Argonne_Seed_seq_processing_June2023

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Code used on the MSU HPCC for QIIME2 workflow

Copy demultiplexed sequences into working space from raw_sequence directory

```
#!/bin/bash -login
#SBATCH --time=03:59:00
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --mem=30G
#SBATCH -- job-name copy_sequences
#SBATCH --mail-user=suleskya@msu.edu
#SBATCH --mail-type=BEGIN, END
###### Job code
cp *.fastq /mnt/research/ShadeLab/WorkingSpace/Sulesky/New_Argonne_Seed_Sequencing/input_data
echo -e "\n `sacct -u suleskya -j $$LURM JOB ID --format=JobID, JobName, Start, End, Elapsed, NCPUS, ReqMem`
scontrol show job $SLURM_JOB_ID
```

submit job: sbatch copy_seqs.sb

Rename files for Figaro to run correctly

rename using the util-linux command "rename"

use module load to load util-linux

```
module spider util-linux
# choose most recent version
module spider util-linux/2.37
# follow instructions to load
module load GCCcore/11.2.0
module load util-linux/2.37
module list #check which packages are loaded
```

Copy sequencing files into figaro_input directory: /mnt/research/ShadeLab/WorkingSpace/Sulesky/New_Argonne_Seed_Sequencing files fil

```
Use 'rename' to make files match illumina naming convention: SampleName_S1_L001_R1_001.fastq Example: > rename G0_ G0 *.fastq this code takes all files that end in *fastq, replaces "G0_" with "G0" had to run various lines to remove extra underscores in sample IDs and add L001 to all rename goes really quickly, \sim1 second per command
```

Figaro for trim parameters

make figaro output folder in workingspace directory /New_Argonne_Seed_Sequencing/figaro go to home directory where figaro is installed to run figaro: cd /mnt/home/suleskya/figaro-master/figaro Run figaro as a job:

nano figaro_argonne_june2023.sb

```
#!/bin/bash -login
######## SBATCH Lines for Resource Request #########
#SBATCH --time=3:59:00
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=1
#SBATCH --mem=64G
#SBATCH -- job-name figaro
#SBATCH -A shade-cole-bonito
#SBATCH --mail-user=suleskya@msu.edu
#SBATCH --mail-type=BEGIN, END
######## Command Lines for Job Running #########
conda activate figaro
python figaro.py -i /mnt/research/ShadeLab/WorkingSpace/Sulesky/New_Argonne_Seed_Sequencing/figaro/figa
conda deactivate
echo -e "\n `sacct -u suleskya -j $$LURM_JOB_ID --format=JobID, JobName, Start, End, Elapsed, NCPUS, ReqMem`
scontrol show job $SLURM_JOB_I
sbatch figaro argonne june2023.sb
check job progress with "sq" command
Once job is finished, check the figure output:
cd
     less trimParameters.json { "trimPosition": [191, 84], "maxExpectedError": [2, 2], "readReten-
     tionPercent": 94.64, "score": 92.63569499836179 },
     control Z to exit "less"
```

Import data into Qiime2 format

Have to rename files in input-data for Qiime2 input with the same method as above

we will truncate the sequences at forward 191 and reverse 84, which will merge 94.64 percent of the reads

(base) -bash-4.2\$ rename G0_ G0. .fastq (base) -bash-4.2\$ rename G1_ G1. .fastq (base) -bash-4.2\$ rename G2_ G2. .fastq (base) -bash-4.2\$ rename Shade_ Shade .fastq (base) -bash-4.2\$ rename _Fina Fina *.fastq (base) -bash-4.2\$ rename _R _L001_R *.fastq

have to zip the fastqs with gzip to .gz (did this in a job, it took 3 hours)

```
#!/bin/bash -login
######## SBATCH Lines for Resource Request ########
#SBATCH --time=3:59:00
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=1
#SBATCH --mem=64G
#SBATCH -- job-name import_data
#SBATCH -A shade-cole-bonito
#SBATCH --mail-user=suleskya@msu.edu
#SBATCH --mail-type=BEGIN, END
######## Command Lines for Job Running ########
conda activate qiime2-2022.8
qiime tools import \
  --type 'SampleData[PairedEndSequencesWithQuality]' \
  --input-path input_data \
  --input-format CasavaOneEightSingleLanePerSampleDirFmt \
  --output-path demux-paired-end.qza
conda deactivate
echo -e "\n `sacct -u suleskya -j $$LURM JOB ID --format=JobID, JobName, Start, End, Elapsed, NCPUS, ReqMem`
scontrol show job $SLURM_JOB_I
```

Successful, saved data as demux-paired-end.qza, can use this file in dada2

Denoise and merge

```
qiime dada2 denoise-paired \
        --i-demultiplexed-seqs demux-paired-end.qza \
        --p-trunc-len-f 191 \
        --p-trunc-len-r 84 \
        --o-table table.qza \
        --o-representative-sequences rep-seqs.qza \
        --o-denoising-stats denoising-stats.qza
qiime metadata tabulate \
  --m-input-file denoising-stats.qza \
  --o-visualization denoising-stats.qzv
qiime feature-table summarize \
  --i-table table.qza \
  --o-visualization table.qzv \
  --m-sample-metadata-file 230126_Shade_Seeds_16s_DG.txt
qiime feature-table tabulate-seqs \
  --i-data rep-seqs.qza \
  --o-visualization rep-seqs.qzv
conda deactivate
echo -e "\n `sacct -u suleskya -j $$LURM_JOB_ID --format=JobID, JobName, Start, End, Elapsed, NCPUS, ReqMem`
scontrol show job $SLURM JOB I
```

job successful! took 12 hr 24 minutes

Taxonomy assignment with silva

Assign taxonomy using Silva

 $Download\ reference\ seqs\ from\ qiime 2.org:\ wget\ https://data.qiime 2.org/2022.8/common/silva-138-99-515-806-nb-classifier.qza$

job: nano classify-silva-taxonomy.sb

```
qiime feature-classifier classify-sklearn \
  --i-classifier silva-138-99-515-806-nb-classifier.qza \
  --i-reads rep-seqs.qza \
  --o-classification taxonomy.qza
qiime metadata tabulate \
  --m-input-file taxonomy.qza \
  --o-visualization taxonomy.qzv
qiime tools export \
  --input-path taxonomy.qza \
  --output-path phyloseq
qiime tools export \
  --input-path table.qza \
  --output-path phyloseq
biom convert \
  -i phyloseq/feature-table.biom \
  -o phyloseq/otu_table.txt \
  --to-tsv
conda deactivate
echo -e "\n `sacct -u suleskya -j $$LURM JOB ID --format=JobID, JobName, Start, End, Elapsed, NCPUS, ReqMem`
scontrol show job $SLURM_JOB_I
Export OTU table to new directory phyloseq qiime tools export
-input-path table.gza
-output-path phyloseq
OTU tables exports as feature-table.biom so convert to .tsv biom convert
```

-i phyloseq/feature-table.biom

-o phyloseq/otu_table.txt

-to-tsv

download otu table, manually change "#OTU ID" column header to "OTUID"

download taxonomy file and manually change Feature ID to OTUID in taxonomy.tsv. change taxonomy and OTU tables to csv format.

these files are now ready to export to R and run using phyloseq

Make phylogenetic tree with de novo alignment

align reads de novo to make multiple sequence alignment (MSA) mask alignment to reduce ambiguity make tree with fasttree root tree to midpoint pipeline below will do all of these steps with default settings and save everything to output directory export makes newick format file for the trees, or .qza can be used in iTOL

```
#!/bin/bash -login
######## SBATCH Lines for Resource Request #########
#SBATCH --time=08:00:00
#SBATCH --nodes=1
#SBATCH --ntasks=1
```

```
\#SBATCH --cpus-per-task=32
#SBATCH --mem=64G
#SBATCH --job-name fasttree
#SBATCH -A shade-cole-bonito
#SBATCH --mail-user=suleskya@msu.edu
#SBATCH --mail-type=BEGIN, END
######## Command Lines for Job Running ########
conda activate qiime2-2022.8
qiime phylogeny align-to-tree-mafft-fasttree \
  --i-sequences rep-seqs.qza \
  --output-dir mafft-fasttree-output
qiime tools export \
  --input-path mafft-fasttree-output/tree.qza \
  --output-path mafft-fasttree-output
qiime tools export \
  --input-path mafft-fasttree-output/rooted_tree.qza \
  --output-path mafft-fasttree-output/exported-rooted-tree/
conda deactivate
echo -e "\n `sacct -u suleskya -j $SLURM_JOB_ID --format=JobID,JobName,Start,End,Elapsed,NCPUS,ReqMem`
scontrol show job $SLURM_JOB_I
```