Complete RNA Virus Genomes Assembled from Murine Cecal Meta
Running title: RNA Viral Genomes from Murine Metatranscriptomes
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Abstract

- ² Efforts to catalog viral diversity in the gut microbiome have largely ignored RNA viruses focused on
- DNA viruses, while RNA viruses remain understudied. To address this, we screened assemblies of
- 4 previously published mouse gut metatranscriptomes for the presence of RNA viruses. We identified
- 5 the complete coding-complete genomes of an Astrovirus and 5 Mitovirus-like viruses.

The viral fraction of the mammalian gut microbiome forms a crucial component in the relationship between microbe and host. Bacterial viruses serve as an important source of genetic diversity and population control for the microbiota, driving its ecology and evolution (1). Mammalian viruses disrupt the gut environment through infection and the response of the host immune system (2). Bacterial and mammalian viruses make significant contributions to host health and disease. Current efforts to describe the diversity of viruses present in the gut have focused on using shotgun metagenomics to identify double-stranded DNA viruses, predominantly bacteriophage and host pathogens (3). However, this method ignores viruses with RNA genomes, which make up a considerable portion of the environmental viromes (4).

We re-analyzed deeply-sequenced metatranscriptome data produced by our lab for the study of 15 microbiome dynamics in a mouse model of Clostridioides difficile infection (5, 6). Briefly, C57Bl/6 mice from a breeding colony we maintain at the University of Michigan were treated with one of three different antibiotics (clindamycin, streptomycin, or cefoperazone). After a 24 hour recovery 18 period, the mice were infected with C. difficile strain 630. Germ free C57Bl/6 mice were also 19 mono-associated with C. difficile strain 630. Cecal contents were removed from each animal 18 hours post infection and frozen for RNA extraction and sequencing. RNA sequences from each 21 sample were trimmed of adapter sequences and low-quality bases using Trimmomatic v0.39 and assembled individually using rnaSPAdes v3.13.1 (7) and concatenated for dereplication, resulting in 70,779 contigs longer than 1 kb. Contigs were then screened against a custom RefSeg database of viral RNA-dependent RNA polymerase (RdRP) protein sequences screened for the presence of 25 RdRP coding sequences using Blast v2.9.0 against a database containing all Viral RefSeq protein sequences annotated as RdRP (screening database available online as described below) with a maximum e-value of 10⁻²⁰, resulting in 29 contigs. RdRP is conserved amongst almost all RNA 28 viruses without a DNA stage in genome replication. These contigs were then annotated with Interproscan v5.39-77.0 (8, 9). We constructed phylogenetic trees from RdRP protein sequences using IQ-TREE v1.6.12 (10). 31

Two classes of RNA viruses were assembled with high coverage with sequences originating from most of the mouse treatment groups, including germ-free mice. First, a 6,811 base-long astrovirus genome (GC 56.6%) was obtained with 1,683.5-fold coverage (Figure 1A). The genome contained 3

predicted open reading frames encoding a capsid, RdRP, and a trypsin-like peptidase, and appears
to be closely related to Murine Astrovirus in Astroviridae. Second, 5 distinct, but closely related
RNA virus genomes, designated Putative Mitovirus JS1 through JS5, ranging in length from 2,309
to 2,447 bases with 4.6 to 16,078.8-fold coverage and average GC content of 46.2% belonged to
a previously undescribed clade of Narnaviridae Narnaviridae adjacent to the Mitoviruses (Figure
1B). These RNA virus genomes will facilitate future studies of RNA virus biology in the murine
microbiome.

Data Availability. The RNA-seq data are available the NCBI Sequence Read Archive (SRA) database under the accession numbers PRJNA354635 (*C. difficile* infected mice) and PRJNA415307 (mock-infected mice). The assembled genomes are available at the National Center for Biotechnology Information (NCBI) GenBank under the accession numbers MN780842-MN780847. All of the scripts and software used to perform this analysis are available at https://github.com/SchlossLab/Stough Mouse RNA Virome MRA 2019.

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51 References

- 1. **Ogilvie LA**, **Jones BV**. 2015. The human gut virome: A multifaceted majority. Frontiers in Microbiology **6**:918. doi:10.3389/fmicb.2015.00918.
- ⁵⁴ 2. Legoff J, Resche-Rigon M, Bouquet J, Robin M, Naccache SN, Mercier-Delarue S,
- Federman S, Samayoa E, Rousseau C, Piron P, Kapel N, Simon F, Socié G, Chiu CY. 2017.
- 56 The eukaryotic gut virome in hematopoietic stem cell transplantation: New clues in enteric
- graft-versus-host disease. Nature Medicine 23:1080–1085. doi:10.1038/nm.4380.
- 3. Garmaeva S, Sinha T, Kurilshikov A, Fu J, Wijmenga C, Zhernakova A. 2019. Studying
- the gut virome in the metagenomic era: Challenges and perspectives. BMC Biology 17:84.
- 60 doi:10.1186/s12915-019-0704-y.
- ⁶¹ 4. Culley A. 2018. New insight into the RNA aquatic virosphere via viromics. Virus Research
- 62 **244**:84–89. doi:10.1016/j.virusres.2017.11.008.
- 5. Jenior ML, Leslie JL, Young VB, Schloss PD. 2017. Clostridium difficile colonizes alternative
- nutrient niches during infection across distinct murine gut microbiomes. mSystems 2:e00063–17.
- 65 doi:10.1128/mSystems.00063-17.
- 66 6. Jenior ML, Leslie JL, Young VB, Schloss PD. 2018. Clostridium difficile alters the structure
- 67 and metabolism of distinct cecal microbiomes during initial infection to promote sustained
- colonization. mSphere **3**:e00261–18. doi:10.1128/mSphere.00261-18.
- 99 7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM,
- Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev
- 71 MA, Pevzner PA. 2012. SPAdes: A new genome assembly algorithm and its applications to
- ₇₂ single-cell sequencing. Journal of Computational Biology: A Journal of Computational Molecular
- 73 Cell Biology **19**:455–477. doi:10.1089/cmb.2012.0021.
- 8. UFBoot2: Improving the ultrafast bootstrap approximation.
- ₇₅ 9. Kalyaanamoorthy S, Minh BQ, Wong TKF, Haeseler A von, Jermiin LS. 2017. ModelFinder:

- Fast model selection for accurate phylogenetic estimates. Nature Methods 14:587–589.
- 77 doi:10.1038/nmeth.4285.
- 10. Nguyen L-T, Schmidt HA, Haeseler A von, Minh BQ. 2015. IQ-TREE: A fast and effective
- 79 stochastic algorithm for estimating maximum-likelihood phylogenies. Molecular Biology and
- 80 Evolution **32**:268–274. doi:10.1093/molbev/msu300.

Figure 1. Phylogenetic trees showing the relatives of the metatranscriptome assembled genomes. Maximum Likelihood phylogenetic trees constructed from RdRP amino acid sequences for (A) Astroviruses and (B) Narnaviruses. Node annotations represent IQTree Ultra-fast Bootstrap statistics, values less than 50% were excluded from the tree. Scale bars are marked in red to the left of each tree. Highlight colors in (B) represent major Narnavirus taxa: Orange - Ourmiaviruses, Pink - Ourmia-like Mycoviruses, Gray - Narnaviruses, Blue - Mitoviruses, Purple - Murine Mitovirus-like viruses, Green - Leviviruses.