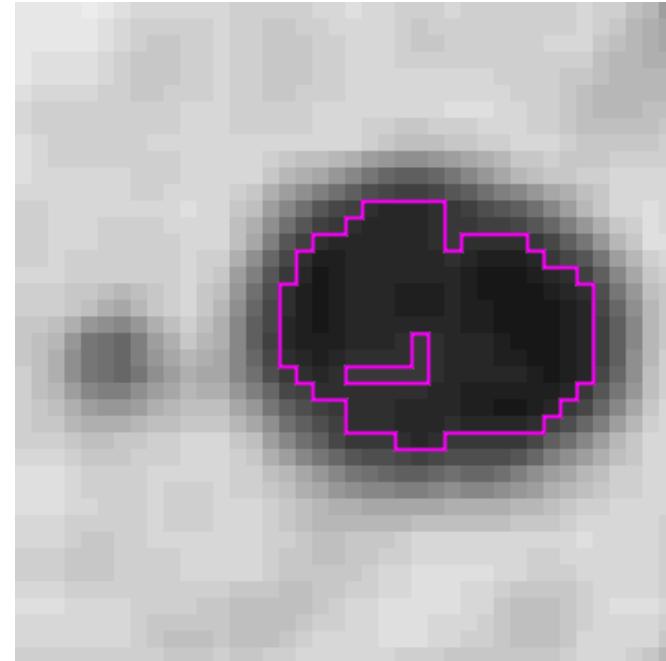
# Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.

$$Var(I) = \sum_{i \in I} g_i - \bar{g}_I$$

$$\bar{g}_I = \frac{\sum_{i \in I} g_i}{n_I}$$

$Var(I)$  ... Variance in image I  
 $g_i$  ... grey value of a pixel i  
 $\bar{g}_I$  ... mean grey value of the whole image I  
 $n_I$  ... number of pixels in Image I



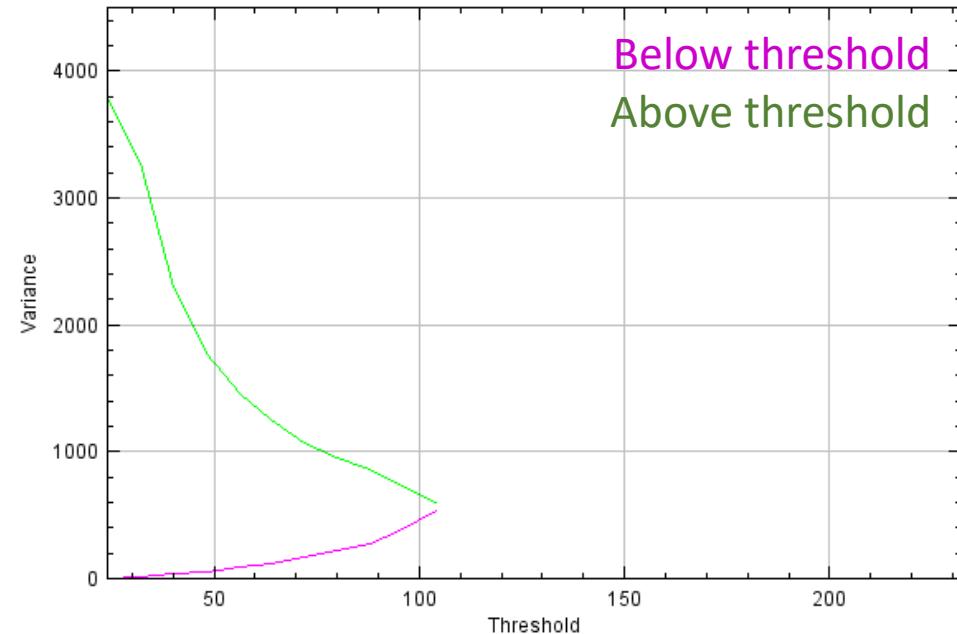
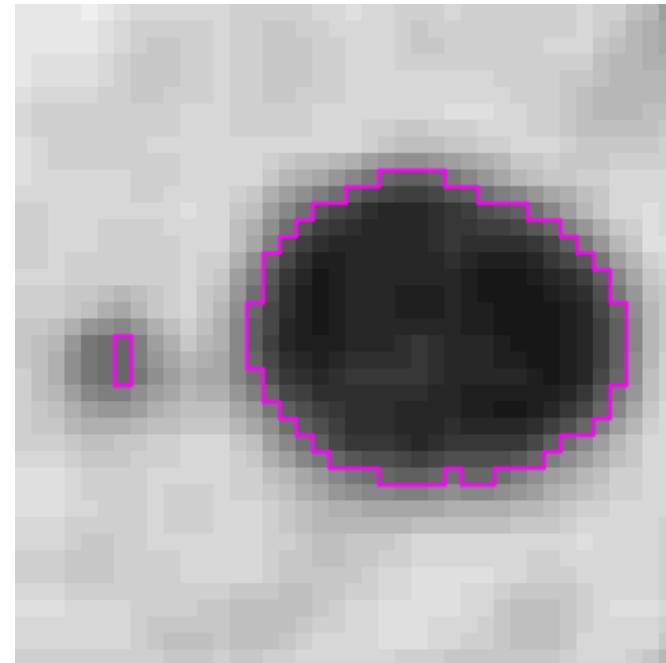
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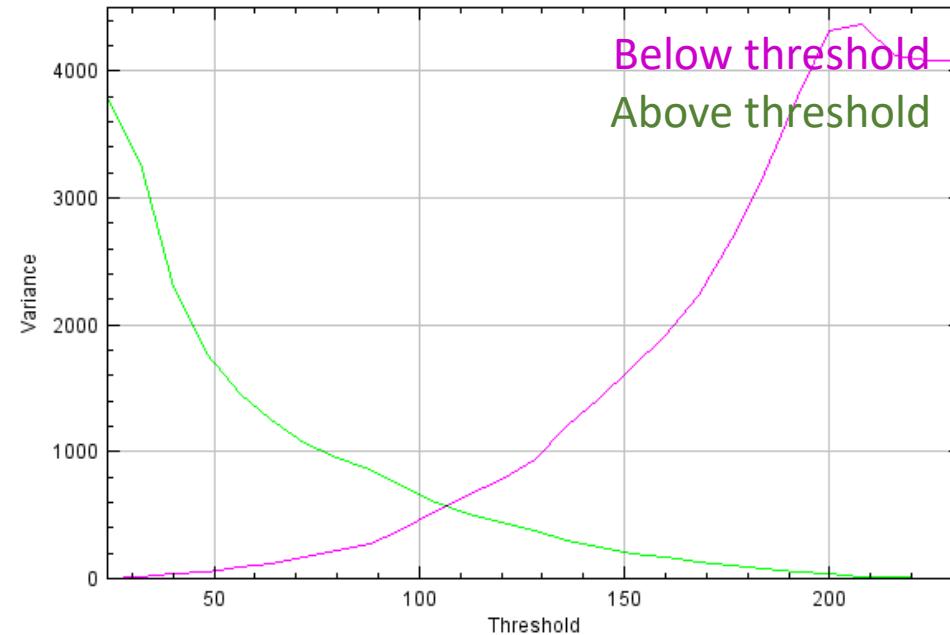
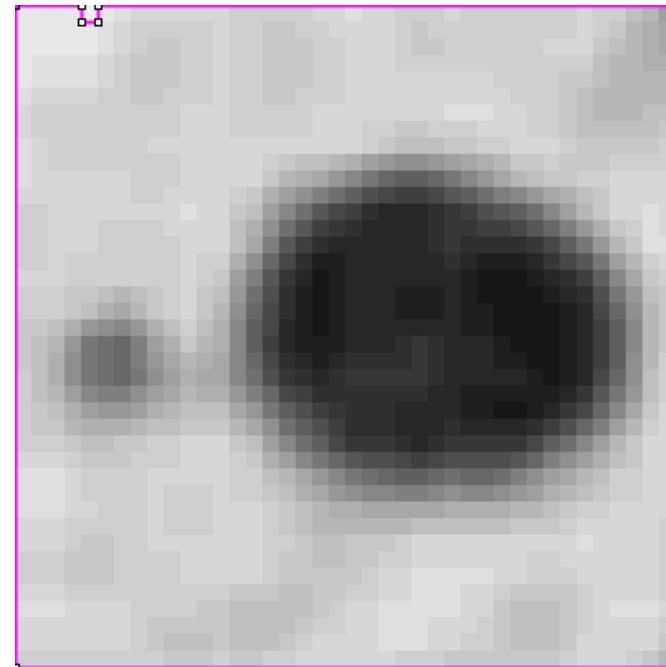
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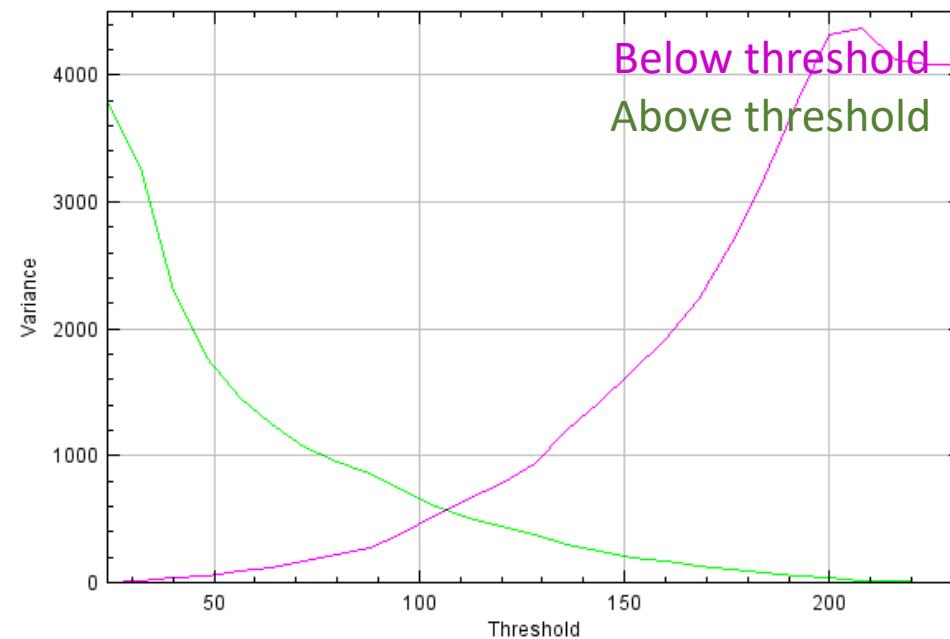
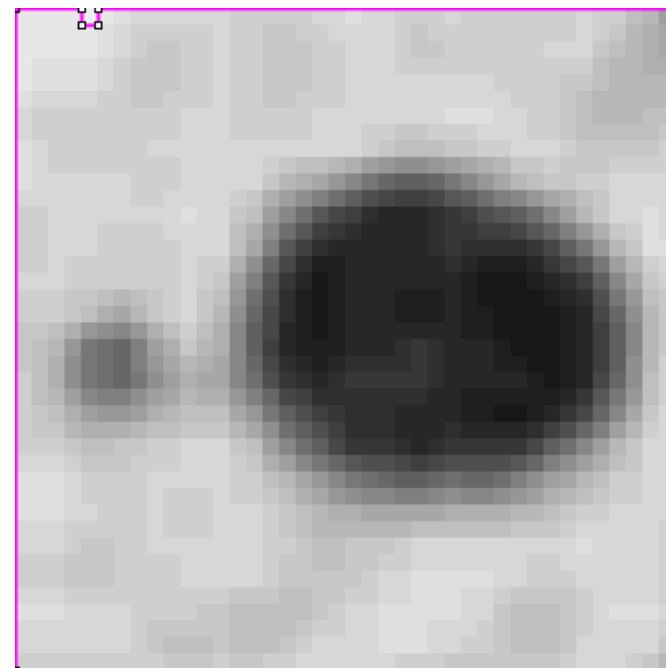
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 $n_I$  ... number of pixels in Image I



# Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance

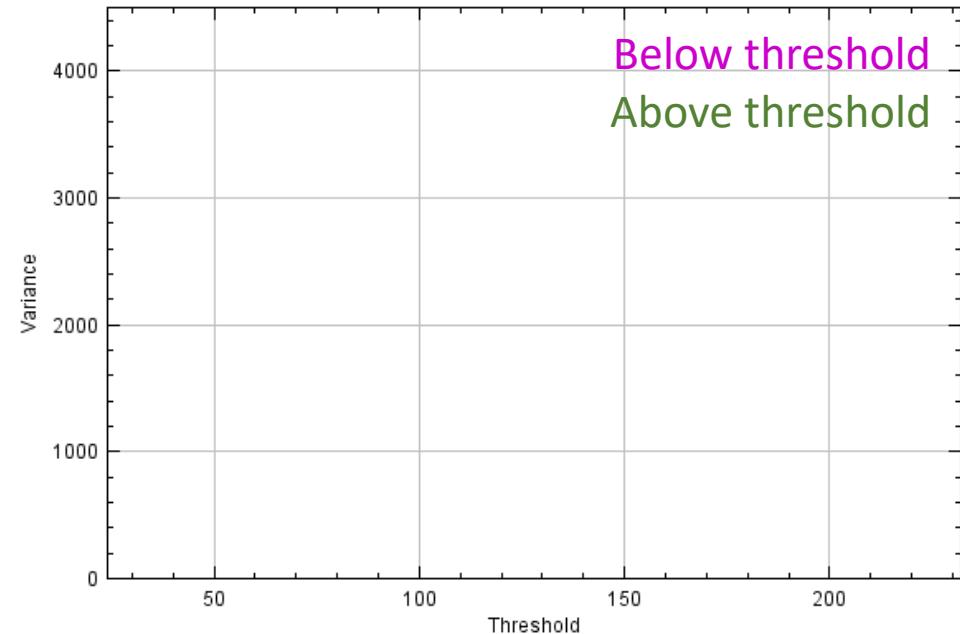
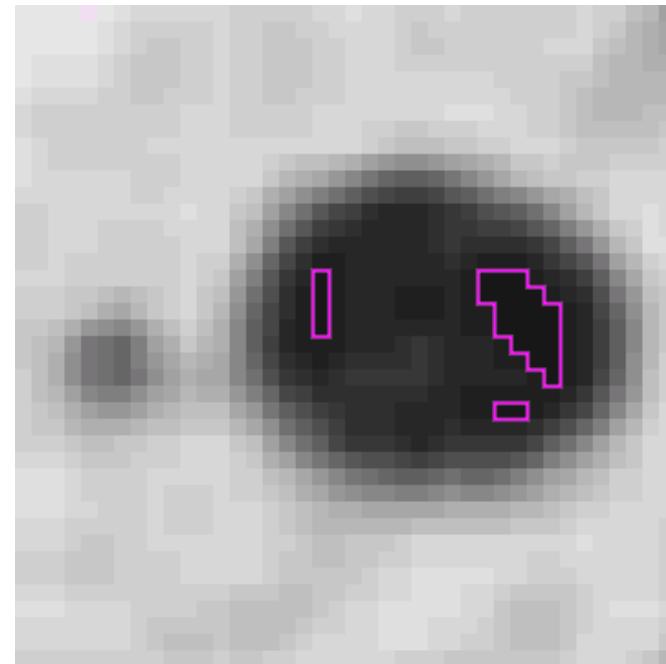
$$Var'(I) = \frac{n_A}{n_I} Var(A) + \frac{n_B}{n_I} Var(B) \quad I = A \cup B$$



# Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance

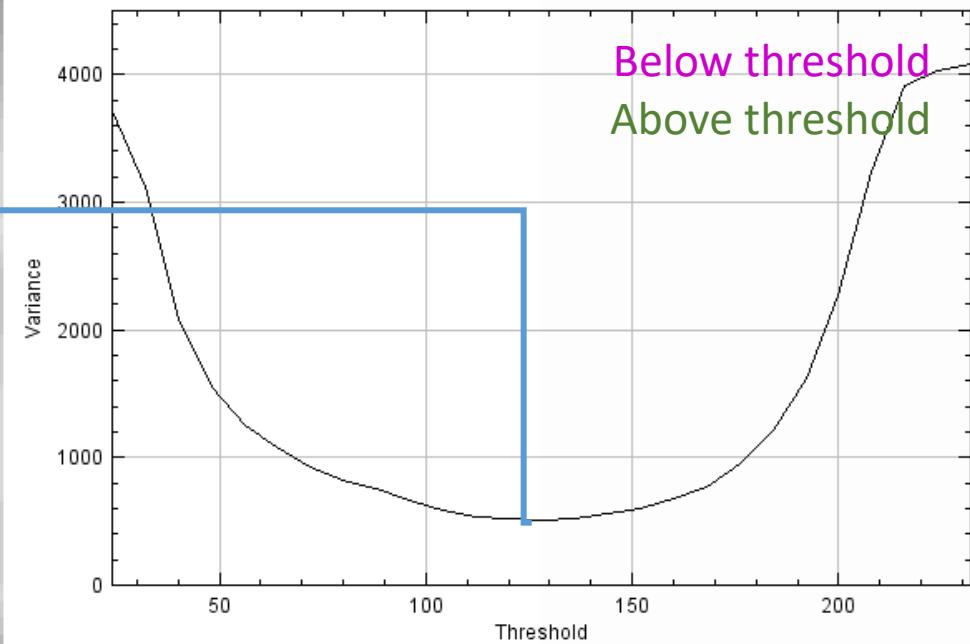
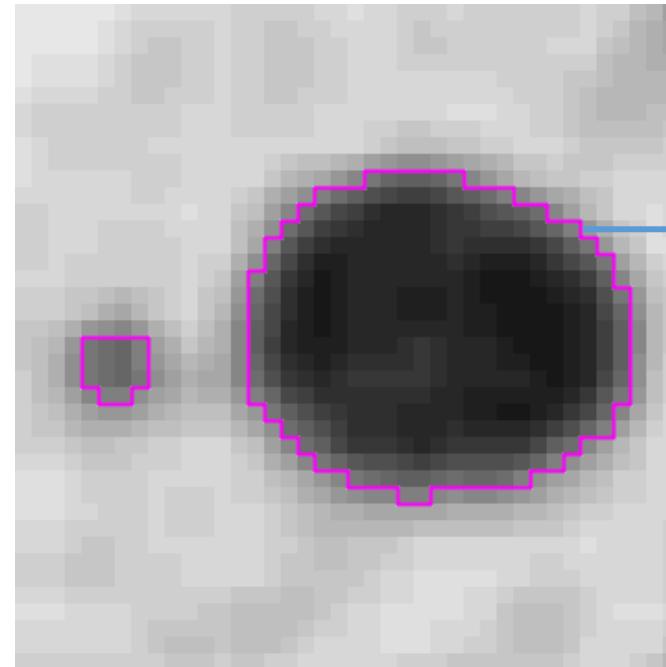
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# Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance

$$Var'(I) = \frac{n_A}{n_I} Var(A) + \frac{n_B}{n_I} Var(B) \quad I = A \cup B$$



# Thresholding: Citing

- Cite the thresholding method of your choice properly

*“We segmented the cell nuclei in the images using Otsu’s thresholding method (Otsu et Al. 1979) implemented in scikit-image (van der Walt et al. 2014).”*

IEEE TRANSACTIONS ON SYSTEMS, MAN, AND CYBERNETICS, VOL. SMC-9, NO. 1, JANUARY 1979

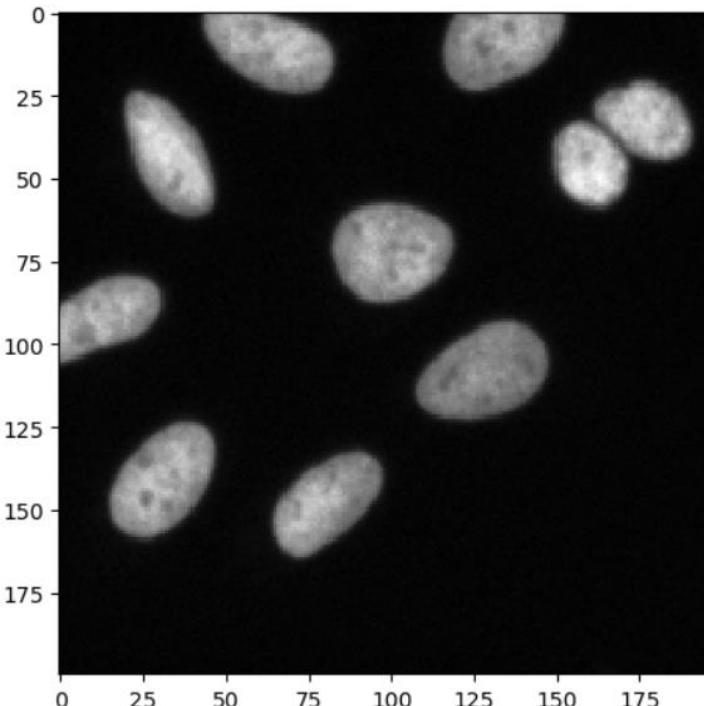
## A Threshold Selection Method from Gray-Level Histograms

NOBUYUKI OTSU

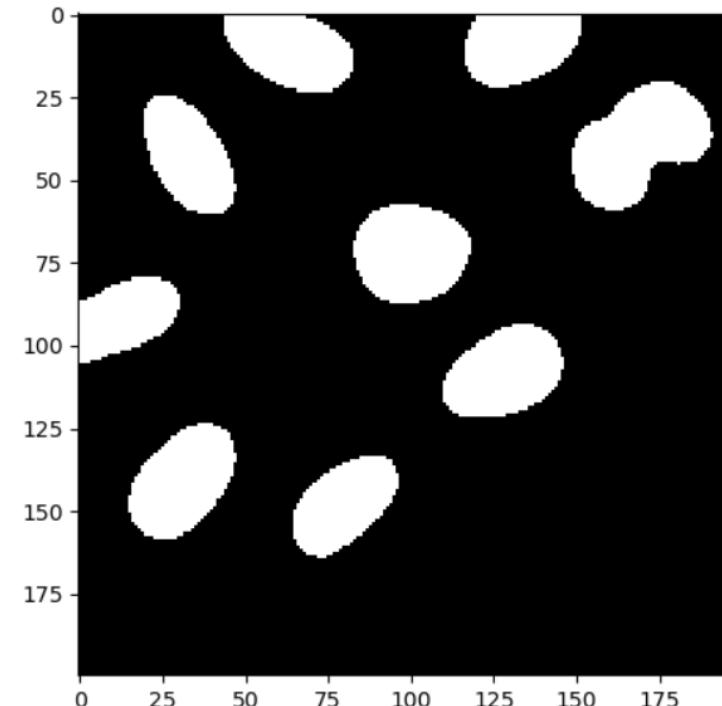
*Abstract*—A nonparametric and unsupervised method of automatic threshold selection for picture segmentation is presented. An optimal threshold is selected by the discriminant criterion, namely, so as to maximize the separability of the resultant classes in gray levels. The procedure is very simple, utilizing only the zeroth- and the first-order cumulative moments of the gray-level histogram. It is straightforward to extend the method to multithreshold problems. Several experimental results are also presented to support the validity of the method.

# Terminology

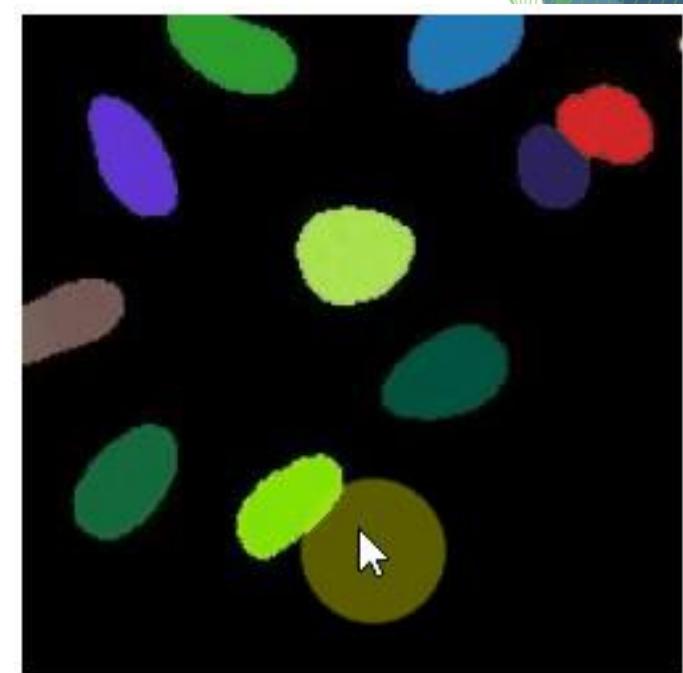
Intensity image



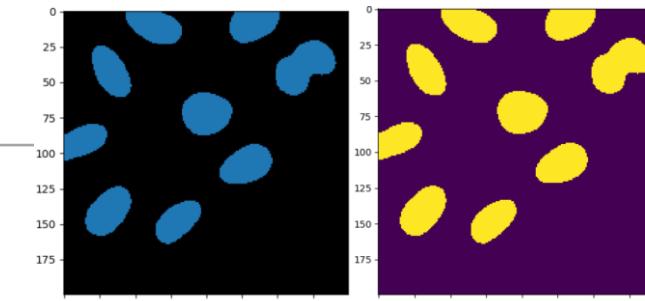
Binary image



Label image

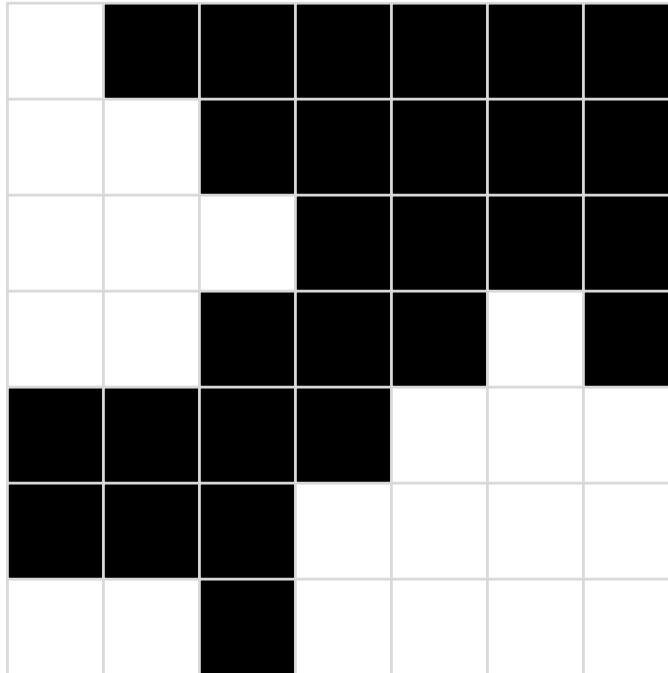


No matter how they are displayed



# Connected component labelling

- In order to allow the computer differentiating objects, connected component analysis (CCA) is used to mark pixels belonging to different objects with different numbers
- Background pixels are marked with 0.
- The maximum intensity of a labelled map corresponds to the number of objects.



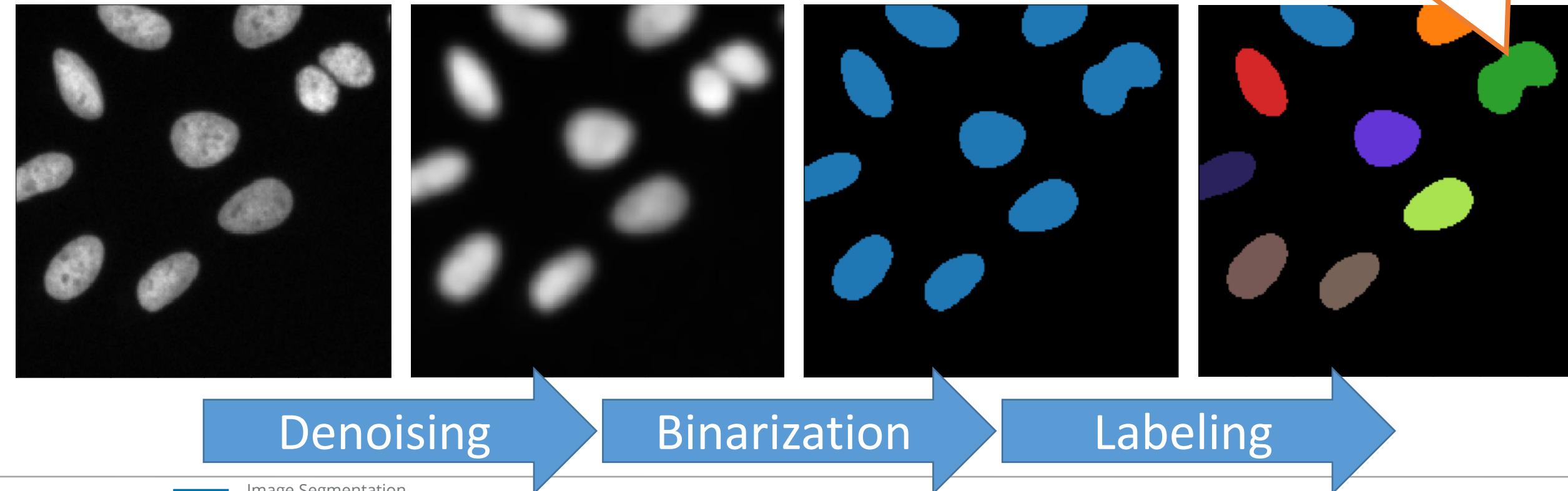
CCA

1	0	0	0	0	0	0
1	1	0	0	0	0	0
1	1	1	0	0	0	0
1	1	0	0	0	3	0
0	0	0	0	3	3	3
0	0	0	3	3	3	3
2	2	0	3	3	3	3

# Common image segmentation workflows

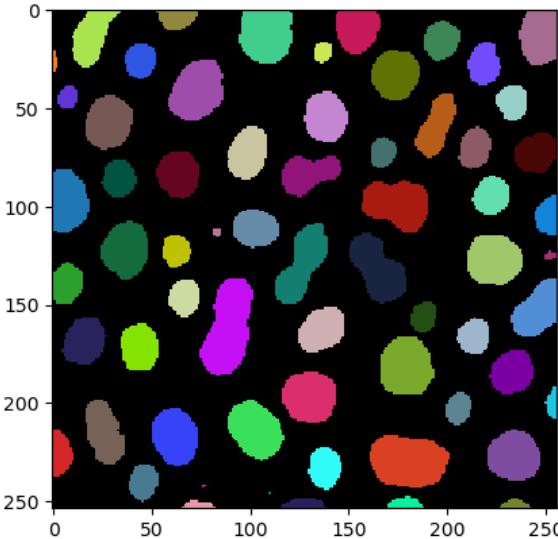
- Presumably the most common segmentation algorithm used for fluorescent microscopy images:
  - Gaussian blur, Otsu's Threshold, Connected Component Labeling

Limitation: Dense objects

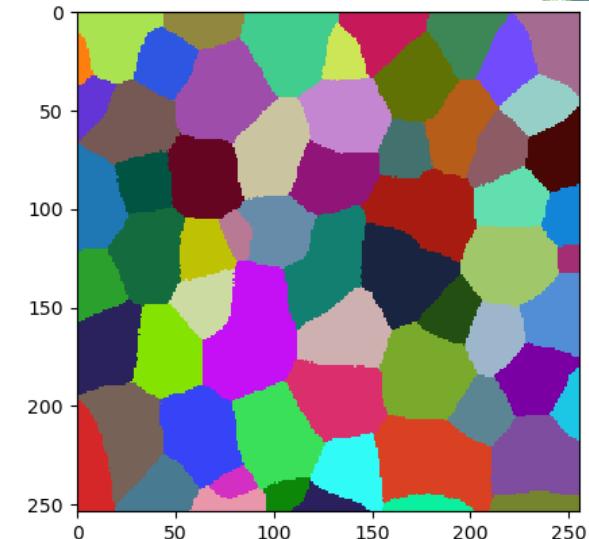
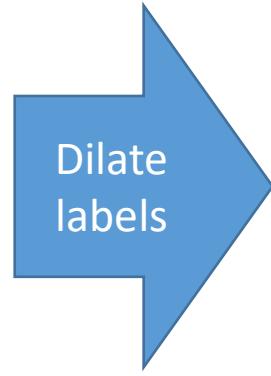
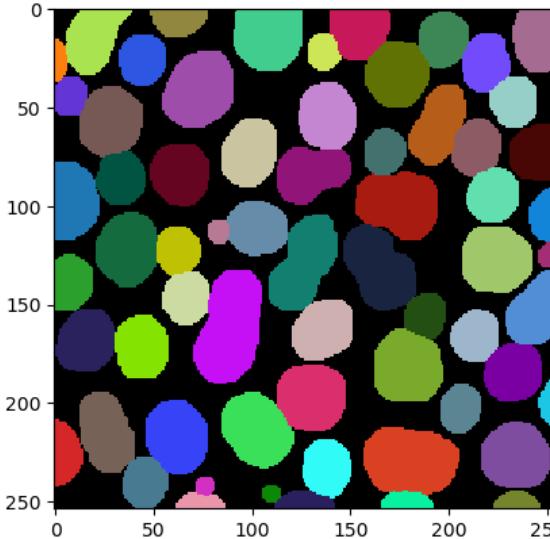
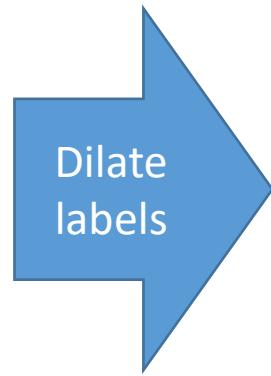


# Voronoi-Tesselation

- For separating objects using spatial constraints (not intensity-based)



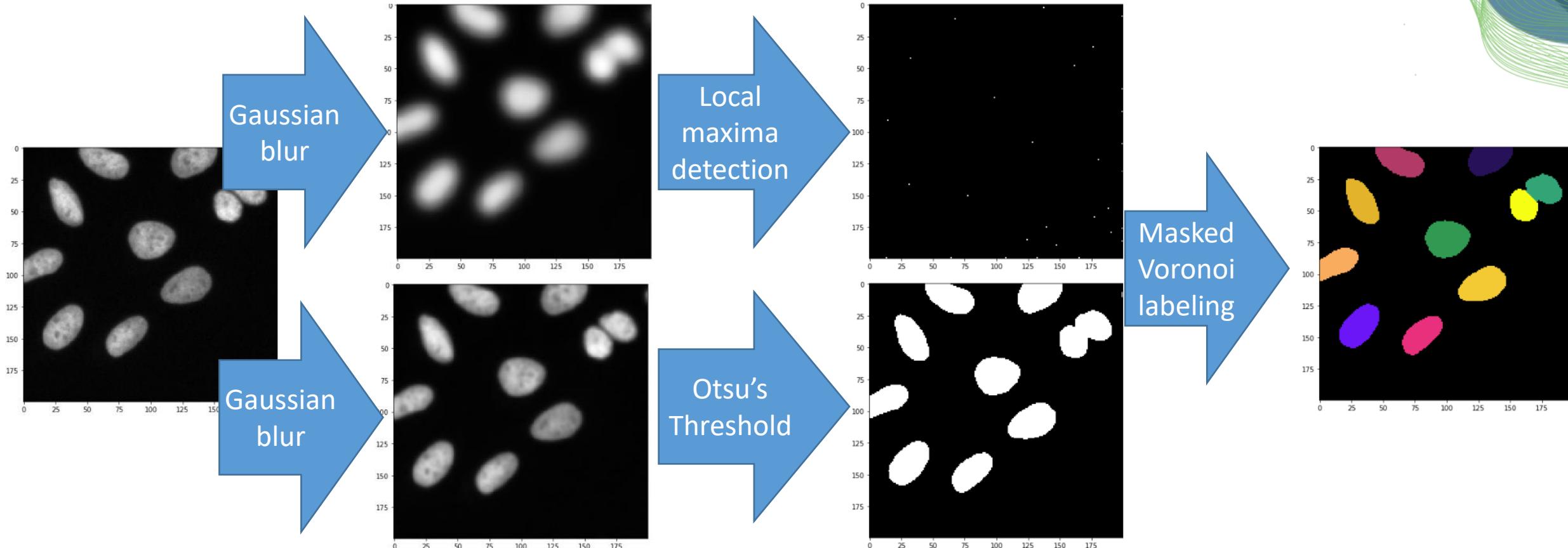
Label-image



Voronoi-label-image

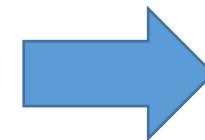
# Common image segmentation workflows

- Combination of Gaussian blur, Otsu's Threshold and Voronoi-labeling



# Voronoi-Otsu-Labeling

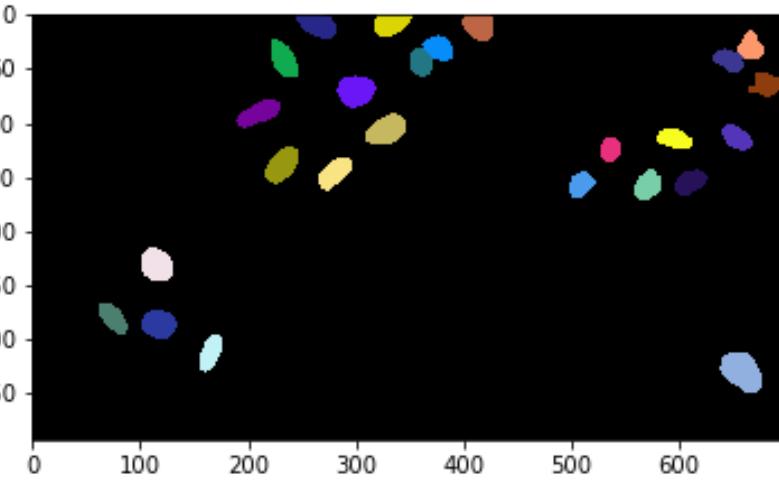
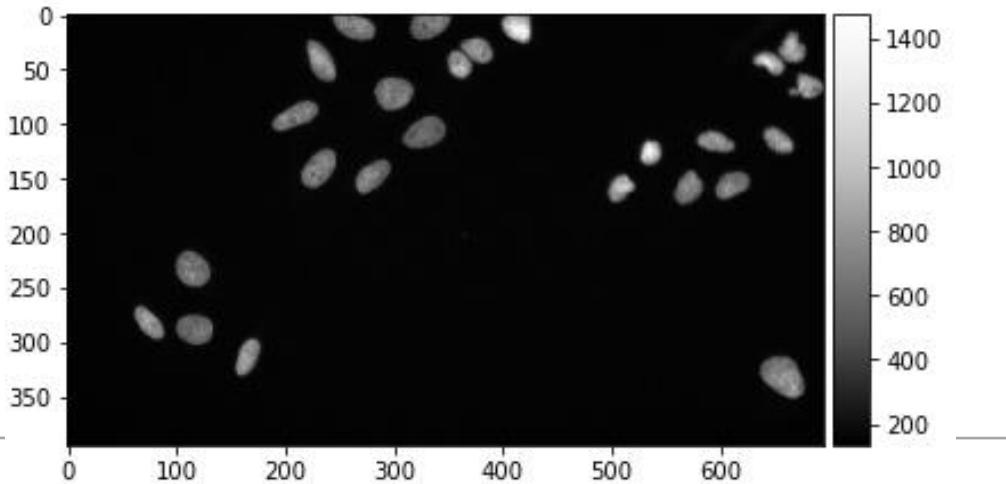
- Gaussian-Blur
- Otsu-Thresholding
- Spot-detection
- Watershed on the binary image



... in a single line of code:

```
segmented = nsbatwm.voronoi_otsu_labeling(input_image,  
                                             spot_sigma=5,  
                                             outline_sigma=1  
)
```

segmented

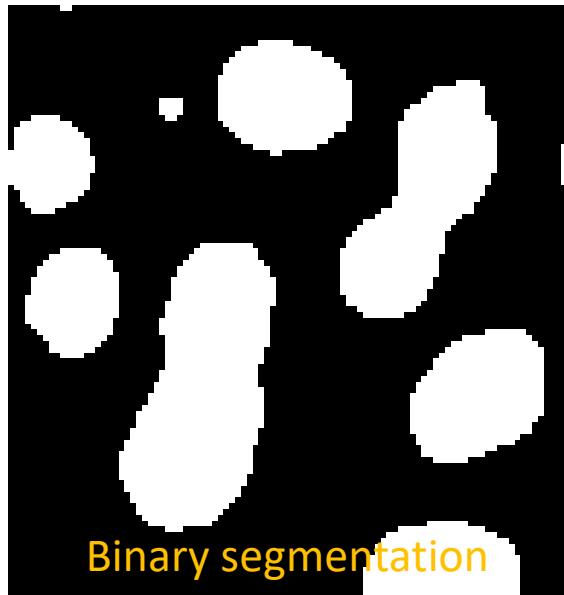


**nsbatwm made image**

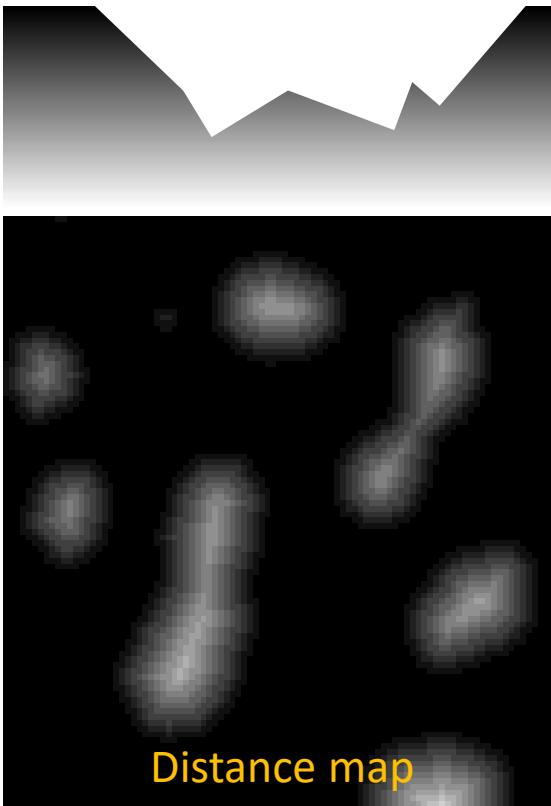
shape	(395, 695)
dtype	int32
size	1.0 MB
min	0
max	25

# Watershed

- The watershed algorithm for binary images allows cutting one object into two where it's reasonable.



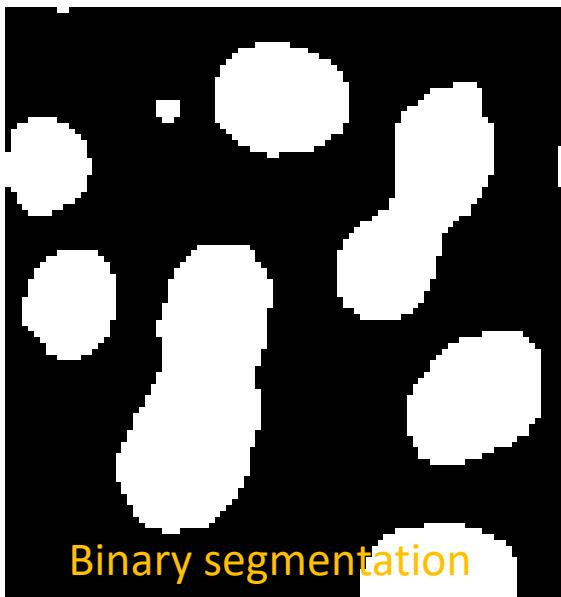
Binary segmentation



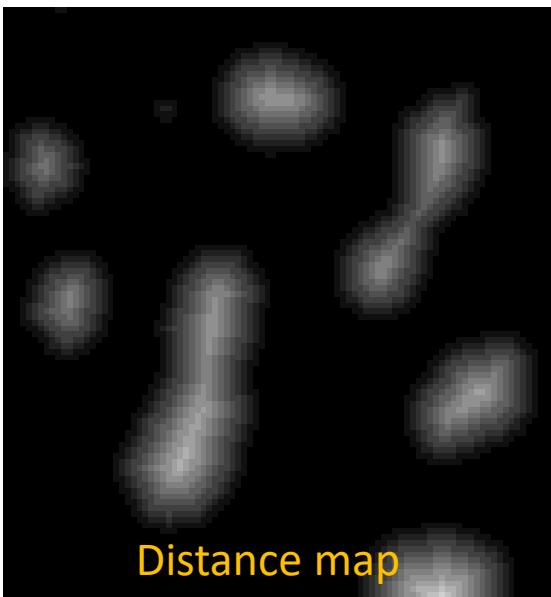
Distance map

# Watershed

- The watershed algorithm for binary images allows cutting one object into two where it's reasonable.



Binary segmentation

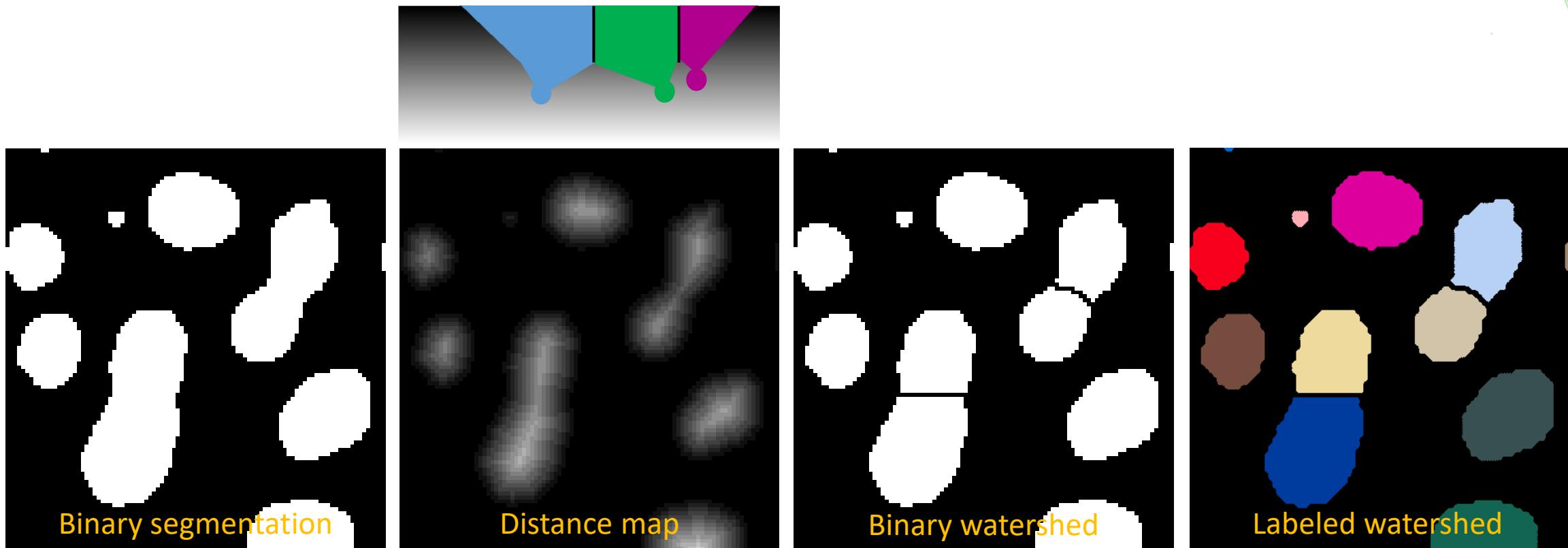


Distance map



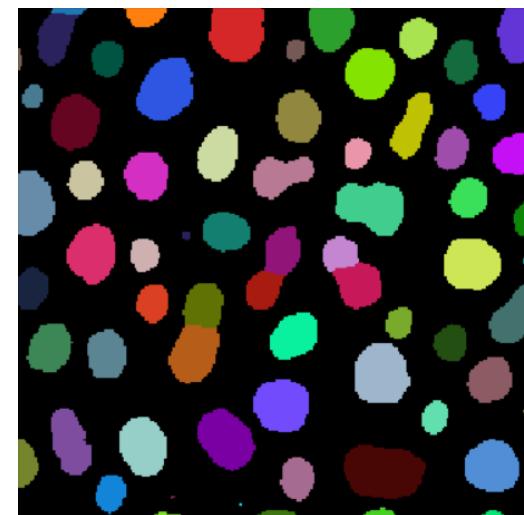
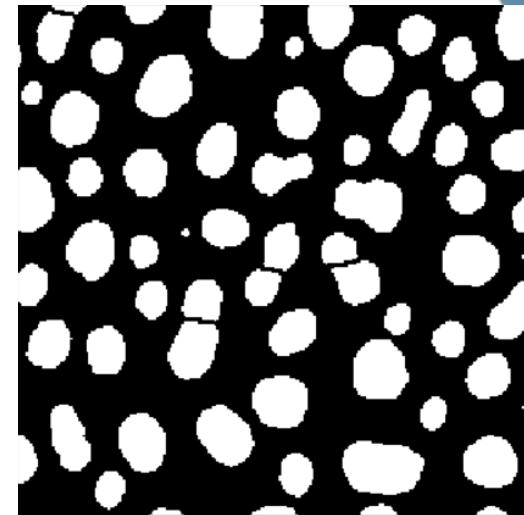
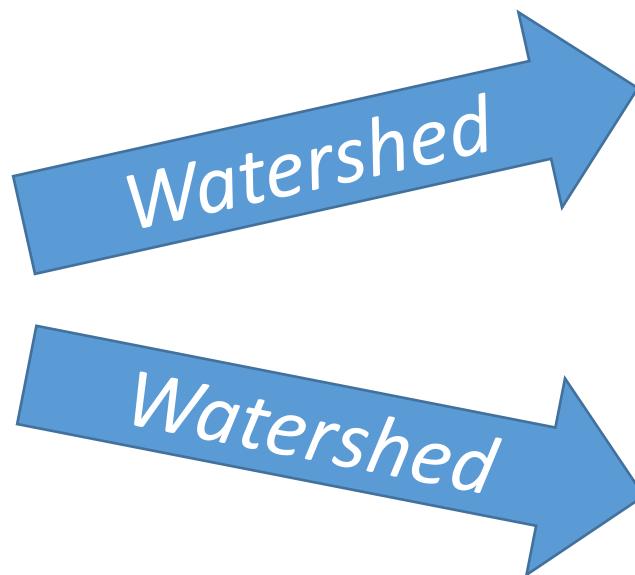
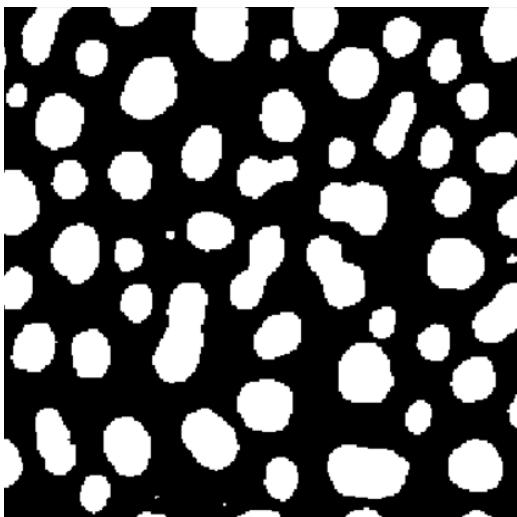
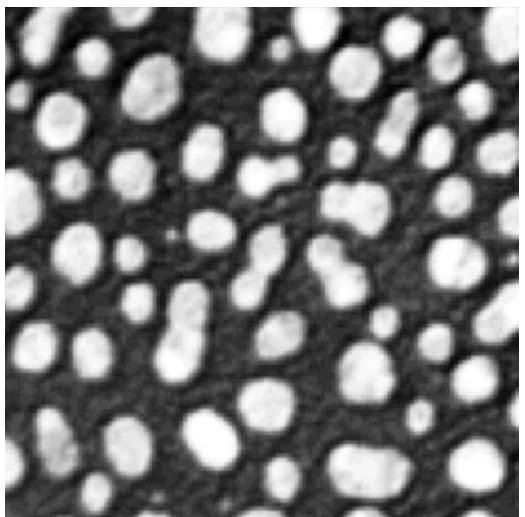
# Watershed

- The watershed algorithm for binary images allows cutting one object into two where it's reasonable.
- The distance-maps are typically made from binary images. It does not take the original image into account!



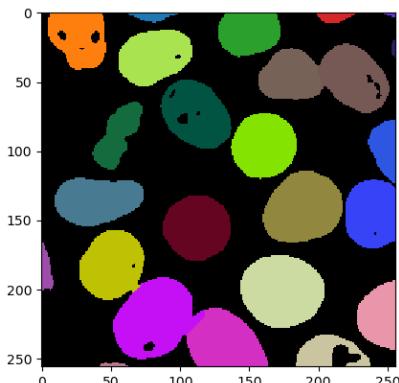
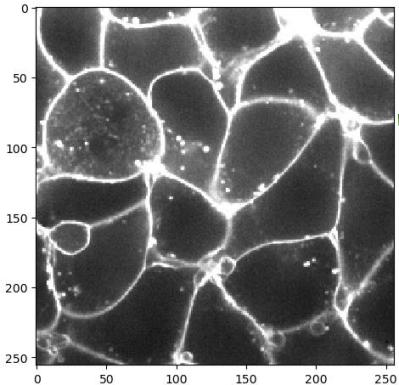
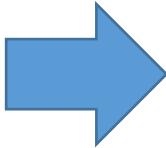
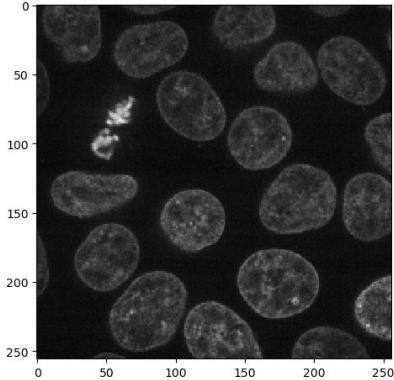
# Watershed use-cases

- Split dense objects



# Watershed use-cases

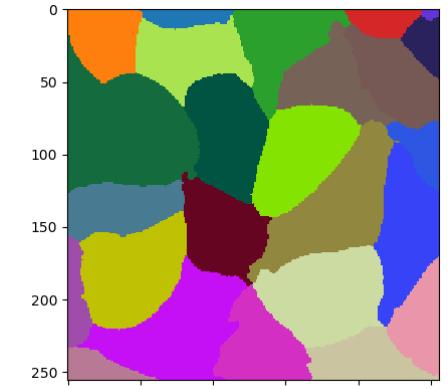
- Seeded watershed: Flood regions from pre-defined seeds
- Example: Flood cells from nuclei positions



```
labeled_cells = seeded_watershed(membrane_channel, labeled_nuclei)
```

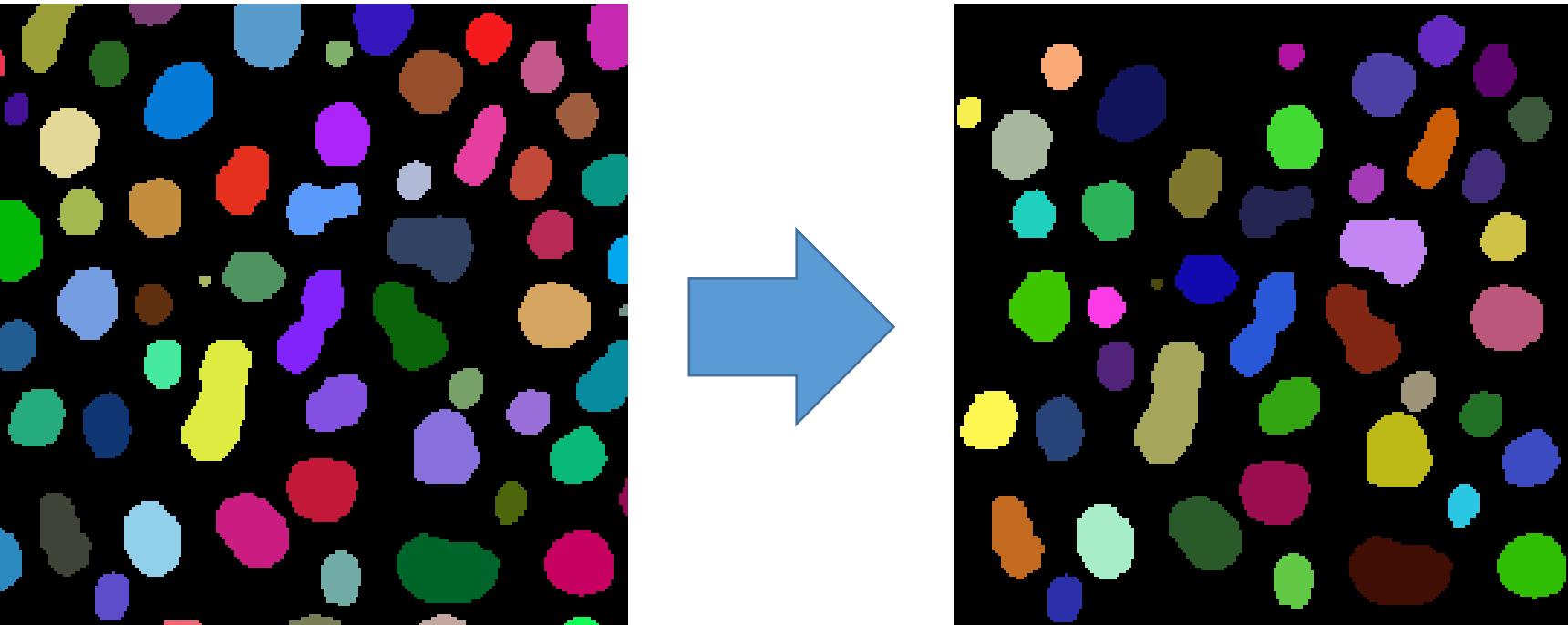
labeled\_cells

“Seeded” watershed



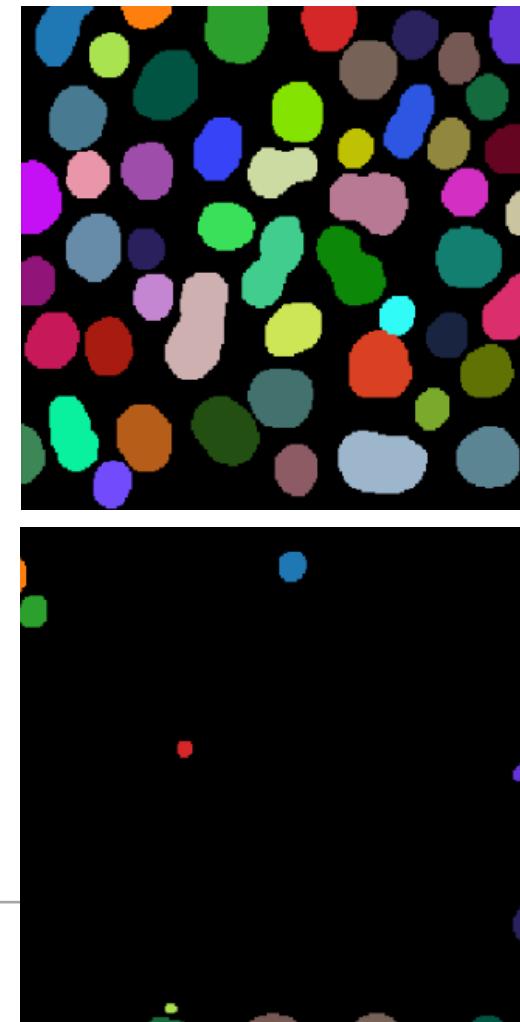
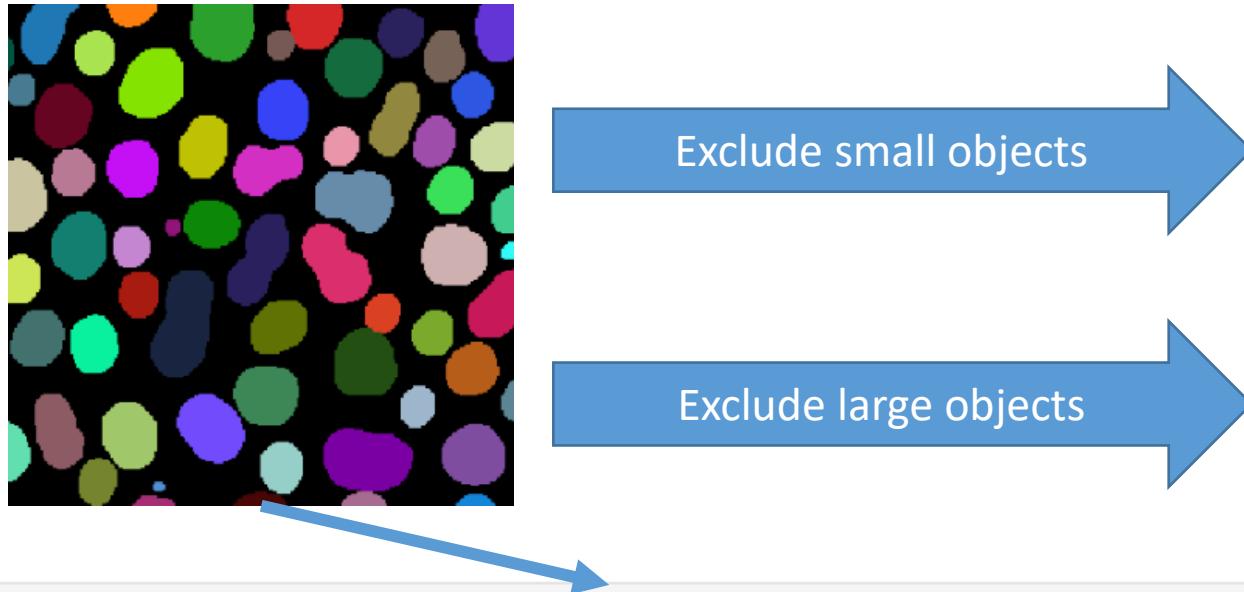
# Label post-processing / selections

- Remove objects at the image border
- Their measurements (shape, size) would be misleading anyway



# Label post-processing / selections

- Excluding small / large objects
- Common correction-step in case segmentations contain noise-related small particles





CENTER FOR SCALABLE DATA ANALYTICS AND  
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# Napari

## Robert Haase

Funded by



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SACHSEN



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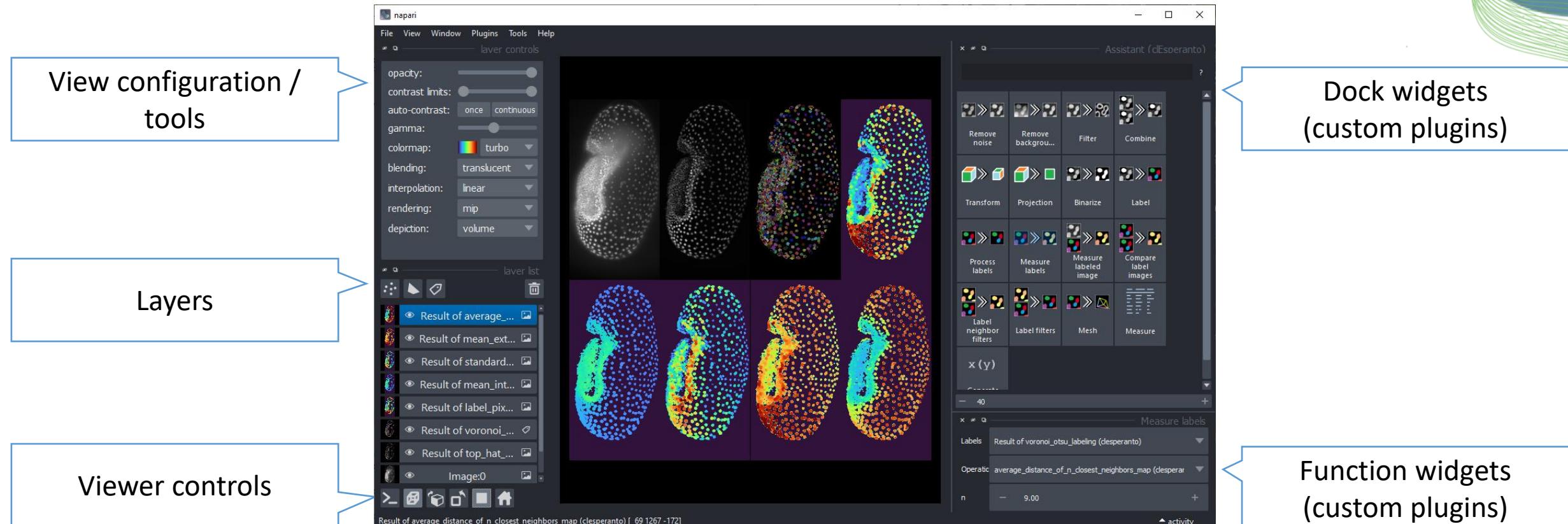
Chan  
Zuckerberg  
Initiative

# Napari

- A viewer for n-dimensional image data written in Python

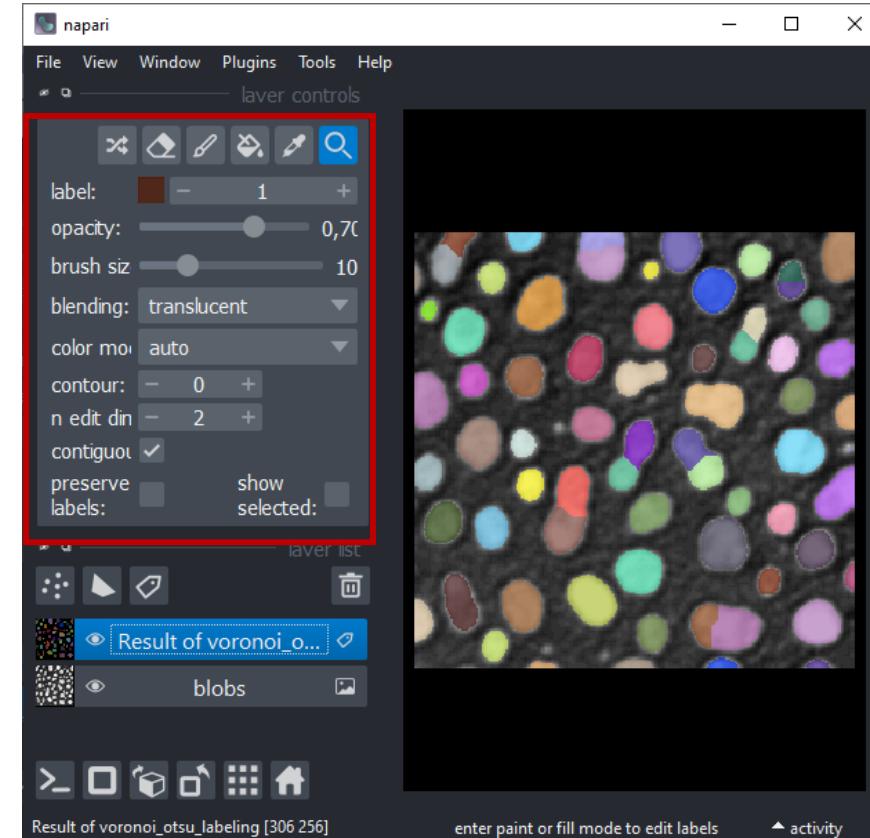
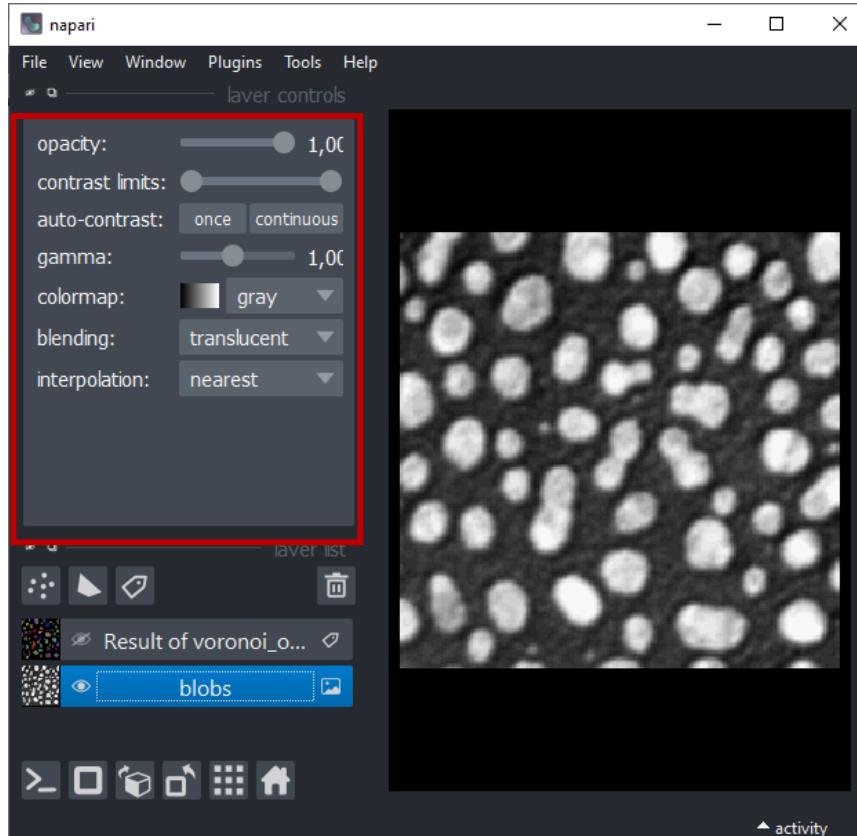


# Napari – Graphical User Interface



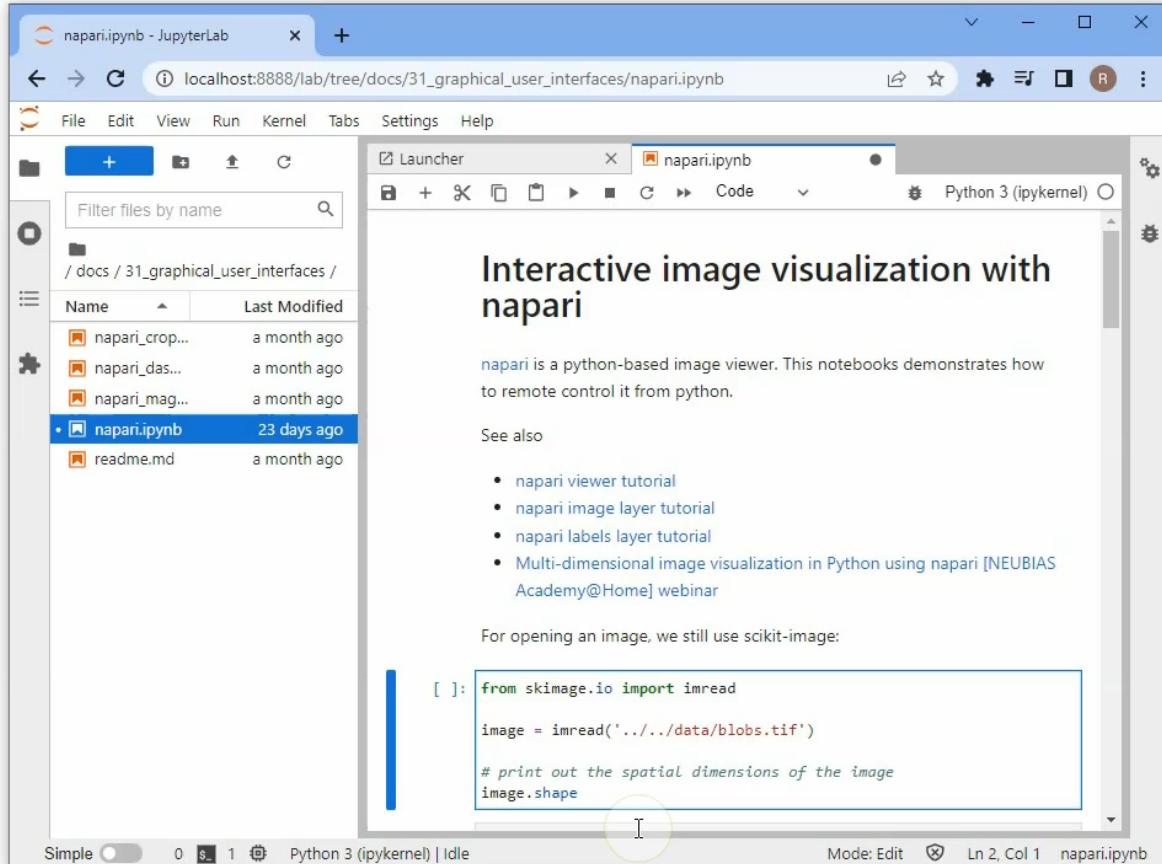
# Napari – Graphical User Interface

- Context / data type dependent tools



# Napari – Python Scripting

- Mixing interactivity and reproducibility



# Napari – Python Scripting

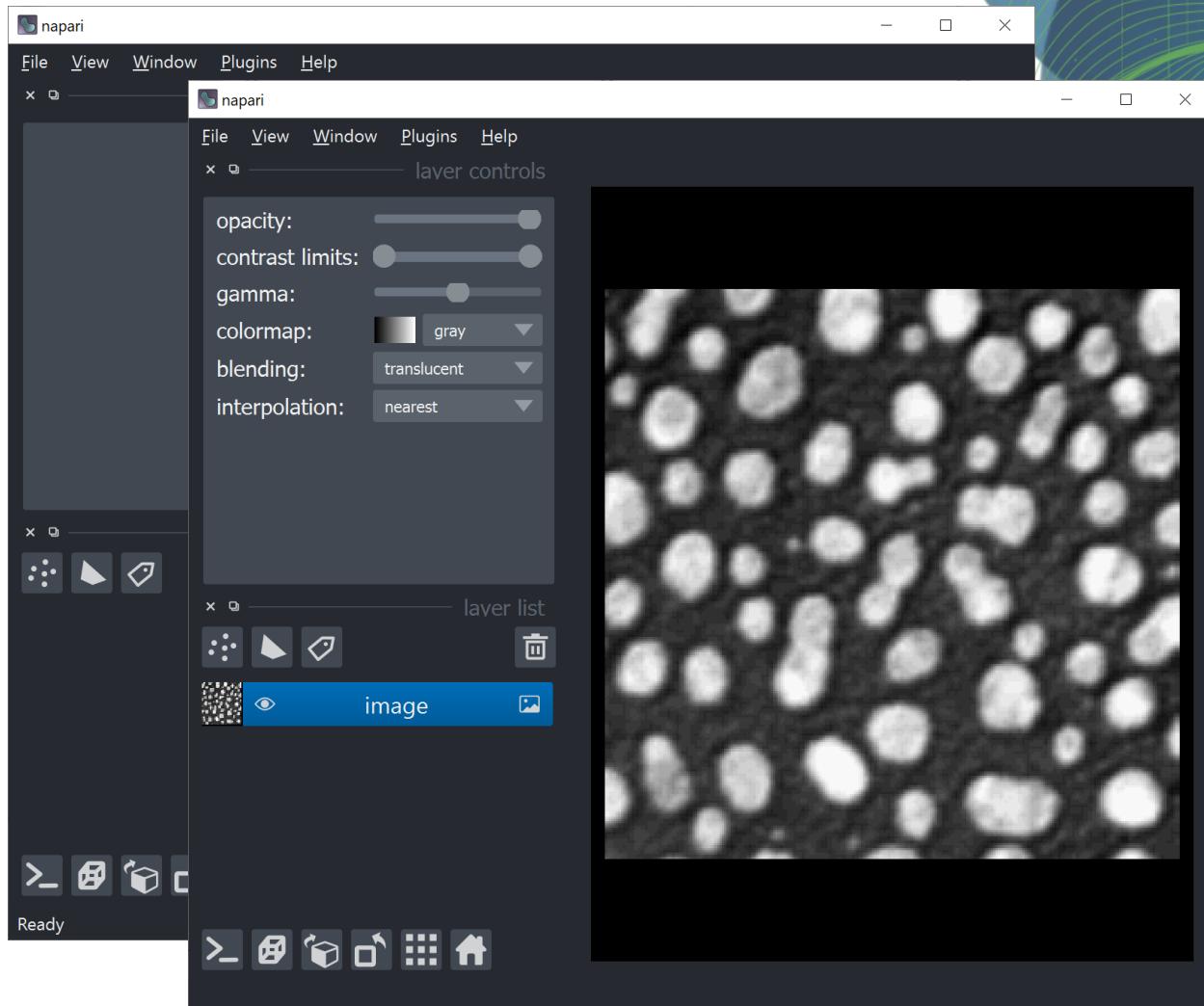
- Initialization

```
import napari
```

```
# Create an empty viewer
viewer = napari.Viewer()
```

- Adding images

```
viewer.add_image(image)
```



# Napari – Python Scripting

- Removing layers

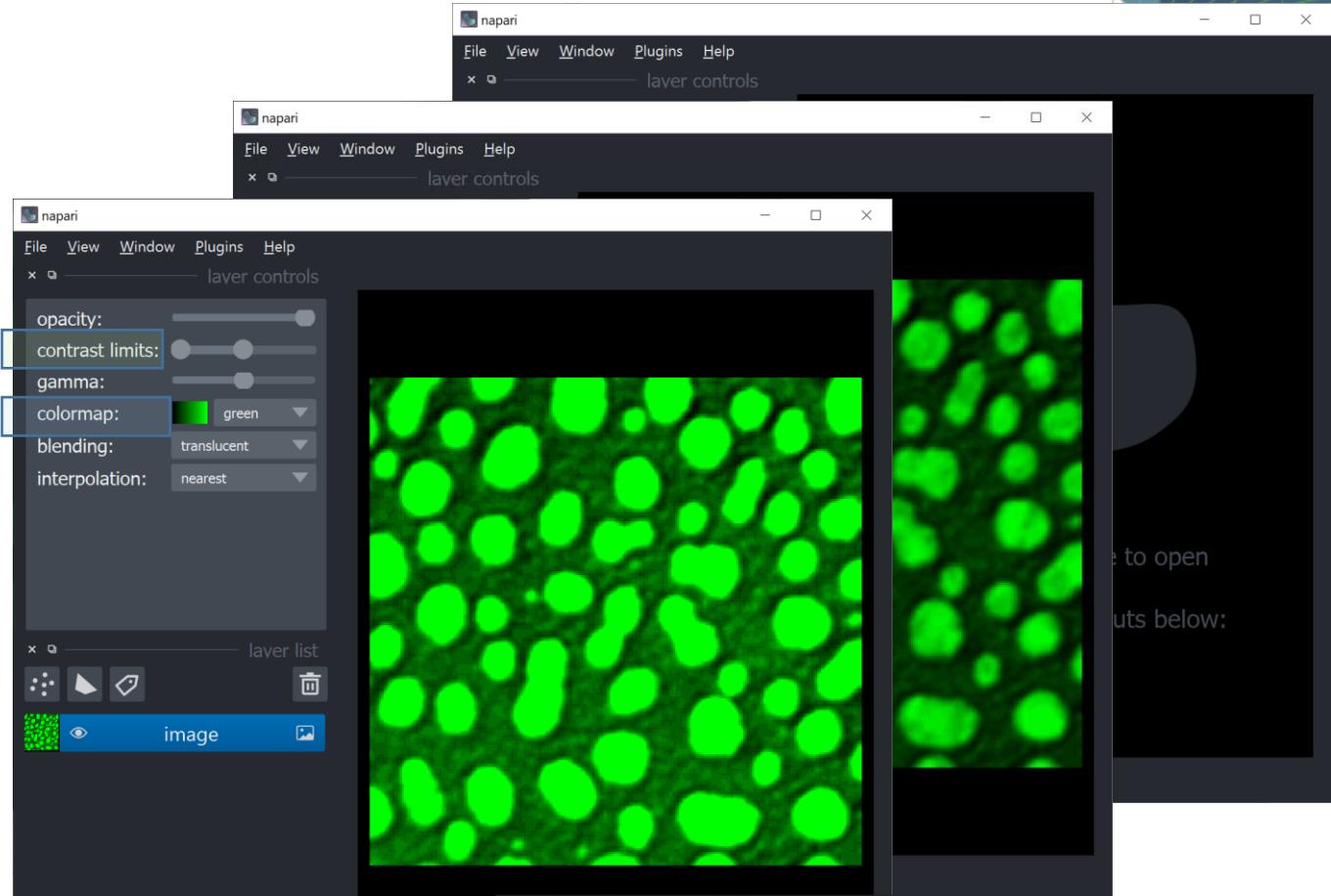
```
for l in viewer.layers:  
    viewer.layers.remove(l)
```

- Modify visualization while adding layers

```
viewer.add_image(image,  
                 colormap='green')
```

- Modify layers after adding

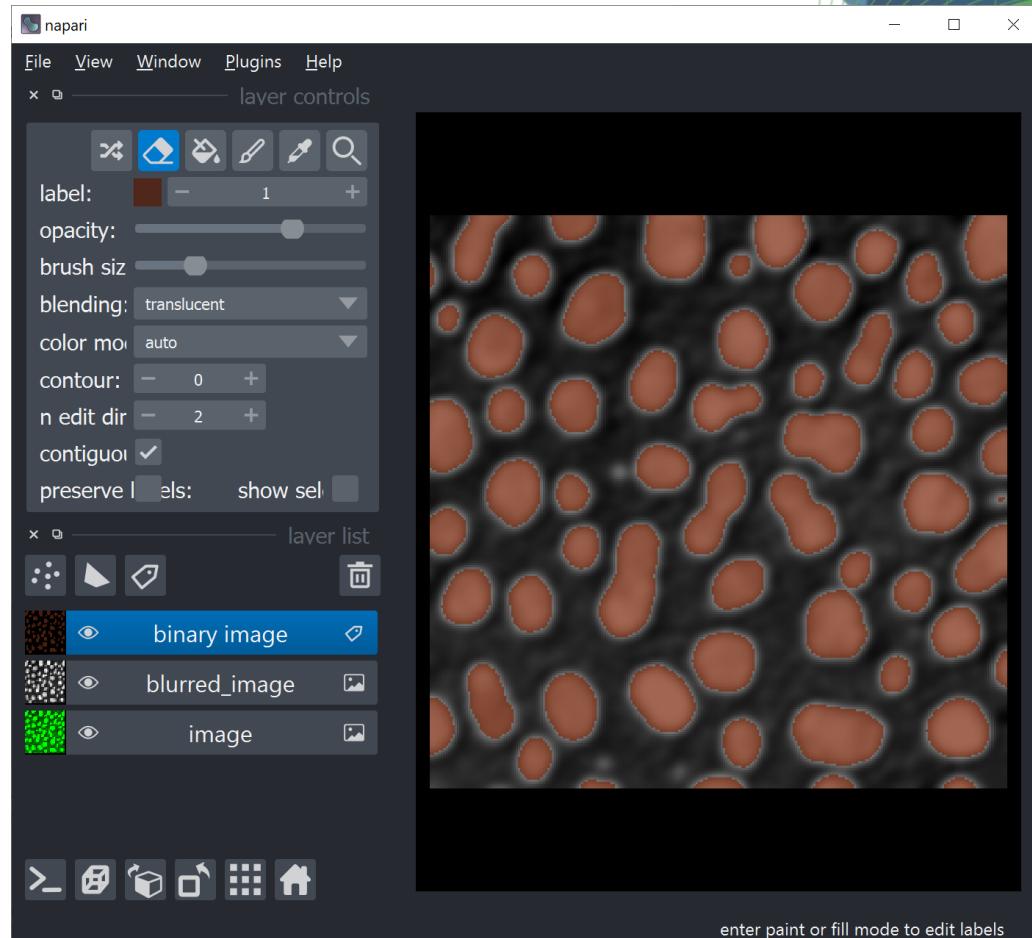
```
layer = viewer.add_image(image)  
layer.colormap = 'green'  
layer.contrast_limits = (0, 128)
```



# Napari – Python Scripting

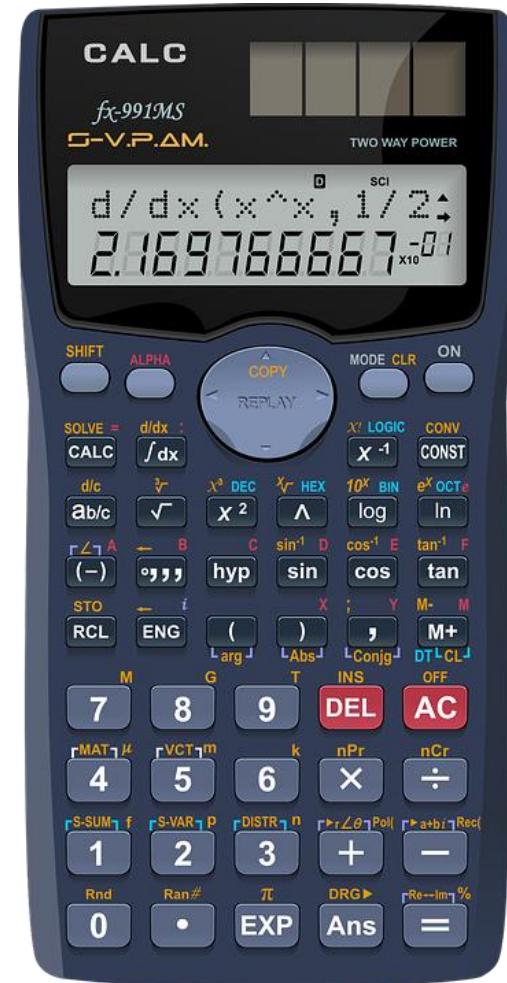
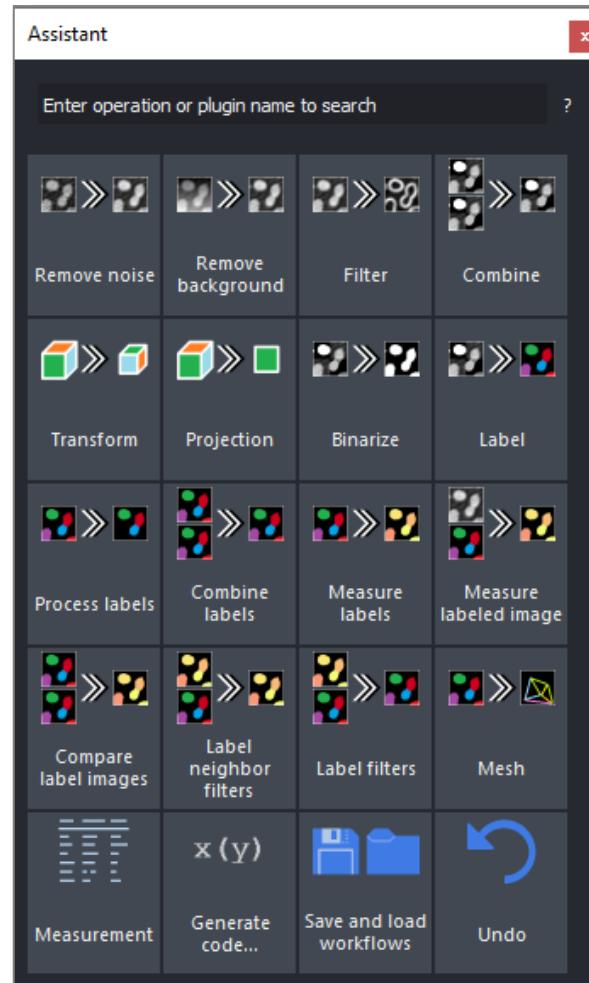
- Binary images and **label** images visualized as label layers

```
from skimage.filters import threshold_otsu  
  
threshold = threshold_otsu(blurred_image)  
  
binary_image = blurred_image > threshold  
  
# Add a new labels layer containing an image  
viewer.add_labels(binary_image)
```



# The Napari Assistant

- A pocket-calculator-like interface to build image analysis workflows



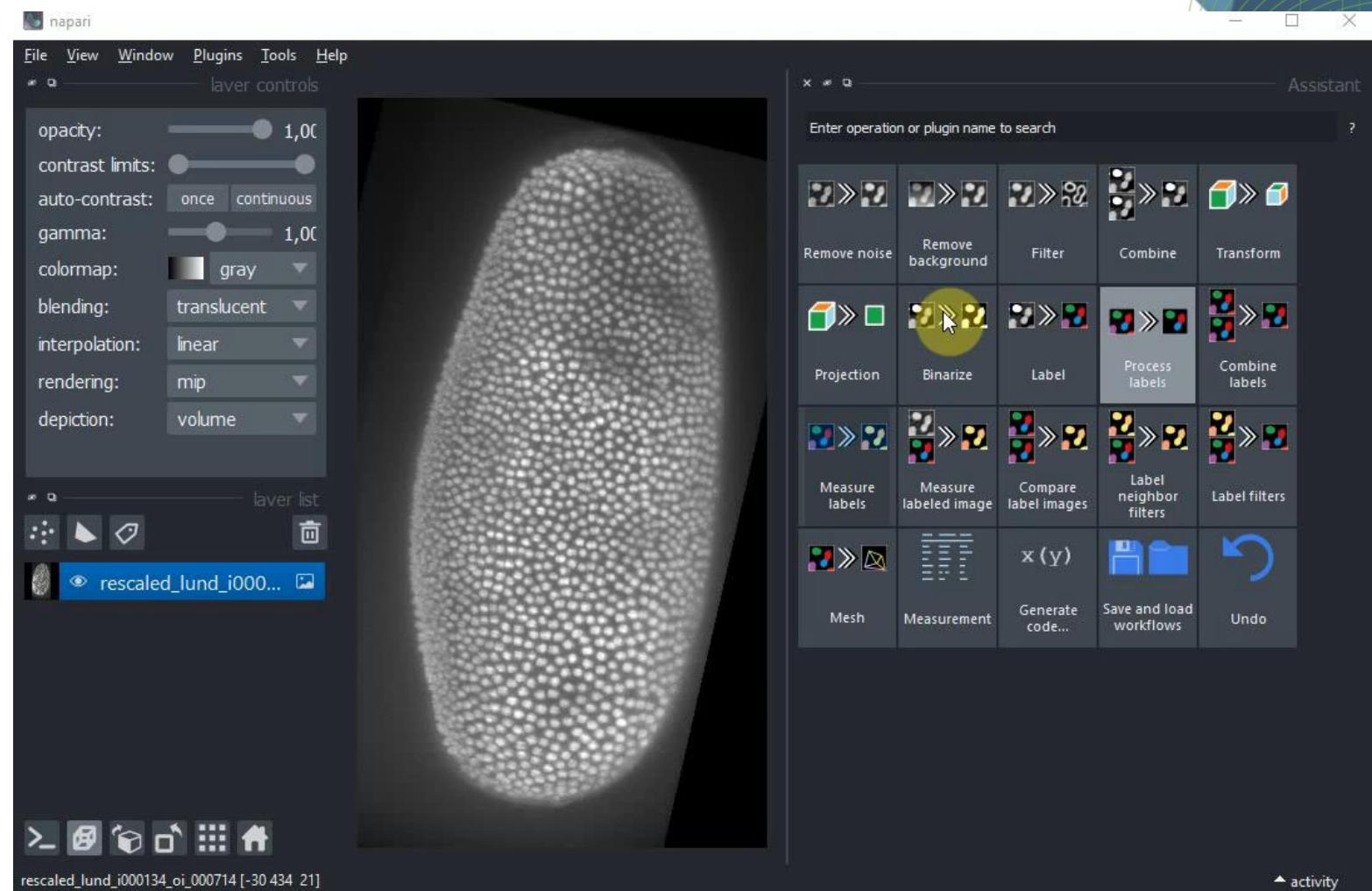
# The Napari Assistant

- Classical image processing operations + advanced tools
- Saving&loading supported
- Undo [redo]
- Hints for next steps
- ...

Big thanks to:

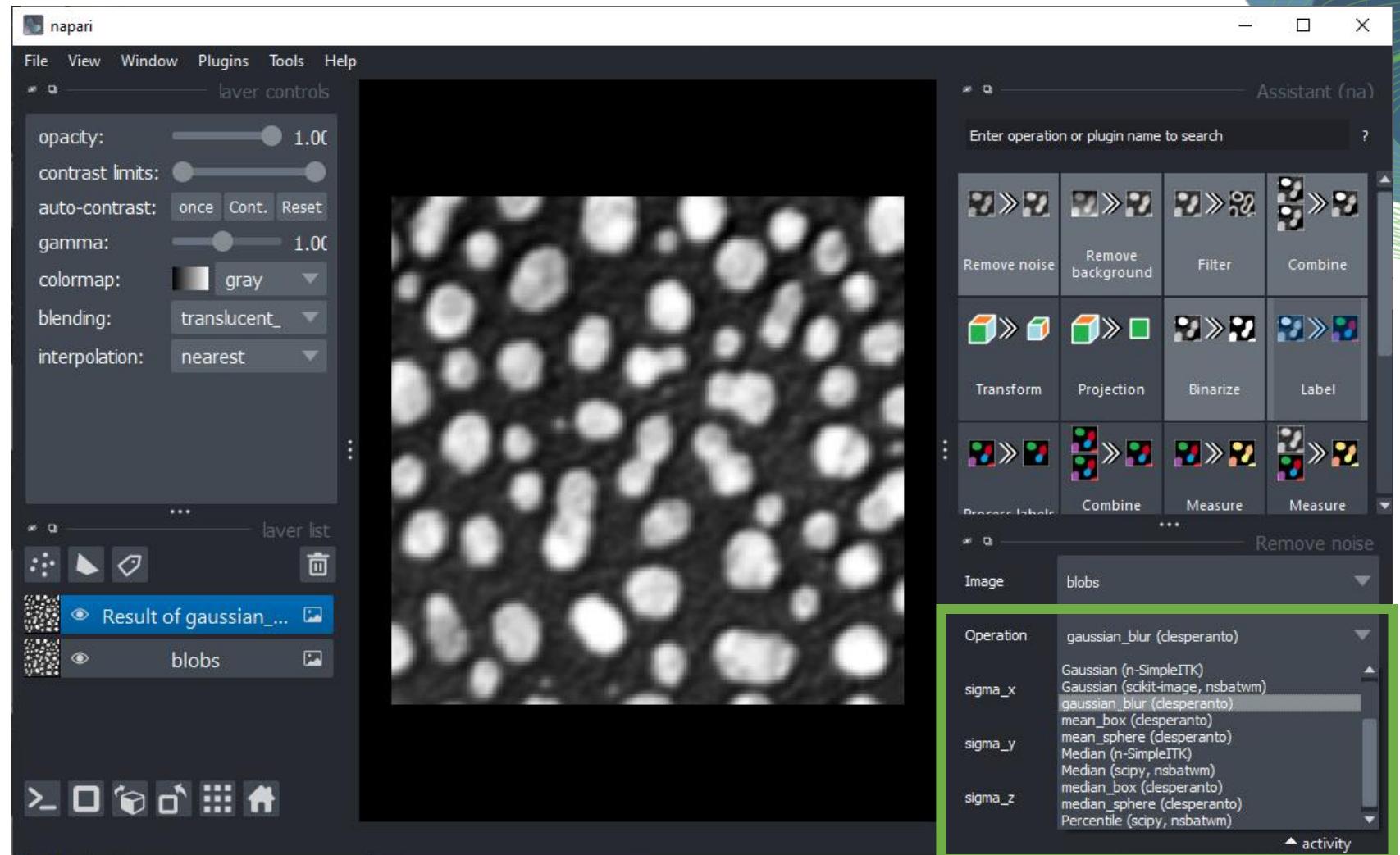


Ryan Savill  
@RyanSavill4



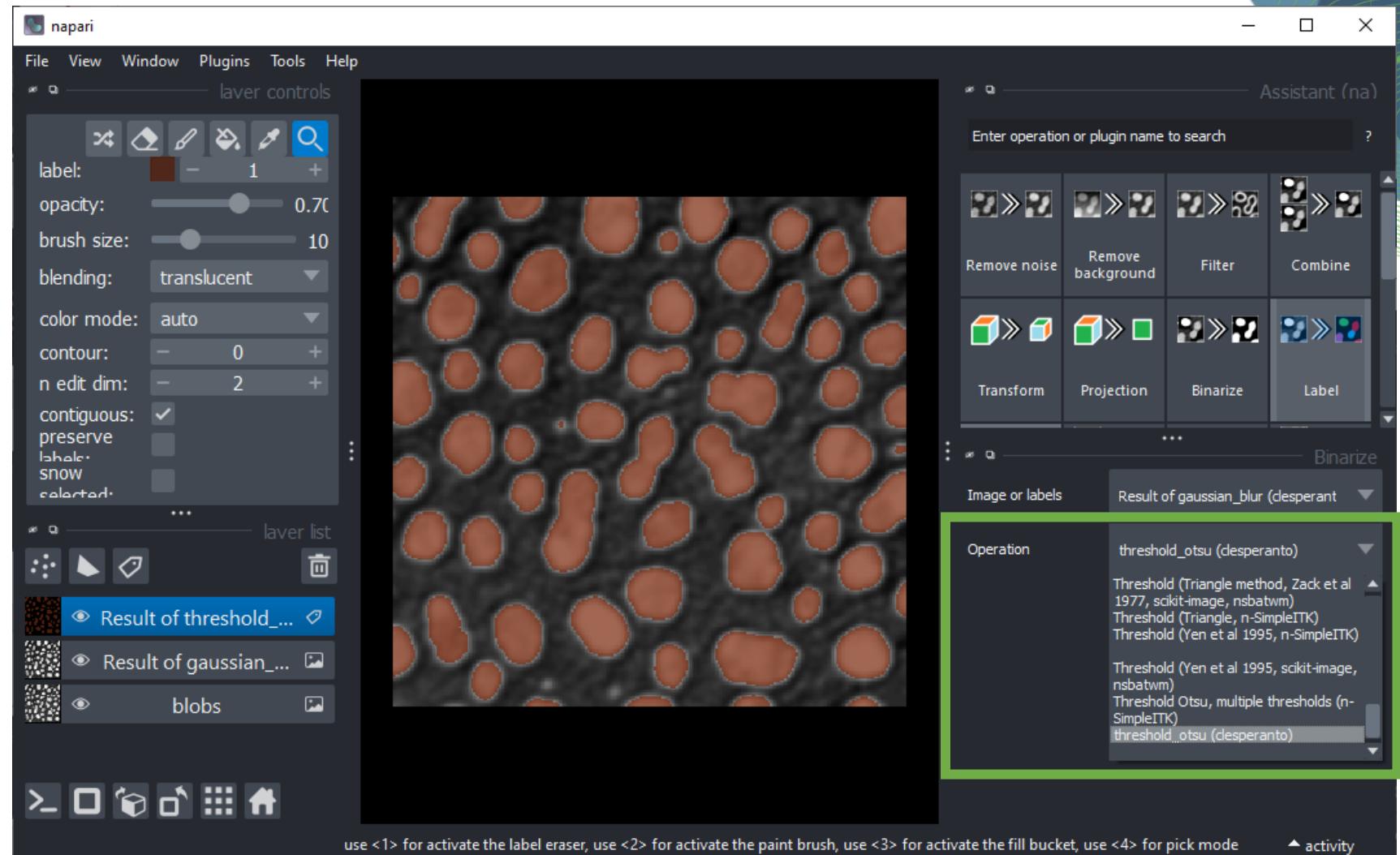
# Workflow building

- Try different algorithms, e.g. for removing noise
- Find them in the pulldown



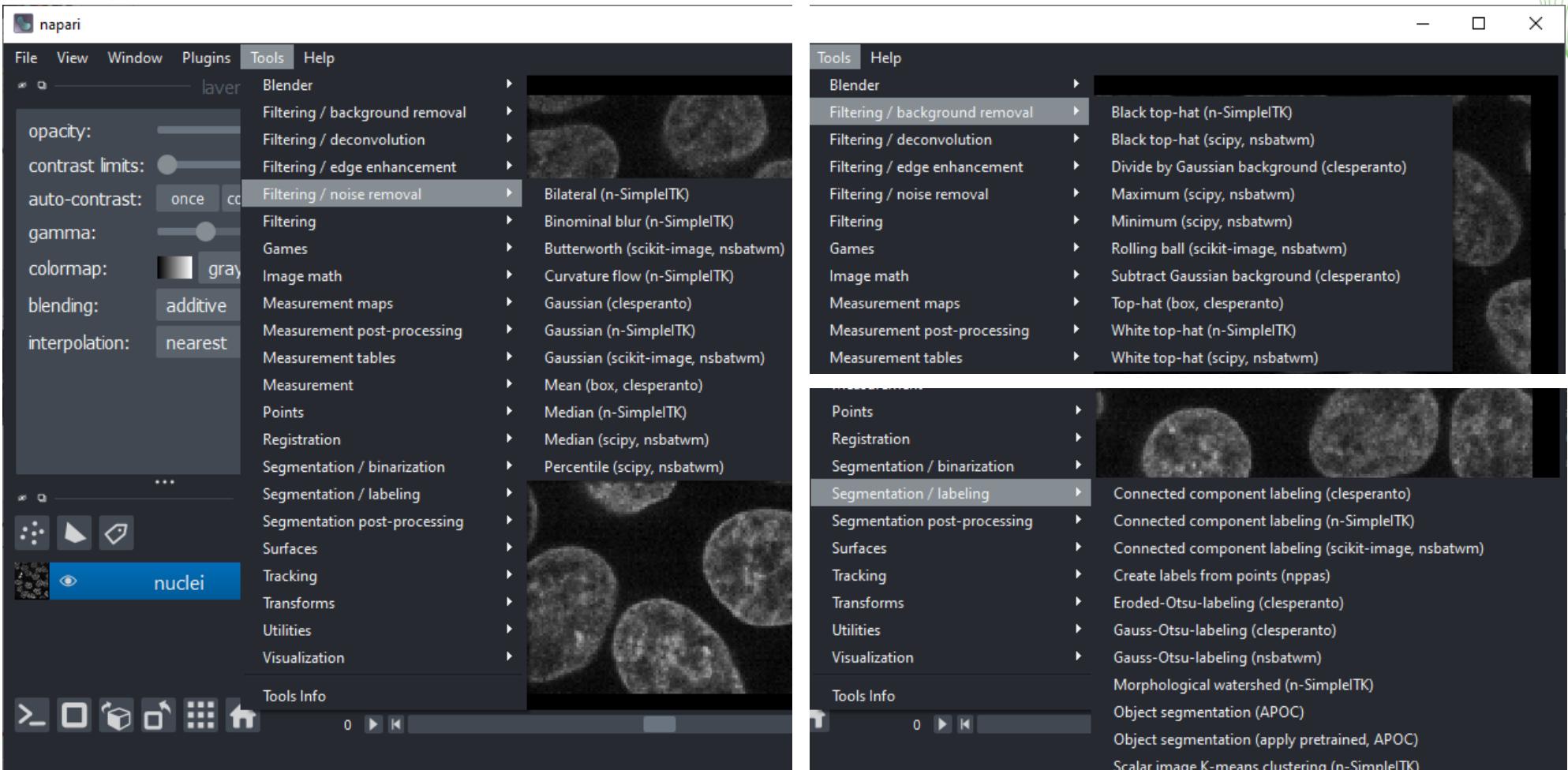
# Workflow building

- Try different binarization algorithms



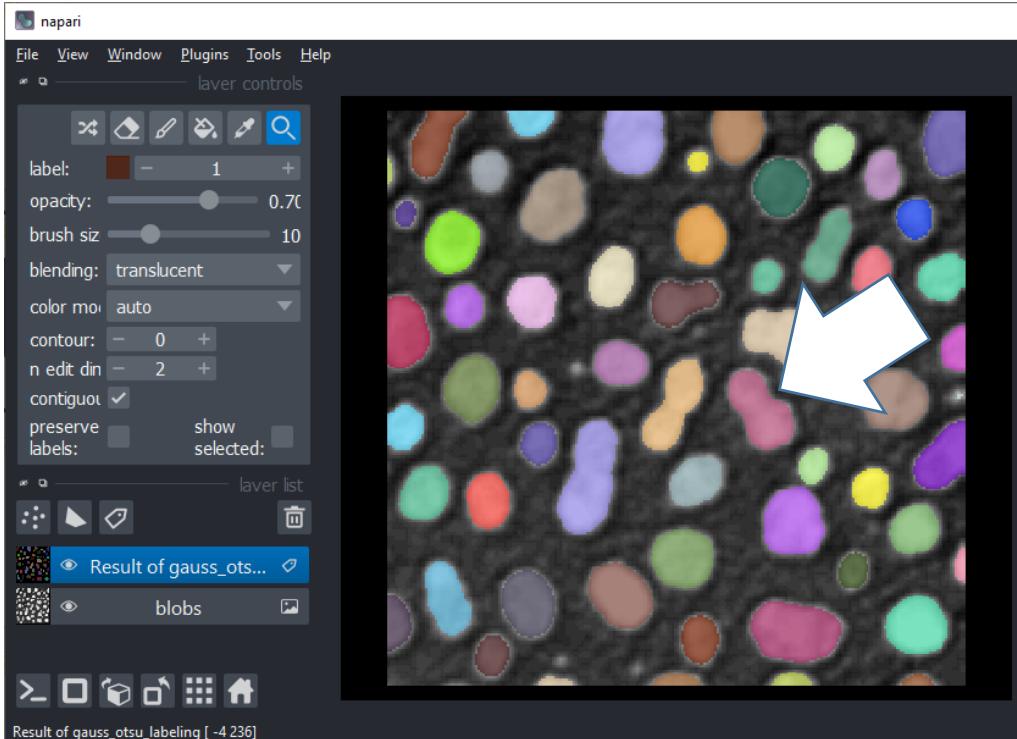
# The Tools menu

- Organized in categories

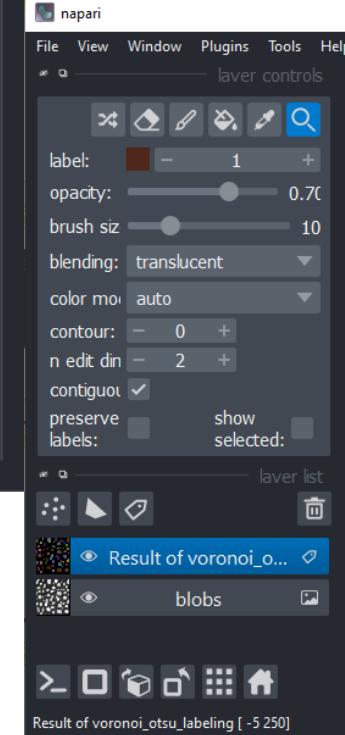
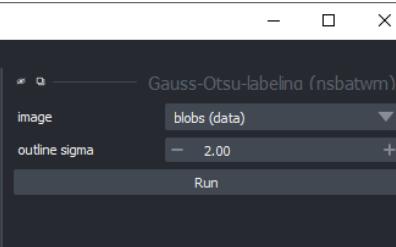


# Short-cuts: Voronoi-Otsu-Labeling

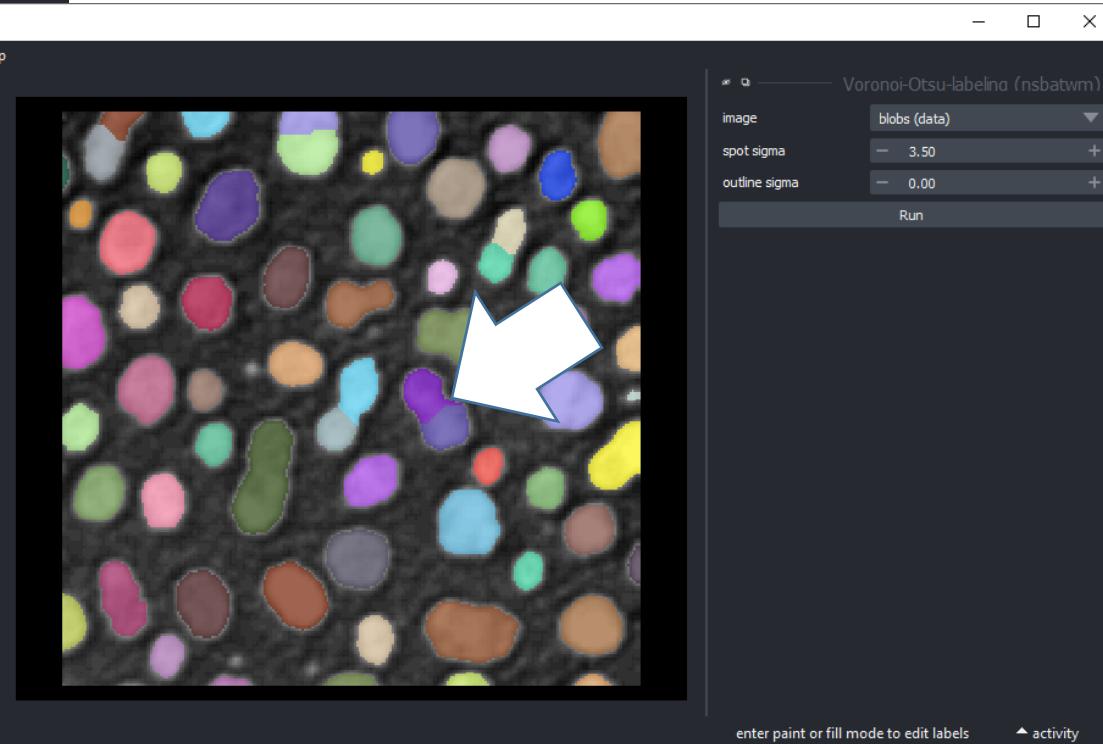
Also check out the Tools > Segmentation / labeling menu



Gauss-Otsu-Labeling

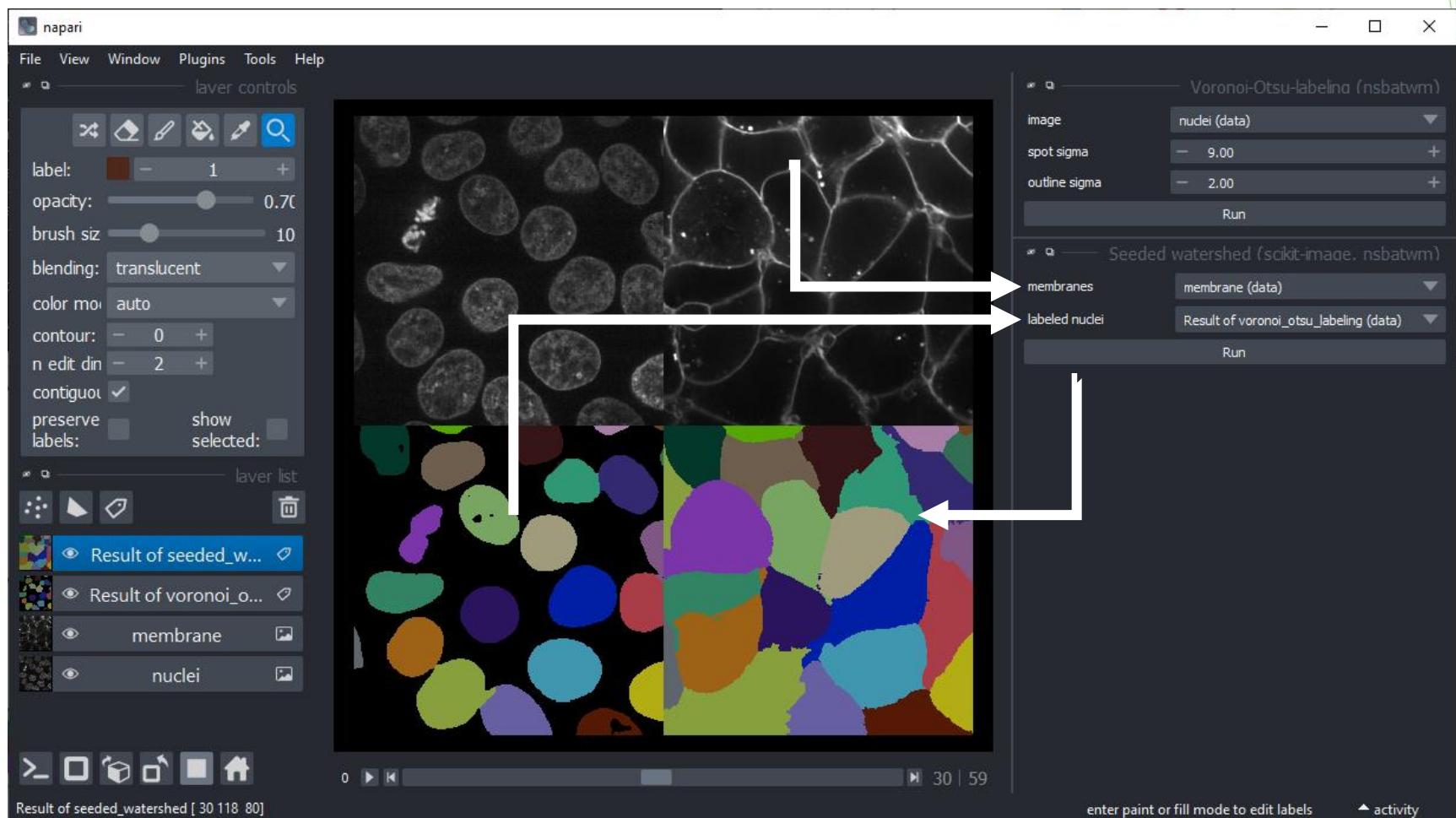


Voronoi-Otsu-Labeling



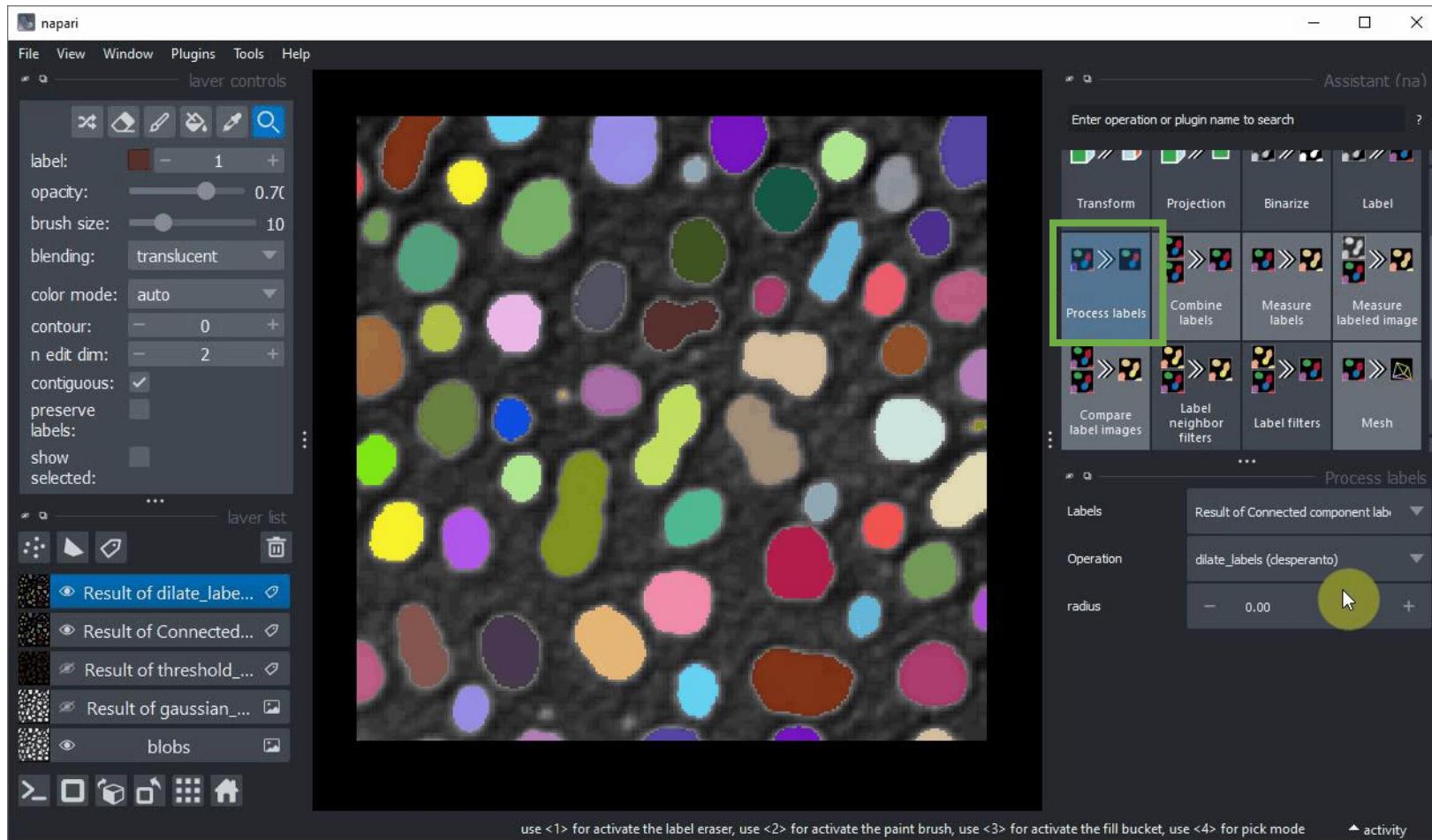
# Watershed

Also check out the Tools > Segmentation / labeling menu



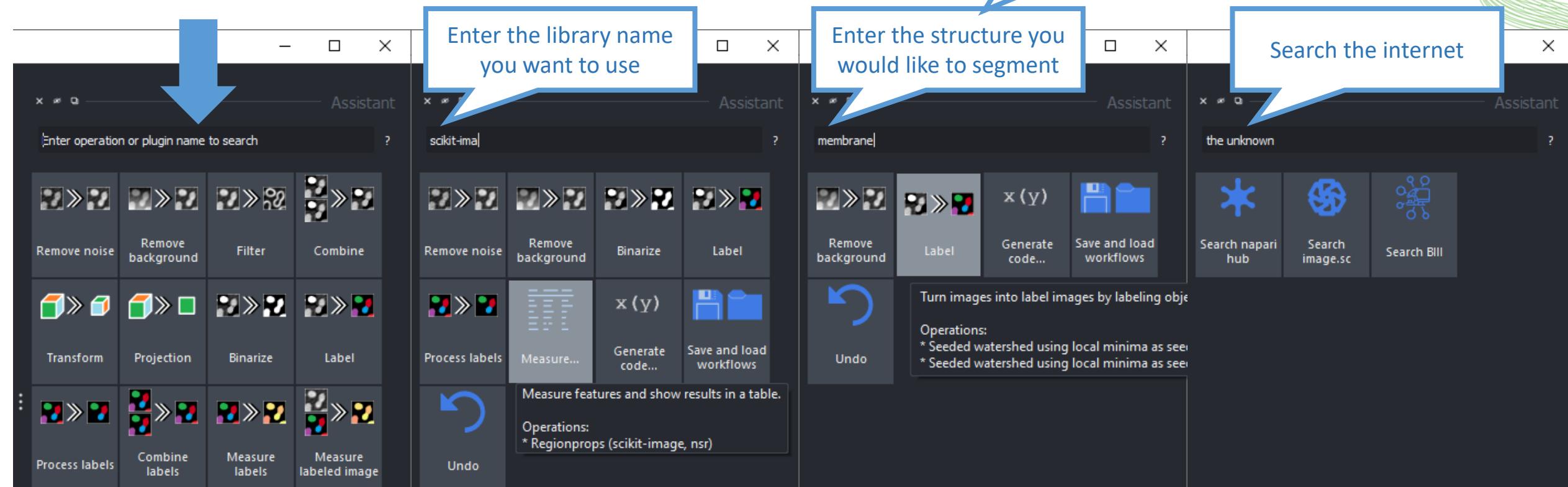
# Label erosion, dilation, opening, closing, ...

- In Napari Assistant: Process labels

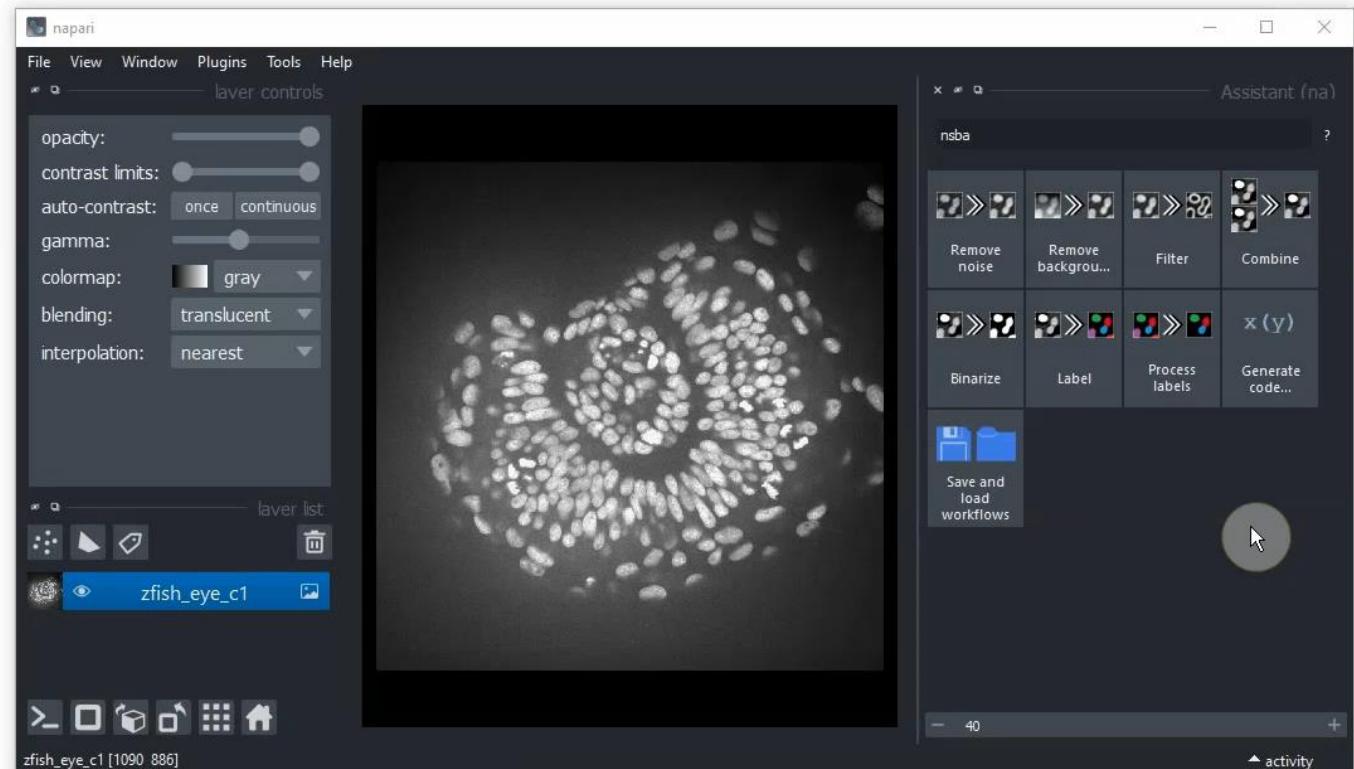


# Browse operations

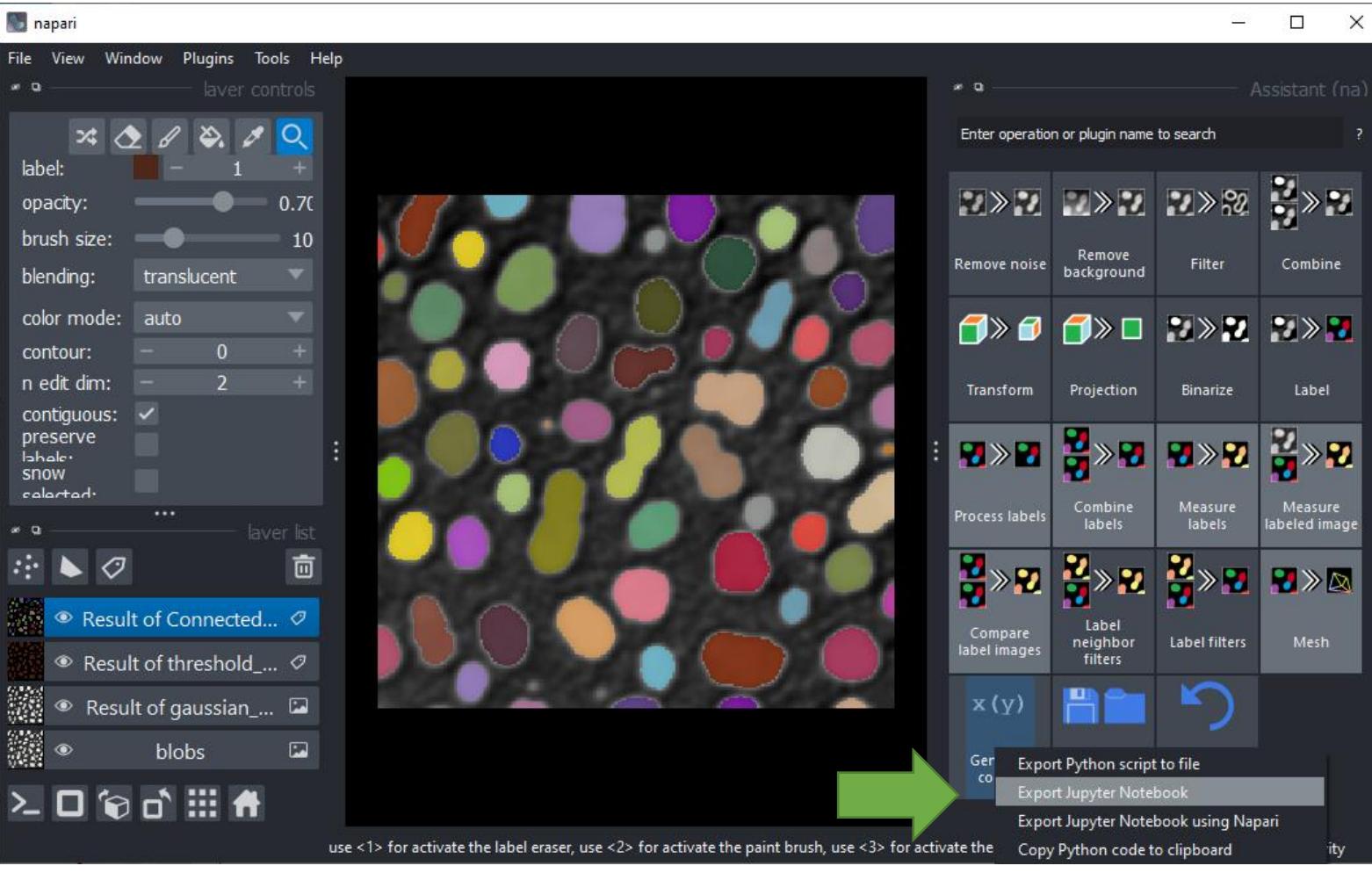
- Use the search...



# Export code to Jupyter Notebooks



# Export code to Jupyter Notebooks



The screenshot shows the napari Assistant application interface. On the left, there is a segmented image of various colored blobs on a black background. On the right, there is a grid of operation icons. A green arrow points from the bottom right towards the export options at the bottom of the interface. The export options include:

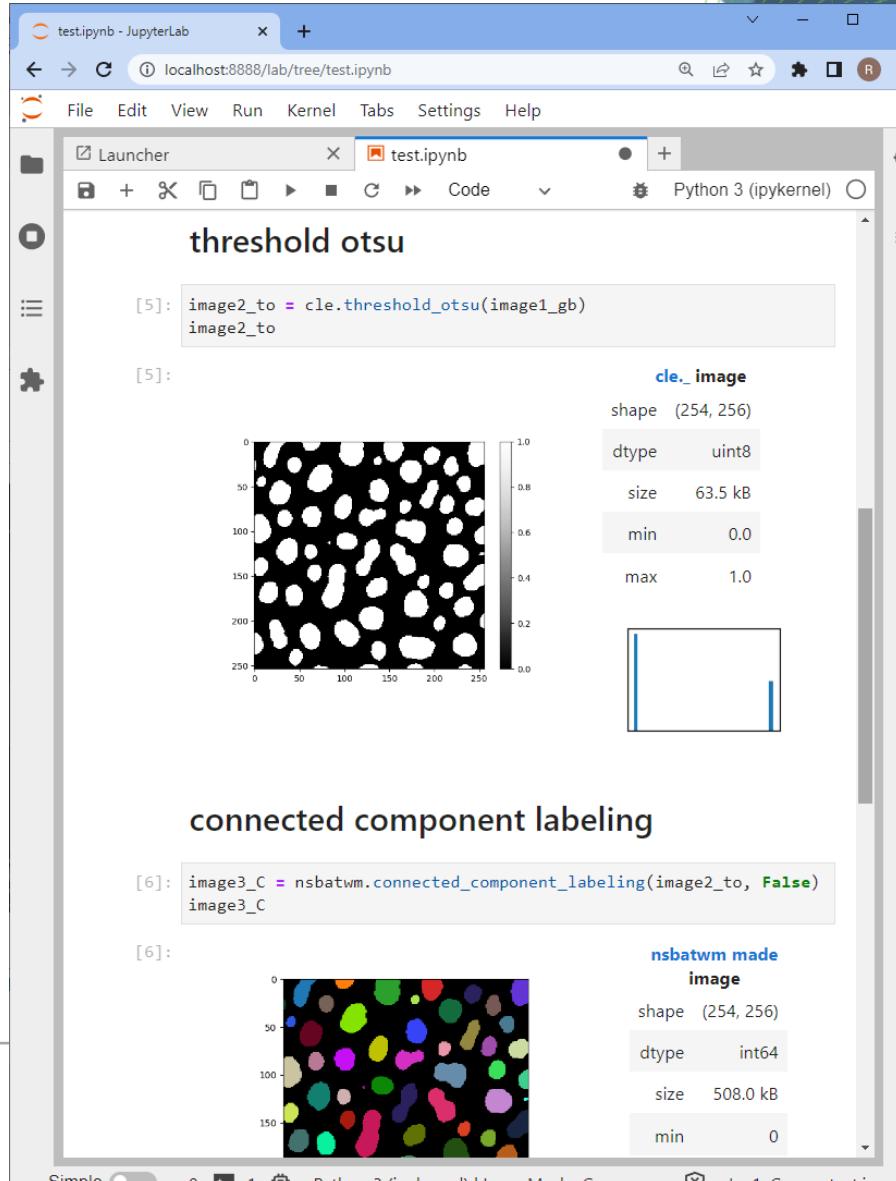
- Export Python script to file
- Export Jupyter Notebook** (highlighted)
- Export Jupyter Notebook using Napari
- Copy Python code to clipboard

On the right side of the image, there is a Jupyter Notebook window titled "test.ipynb - JupyterLab". It contains the following code and output:

```
threshold otsu
[5]: image2_to = cle.threshold_otsu(image1_gb)
image2_to
```

Output:

```
cle._image
shape (254, 256)
dtype uint8
size 63.5 kB
min 0.0
max 1.0
```



```
connected component labeling
[6]: image3_C = nsbatwm.connected_component_labeling(image2_to, False)
image3_C
```

Output:

```
nsbatwm made
image
shape (254, 256)
dtype int64
size 508.0 kB
min 0
```

At the bottom of the slide, there is a footer with the ScaDS.AI logo and some text:

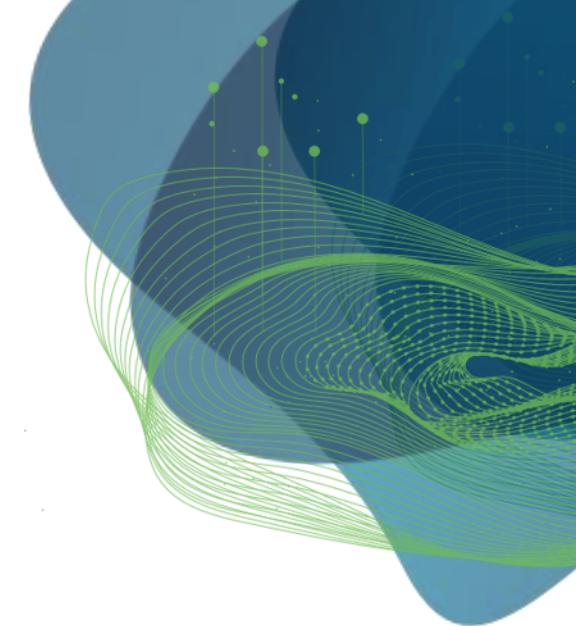
Robert Haase  
@haesleinhuepf  
BIDS Lecture 4/14  
April 23rd 2024

<https://github.com/haesleinhuepf/napari-assistant>  
#code-generation



DRESDEN LEIPZIG

CENTER FOR SCALABLE DATA ANALYTICS  
AND ARTIFICIAL INTELLIGENCE

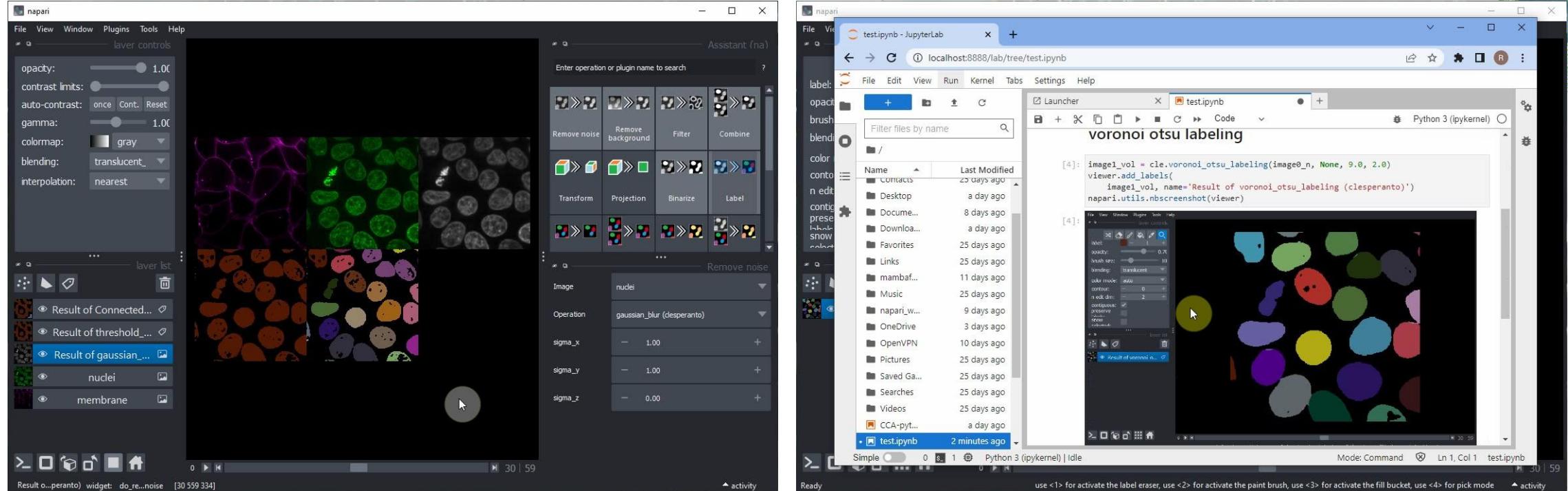


# Exercises

Robert Haase

# Napari - Exercises

- Start napari from the terminal!
- Follow the instructions to set up a workflow and export a Jupyter notebook



[https://github.com/ScaDS/BIDS-lecture-2024/blob/main/04b\\_napari\\_notebooks/napari-assistant.md](https://github.com/ScaDS/BIDS-lecture-2024/blob/main/04b_napari_notebooks/napari-assistant.md)

[https://github.com/ScaDS/BIDS-lecture-2024/blob/main/04b\\_napari\\_notebooks/notebook\\_export.md](https://github.com/ScaDS/BIDS-lecture-2024/blob/main/04b_napari_notebooks/notebook_export.md)

# Napari - Exercises

- Start using napari from Python

The image shows two JupyterLab windows side-by-side, both titled "napari\_intro.ipynb - JupyterLab".

**Left Window:** The title bar says "Opening the napari Viewer". It contains the following code and output:

```
[2]: import napari  
Now, we can open the viewer with the following command:  
[3]: viewer = napari.Viewer()  
Napari should open in a separated window. Some warning messages in the cell above are normal.  
Let's show a screenshot of the viewer here. We pass the variable viewer to the function.  
[4]: napari.utils.nbscreenshot(viewer)
```

The output shows a screenshot of the Napari viewer window, which is dark with a single blue segmented region.

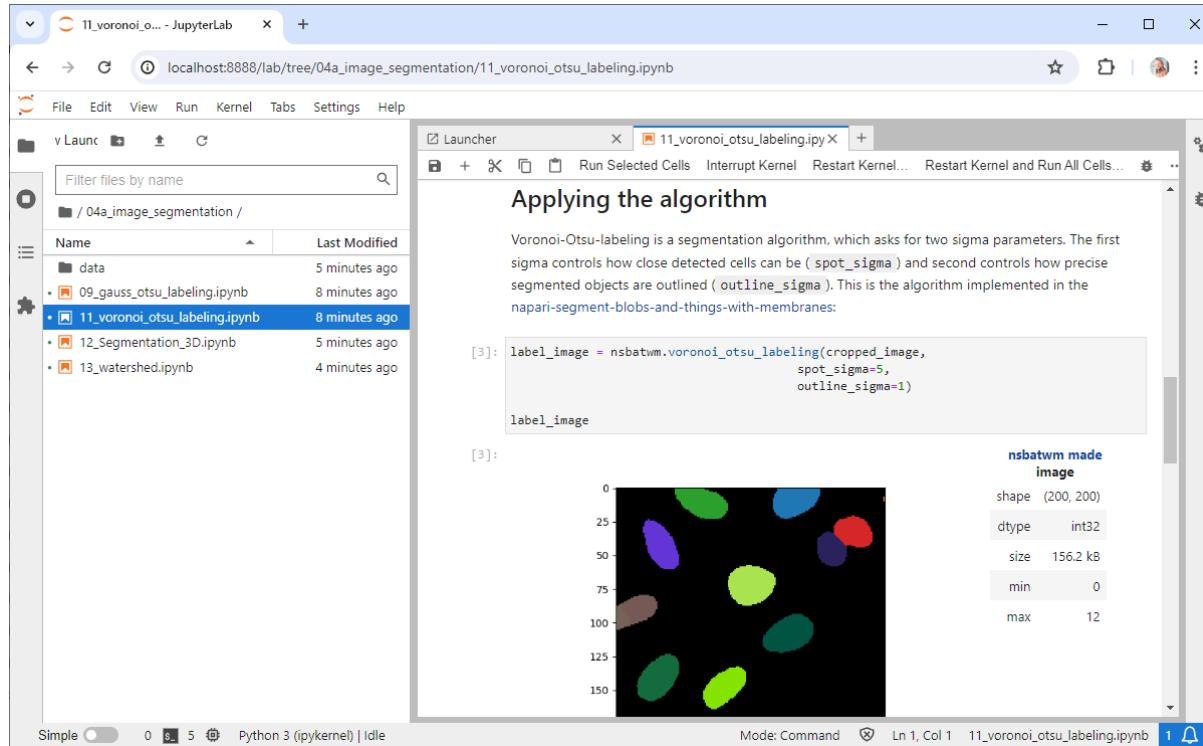
**Right Window:** The title bar says "Segmentation visualization". It contains the following code and output:

```
[13]: blurred = gaussian(mri, sigma=5)  
binary_image = blurred > threshold_otsu(blurred)  
viewer.add_labels(binary_image)  
napari.utils.nbscreenshot(viewer)
```

The output shows a screenshot of the Napari viewer window displaying a grayscale MRI scan with a red segmented region overlay.

# Image segmentation exercises

- Try out segmentation algorithms and apply them to other datasets



localhost:8888/lab/tree/04a\_image\_segmentation/11\_voronoi\_otsu\_labeling.ipynb

File Edit View Run Kernel Tabs Settings Help

Applying the algorithm

Voronoi-Otsu-labeling is a segmentation algorithm, which asks for two sigma parameters. The first sigma controls how close detected cells can be (spot\_sigma) and second controls how precise segmented objects are outlined (outline\_sigma). This is the algorithm implemented in the napari-segment-blobs-and-things-with-membranes:

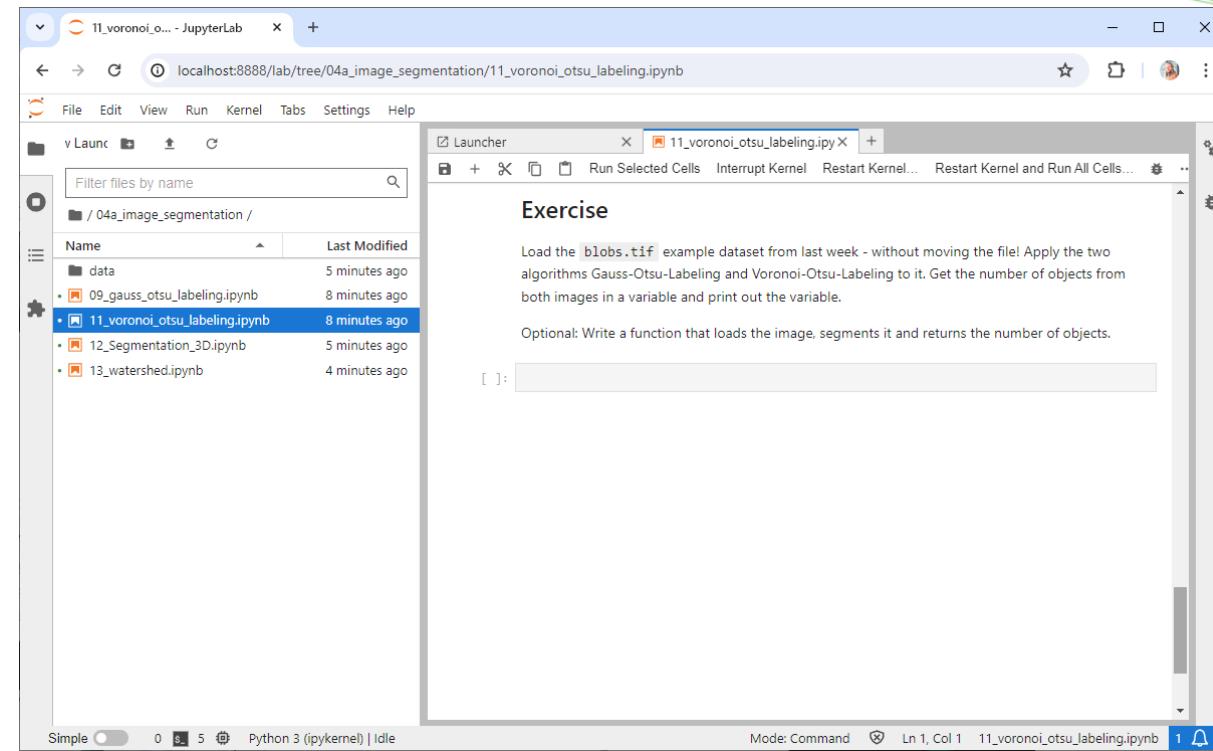
```
[3]: label_image = nsbatwm.voronoi_otsu_labeling(cropped_image,
                                                spot_sigma=5,
                                                outline_sigma=1)
```

label\_image

nsbatwm made image  
shape (200, 200)  
dtype int32  
size 156.2 kB  
min 0  
max 12

0 25 50 75 100 125 150

Mode: Command Ln 1, Col 1 11\_voronoi\_otsu\_labeling.ipynb 1



localhost:8888/lab/tree/04a\_image\_segmentation/11\_voronoi\_otsu\_labeling.ipynb

File Edit View Run Kernel Tabs Settings Help

Exercise

Load the blobs.tif example dataset from last week - without moving the file! Apply the two algorithms Gauss-Otsu-Labeling and Voronoi-Otsu-Labeling to it. Get the number of objects from both images in a variable and print out the variable.

Optional: Write a function that loads the image, segments it and returns the number of objects.

```
[ ]:
```

Simple 0 5 Python 3 (ipykernel) | Idle Mode: Command Ln 1, Col 1 11\_voronoi\_otsu\_labeling.ipynb 1