Positron Emission Tomography Imaging of Amphetamine-Induced Dopamine Release in the Human Cortex: A Comparative Evaluation of the High Affinity dopamine D_{2/3} Radiotracers [¹¹C]FLB 457 and [¹¹C]Fallypride

RAJESH NARENDRAN,^{1,2*} W. GORDON FRANKLE,^{1,2} N. SCOTT MASON,¹ EUGENII A. RABINER,³ ROGER N. GUNN,³ GRAHAM E. SEARLE,³ SHIVANGI VORA,¹ MARALEE LITSCHGE,¹ STEVE KENDRO,¹ THOMAS B. COOPER,⁴ CHESTER A. MATHIS,¹ AND MARC LARUELLE^{3,5}

¹Department of Radiology, University of Pittsburgh, Pittsburgh, Pennsylvania
 ²Department of Psychiatry, University of Pittsburgh, Pittsburgh, Pennsylvania
 ³Clinical Imaging Center, GlaxoSmithKline, London, United Kingdom
 ⁴Department of Psychiatry, Columbia University, New York, New York
 ⁵Department of Neurosciences, Imperial College London, United Kingdom

KEY WORDS PET; dopamine; [11C]FLB 457; [11C]fallypride; amphetamine and human cortex

ABSTRACT The use of PET and SPECT endogenous competition binding techniques has contributed to the understanding of the role of dopamine in several neuropsychiatric disorders. An important limitation of these imaging studies is the fact that measurements of acute changes in synaptic dopamine have been restricted to the striatum. The ligands previously used, such as [11C]raclopride and [123I]IBZM, do not provide sufficient signal to noise ratio to quantify D2 receptors in extrastriatal areas, such as cortex, where the concentration of D₂ receptors is much lower than in the striatum. Given the importance of cortical DA function in cognition, a method to measure cortical dopamine function in humans would be highly desirable. The goal of this study was to compare the ability of two high affinity DA D₂ radioligands [11C]FLB 457 and [11C]fallypride to measure amphetamine-induced changes in DA transmission in the human cortex. D2 receptor availability was measured in the cortical regions of interest with PET in 12 healthy volunteers under control and postamphetamine conditions (0.5 mg kg⁻¹, oral), using both [¹¹C]FLB 457 and [¹¹C]fallypride (four scans per subjects). Kinetic modeling with an arterial input function was used to derive the binding potential (BP_{ND}) in eight cortical regions. Under controlled conditions, [11C]FLB 457 BP_{ND} was 30–70% higher compared with [11C]fallypride BP_{ND} in cortical regions. Amphetamine induced DA release led to a significant decrease in [11C]FLB 457 BP_{ND} in five out the eight cortical regions evaluated. In contrast, no significant decrease in [11C]fallypride BP_{ND} was detected in cortex following amphetamine. The difference between [11C]FLB 457 and [11C]fallypride ability to detect changes in the cortical D₂ receptor availability following amphetamine is related to the higher signal to noise ratio provided by [11C]FLB 457. These findings suggest that [11C]FLB 457 is superior to [11C] fallypride for measurement of changes in cortical synaptic dopamine. **Synapse 63:447–461, 2009.** © 2009 Wiley-Liss, Inc.

INTRODUCTION

Over the last few years, several groups have demonstrated that PET and SPECT neuroreceptor imaging techniques can be used, not only to measure

Received 16 August 2008; Accepted 20 September 2008

DOI 10.1002/syn.20628

Published online in Wiley InterScience (www.interscience.wiley.com).

Contract grant sponsor: GlaxoSmithKline, plc.

^{*}Correspondence to: Raj Narendran, MD, University of Pittsburgh, UPMC Presbyterian, PET Facility, B938, 200 Lothrop Street, Pittsburgh, PA 15213, USA. E-mail: narendranr@upmc.edu

receptor parameters but also to detect acute fluctuations in the concentration of endogenous transmitters in the vicinity of the receptors (for review, see Laruelle, 2000). For example, the displacement of the $D_{2/3}$ (hereafter referred to as D_2) receptor antagonists [11C]raclopride and [123I]IBZM following acute administration of amphetamine has been well validated as a noninvasive measure of the change in extracellular DA concentration induced by the challenge drug (Breier et al., 1997; Carson et al., 1997; Endres et al., 1997; Kegeles et al., 1999; Laruelle et al., 1995, 1997; Villemagne et al., 1999). Using these endogenous competition PET techniques, several groups have reported abnormal dopamine transmission in clinical populations such as schizophrenia (Breier et al., 1997; Laruelle et al., 1996) and addictive disorders (Malison et al., 1999; Martinez et al., 2005, 2007; Volkow et al., 1997). An important limitation of the use of [123I]IBZM and [11C]raclopride in these studies is the fact that measurements of D2 receptors and amphetamine-induced DA release were restricted mostly to the striatum because of their relatively low signal to noise ratio in the extrastriatal regions.

Since the introduction of the high affinity D2 receptor PET radiotracers [11C]FLB 457 (Halldin et al., 1995; in vitro KD = 0.018 nM) and [18 F]fallypride (Mukherjee et al., 1995, 2004; in vitro KD = 0.030 nM), several groups have confirmed their increased signal to noise ratio relative to [11C]raclopride and $[^{1\bar{2}\bar{3}}I]IBZM$ and reported on their ability to measure D₂ receptors in several extrastriatal regions of interest (Olsson et al., 2004; Slifstein et al., 2004a; Suhara et al., 1999). The question of whether the signal to noise ratio of the radiotracers [11C]FLB 457 and [¹⁸F]fallypride is sufficient to image amphetamineinduced dopamine transmission in relatively low D₂ receptor density prefrontal cortical regions of interest, such as the dorsolateral prefrontal cortex (DLPFC), orbital frontal cortex (OFC), medial prefrontal cortex (MPFC), and anterior cingulate cortex (ACC), is still unresolved in the literature.

The advantage of [18F]fallypride compared with [11C]FLB 457 to image DA transmission is primarily based on the fact that it is labeled with the relatively slowly decaying isotope F-18 and its uptake and washout kinetics are relatively fast compared with [11C]FLB 457: the combination of both factors allows for the measurement of D2 receptor BP in both striatal and extrastriatal areas (Cropley et al., 2008; Mukherjee et al., 1997; Price et al., 1997; Riccardi et al., 2005; Slifstein et al., 2007a). Nevertheless, the use of [18F]fallypride to measure amphetamine induced DA release in the cortex is impaired by its relatively low signal to noise ratio in the cortical ROIs (BP_{ND} \leq 0.5; Cropley et al., 2008; Riccardi et al., 2005; Slifstein et al., 2007a). Furthermore, the use of F-18 makes it impossible to perform a baseline

scan and postamphetamine scan the same day, a paradigm typically used with C-11 labeled radiotracers. The acquisition of baseline and postamphetamine scans on two different days increases the within-subject variability for the radiotracer and contributes to a lower sensitivity for the technique to detect amphetamine-induced changes in BP_{ND}. In contrast, [11C]FLB 457 is labeled with the rapidly decaying isotope C-11 and its kinetic of uptake is slower. Therefore, ¹¹C]FLB 457 can only be used for quantification of D₂ receptor parameters in extrastriatal areas. Several groups have attempted to use [11C]FLB 457 to study stimulant-induced DA release in the extrastriatal regions and reported inconsistent results (Aalto et al., 2005; Chou et al., 2000; Montgomery et al., 2006; Okauchi et al., 2001; Tsukada et al., 2005). The major limitations of these studies with respect to the vulnerability of the in vivo binding of [11C]FLB 457 following a psychostimulant challenge can be summarized as follows:

- 1. Administration of pharmacological (or nontracer) doses of [¹¹C]FLB 457 in both human and nonhuman primate PET studies. This limitation is based on previous studies, which demonstrate that the injected mass of [¹¹C]FLB 457 should be restricted to less than 1.6 nmoles (~0.6 μg) per 70 kg human subject to accomplish less than 5% of occupancy at the D₂ receptors (Olsson et al., 2004; Sudo et al., 2001). A review of the published literatures suggests that very few investigations (3 out of 16 studies, Table V) have been performed with such a strict restriction on injected mass (Ito et al., 2001; Sudo et al., 2001; Suhara et al., 2002).
- 2. The failure to quantify amphetamine (or stimulant)-induced changes in the cerebellum distribution volume ($V_{\rm T\ CER}$ or $V_{\rm ND}$ for binding in the cerebellum or reference region is taken as a measure of nonspecfic binding) by using reference region analysis instead of kinetic analysis with arterial input function (3 out of 16 studies, Table V), may have led to an underestimation of change in BP_{ND} in the regions of interest. This issue has been underscored by several recent studies demonstrating a decrease in [11 C]FLB 457 $V_{\rm TCER}$ following a psychostimulant or D₂ antagonist drug challenge (Ahmad et al., 2006; Asselin et al., 2007; Montgomery et al., 2006).
- 3. Scanning protocols which collect data for <90 min may not be sufficient for robust quantification of the $BP_{\rm ND}$ in the extrastriatal regions (2 out of 16 studies, Table V).

In this study, the authors compared the in vivo binding characteristics of [¹¹C]FLB 457 and [¹¹C]fallypride in the same healthy human subjects before and after an amphetamine challenge. The choice of [¹¹C]fallypride (a novel first-in-human radio-

ligand with the C-11 label) rather than [¹⁸F]fallypride was based on the fact that we were interested in measuring only cortical binding and it allowed for two PET scans (a baseline and postamphetamine) to be performed the same day.

The primary objectives of this human PET study were as follows:

- 1. To assess the signal to noise ratio (BP_{ND}) of $[^{11}C]FLB$ 457 and $[^{11}C]fallypride$ in the cortical regions of interest.
- 2. To assess the vulnerability of [¹¹C]FLB 457 and [¹¹C]fallypride binding to endogenous competition by DA following an acute amphetamine challenge (0.5 mg kg⁻¹) in the same subjects in the cortical regions of interest.

METHODS General design

The study was approved by the Institutional Review Board of the University of Pittsburgh Medical Center. A total of 48 PET scans were acquired for this study in 12 healthy control subjects over 24 experimental sessions. Each experimental session included two PET scans: a baseline scan and a postamphetamine scan with the same radiotracer. All subjects returned for a second experimental session in a minimum of one week (but no longer than 3 weeks) identical to the first, but with the other radiotracer (a total of four scans per subject). The sequence of the radiotracers was counterbalanced across subjects-half received two scans with [11C]FLB 457 during the first experimental session, whereas the other half received two scans with [11C]fallypride during the first experimental session—to prevent bias in the between radiotracer comparison. The postamphetamine scan occurred 3 h following the administration of 0.5 mg kg⁻¹ oral d-amphetamine (dexedrine). The timing was based on the time for d-amphetamine to reach $C_{\rm max}$ and remain at a relatively constant level in the plasma (Angrist et al., 1987; Cropley et al., 2008; Riccardi et al., 2005).

Radiolabeling

Radiolabeling of [¹¹C]FLB 457 and [¹¹C]fallypride were performed using previously published procedures (Halldin et al., 1995; Mukherjee et al., 2004).

PET protocol

Imaging experiments were conducted on the ECAT EXACT HR+ consistent with previously described image acquisition protocols (Abi-Dargham et al., 2000). Briefly, following completion of a transmission scan (\sim 10 min) for attenuation correction of the emis-

sion data, subjects received either an intravenous injection of [$^{11}\mathrm{C}$]FLB 457 or [$^{11}\mathrm{C}$]fallypride as a bolus over 20 s. Based on previously published studies, the maximum injected mass for [$^{11}\mathrm{C}$]FLB 457 and [$^{11}\mathrm{C}$]fallypride was restricted to 0.6 and 1.2 µg, respectively (Cropley et al., 2008; Riccardi et al., 2005; Sudo et al., 2001). Emission data were collected for 90 min. The postamphetamine scan was performed 3 h after the administration of 0.5 mg kg $^{-1}$ PO of amphetamine.

Input function measurement

Following radiotracer injection, arterial samples were collected manually approximately every 6 s for the first 2 min and thereafter at longer intervals. A total of 35 samples were obtained per scan. Following centrifugation, plasma was collected in 200 μ l aliquots and activities were counted in a gamma counter.

To determine the plasma activity representing unmetabolized parent compound ([11C]FLB 457 or [11C]fallypride) six samples (collected at 2, 10, 20, 40, 60, and 75 min) were further processed using HPLC methods described previously (Olsson et al., 1999; Slifstein et al., 2004b). Briefly, the supernatant obtained after centrifugation was deproteinized with acetonitrile. The acetonitrile-plasma mixture was then analyzed by reverse-phase HPLC methods. [11C]FLB 457 analyses were performed on a Prodigy ODS-3, (5 μ , 4.6 \times 250 mm) eluted at 2.0 ml/min with 23/77 acetonitrile/ammonium formate buffer (pH = 4.2). [11C]Fallypride analyses were performed on a Prodigy ODS-3, (5 μ , 4.6 \times 250 mm) eluted at 2.0 ml/ min with 45/55 acetonitrile/ammonium formate buffer (pH 4.2). The radioactivity corresponding to the parent compound was then expressed as the percentage of the total radioactivity in the sample.

For [11 C]FLB 457 the six measured parent fractions were fitted using a Hill model (Gunn et al., 1998; Hill, 1910; Wu et al., 2007). For [11 C]fallypride the parent fractions were fitted to the sum of two exponentials. The smallest exponential of the fraction of the parent curve, $\lambda_{\rm par}$, was constrained to the difference between $\lambda_{\rm cer}$ (the terminal rate of washout of the cerebellar activity) and $\lambda_{\rm tot}$ (the smallest elimination rate constant of the total plasma) (Abi-Dargham et al., 1999).

The input function was then calculated as the product of total counts and interpolated parent fraction at each time point. The measured input function values were fitted to a sum of three exponentials from the time of peak plasma activity and the fitted values were used as the input to the kinetic analysis. The clearance of the parent compound (L/h) was calculated as the ratio of the injected dose to the area under the curve of the input function (Abi-Dargham et al., 1994).

For the determination of the plasma free fraction $(f_{\rm P})$, triplicate aliquots of plasma collected prior to injection were mixed with the radiotracer, pipetted into ultrafiltration units (Amicon Centrifree; Millipore, Bedford, MA), and centrifuged at room temperature (30 min at 6000 rpm). At the end of centrifugation, the plasma and ultrafiltrate activities were counted in a gamma counter, and $f_{\rm P}$ was calculated as the ratio of activity in the ultrafiltrate to total activity (Gandelman et al., 1994). Triplicate aliquots of saline solution mixed with the radiotracer were also processed to determine the filter retention of the free tracer

In the postamphetamine condition, amphetamine plasma levels were measured in three arterial samples obtained at time 0, 45, and 90 min relative to the PET scan as previously described (Reimer et al., 1993). These data ensured that differences in plasma amphetamine concentration did not bias the radiotracer comparison.

MRI acquisition and segmentation procedures

To provide an anatomical framework for analysis of the PET data, MRI scans were obtained using a 1.5 T GE Medical Systems (Milwaukee, WI) Signa Scanner and a 3D spoiled gradient recalled sequence. MRI segmentation was performed using the FAST automated segmentation tool (Zhang et al., 2001) implemented in the FMRIB Software Library (v4.0, Smith et al., 2004).

Image analysis

PET data were reconstructed using filtered back-projection (Fourier rebinning/2D backprojection, 3 mm Hann filter) and corrected for photon attenuation (⁶⁸Ge/⁶⁸Ga rods), scatter (Watson, 2000), and radioactive decay. Reconstructed image files were then processed with the image analysis software MEDx (Sensor Systems, Sterling, Virginia) and SPM2 (www.fil.ion.ucl.ac.uk/spm). Frame-to-frame motion correction for head movement and MR-PET image alignment were performed using a mutual information algorithm implemented in SPM2.

Time activity curves were generated for the eight cortical regions of interest using the criteria and methods outlined in (Abi-Dargham et al., 2000, 2002). Sampled cortical regions (n=8) included the medial temporal lobe MTL (which included the hippocampus, amygdala, parahippocampal gyri, and entorrhinal cortex), anterior cingulate cortex (ACC), dorsolateral prefrontal cortex (DLPFC), orbital frontal cortex (OFC), medial prefrontal cortex (MPFC), temporal cortex (TC), parietal cortex (PC), and occipital cortex (OC). The cerebellum was subsampled in 15 consecutive coronal MRI slices caudal to the cerebellar penduncle and used as a reference region. The subsampling

included only the gray matter and excluded the vermis, white matter, and the cerebro-cerebellar fissure.

For bilateral regions, right and left values were averaged. The contribution of plasma total activity to the regional activity was calculated assuming a 5% blood volume in the regions of interest (Mintun et al., 1984) and tissue activities were calculated as the total regional activities minus the plasma contribution.

Derivation of binding parameters

We denote here the outcome variables using the recently issued consensus nomenclature for in vivo imaging of reversibly binding radioligands (for details refer to Innis et al., 2007).

The regional tissue distribution volume $(V_T, \text{ ml cm}^{-3})$ was defined as the ratio of the ligand concentration in a region $(C_T, \mu\text{Ci g}^{-1})$ to the concentration of unmetabolized ligand in arterial plasma $(C_P, \mu\text{Ci ml}^{-1})$ at equilibrium,

$$V_{\rm T} = \frac{C_{\rm T}}{C_{\rm P}} \tag{1}$$

In the cerebellum, a region with negligible D_2 receptor density, V_T includes the free and nonspecific binding, and V_T in the cerebellum ($V_{T \ CER}$) is equal to the nondisplaceable distribution volume (V_{ND}). The region of interest V_T ($V_{T \ ROI}$) includes V_{ND} and the specific binding distribution volume or BP_P (binding potential in which specific binding is compared with total plasma concentration). Assuming that V_{ND} is equal in both regions, BP_P was derived as the difference between $V_{T \ ROI}$ and $V_{T \ CER}$.

Under the assumption of a competitive interaction between the radiotracer and DA, $BP_{\rm P}$ is related to receptor parameters and endogenous DA by

$$V_{\mathrm{TROI}} - V_{\mathrm{TCER}} = \mathrm{BP_P} = f_{\mathrm{P}} \times \frac{B_{\mathrm{max}}}{K_{\mathrm{D}}'} = f_{\mathrm{P}} \times \frac{B_{\mathrm{max}}}{K_{\mathrm{D}} \left(1 + F_{\mathrm{DA}} \middle/ K_{\mathrm{I}}\right)} \tag{2}$$

where $f_{\rm P}$ is defined as free fraction of radioligand in the plasma, $B_{\rm max}$ is the concentration of $\rm D_2$ receptors(nM/g of tissue), $K_{\rm D}$ is the in vivo equilibrium dissociation constant of the radiotracer (nM/ml of brain water), $K'_{\rm D}$ is $K_{\rm D}$ in the presence of the competitor DA (nM/ml of brain water), $F_{\rm DA}$ is the free concentration of endogenous DA in the vicinity of the receptors, and $K_{\rm I}$ is the inhibition constant of DA for the binding of the radiotracer (Laruelle et al., 1994, 1998).

A preferred outcome measure for endogenous competition experiments is the $BP_{\rm ND}$ (binding potential in which specific binding is compared with nondisplaceable uptake). $BP_{\rm ND}$ was calculated as the ratio of $BP_{\rm P}$ to $V_{\rm T~CER}.$ $BP_{\rm ND}$ is related to receptor parameters and endogenous DA by

TABLE I. Base	eline scan parameters	nd plasma ana	alysis (n = 12 subjects)
---------------	-----------------------	---------------	--------------------------

	<u></u>	[¹¹ C]fallypride		
ne Postamphetamir	ne Baseline	Postamphetamine		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	5.0 ± 0.6 2105 ± 638 0.9 ± 0.2 13.8 ± 5.0 34 ± 6 1.02 ± 0.34 14.0 ± 4.56	4.7 ± 0.7 1640 ± 417^{a} 1.1 ± 0.1^{a} 13.3 ± 5.5 36 ± 8 1.02 ± 0.31 13.8 ± 6.2 76.2 ± 10.3 68.3 ± 7.6		
1	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		

 $^{^{\}mathrm{a}}P < 0.05$, paired t-test, baseline compared with postamphetamine.

$$\frac{V_{\text{T ROI}} - V_{\text{T CER}}}{V_{\text{T CER}}} = \text{BP}_{\text{ND}} = f_{\text{ND}} \times \frac{B_{\text{max}}}{K_{\text{D}}'} = f_{\text{ND}} \times \frac{B_{\text{max}}}{K_{\text{D}} \left(1 + F_{\text{DA}} / K_{\text{I}}\right)} \tag{3}$$

where $f_{\rm ND}$ is the free fraction in the nonspecific distribution volume of the brain ($f_{\rm ND}=f_{\rm P}/V_{\rm ND}$) (Laruelle et al., 1994). The change in BP_{ND} elicited by amphetamine (Δ BP_{ND}) was calculated as the difference between BP_{ND} measured in the postamphetamine condition (BP_{ND_{AMPH}}) and BP_{ND} measured in the baseline condition on that day (BP_{ND_{BASE}}) and expressed as a percentage of BP_{ND_{BASE}}:

$$\Delta BP_{ND} = 100 \times \frac{BP_{ND_{AMPH}} - BP_{ND_{BASE}}}{BP_{ND_{BASE}}} \tag{4} \label{eq:delta_BP_ND}$$

Equation 3 demonstrates that the increase in $F_{\rm DA}$ resulting from the challenge translates into a decrease in ${\rm BP_{ND}}$. Equation 3 also indicates that the attribution of changes in ${\rm BP_{ND}}$ solely to changes in $F_{\rm DA}$ requires the assumption that all other parameters of this equation remain constant, including the nonspecific binding $(f_{\rm ND})$. $\Delta {\rm BP_{ND}}$ is generally preferred to $\Delta {\rm BP_{P}}$ to measure the effect of amphetamine, because the test–retest reproducibility of ${\rm BP_{ND}}$ is generally better than that of ${\rm BP_{P}}$ The effect of amphetamine on plasma clearance $(C_{\rm L})$, $f_{\rm P}$, $f_{\rm ND}$, and $V_{\rm ND}$ was also expressed relative to the preamphetamine value measured the same day.

Derivation of [11 C]FLB 457 and [11 C]fallypride $V_{\rm T}$ was performed using kinetic analysis and the arterial input function. A two-tissue compartment model (2TCM) described the cerebellum and regions of interest for both tracers (Montgomery et al., 2006; Mukherjee et al., 2002; Olsson et al., 1999).

Statistical analysis

Data were analyzed with repeated measure ANOVA (RM ANOVA) and paired *t*-test as specified. For the regions of interest a false discovery rate (FDR) correc-

tion with $\alpha = 0.05$ was applied to correct for multiple comparisons (Benjamini and Hochberg, 1995).

RESULTS Demographics

Twelve subjects (5 males/7 females; 1 AA/11 C) participated in the study. The mean age of the subjects was 28 ± 10 (range 19–48). The mean BMI of the subjects was 24 ± 3 (range 19–27). All twelve subjects who participated in the study were nonsmokers.

Baseline scan parameters

Injected dose and mass

The mean injected dose, mass, and specific activity at the time of injection for the baseline and postamphetamine condition for both radiotracers [¹¹C]FLB 457 and [¹¹C]fallypride are listed in Table I.

No significant differences were observed between the baseline and postamphetamine condition in injected dose and injected mass for [¹¹C]FLB 457.

No significant differences were observed between the baseline and postamphetamine condition in injected radiation dose for $[^{11}\mathrm{C}]$ fallypride. A slightly but significantly higher mass dose of $[^{11}\mathrm{C}]$ fallypride was injected during the postamphetamine condition $(1.1~\pm~0.1)$ relative to the baseline condition $(0.9~\pm~0.1)$. Nevertheless, as the total injected mass was $<1.2~\mu\mathrm{g}$ (mass required for less than 5% of occupancy D_2 receptor, tracer dose; unpublished data from Columbia University, NY, NY) in both conditions (baseline and postamphetamine) it is unlikely to have had a significant impact on the measurement of $[^{11}\mathrm{C}]$ fallypride ΔBP_{ND} following amphetamine.

Plasma analysis

Clearance

Under baseline conditions, [11 C]FLB 457 plasma $C_{\rm L}$ was significantly faster than [11 C]fallypride plasma $C_{\rm L}$ (RM ANOVA, P < 0.001). Amphetamine did not significantly alter the plasma $C_{\rm L}$ for [11 C]FLB 457 or [11 C]fallypride (individual values listed in Table I).

Free fraction in plasma

Under baseline conditions, [11 C]FLB 457 f_P was significantly higher than [11 C]fallypride f_P (RM ANOVA, P < 0.001). Amphetamine did not significantly alter f_P for [11 C]FLB 457 or [11 C]fallypride (Table I).

Amphetamine plasma levels

No significant differences in the amphetamine plasma levels were observed between the postamphetamine [11 C]FLB 457 and [11 C]fallypride scans (RM ANOVA, P=0.72). In addition, the amphetamine plasma levels were relatively stable throughout the entire duration of the postamphetamine scan for both radioligands (Table I).

Regions of interest volumes

The volumes of the eight cortical regions of interest and reference region are provided in Table II.

Reference region analysis

Cerebellum distribution volume

Under baseline conditions, [$^{11}\mathrm{C}$]FLB 457 V_ND was significantly higher than [$^{11}\mathrm{C}$]fallypride V_ND (RM ANOVA, P<0.001). Amphetamine did not significantly alter the V_ND for [$^{11}\mathrm{C}$]FLB 457 or [$^{11}\mathrm{C}$]fallypride (Table I)

Free fraction in nondisplaceable compartment (f_{ND})

Under baseline conditions, [11 C]FLB 457 $f_{\rm ND}$ was not significantly different than [11 C]fallypride $f_{\rm ND}$ (RM ANOVA, P=0.12). Amphetamine did not signifi-

TABLE II. Regional volumes

Region	Volume (mm ³)
Medial temporal lobe	$15,383 \pm 3386$
Anterior cingulate cortex	3945 ± 1231
Dorsolateral prefrontal cortex	$21,451 \pm 4886$
Orbital frontal cortex	9640 ± 2661
Medial prefrontal cortex	4779 ± 1227
Temporal cortex	$37,099 \pm 4854$
Parietal cortex	$74,053 \pm 15,018$
Occipital cortex	$53,459 \pm 14,564$
Cerebellum	$21,640 \pm 7586$

cantly alter the $f_{\rm ND}$ for [$^{11}{\rm C}$]FLB 457 or [$^{11}{\rm C}$]fallypride (Table I). The lack of any change in $f_{\rm ND}$ following amphetamine for both radioligands validated our assumptions for using $\Delta {\rm BP}_{\rm ND}$ as an outcome measure to detect changes in free dopamine concentrations following amphetamine ($F_{\rm DA}$, see Eq. 3).

Region of interest analysis: binding potential $\mathrm{BP}_{\mathrm{ND}}$

The values of $BP_{\rm ND}$ and $\Delta BP_{\rm ND}$ for each region are given in Table III.

Comparison of baseline [11C]FLB 457 and [11C]fallypride BP_{ND}

Under baseline conditions, [11 C]FLB 457 BP_{ND} was significantly higher than [11 C]fallypride BP_{ND} (RM ANOVA, BP_{ND} as dependent variable; tracer factor, P = 0.012; tracer \times region interaction, P = 0.03; see Fig. 1)

Amphetamine-induced changes in $[^{11}C]FLB$ 457 BP_{ND}

Amphetamine produced a statistically significant decrease in BP $_{\rm ND}$ in the MTL, ACC, DLPFC, MPFC, and PC (paired t-tests, P < 0.05; see Table III, Figs. 2 and 3). These results remained significant after correction for multiple comparisons using the false discovery rate. Nonsignificant decreases in BP $_{\rm ND}$ were observed in the remaining regions (OFC, TC, and OCC).

The regional differences in ΔBP_{ND} were evaluated with repeated measures ANOVA, with regional ΔBP_{ND} as repeated factor. No significant between-region differences in ΔBP_{ND} were observed (RM ANOVA, P=0.5)

Amphetamine-induced changes in $[^{11}C]$ fallypride BP_{ND}

The values of BP_{ND} and ΔBP_{ND} for each region are given in Table III and Fig. 3). Amphetamine produced a statistically significant decrease in BP_{ND} only in the

TABLE III. Effect of amphetamine on [11 C]FLB 457 and [11 C]fallypride BP $_{ND}$

	$[^{11}\mathrm{C}]\mathrm{FLB}$ 457 $\mathrm{BP_{ND}}$			$[^{11}{ m C}]$ fallypride B ${ m P}_{ m ND}$				
Region	Baseline	Postamphetamine	Difference (%)	P values	Baseline	Postamphetamine	Difference (%)	P values
Medial temporal lobe Anterior cingulate cortex Dorsolateral prefrontal cortex Orbital frontal cortex Medial prefrontal cortex Temporal cortex Parietal cortex	$\begin{array}{c} 1.51 \pm 0.28 \\ 0.94 \pm 0.37 \\ 0.58 \pm 0.32 \\ 0.67 \pm 0.34 \\ 0.78 \pm 0.35 \\ 1.72 \pm 0.59 \\ 0.57 \pm 0.32 \end{array}$	0.61 ± 0.29 0.69 ± 0.30 1.63 ± 0.49	-7.0 ± 6.1 -8.1 ± 8.2 -12.6 ± 15.2 -5.1 ± 21.6 -11.3 ± 14.3 -4.2 ± 9.1 -12.4 ± 13.4	0.005^{a} 0.024^{a} 0.011^{a} 0.224 0.021^{a} 0.146 0.005^{a}	1.16 ± 0.17 0.61 ± 0.24 0.36 ± 0.19 0.44 ± 0.19 0.47 ± 0.20 1.12 ± 0.29 0.33 ± 0.19	$\begin{array}{c} 0.31 \pm 0.19 \\ 0.40 \pm 0.22 \end{array}$	-4.4 ± 11.8 -8.7 ± 21.2 -9.7 ± 25.3 -8.0 ± 27.4 -6.9 ± 17.6 -7.5 ± 10.9 -10.0 ± 33.4	0.216 0.053 0.246 0.166 0.048
Occipital cortex	0.57 ± 0.32 0.51 ± 0.26		-12.4 ± 13.4 -4.7 ± 19.6	0.005	0.33 ± 0.19 0.32 ± 0.18	0.32 ± 0.23 0.31 ± 0.23	-8.7 ± 30.0	

Values are mean \pm SD; n = 12 subjects. P values indicate paired t-tests.

^aSignificant following multiple corrections using FDR.

TC (paired t-tests, P < 0.048). This result was not significant after correction for multiple comparisons using the false discovery rate (Table III).

No significant differences were observed between [11 C]FLB 457 Δ BP $_{ND}$ and [11 C]fallypride Δ BP $_{ND}$ (RM ANOVA; Δ BP $_{ND}$ as dependent variable; tracer factor, P=0.97).

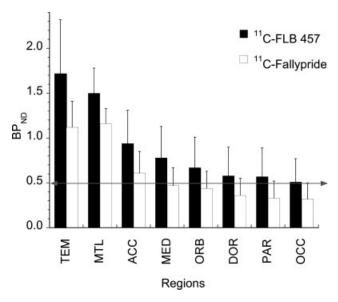


Fig. 1. Comparison of the baseline BP_{ND} for [11 C]FLB 457 and [11 C]fallypride (n=12 subjects). The [11 C]FLB 457 BP_{ND} is 1.6-fold (average across eight ROIs) greater than [11 C]fallypride BP_{ND}. Note: gray arrow illustrates [11 C]FLB 457 BP_{ND} > 0.5 in all ROIs evaluated in this study. Regions include TEM (Temporal cortex); MTL (medial temporal lobe); ACC (anterior cingulate cortex); MPFC (medial prefrontal cortex); ORB (orbital frontal cortex); DOR (dorsolateral prefrontal cortex); PAR (Parietal cortex); and OCC (Occipital cortex).

Region of interest analysis: binding potential BP_P

Comparison of baseline [11C]FLB 457 and [11C]fallypride BP_{ND}

Table IV lists the values of [11 C]FLB 457 and [11 C]fallypride BP_P under baseline and postamphetamine conditions. Under baseline conditions, [11 C]FLB 457 BP_P was significantly higher than [11 C]fallypride BP_{ND} (RM ANOVA, BP_{ND} as dependent variable; tracer factor, P < 0.0001; tracer × region interaction, P < 0.0001).

Amphetamine-induced changes in $[^{11}C]FLB$ 457 BP_P

The values of BP_P and Δ BP_P for each region are given in Table IV. Amphetamine produced a statistically significant decrease in BP_P only in the PC (paired *t*-tests, P < 0.05; Table IV). This result was not significant after correction for multiple comparisons using the false discovery rate.

Amphetamine-induced changes in [11C]fallypride BP_P

The values of BP_P and ΔBP_P for each region are given in Table IV. Amphetamine did not produce a statistically significant decrease in BP_P in any of the regions (paired *t*-tests, P < 0.05; Table IV).

Amphetamine induced decrease in [11 C]FLB 457 or [11 C]fallypride BP_P or BP_{ND} in the cortical regions of interest were not significantly correlated with the subjects' age or plasma amphetamine levels achieved during the scan (P > 0.05, Pearson Correlation Coefficient).

In addition, the order of the sequence (first or second) in which subjects received radiotracer had no

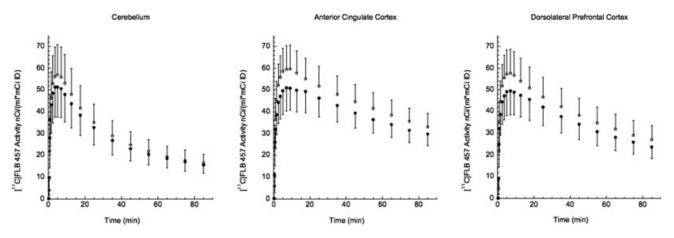


Fig. 2. Amphetamine-induced displacement of [11 C]FLB 457; mean \pm SD, baseline (open circles, n=12 subjects) and postamphetamine (closed circles, n=12 subjects) [11 C]FLB 457 time activity curves (TAC) normalized to injected doses are shown for Cerebellum (reference region), Anterior Cingulate Cortex (ACC) and Dorsolateral prefrontal cortex (DLPFC). Note the significant decreases in TAC following amphetamine in ACC and DLPFC but not in cerebellum.

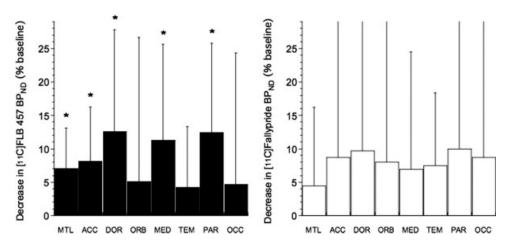


Fig. 3. Mean \pm SD decrease in [\$^{11}\$C]FLB 457 (black bars) and [\$^{11}\$C]fallypride (white bars) D₂ receptor BP_{ND} in eight cortical regions of interest in 12 healthy subjects after the administration of 0.5 mg kg $^{-1}$ amphetamine. * represents significant decrease (\$P < 0.05\$) following the FDR correction for multiple comparisons.

TABLE IV. Effect of amphetamine on [11C]FLB 457 and [11C]fallypride BPP

$[^{11}\mathrm{C}]\mathrm{FLB}$ 457 $\mathrm{BP_P}$				[11C]fallypride BP _P			
Baseline	Post amphetamine	Difference (%)	P Values	Baseline	Post amphetamine	Difference (%)	P Values
8.60 ± 2.26	7.88 ± 1.72	-6.6 ± 12.4	0.112	1.17 ± 0.38	1.12 ± 0.39	-4.5 ± 11.6	0.266
							0.073 0.129
		-12.0 ± 19.0 -5.6 ± 20.5			0.31 ± 0.19 0.39 ± 0.22	-9.9 ± 25.4 -8.8 ± 24.4	0.129
4.40 ± 2.17	3.76 ± 1.47	-10.3 ± 21.3	0.111	0.46 ± 0.20	0.42 ± 0.19	-7.6 ± 13.8	0.091
	9.11 ± 2.67	-3.1 ± 19.4			1.04 ± 0.39	-7.4 ± 12.3	0.135
3.23 ± 1.99 2.86 ± 1.56	2.68 ± 1.34 2.57 ± 1.21	$-11.9 \pm 17.5 -4.5 \pm 21.6$	$0.037 \\ 0.107$	$0.33 \pm 0.20 \\ 0.32 \pm 0.17$	0.30 ± 0.18 0.29 ± 0.18	-11.1 ± 30.1 -9.7 ± 28.0	$0.196 \\ 0.256$
	8.60 ± 2.26 5.32 ± 2.26 3.27 ± 2.04 3.78 ± 2.08 4.40 ± 2.17 9.73 ± 3.63 3.23 ± 1.99	$\begin{array}{c} \text{Baseline} & \begin{array}{c} \text{Post} \\ \text{amphetamine} \end{array} \\ \hline 8.60 \pm 2.26 & 7.88 \pm 1.72 \\ 5.32 \pm 2.26 & 4.71 \pm 1.41 \\ 3.27 \pm 2.04 & 2.73 \pm 1.37 \\ 3.78 \pm 2.08 & 3.43 \pm 1.62 \\ 4.40 \pm 2.17 & 3.76 \pm 1.47 \\ 9.73 \pm 3.63 & 9.11 \pm 2.67 \\ 3.23 \pm 1.99 & 2.68 \pm 1.34 \\ \hline \end{array}$	$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$				

Values are mean \pm SD; n = 12 subjects. P values indicate paired t-tests.

^cSignificant following multiple corrections using FDR.

significant effect on amphetamine-induced reduction in BP $_{\rm ND}$ or BP $_{\rm P}$ for [$^{11}{\rm C}$]FLB 457 or [$^{11}{\rm C}$]fallypride (RM ANOVA, P>0.05).

DISCUSSION

The fundamental aim of this investigation was to compare the in vivo specific binding of [11C]FLB 457 and [11C]fallypride in terms of vulnerability to endogenous competition by DA. Endogenous competition experiments were performed in the same human subjects, under carefully controlled conditions, and in counterbalanced sequence. The results demonstrate that in vivo [11C]FLB 457 has a higher signal to noise ratio than [11C]fallypride, a finding consistent with the previously published human PET studies performed with both radioligands (Table V). These results also demonstrate for the first time, unequivocally, that this enhanced signal to noise ratio allows for the measurement of amphetamine induced DA release in the human cortex with [11C]FLB 457 and PET. This observation is also consistent with the previous studies in the literature that have reported

more success in measuring changes in cortical DA transmission with [\$^{11}\$C]FLB 457 (Aalto et al., 2005; Montgomery et al., 2006) than with [\$^{18}\$F]fallypride (Cropley et al., 2008; Riccardi et al., 2005; Slifstein et al., 2007a).

To summarize, our observation of amphetamineinduced DA release reduction of in vivo binding of [11C]FLB 457 in the human prefrontal cortex is consistent with previous human studies, in which the binding of this radioligand have been reported to be vulnerable to endogenous competition by DA in the frontal as well as other cortical regions of interest following an acute cognitive (Aalto et al., 2005) or methylphenidate challenge (Montgomery et al., 2006). On the other hand, consistent with the inability to reliably measure amphetamine-induced displacement of [¹¹C]fallypride in the human cortex in this study are two out of three investigations with [18F]fallypride reporting that [18F]fallypride cannot be used to measure amphetamine-induced DA release (greater than 5% decrease in radioligand binding that is statistically significant) in the cortical regions of interest because of its relatively low signal to noise ratio in

TABLE V. Comparison of in vivo binding parameters of [IIC]FLB 457 and [18F]/[IIC]fallypride in published human studies

Study	$\mathrm{HC}\left(n\right)$	Mass (nmoles)	Duration (min)	Analysis	Modeling	$\begin{array}{c} Prefrontal \\ cortex \ BP_{ND} \end{array}$	Anterior cingulate $\mathrm{BP}_{\mathrm{ND}}$
[11CIFLB 457							
Ito et al., 2001	7	0.7 - 1.52	70	SUB	KIN	1.0 ± 0.2	0.9 ± 0.2
Sudo et al., 2001	8	0.5 - 1.7	90	SUB	SRTM	1.0 ± 0.2	_
Suhara et al., 2002	18	0.5 - 1.67	80	SUB	SRTM	1.0 ± 0.2	1.2 ± 0.2
Olsson et al., 1999	8	2.4 - 4.6	60	ROI	KIN	0.7 ± 0.2	0.8 ± 0.2
Vilkman et al., 2000	7	3.7 ± 1.4	70	ROI	GRA	0.5 ± 0.1	
Olsson et al., 2004	10	2.4 - 4.6	60	ROI	SRTM	0.7 ± 0.1	0.8 ± 0.2
Talvik et al., 2003	8	1.9 ± 0.4	60	ROI	SRTM	0.7 ± 0.2	0.9 ± 0.4
Aalto et al., 2005	12	2.2 - 2.6	70	ROI	SRTM	0.6 ± 0.2	0.9 ± 0.2
Kemppainen et al., 2003	11	2.2 - 4.8	70	ROI	SRTM	0.4 ± 0.1	0.4 ± 0.1
Kaasinen et al., 2001	12^{a}	3.5 ± 1.2	70	ROI	SRTM	0.3 ± 0.1	0.4 ± 0.1
Kaasinen et al., 2001	$12^{ m b}$	3.6 ± 1.1	70	ROI	SRTM	0.5 ± 0.1	0.6 ± 0.1
Kaasinen et al., 2000a	20	3.3 ± 1.0	70	ROI	SRTM	0.4 ± 0.1	0.5 ± 0.1
Hagelberg et al., 2004	8	2.6 ± 0.4	70	ROI	SRTM	0.8 ± 0.2	1.0 ± 0.2
Hagelberg et al., 2004	8	3.8 ± 1.5	70	ROI	SRTM	0.7 ± 0.6	0.9 ± 0.2
Okubo et al., 1999	13	1.6 - 5.0	65	ROI	SRTM	0.5 ± 0.2	0.9 ± 0.4
Montgomery et al., 2006	12	2.7 - 10.8	90	ROI	KIN	0.7 ± 0.1	0.9 ± 0.3
This study	12	0.4 ± 0.1	90	SEG	KIN	0.6 ± 0.3^{c}	0.9 ± 0.3
Average BP_{ND}						0.7 ± 0.2	0.8 ± 0.2
[18F]Fallypride							
Slifstein et al., 2007a	15	< 3.3	240	SEG	KIN	0.4 ± 0.2	0.5 ± 0.1
Mukherjee et al., 2002	6	1.8 - 2.5	180	ROI	GRA	0.3 ± 0.1	_
Riccardi et al., 2005	14	< 2.0	210	ROI	SRTM	< 0.5	_
Cropley et al., 2008	14	< 3.3	300	SEG	SRTM	$0.6\pm0.1^{ m d}$	0.5 ± 0.1
This study	12	0.9 ± 0.2	90	SEG	KIN	0.4 ± 0.2^{c}	0.6 ± 0.2
Average $\tilde{\mathrm{B}}\mathrm{P}_{\mathrm{ND}}$						0.4 ± 0.2	0.5 ± 0.1

V_{ND}, nonspecific binding; BP_{ND}, binding potential; ROI, full region of interest delineated; SEG, segmentation of gray matter pixels in region of interest; SUB, subsampling of region interest with small circles; KIN, kinetic analysis; SRTM, simplified reference tissue method; GRA, logan graphical method. "Males.

these regions (Riccardi et al., 2005; Slifstein et al., 2007a). The third study by Cropley et al. reported a statistically significant decrease of [18F]fallypride binding $(-13\% \pm 4\%)$ in the medial OFC but not in the temporal cortex. Other cortical regions such as the DLPFC, MPFC, and ACC were not evaluated in this study either due to relatively low binding potential $(BP_{ND} < 0.5)$ or poor reproducibility for BP_{ND} (Cropley et al., 2008). Thus, the success of [11C]FLB 457 and failure of [11C]fallypride to measure amphetamineinduced DA release in the human cortex was somewhat predicted by the published literature. The choice of [18F]fallypride over [11C]FLB 457 as a preferred tool to measure amphetamine-induced DA transmission was primarily driven by its promise to evaluate both the high binding striatal and low binding extrastriatal regions of interest such as the cortex at the same time. Unfortunately, the results of this study as well as other recent investigations with [18F]fallypride only confirm the fact that measurement of amphetamine induced DA release in both striatum and cortex with one tracer is not currently feasible.

Methodological considerations

The study was designed to establish the amphetamine effect in the cortical regions of interest for both [11C]FLB 457 and [11C]fallypride. Because of the between-subject variability in the magnitude of this effect, it was important to study the same subject following the same dose of amphetamine with both radioligands. Although previous results are mixed with respect to sensitization of dopamine transmisfollowing repeated amphetamine exposure (Boileau et al., 2006; Kegeles et al., 1999), this requirement induced the concern that tolerance or sensitization to the amphetamine effect might develop during the course of the study. To alleviate this concern, the radiotracers were administered in counterbalanced fashion. Our results demonstrated a statistically significant decrease in 5/8 ROIs for [11C]FLB 457 but not for [11C]falypride (other than TC, which failed to survive correction for multiple comparisons). This is likely due to the increased variance in amphetamine-induced ΔBP_{ND} for [¹¹C]fallypride, which was caused by its relatively low signal to noise ratio in the cortical regions of interest (BP_{ND} < 0.5 in DLPFC, OFC, MPFC, PC, and OC).

Other issues that are significant and will need to be addressed in the validation of the use of [11C]FLB 457 to measure cortical DA transmission are discussed below.

FLB 457 occupancy

The paradigm we used to study amphetamine induced DA release included two administrations of [11C]FLB 457 in the same day 3-h apart. An issue that needs further evaluation is the contribution of the carryover mass of FLB 457 from the first scan to

^cWeighted average of DLPFC, MPFC, and OFC. ^dMedial orbital frontal cortex.

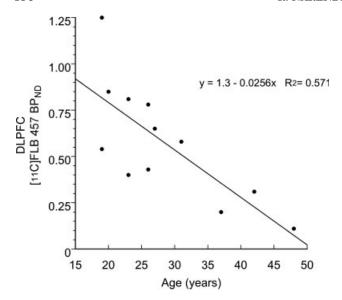


Fig. 4. Effect of age on cortical [11 C]FLB 457 BP_{ND}. Note significant decrease in baseline DLPFC BP_{ND} values in relatively older subjects compared with those less than 35 years of age ($\rho = -0.76$; P < 0.005).

the second scan and its impact on ΔBP_{ND} . In general, this particular issue is less of a problem when radioligands such as [11C]raclopride, which have relatively low affinity and fast washout kinetics are used to measure amphetamine-induced DA-release. Nevertheless, there are significant advantages to performing two PET scans (such as baseline and postamphetamine) on the same day as this technique yields better within-subject reproducibility. In addition, completing all study procedures in a single day is less invasive (requiring only one A-line, IV, etc.) and less of time commitment for the subjects. This issue was debated extensively in the designing of this study. Our decision to pursue a single day study design (3-h apart) was based on a previous test-retest study that did not detect any carry over mass-induced decreases in [11C]FLB 457 BP_{ND} in the frontal and temporal cortex when subjects underwent the test and retest scans 3-h apart (Vilkman et al., 2000). In addition, in a counterbalanced study evaluating DA release following a cognitive task in which subjects received three [11C]FLB 457 scans the same day, injected mass was not correlated with decrease in [11C]FLB 457 BP_{ND} (Aalto et al., 2005). Consistent with these previous reports we observed no correlation between either the injected mass of Scan 1 or the total injected mass (Scan 1 + 2) and decrease in [11 C]FLB 457 BP $_{ND}$ in the postamphetamine scan. This was not unexpected as the carrier mass for [11C]FLB 457 in the above studies were restricted to 0.6 µg (1.6 nmoles), a tracer dose at which it occupies less than 5% of D2 receptors in vivo (Sudo et al., 2001). Also, consistent with this report, an estimate of the in vivo D₂ receptor occupancy of [11C]FLB 457 during the peak uptake of the radioligand in the DLPFC suggested 6% ± 2% in the studies in this report (DLPFC D2 receptor occupancy was derived as $100 \times F/(K_D + F)$, where the free concentration of radioligand in the brain, F was obtained by multiplying the measured concentration of [¹¹C]FLB 457 in the cerebellum at the time of peak specific binding in DLPFC by f_{ND} ; and in vivo K_D value of 0.09 nM for [11C]FLB 457 in the DLPFC was used as reported by (Suhara et al., 1999). Given that the relatively low occupancy observed at peak uptake of the radioligand (6%) is followed by washout of the radiotracer, it is highly unlikely that the mass carried over contributed significantly to the amphetamineinduced decrease in BP_{ND} measured in the second scan, which was administered 3-h postamphetamine.

Increased between-subject variance in prefrontal cortical [¹¹C]FLB 457 BP_{ND}

To reliably and robustly measure amphetamineinduced DA release it is desirable that the specific signal be at least 50% of the nonspecific signal, i.e., $\mathrm{BP_{ND}} > 0.5$ (for review, see Laruelle, 2000; Slifstein et al., 2004b). As evidenced in our data, a relatively low baseline [11C]FLB 457 BP_{ND} with high variability as was observed in the DLPFC (0.58 ± 0.32), OFC (0.67 ± 0.34) , and MPFC (0.78 ± 0.35) could be a problem in measuring amphetamine effect in some subjects in these regions. In our data set this variance in baseline BP_{ND} in the prefrontal cortical regions was driven by age (DLPFC $\rho = -0.76$, P =0.005; OFC $\rho = -0.69$, P = 0.013; and MPFC $\rho =$ -0.67, P = 0.018; where ρ is Pearson Correlation Coefficient; see Fig. 4 in which the DLPFC is shown as an example). This observation is consistent with previous [11C]FLB 457 studies demonstrating a significant age related decrease in cortical D2 receptor BP_{ND} that is most pronounced in the frontal cortex $(\sim 12\%$ per decade) (Inoue et al., 2001; Kaasinen et al., 2000b, 2002). When we restricted the data to subjects <35 years of age (n = 9 subjects), the [11C]FLB 457 BP_{ND} values in the DLPFC, OFC, and MPFC were 0.70 \pm 0.26, 0.80 \pm 0.27, and 0.91 \pm 0.30, respectively. These values are comparable to the striatal BP_{ND} (0.7 \pm 0.1) values of the SPECT D_2 radiotracer [123I]IBZM that has been widely used to measure amphetamine-induced DA release in clinical populations such as schizophrenia (Laruelle et al., 1996) and cocaine addiction (Malison et al., 1999). Thus, our relatively small dataset (n = 12 subjects)suggests that the use of [11C]FLB 457 to measure amphetamine-induced DA release in the prefrontal cortex is likely to be successful only in young adult subjects (<35 years of age). Further studies are necessary to confirm this preliminary observation in a larger representative sample.

Displacement of [11C]FLB 457 in OFC

In our studies amphetamine failed to induce displacement by DA of [11C]FLB 457 in the OFC (-5% \pm 21%). This result was in contrast to the robust displacements (DLPFC [$^{11}\text{C}]\text{FLB}$ 457 ΔBP_{ND} -13% \pm 15%; MPFC [11 C]FLB 457 Δ BP_{ND} $-11\% \pm 14\%$) observed in the other two prefrontal cortical regions of interest. Nevertheless, this is likely to be a significant issue because the study of DA transmission in the OFC is of interest in addiction (Winstanley, 2007). One possible explanation for not detecting an amphetamine effect in the OFC is that [11C]FLB 457 binding is associated with poor within-subject reproducibility in this particular region. A second possible explanation is that amphetamine leads to less displacement of [11C]FLB 457 binding in the OFC relative to the DLPFC in healthy controls. Future test-retest studies as well as expansion of our current study cohort are likely to provide us with the information necessary to further understand this discrepant observation in the OFC.

Magnitude of [11C]FLB 457 in the prefrontal cortical regions (DLPFC and MPFC)

Another interesting observation was that the magnitude of the amphetamine-induced displacement of [11 C]FLB 457 in the DLPFC and MPFC (\sim 12%) was comparable to that previously reported in the striatum ($\sim 10\%$) with other D_2 antagonist radiotracers (for review, see Laruelle, 2000). Although these data appear discrepant because the striatum receives a greater DA projection from the VTA/substantia nigra, these data are consistent with previous [11C]FLB 457 and [18F]fallypride studies, which have reported displacement values in the range of 10-15% in regions such as the hippocampus, ACC, DLPFC, and MPFC following amphetamine administration or cognitive stimulation (Aalto et al., 2005; Cropley et al., 2008; Slifstein et al., 2007a). A possible reason for comparable decreases in the striatum and DLPFC/MPFC binding following amphetamine is due to the difference in baseline DA levels between these regions. This hypothesis is consistent with combined microdialysis-PET studies, which have demonstrated that increases in endogenous DA following acute amphetamine challenge leads to a relatively larger change in [¹⁸F]fallypride BP_{ND} in the cingulate cortex than in the striatum (Slifstein et al., 2007b). In other words, the in vivo binding of the radioligand is more vulnerable or sensitive to endogenous competition by DA in the cortex than in the striatum because of less DA at baseline in the cortex. Combined PET-microdialysis studies with [11C]FLB 457 in nonhuman primates are likely to further our understanding of these mechanistic differences between amphetamine-induced DA transmission in the striatum and cortex.

Use of oral rather than intravenous d-amphetamine (amphetamine) as a challenge

Several groups have recently demonstrated the safety and effectiveness of oral amphetamine (dexedrine, 0.5 mg kg⁻¹) to measure amphetamine-induced DA release with PET (Cardenas et al., 2004; Cropley et al., 2008; Riccardi et al., 2005; Willeit et al., 2008). In contrast to intravenous amphetamine (0.3 mg kg⁻¹), which can lead to high fluctuations in vital signs and EKG parameters, oral amphetamine has been shown to have less effect on these parameters. For example, the mean peak systolic blood pressure following 0.5 mg kg⁻¹ oral amphetamine (n = 24 administrations) in this study was 160 \pm 13 mm of Hg, as opposed to 180 \pm 8 following 0.3 mg kg⁻¹ intravenous amphetamine (n = 24 administrations, healthy subjects) in Martinez et al. (2007).

In a recent study of healthy controls (Ziolko et al., 2007; n = 8, healthy women subjects, 27 ± 5 years of age), we were able to successfully displace the binding of [11C]raclopride in striatal subdivisions following oral amphetamine 0.5 mg kg⁻¹ ([¹¹C]raclopride ΔBP_{ND} ; RM ANOVA, F = 31.3, df = 4, 13; P <0.0001). Of significant interest is the fact that the displacements in the three functional subdivisions of the striatum (limbic striatum, LST—15% ± 8%; associative striatum, AST—10% ± 5%, and sensorimotor striatum, SMST $-17\% \pm 5\%$) were comparable to that reported in a previous [11C]raclopride PET study, which used intravenous amphetamine 0.3 mg kg⁻¹ (LST $-15\% \pm 10\%$; AST $-8\% \pm 7\%$; and SMST -16± 9 Martinez et al., 2003). These data clearly demonstrate that oral amphetamine is as effective as intravenous amphetamine in measuring amphetamineinduced DA release with PET.

Higher plasma variance in amphetamine levels between subjects has been mentioned as a criticism for using oral rather than intravenous amphetamine to measure amphetamine-induced DA release with PET. PET studies in which higher plasma variance have been observed following oral amphetamine have typically scanned subjects after 1-2 h (Leyton et al., 2002; Willeit et al., 2008), as opposed to 3 h following amphetamine (Cropley et al., 2008; Riccardi et al., 2005). This is not surprising as pharmacokinetic studies have shown that the time taken for d-amphetamine to reach plasma C_{max} and remain relatively stable is 3-4 h (Angrist et al., 1987). Thus, in the current study subjects were scanned 3 h following amphetamine administration. Another factor that may have driven the variance in plasma amphetamine levels in these studies is the effect of food on gastrointestinal (GI) absorption kinetics of d-amphetamine. In the above studies, subjects were not allowed to eat solid foods 5 h before and 3 h after the dose of amphetamine to reduce the variance in GI

TABLE VI. Comparison of in vivo affinity of [11C]FLB 457 and [18F]fallypride

Study	Species	In vivo K'_{D} (nM)	In vivo $K_{\rm D}$ (nM)	Method
[11C]FLB				
Olsson et al., 2004	Human $(n = 10)$	$0.4\ (0.27-0.43)$		TEA
Suhara et al., 1999	Human $(n = 8)$	0.9(0.7-1.1)		TEA
Delforge et al., 2001	Baboon $(n = 4)$	0.3 (0.16-0.40)		MIA
Average	,	0.53	0.06^{a}	
[18F]Fallypride				
Christian et al., 2004	Rhesus $(n = 3)$	0.7(0.38-1)		MIA
Slifstein et al., 2004a	Baboon $(n = 3)$	1.3	0.2 (0.17 to 0.22)	NLKM, TEA
Average	Basson (it o)	1.0	0.14 ^a	1,222,1, 1211

 K_D is uncorrected for free fraction of radiotracer in the brain (f_{ND}) . The corrected value provides in vivo K_D val-

absorption kinetics. The between-subject variance in amphetamine plasma levels in our data (for example, at the time of start of the postamphetamine [11C]FLB 457 the variance was 72 ± 7 ng ml⁻¹; for more detailed data refer to Table I) is no different than that reported following intravenous amphetamine 0.3 $mg kg^{-1} (40 \pm 13 ng ml^{-1})$ in Martinez et al. 2003. Thus future studies could propose to use the relatively safer and equally effective oral amphetamine paradigm to study subcortical and cortical DA transmission in clinical populations.

The use of cerebellum as a reference region for [11C]FLB 457

Some, but not all (Farde et al., 1997; Olsson et al., 1999), studies suggest 11-75% of the binding of [11C]FLB 457 in the cerebellum is specific to D₂ receptors (Ahmad et al., 2006; Asselin et al., 2007; Delforge et al., 1999; Montgomery et al., 2006). We did not observe any change in [11 C]FLB 457 $V_{\rm T}$ CER before and after amphetamine (baseline $V_{
m T~CER}$ 5.7 \pm 1.2; postamphetamine $V_{\rm T~CER}$ 5.7 \pm 1.1; n=12 subjects). The nonspecific binding in the cerebellum was comparable to that observed in the pons (baseline $V_{\mathrm{T\ PONS}}$ 6.3 ± 1.2 ; postamphetamine $V_{\rm T~PONS}$ 6.1 ± 1.2 ; n =12 subjects), another brain region, which is relatively devoid of D₂ receptors (Hall et al., 1996). One possible reason for the discordant reports in the literature may be related to the differences in delineating the cerebellum such as including the vermis, which has been shown to receive DA input (Melchitzky and Lewis, 2000), or including the cerebro-cerebellar fissue, which may be more prone to spillage (or partial voluming) of signal from the cortex. We performed a subsampling of the cerebellum that carefully avoided both these subregions (vermis and cerebro-cerebellar fissure) and white matter (which may have different blood flow characteristics than the gray matter) Nevertheless, consistent with previous autoradiographic studies demonstrating near negligible D2 receptor binding in the cerebellum (0.1% of that observed in the putamen) for [123I]epidepride, the iodinated SPECT derivative of FLB 457 (Hall et al., 1996), our data supports the continued use of the cerebellum as a reference region for [11C]FLB 457.

In vivo K_D for [11C]FLB 457 and [11C]fallypride

The data from our results also suggests that the in vivo K_D of [11C]FLB 457 is 2-fold lower than that of [¹¹C]fallypride. This value is derived based on the following equation:

$$\frac{[^{11}C]FLB457BP_{F}}{[^{11}C]fallyprideBP_{F}} = \frac{[^{11}C]fallyprideK_{D}}{[^{11}C]FLB457K_{D}} = \frac{8.6}{4.3} = 2 \quad (5)$$

where BPF is specific binding compared with free plasma concentration calculated as BP_P/f_P. [11C]FLB 457 and [11C]fallypride BP_F values in Eq. 5 are the average values across the eight regions of interest evaluated.

Approximately 2-fold lower in vivo $K_{\rm D}$ for [11C]FLB 457 (0.06 nM) relative to [11C]fallypride (0.14 nM) is consistent with the in vivo K_D values published in the literature for both these radioligands (Table VI). This enhanced affinity is what leads to the increased signal to noise ratio associated with [11C]FLB 457 compared with [11C]fallypride in the cortical regions of interest (Table V).

Another point worthy of consideration is the fact that there were differences in the variability but not the magnitude of displacement as measured with [11C]FLB 457 and [11C]fallypride following an identical dose of amphetamine (as shown in Fig. 3). This observation of lack of differences in the magnitude of displacement of these two radiotracers of different affinity is yet another illustration of the fact that the affinity of the radiotracer does not impact on its vulnerability to endogenous competition by DA, given DA levels are constant and the radioligand is administered at tracer dose (for detailed discussion refer to Laruelle, 2000).

The average published in vivo K_D for both radioligands were corrected for $f_{\rm ND}$ using values from Table I ([$^{11}{\rm C}$]FLB 457 = 10.8% and [$^{11}{\rm C}$]fallypride = 14.0%).

CONCLUSIONS

Despite the methodological considerations discussed in the previous section, our results clearly demonstrate the superiority of [\$^{11}C\$]FLB 457 over [\$^{11}C\$]fallypride to measure amphetamine-induced DA release in the human cortex with PET. Further validation of [\$^{11}C\$]FLB 457 as a tool to image amphetamine-induced DA release is likely to allow for the measurement of in vivo DA transmission in the prefrontal cortex, which is of extreme interest to the study of several neuropsychiatric disorders such as schizophrenia, addiction, Parkinson's disease, and ADHD.

ACKNOWLEDGMENTS

The authors thank members of the PET Facility Staff who carried out the acquisition of PET data and care of all subjects during PET procedures. The authors also acknowledge the editorial assistance provided by Maureen A. May, BS.

REFERENCES

- Aalto S, Bruck A, Laine M, Nagren K, Rinne JO. 2005. Frontal and temporal dopamine release during working memory and attention tasks in healthy humans: A positron emission tomography study using the high-affinity dopamine D2 receptor ligand [11C]FLB 457. J Neurosci 25:2471–2477.
- Abi-Dargham A, Laruelle M, Seibyl J, Rattner Z, Baldwin RM, Zoghbi SS, Zea-Ponce Y, Bremner JD, Hyde TM, Charney DS, Hoffer PB, Innis RB. 1994. SPECT measurement of benzodiazepine receptors in human brain with [123-I]iomazenil: Kinetic and equilibrium paradigms. J Nucl Med 35:228–238.
- Abi-Dargham A, Martinez D, Mawlawi O, Simpson N, Hwang DR, Slifstein M, Anjilvel S, Pidcock J, Guo NN, Lombardo I, Mann JJ, Van Heertum R, Foged C, Halldin C, Laruelle M. 2000. Measurement of striatal and extrastriatal dopamine D1 receptor binding potential with [11C]NNC 112 in humans: Validation and reproducibility. J Cereb Blood Flow Metab 20:225–243.
- Abi-Dargham A, Mawlawi O, Lombardo I, Gil R, Martinez D, Huang Y, Hwang DR, Keilp J, Kochan L, Van Heertum R, Gorman JM, Laruelle M. 2002. Prefrontal dopamine D1 receptors and working memory in schizophrenia. J Neurosci 22:3708–3719.

 Abi-Dargham A, Simpson N, Kegeles L, Parsey R, Hwang DR, Anjil-
- Abi-Dargham A, Simpson N, Kegeles L, Parsey R, Hwang DR, Anjilvel S, Zea-Ponce Y, Lombardo I, Van Heertum R, Mann JJ, Foged C, Halldin C, Laruelle M. 1999. PET studies of binding competition between endogenous dopamine and the D1 radiotracer [11C]NNC 756. Synapse 32:93–109.
- Ahmad R, Hirani E, Grasby PM, Hume SP. 2006. Effect of reduction in endogenous dopamine on extrastriatal binding of [11C]FLB 457 in rat brain—An ex vivo study. Synapse 59:162–172.
- Angrist B, Corwin J, Bartlik B, Cooper T. 1987. Early pharmacokinetics and clinical effects of oral D-amphetamine in normal subjects. Biol Psychiatry 22:1357–1368.
- Asselin MC, Montgomery AJ, Grasby PM, Hume SP. 2007. Quantification of PET studies with the very high-affinity dopamine D2/D3 receptor ligand [11C]FLB 457: Re-evaluation of the validity of using a cerebellar reference region. J Cereb Blood Flow Metab 27:378-392
- Benjamini Y, Hochberg Y. 1995. Controlling the false discovery rate: A practical and powerful approach to multiple testing. J R Stat Soc Ser B 57:289-300.
- Boileau I, Dagher A, Leyton M, Gunn RN, Baker GB, Diksic M, Benkelfat C. 2006. Modeling sensitization to stimulants in humans: An [11C]raclopride/positron emission tomography study in healthy men. Arch Gen Psychiatry 63:1386–1395.
- Breier A, Su TP, Saunders R, Čarson ŘE, Kolachana BS, deBartolomeis A, Weinberger DR, Weisenfeld N, Malhotra AK, Eckelman WC, Pickar D. 1997. Schizophrenia is associated with elevated amphetamine-induced synaptic dopamine concentrations: Evidence from a novel positron emission tomography method. Proc Natl Acad Sci USA 94:2569–2574.

- Cardenas L, Houle S, Kapur S, Busto UE. 2004. Oral D-amphetamine causes prolonged displacement of [11C]raclopride as measured by PET. Synapse 51:27–31.
- Carson RE, Breier A, deBartolomeis A, Saunders RC, Su TP, Schmall B, Der MG, Pickar D, Eckelman WC. 1997. Quantification of amphetamine-induced changes in [C-11]raclopride binding with continuous infusion. J Cereb Blood Flow Metab 17:437–447.
- Chou YH, Halldin C, Farde L. 2000. Effect of amphetamine on extrastriatal D2 dopamine receptor binding in the primate brain: A PET study. Synapse 38:138–143.
- Christian BT, Narayanan T, Shi B, Morris ED, Mantil J, Mukherjee J. 2004. Measuring the in vivo binding parameters of [18F]-fall-ypride in monkeys using a PET multiple-injection protocol. J Cereb Blood Flow Metab 24:309–322.
- Cropley VL, Innis RB, Nathan PJ, Brown AK, Sangare JL, Lerner A, Ryu YH, Sprague KE, Pike VW, Fujita M. 2008. Small effect of dopamine release and no effect of dopamine depletion on [(18)F]fallypride binding in healthy humans. Synapse 62:399–408.
- Delforge J, Bottlaender M, Loc'h C, Dolle F, Syrota A. 2001. Parametric images of the extrastriatal D2 receptor density obtained using a high-affinity ligand (FLB 457) and a double-saturation method. J Cereb Blood Flow Metab 21:1493–1503. Delforge J, Bottlaender M, Loc'h C, Guenther I, Fuseau C, Bend-
- Delforge J, Bottlaender M, Loc'h C, Guenther I, Fuseau C, Bendriem B, Syrota A, Maziere B. 1999. Quantitation of extrastriatal D2 receptors using a very high-affinity ligand (FLB 457) and the multi-injection approach. J Cereb Blood Flow Metab 19:533–546.
- Endres CJ, Kolachana BS, Saunders RC, Su T, Weinberger D, Breier A, Eckelman WC, Carson RE. 1997. Kinetic modeling of [C-11]raclopride: Combined PET-microdialysis studies. J Cereb Blood Flow Metab 17:932–942.
- Farde L, Suhara T, Nyberg S, Karlsson P, Nakashima Y, Hietala J, Halldin C. 1997. A PET study of [C-11]FLB 457 binding to extrastriatal D-2-dopamine receptors in healthy subjects and antipsychotic drug-treated patients. Psychopharmacology 133:396–404.
- Gandelman MS, Baldwin RM, Zoghbi SS, Zea-Ponce Y, Innis RB. 1994. Evaluation of ultrafiltration for the free fraction determination of single photon emission computerized tomography (SPECT) radiotracers: β-CIT. IBF and iomazenil. J Pharm Sci 83:1014–1019
- Gunn RN, Sargent PA, Bench CJ, Rabiner EA, Osman S, Pike VW, Hume SP, Grasby PM, Lammertsma AA. 1998. Tracer kinetic modeling of the 5-HT1A receptor ligand [carbonyl-11C]WAY-100635 for PET. Neuroimage 8:426-440.
- Hagelberg N, Aalto S, Kajander J, Oikonen V, Hinkka S, Nagren K, Hietala J, Scheinin H. 2004. Alfentanil increases cortical dopamine D2/D3 receptor binding in healthy subjects. Pain 109:86–93.
- Hall H, Farde L, Halldin C, Hurd YL, Pauli S, Sedvall G. 1996. Autoradiographic localization of extrastriatal D2-dopamine receptors in the human brain using [125I]epidepride. Synapse 23:115–123.
- Halldin C, Farde L, Hogberg T, Mohell N, Hall H, Suhara T, Karlsson P, Nakashima Y, Swahn CG. 1995. Carbon-11-FLB 457: A radioligand for extrastriatal D2 dopamine receptors. J Nucl Med 36:1275–1281.
- Hill AV. 1910. The possible effects of the aggregation of the molecules of haemoglobin on its disassociation curves. Phys Med Biol 1991:749–761.
- Innis R, Cunningham VJ, Delforge J, Fujioka K, Gjedde A, Gunn R, Holden JE, Houle S, Huang SC, Ichise M, Hidehiro I, Hiroshi I, Yuichi K, Koeppe RA, Knudsen GM, Juhani K, Lammertsma AA, Laruelle M, Logan J, Maguire RP, Mintun M, Morris ED, Parsey RV, Price CJ, M S, Sossi V, Suhara T, Votaw JR, DF W, Carson RE. 2007. Consensus nomenclature for in vivo imaging of reversibly binding radioligands. J Cereb Blood Flow Metab 27:1533–1539
- Inoue M, Suhara T, Sudo Y, Okubo Y, Yasuno F, Kishimoto T, Yoshikawa K, Tanada S. 2001. Age-related reduction of extrastriatal dopamine D2 receptor measured by PET. Life Sci 69: 1079–1084.
- Ito H, Sudo Y, Suhara T, Okubo Y, Halldin C, Farde L. 2001. Error analysis for quantification of [(11)C]FLB 457 binding to extrastriatal D(2) dopamine receptors in the human brain. Neuroimage 13:531–539.
- Kaasinen V, Kemppainen N, Nagren K, Helenius H, Kurki T, Rinne JO. 2002. Age-related loss of extrastriatal dopamine D(2) -like receptors in women. J Neurochem 81:1005–1010.
- Kaasinen V, Nagren K, Hietala J, Farde L, Rinne JO. 2001. Sex differences in extrastriatal dopamine d(2)-like receptors in the human brain. Am J Psychiatry 158:308–311.
- Kaasinen V, Nagren K, Hietala J, Oikonen V, Vilkman H, Farde L, Halldin C, Rinne JO. 2000a. Extrastriatal dopamine D2 and D3 receptors in early and advanced Parkinson's disease. Neurology 54:1482–1487.

- Kaasinen V, Vilkman H, Hietala J, Nagren K, Helenius H, Olsson H, Farde L, Rinne J. 2000b. Age-related dopamine D2/D3 receptor loss in extrastriatal regions of the human brain. Neurobiol Aging 21:683–688.
- Kegeles LS, Zea-Ponce Y, Abi-Dargham A, Rodenhiser J, Wang T, Weiss R, Van Heertum RL, Mann JJ, Laruelle M. 1999. Stability of [1231]IBZM SPECT measurement of amphetamine-induced striatal dopamine release in humans. Synapse 31:302–308.
- Kemppainen N, Laine M, Laakso MP, Kaasinen V, Nagren K, Vahlberg T, Kurki T, Rinne JO. 2003. Hippocampal dopamine D2 receptors correlate with memory functions in Alzheimer's disease. Eur J Neurosci 18:149–154.
- Laruelle M. 2000. Imaging synaptic neurotransmission with in vivo binding competition techniques: A critical review. J Cereb Blood Flow Metab 20:423–451.
- Laruelle M, Abi-Dargham A, Innis RB. 1998.Imaging receptor occupancy by endogenous transmitters in humans. In: Carson R, Daube-Whiterspoon ME, Herscovith P, editors. Quantitative functional brain imaging with positron emission tomography. San Diego: Academic Press. 455–462.
- Laruelle M, Abi-Dargham A, van Dyck CH, Gil R, De Souza CD, Erdos J, McCance E, Rosenblatt W, Fingado C, Zoghbi SS, Baldwin RM, Seibyl JP, Krystal JH, Charney DS, Innis RB. 1996. Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug free schizophrenic subjects. Proc Natl Acad Sci USA 93:9235–9240.

Laruelle M, Abi-Dargham A, van Dyck CH, Rosenblatt W, Zea-Ponce Y, Zoghbi SS, Baldwin RM, Charney DS, Hoffer PB, Kung HF, Innis RB. 1995. SPECT imaging of striatal dopamine release after amphetamine challenge. J Nucl Med 36:1182–1190.

- Laruelle M, Iyer RN, Al-Tikriti MS, Zea-Ponce Y, Malison R, Zoghbi SS, Baldwin RM, Kung HF, Charney DS, Hoffer PB, Innis RB, Bradberry CW. 1997. Microdialysis and SPECT measurements of amphetamine-induced dopamine release in nonhuman primates. Synapse 25:1–14.
- Laruelle M, van Dyck C, Abi-Dargham A, Zea-Ponce Y, Zoghbi SS, Charney DS, Baldwin RM, Hoffer PB, Kung HF, Innis RB. 1994. Compartmental modeling of iodine-123-iodobenzofuran binding to dopamine D₂ receptors in healthy subjects. J Nucl Med 35:743–754.
- Leyton M, Boileau I, Benkelfat C, Diksic M, Baker G, Dagher A. 2002. Amphetamine-induced increases in extracellular dopamine, drug wanting, and novelty seeking: A PET/[11C]raclopride study in healthy men. Neuropsychopharmacology 27:1027–1035.
- Malison RT, Mechanic KY, Klummp H. 1999. Reduced amphetamine-stimulated dopamine release in cocaine addicts as measured by [123I]IBZM SPECT. J Nucl Med 40:110.
- Martinez D, Gil R, Slifstein M, Hwang DR, Huang Y, Perez A, Kegeles L, Talbot P, Evans S, Krystal J, Laruelle M, Abi-Dargham A. 2005. Alcohol dependence is associated with blunted dopamine transmission in the ventral striatum. Biol Psychiatry 58:779–786.
- Martinez D, Narendran R, Foltin RW, Slifstein M, Hwang DR, Broft A, Huang Y, Cooper TB, Fischman MW, Kleber HD, Laruelle M. 2007. Amphetamine-induced dopamine release: Markedly blunted in cocaine dependence and predictive of the choice to self-administer cocaine. Am J Psychiatry 164:622–629.
- Martinez D, Slifstein M, Broft A, Mawlawi O, Hwang DR, Huang Y, Cooper T, Kegeles L, Zarahn E, Abi-Dargham A, Haber SN, Laruelle M. 2003. Imaging human mesolimbic dopamine transmission with positron emission tomography. II. Amphetamine-induced dopamine release in the functional subdivisions of the striatum. J Cereb Blood Flow Metab 23:285–300
- J Cereb Blood Flow Metab 23:285–300.

 Melchitzky DS, Lewis DA. 2000. Tyrosine hydroxylase- and dopamine transporter-immunoreactive axons in the primate cerebellum. Evidence for a lobular- and laminar-specific dopamine innervation. Neuropsychopharmacology 22:466-472
- vation. Neuropsychopharmacology 22:466–472.

 Mintun MA, Raichle ME, Kilbourn MR, Wooten GF, Welch MJ. 1984. A quantitative model for the in vivo assessment of drug binding sites with positron emission tomography. Ann Neurol 15:217–227.
- Montgomery AJ, Asselin MC, Farde L, Grasby PM. 2007. Measurement of methylphenidate-induced change in extrastriatal dopamine concentration using [(11)C]FLB 457 PET. J Cereb Blood Flow Metab 2:369–377.
- Mukherjee J, Christian BT, Dunigan KA, Shi B, Narayanan TK, Satter M, Mantil J. 2002. Brain imaging of 18F-fallypride in normal volunteers: Blood analysis, distribution, test-retest studies, and preliminary assessment of sensitivity to aging effects on dopamine D-2/D-3 receptors. Synapse 46:170–188.

 Mukherjee J, Shi B, Christian BT, Chattopadhyay S, Narayanan
- Mukherjee J, Shi B, Christian BT, Chattopadhyay S, Narayanan TK. 2004. 11C-Fallypride: Radiosynthesis and preliminary evaluation of a novel dopamine D2/D3 receptor PET radiotracer in non-human primate brain. Bioorg Med Chem 12:95–102.

- Mukherjee J, Yang ZY, Das MK, Brown T. 1995. Fluorinated benzamide neuroleptics—III. Development of (S)-N-[(1-allyl-2-pyrrolidinyl)methyl]-5-(3-[18F]fluoropropyl)-2,3-dimethoxybenzamide as an improved dopamine D-2 receptor tracer. Nucl Med Biol 22: 283—296.
- Mukherjee J, Yang ZY, Lew R, Brown T, Kronmal S, Cooper MD, Seiden LS. 1997. Evaluation of d-amphetamine effects on the binding of dopamine D-2 receptor radioligand, F-18-fallypride in nonhuman primates using positron emission tomography. Synapse 27:1_13
- Okauchi T, Suhara T, Maeda J, Kawabe K, Obayashi S, Suzuki K. 2001. Effect of endogenous dopamine on endogenous dopamine on extrastriated [(11)C]FLB 457 binding measured by PET. Synapse 41:87–95.
- Okubo Y, Olsson H, Ito H, Lofti M, Suhara T, Halldin C, Farde L. 1999. PET mapping of extrastriatal D2-like dopamine receptors in the human brain using an anatomic standardization technique and [11C]FLB 457. Neuroimage 10:666-674.
- and [11C]FLB 457. Neuroimage 10:666–674.
 Olsson H, Halldin C, Farde L. 2004. Differentiation of extrastriatal dopamine D2 receptor density and affinity in the human brain using PET. Neuroimage 22:794–803.
- Olsson H, Halldin C, Swahn CG, Farde L. 1999. Quantification of [11C]FLB 457 binding to extrastriatal dopamine receptors in the human brain. J Cereb Blood Flow Metab 19:1164–1173.
- Price JC, Mason S, Lopresti B, Holt D, Simpson NR, Drevets W, Smith GS, Mathis CA. 1997. PET measurements of endogenous neurotransmitter activity using high and low affinity radiotracers. Neuroimage 5:B77.
- Reimer ML, Mamer OA, Zavitsanos AP, Siddiqui AW, Dadgar D. 1993. Determination of amphetamine, methamphetamine and desmethyldeprenyl in human plasma by gas chromatography/negative ion chemical ionization mass spectrometry. Biol Mass Spectrom 22:235–242.
- Riccardi P, Li R, Ansari MS, Zald D, Park S, Dawant B, Anderson S, Doop M, Woodward N, Schoenberg E, Schmidt D, Baldwin R, Kessler R. 2006. Amphetamine-induced displacement of [(18)F] fallypride in striatum and extrastriatal regions in humans. Neuropsychopharmacology 31:1016–1026.
- Slifstein M, Hwang DR, Huang Y, Guo N, Sudo Y, Narendran R, Talbot P, Laruelle M. 2004a. In vivo affinity of [(18)F]fallypride for striatal and extrastriatal dopamine D(2) receptors in nonhuman primates. Psychopharmacology (Berl) 175:974-286
- man primates. Psychopharmacology (Berl) 175:274–286. Slifstein M, Kegeles L, Hackett E, Castrillon J, Gonzales R, Bae J, Laruelle M, Abi-Dargham A. 2007a. Effect of amphetamine challenge on the binding of [18F]fallypride in the striatum and extrastriatal brain regions: A study in healthy human volunteers. Paper Presented at the 46th Annual Meeting of the American College of Neuropsychopharmacology, Boca Raton, FL.
- Slifstein M, Moore H, Kegeles L, Duvall M, Xiaoyan X, Hackett E, Castrillon J, Kambalov O, Scher E, Laruelle M, Abi-Dargham A. 2007b. Correlation between PET and microdialysis measurements of amphetamine induced DA release in striatal and extrastriatal brain regions in baboons. 48 (Suppl 2):113.
- Slifstein M, Narendran R, Hwang DR, Sudo Y, Talbot PS, Huang Y, Laruelle M. 2004b. Effect of amphetamine on [(18)F]fallypride in vivo binding to D(2) receptors in striatal and extrastriatal regions of the primate brain: Single bolus and bolus plus constant infusion studies. Synapse 54:46–63.
- Smith SM, Jenkinson M, Woolrich MW, Beckmann CF, Behrens TE, Johansen-Berg H, Bannister PR, De Luca M, Drobnjak I, Flitney DE, Niazy RK, Saunders J, Vickers J, Zhang Y, De Stefano N, Brady JM, Matthews PM. 2004. Advances in functional and structural MR image analysis and implementation as FSL. Neuroimage 23 (Suppl 1):S208—S219
- image 23 (Suppl 1):S208—S219.
 Sudo Y, Suhara T, Inoue M, Ito H, Suzuki K, Saijo T, Halldin C, Farde L. 2001. Reproducibility of [11 C]FLB 457 binding in extrastriatal regions. Nucl Med Commun 22:1215—1221.
- Suhara T, Okubo Y, Yasuno F, Sudo Y, Inoue M, Ichimiya T, Nakashima Y, Nakayama K, Tanada S, Suzuki K, Halldin C, Farde L. 2002. Decreased dopamine D2 receptor binding in the anterior cingulate cortex in schizophrenia. Arch Gen Psychiatry 59:25–30.
- Suhara T, Sudo Y, Okauchi T, Maeda J, Kawabe K, Suzuki K, Okubo Y, Nakashima Y, Ito H, Tanada S, Halldin C, Farde L. 1999. Extrastriatal dopamine D2 receptor density and affinity in the human brain measured by 3D PET. Int J Neuropsychopharmacol 2:73–82.
- Talvik M, Nordstrom AL, Olsson H, Halldin C, Farde L. 2003. Decreased thalamic D2/D3 receptor binding in drug-naive patients with schizophrenia: A PET study with [11C]FLB 457. Int J Neuropsychopharmacol 6:361–370.
- Tsukada H, Miyasato K, Nishiyama S, Fukumoto D, Kakiuchi T, Domino EF. 2005. Nicotine normalizes increased prefrontal cortical dopamine D1 receptor binding and decreased working memory

performance produced by repeated pretreatment with MK-801: A PET study in conscious monkeys. Neuropsychopharmacology 30:

Vilkman H, Kajander J, Nagren K, Oikonen V, Syvalahti E, Hietala J. 2000. Measurement of extrastriatal D2-like receptor binding with [11C]FLB 457-a test-retest analysis. Eur J Nucl Med 27:1666-1673.

Villemagne VL, Wong DF, Yokoi F, Stephane M, Rice KC, Matecka D, Clough DJ, Dannals RF, Rothman RB. 1999. GBR12909 attenuates amphetamine-induced striatal dopamine release as measured by [(11)C]raclopride continuous infusion PET scans. Synapse 33:268–273.

Volkow ND, Wang GJ, Fowler JS, Logan J, Gatley SJ, Hitzemann R, Chen AD, Dewey SL, Pappas N. 1997. Decreased striatal dopaminergic responsiveness in detoxified cocaine-dependent subjects.

Nature 386:830-833.

Watson C. 2000. New, faster, image-based scatter correction for 3D

PET. IEEE Trans Nucl Sci 47:1587–1594. Willeit M, Ginovart N, Graff A, Rusjan P, Vitcu I, Houle S, Seeman P, Wilson AA, Kapur S. 2008. First human evidence of d-ampheta-

mine induced displacement of a D2/3 agonist radioligand: A [11C]-(+)-PHNO positron emission tomography study. Neuropsychopharmacology 33:279–289.

Winstanley CA. 2007. The orbitofrontal cortex, impulsivity, and addiction: Probing orbitofrontal dysfunction at the neural, neurochemical, and molecular level. Ann NY Acad Sci 1121:639-

Wu S, Ogden RT, Mann JJ, Parsey RV. 2007. Optimal metabolite curve fitting for kinetic modeling of 11C-WAY-100635. J Nucl Med 48:926-931.

Zhang Y, Brady M, Smith S. 2001. Segmentation of brain MR images through a hidden Markov random field model and the expectation-maximization algorithm. IEEE Trans Med Imaging 20: 45–57.

Ziolko S, Narendran R, Becker C, Carmichael O, Frankle WG, Kaye W, Price CJ. 2007. A comparison of automated, fully, deformable atlas-based segmentation and manually drawn striatal volumes of interest as applied to PET scans. Neuroimage S79: 320T-PM.