BreedR Overview

Facundo Muñoz

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Intro

What is breedR

- R-package implementing **statistical models** specifically suited for forest genetic resources analysts.
- Ultimately Mixed Models, but not necessarily easy to implement and use
- **breedR** acts as an **interface** which provides the means to:
  1. **Combine** any number of these models as **components** of a larger model
  2. Compute automatically **incidence** and **covariance matrices** from a few input parameters
  3. **Fit** the model
  4. Plot data and results, and perform **model diagnostics**

Installation

- Project web page http://famuvie.github.io/breedR/
  - Set up this URL as a package repository in `.Rprofile` (detailed instructions on the web)
  - `install.packages('breedR')`
  - Not possible to use `CRAN` due to closed-source `BLUPF90` programs
- GitHub dev-site https://github.com/famuvie/breedR
  - `if( !require(devtools) ) install.packages('devtools')`

  - `devtools::install_github('famuvie/breedR')`

Where to find help

- Package's help: `help(package = breedR)`
  - Help pages `?remlf90`
  - Code demos `demo(topic, package = 'breedR')` (omit `topic` for a list)
  - Vignettes `vignette(package = 'breedR')` (pkg and wiki)
- Wiki pages
  - Guides, tutorials, FAQ
- Mailing list http://groups.google.com/group/breedr
  - Questions and debates about usage and interface
- Issues page
  - Bug reports
  - Feature requests

License

Figure 1: GPL-3

- **breedR** is FOSS. Licensed GPL-3
  - `RShowDoc('LICENSE', package = 'breedR')`
- You can **use** and **distribute breedR** for any purpose
- You can **modify** it to suit your needs
  - we encourage to!
  - please consider contributing your improvements
  - you can **distribute** your modified version under the GPL
- However, **breedR** makes (intensive) use of the `BLUPF90` suite of Fortran programs, which are for *free* but not **free** (remember CRAN?)

Roadmap | Future developments

- Bayesian inference
- Multi-trait support
- Genotype×Environment interaction
- Support for longitudinal data

Functionality

Inference

Frequentist

- Currently, only **frequentist inference** is supported via REML estimation of variance components.
- The function `remlf90()`, provides an interface to both `REMLF90` and `AIREMLF90` functions in the `BLUPF90` suite of Fortran programs.
- Type `?remlf90` for details on the syntax

Bayesian

- It's on the roadmap for the next year
- Will use a gibbs sampler from `BLUPF90`, and possibly also `INLA`
- The **interface** will change a bit, separating the model specification from the fit

Linear Mixed Models with unstructured random effects

Example dataset

self	dad	mum	gen	gg	bl	phe_X	x	y	fam
69	0	64	1	14	13	15.756	0	0	64
70	0	41	1	4	13	11.141	3	0	41
71	0	56	1	14	13	19.258	6	0	56
72	0	55	1	14	13	4.775	9	0	55
73	0	22	1	8	13	19.099	12	0	22
74	0	50	1	14	13	19.258	15	0	50

```
## 'data.frame':    1021 obs. of  10 variables:
##  $ self : int  69 70 71 72 73 74 75 76 77 78 ...
##  $ dad  : int  0 0 0 0 0 0 0 0 0 4 ...
##  $ mum  : int  64 41 56 55 22 50 67 59 49 8 ...
##  $ gen  : Factor w/ 1 level "1": 1 1 1 1 1 1 1 1 1 1 ...
##  $ gg   : Factor w/ 14 levels "1","2","3","4",..: 14 4 14 14 8 14 14 14 14 11 ...
##  $ bl   : Factor w/ 15 levels "1","2","3","4",..: 13 13 13 13 13 13 13 13 9 9 ...
##  $ phe_X: num  15.76 11.14 19.26 4.78 19.1 ...
##  $ x    : int  0 3 6 9 12 15 18 21 24 27 ...
##  $ y    : int  0 0 0 0 0 0 0 0 0 0 ...
##  $ fam  : Factor w/ 63 levels "6","7","8","9",..: 59 36 51 50 17 45 62 54 44 3 ...
```

A simple Provenance Test

Specify the *genetic group* `gg` as an **unstructured random effect** using the standard `formulas` in `R`

$$\begin{aligned}\text{phe}_X =& \mu + Z\text{gg} + \varepsilon \\ \text{gg} \sim& N(0, \sigma^2_{\text{gg}}) \\ \varepsilon \sim& N(0, \sigma^2_{\varepsilon})\end{aligned}$$

```
res <- remlf90(fixed  = phe_X ~ 1,
               random = ~ gg,
               data   = globulus)
```

```
## Using default initial variances given by default_initial_variance()
## See ?breedR.getOption.
```

Initial variances specification

To avoid the notification, initial values for *all* the variance components must be made explicit using the argument `var.ini`:

```
res <- remlf90(fixed = phe_X ~ 1,
               random = ~ gg,
               var.ini = list(gg = 2, resid = 10),
               data = globulus)
```

Although in most cases the results will not change at all, we encourage to give explicit initial values for variance components. Specially when some estimate can be artifact. This is also useful for checking sensitivity to initial values.

Exploring the results

```
summary(res)
```

```
## Formula: phe_X ~ 0 + Intercept + gg
##    Data: globulus
##   AIC  BIC logLik
##  5864 5874  -2930
##
## Parameters of special components:
##
##
## Variance components:
##          Estimated variances   S.E.
## gg                     2.857 1.3584
## Residual              17.695 0.7888
##
## Fixed effects:
##            value   s.e.
## Intercept 14.799 0.4911
```

- Note that `AI-REML` has been used by default.
- You can also specify `method = 'em'`.
- Learn about the difference.

Further *extractor* functions

```
fixef(res)
```

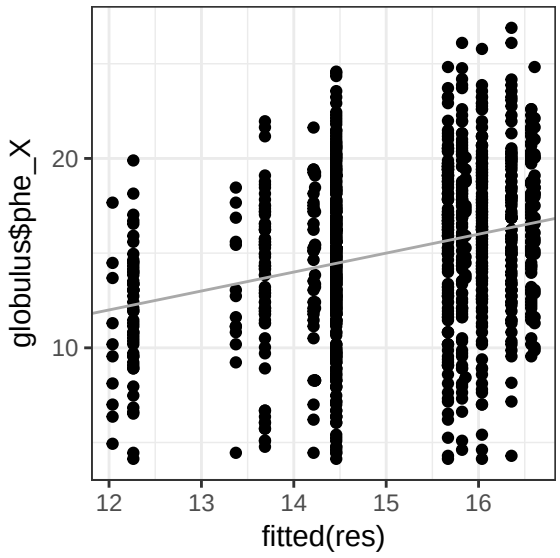
```
## $Intercept
##      value      s.e.
## 1 14.79913 0.4910931
```

```
ranef(res)
```

```
## $gg
##          value      s.e.
## 1  -1.1113031 0.6582245
## 2  -0.5850024 0.8241561
## 3   1.2381743 0.6017957
## 4  -2.5360692 0.7047331
## 5   1.0223492 0.6298409
## 6  -2.7605955 1.0884704
## 7  -0.5691183 0.9776411
## 8   0.8700425 0.5933964
## 9   1.5572484 0.6381498
## 10 -1.4262287 0.9961138
## 11  1.7715256 0.6527002
## 12  1.8079958 0.8241561
## 13  1.0604393 0.9776411
## 14 -0.3394577 0.5380184
```

Further *extractor* functions

```
qplot(
      fitted(res),
      globulus$phe_X) +
  geom_abline(intercept = 0,
              slope = 1,
              col = 'darkgrey')
```



```r
str(resid(res))
```

```
##  Named num [1:1021] 1.3 -1.12 4.8 -9.68 3.43 ...
##  - attr(*, "names")= chr [1:1021] "1" "2" "3" "4" ...
```

```r
extractAIC(res)
```

```
## [1] 5863.716
```

```r
logLik(res)
```

```
## 'log Lik.' -2929.858 (df=2)
```

Hierarchical and Factorial models

- In globulus, the **family** (`mum`) is *nested* within the **provenance** (gg)
- This is a matter of codification:

Nested factors

gg	mum
A	1
A	2
B	3
B	4

Crossed factors

gg	mum
A	1
A	2
B	1
B	2

Model specification

- Otherwise, in both cases we specify the model in the same way:

```r
random = ~ gg + factor(mum)  # note that mum is numeric
```

- Furthermore, this approach can handle unbalanced and mixed designs

Interactions

- Standard `R` notation:

```r
random = ~ gg * factor(mum)
```

- Not available yet (feature request?)
- Workaround: build the interaction variable manually
- Example: `gg` and `block` are crossed factors

```
dat <- transform(globulus,
                 interaction = factor(gg:bl))
random = ~ gg + bl + interaction
```

Exercise | Hierarchical and Factorial models

1. Use `remlf90()` and the globulus dataset to fit
   - a hierarchical model using `mum` **within** `gg`
   - a factorial model using `gg` and `bl`
2. Explore the results with `summary()`
   - is the family (`mum`) effect **relevant**?
   - is there any evidence of interaction between `gg` and `bl`?

Hierarchical and Factorial models #1 | Fitting models

```
res.h <- remlf90(fixed = phe_X ~ 1,
                 random = ~ factor(mum) + gg,
                 data = globulus)
```

```
# Interaction variable
globulus.f <- transform(globulus,
                        gg_bl = factor(gg:bl))

res.f <- remlf90(fixed = phe_X ~ 1,
                 random = ~ gg + bl + gg_bl,
                 data = globulus.f)
```

Hierarchical and Factorial models #2 | Hierarchical model

- The family effect is not very **important**, in terms of explained variance
- However, the model is a bit better with it (AIC, logLik)

```
summary(res)
```

```
## Formula: phe_X ~ 0 + Intercept + gg
##    Data: globulus
##   AIC  BIC logLik
##  5864 5874  -2930
##
## Parameters of special components:
##
##
## Variance components:
##          Estimated variances   S.E.
## gg                     2.857 1.3584
## Residual              17.695 0.7888
##
## Fixed effects:
##            value   s.e.
## Intercept 14.799 0.4911
```

```
summary(res.h)
```

```
## Formula: phe_X ~ 0 + Intercept + factor(mum) + gg
##    Data: globulus
##   AIC  BIC logLik
##  5857 5872  -2926
##
## Parameters of special components:
##
##
## Variance components:
##             Estimated variances   S.E.
## factor(mum)              0.8955 0.4177
## gg                       2.0540 1.1706
## Residual                17.0770 0.7819
##
## Fixed effects:
##            value   s.e.
## Intercept 14.973 0.4702
```

Hierarchical and Factorial models #3 | Factorial model

- Looks like the interaction between **block** and **provenance** is negligible
- (apart from the fact that it makes no sense at all, and shuld not have been even considered in the first place)
- compare with the model without interaction

```
summary(res.f)
```

```
## Formula: phe_X ~ 0 + Intercept + gg + bl + gg_bl
##    Data: globulus.f
##   AIC  BIC logLik
##  5752 5772  -2872
##
## Parameters of special components:
##
##
## Variance components:
##          Estimated variances   S.E.
## gg                   3.10970 1.4329
## bl                   2.57280 1.0606
## gg_bl                0.02912 0.2713
## Residual            15.19800 0.7159
##
## Fixed effects:
##            value  s.e.
## Intercept 14.764 0.653
```

```
## result without interaction
res.f0 <- remlf90(fixed  = phe_X ~ 1,
                  random = ~ gg + bl,
                  data = globulus)
paste('AIC:', round(extractAIC(res.f0)),
      'logLik:', round(logLik(res.f0)))
```

```
## [1] "AIC: 5750 logLik: -2872"
```

Additive Genetic Effect

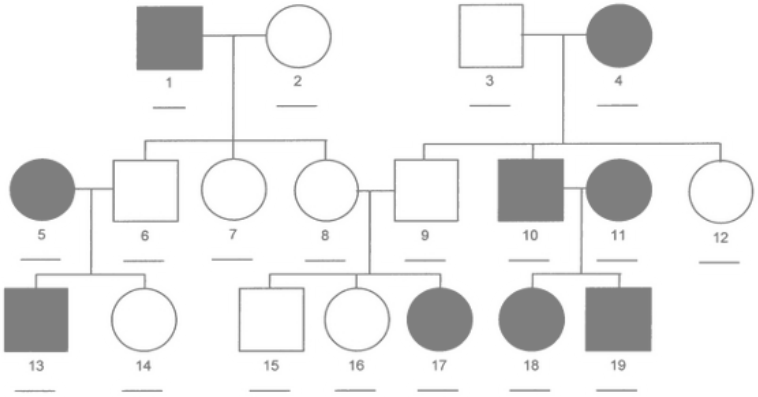


Figure 2: pedigree

What is an additive genetic effect

- Random effect at **individual level**
- Based on a **pedigree**
- BLUP of **Breeding Values** from own and relatives' phenotypes
- Represents the **additive component** of the genetic value
- More general:
- family effect is a particular case
- accounts for more than one generation
- mixed relationships
- More flexible: allows to select individuals within families

Specifying a *pedigree*

- A 3-column `data.frame` or `matrix` with the codes for each individual and its parents
- A **family** effect is easily translated into a pedigree:
  - use the **family code** as the identification of a fictitious **mother**
  - use `0` or `NA` as codes for the **unknown fathers**

self	dad	mum
69	0	64
70	0	41
71	0	56
72	0	55
73	0	22
74	0	50

Fitting an *animal model*

```
res.animal <- remlf90(fixed  = phe_X ~ 1,
                      random = ~ gg,
                      genetic = list(model = 'add_animal',
                                     pedigree = globulus[, 1:3],
                                     id = 'self'),
                      data = globulus)
```

Animal model: results

- gg explains almost the same amount of phenotypic variability
- The (additive) genetic effect explains **part** of the formerly residual variance
- The **heritability** is computed automatically as

$$h^2 = \frac{\sigma_a^2}{\sigma_a^2 + \sigma_{gg}^2 + \sigma^2}$$

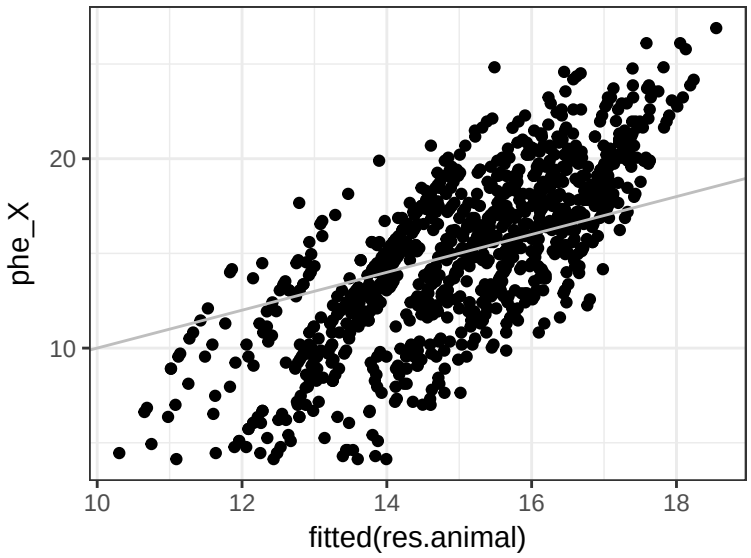
```
summary(res.animal)
```

```
## Formula: phe_X ~ 0 + Intercept + gg + pedigree
##    Data: globulus
##   AIC  BIC logLik
##  5857 5872  -2926
##
## Parameters of special components:
##
##
## Variance components:
##          Estimated variances  S.E.
## gg                     2.356 1.249
## genetic                3.632 1.649
## Residual              14.271 1.561
##
##              Estimate    S.E.
## Heritability   0.1795 0.08253
##
## Fixed effects:
##            value s.e.
## Intercept 14.797 0.47
```

Extracting Predicted Breeding Values

```
## Predicted Breeding Values
# for the full pedigree first, and for the observed individuals
# by matrix multiplication with the incidence matrix
PBV.full <- ranef(res.animal)$genetic
PBV <- model.matrix(res.animal)$genetic %*% PBV.full

# Predicted genetic values vs.
```

```r
# phenotype.
# Note: fitted = mu + PBV
qplot(fitted(res.animal), phe_X,
      data = globulus) +
  geom_abline(intercept = 0,
              slope = 1,
              col = 'gray')
```



Handling pedigrees

- The pedigree needs to meet certain conditions
- If it does not, **breedR** automatically completes, recodes and sorts
- If recoding is necessary, **breedR** issues a warning because you need to be careful when retrieving results
- See this guide for more details

Spatial autocorrelation

What is spatial autocorrelation

- The **residuals** of any LMM must be **noise**
- However, most times there are **environmental factors** that affect the response
- This causes that observations that are close to each other **tend** to be more similar that observations that are far away
- This is called **spatial autocorrelation**
- It may affect both the estimations and their accuracy
- This is why experiments are randomized into spatial **blocks**

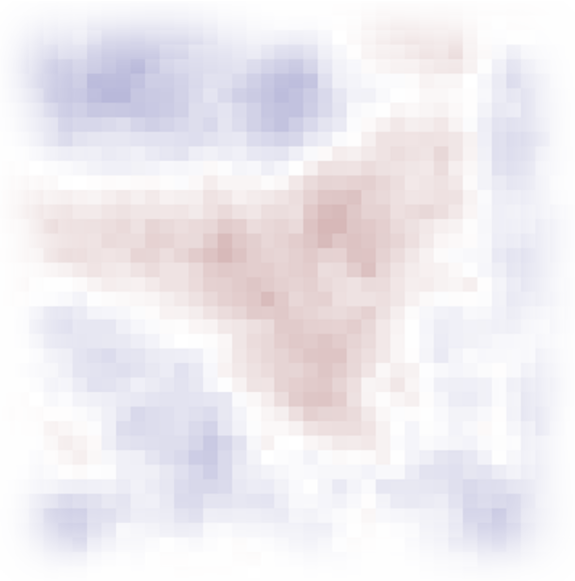
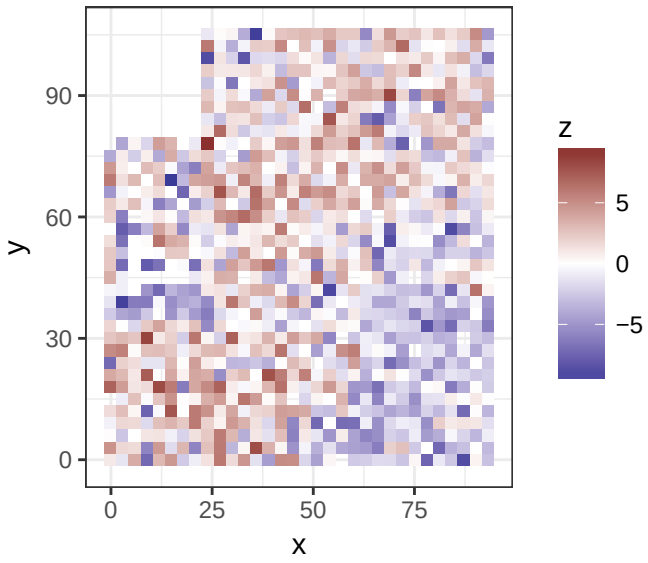


Figure 3: spatial

Diagnosing spatial autocorrelation | residuals spatial plot

- You can `plot()` the spatial arrangement of various model components (e.g. residuals)
- Look like **independent** gaussian observations (i.e. noise)?
- Do you see any **signal** in the background?

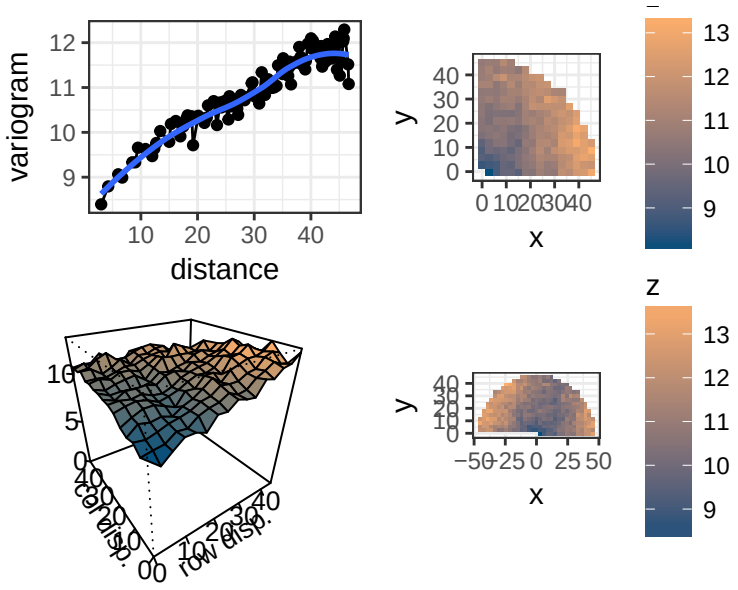
```
## Since coordinates have not
## been passed before they
## must be provided explicitly.
coordinates(res.animal) <-
  globulus[, c('x', 'y')]
plot(res.animal, 'resid')
```



Diagnosing spatial autocorrelation | variograms of residuals

- Plot the **variogram of residuals** with `variogram()`

```
variogram(res.animal)
```

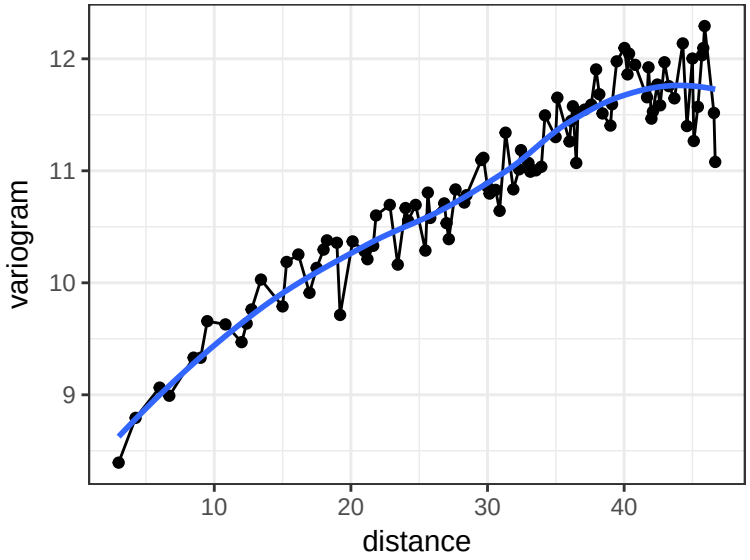


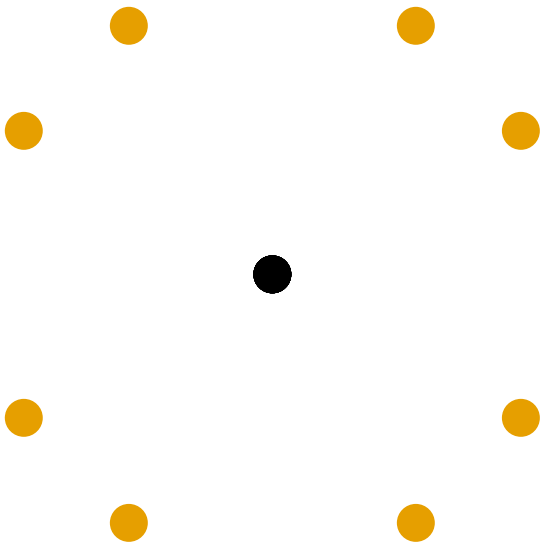
Interpreting the variograms

- Isotropic variogram:

$$\gamma(h) = \frac{1}{2} V[Z(\mathbf{u}) - Z(\mathbf{v})], \quad \text{dist}(\mathbf{u}, \mathbf{v}) = h$$

The **empirical** isotropic variogram is built by aggregating **all the pairs** of points separated by h, **no matter the direction**.



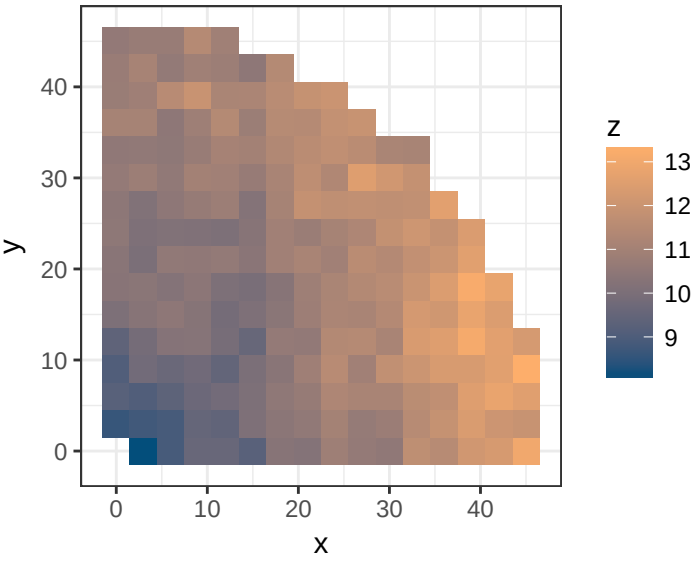


Interpreting the variograms

- Row/Column variogram:

$$\gamma(x, y) = \frac{1}{2} V[Z(\mathbf{u}) - Z(\mathbf{v})], \quad \text{dist}(\mathbf{u}, \mathbf{v}) = (x, y)$$

The **empirical** row/col variogram is built by aggregating **all the pairs** of points separated by exactly x rows and y columns.

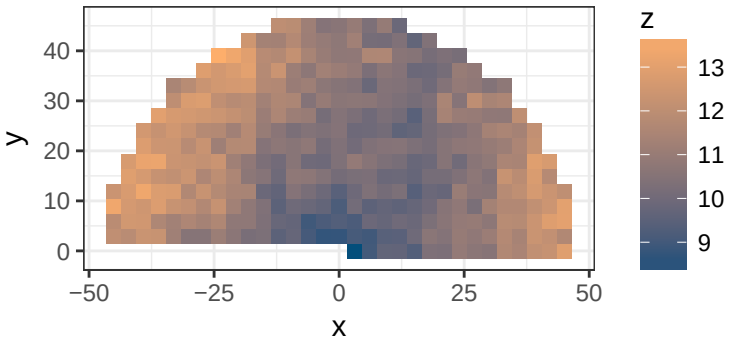


Interpreting the variograms

- Anisotropic variogram:

$$\gamma(\mathbf{x}) = \frac{1}{2} V[Z(\mathbf{u}) - Z(\mathbf{v})], \quad \mathbf{u} = \mathbf{v} \pm \mathbf{x}$$

The **empirical** anisotropic variogram is built by aggregating **all the pairs** of points **in the same direction** separated by $|\mathbf{x}|$.



Accounting for spatial autocorrelation

- Include an explicit **spatial effect** in the model
- I.e., a **random effect** with a specific covariance structure that reflects the spatial relationship between individuals
- The **block** effect, is a very particular case:
  - It is designed from the begining, possibly using prior knowledge
  - Introduces **independent** effects between blocks
  - Most neighbours are within the same block (i.e. share the same effect)

The `blocks` model

```
# The genetic component (DRY)
gen.globulus <- list(model    = 'add_animal',
                     pedigree = globulus[, 1:3],
                     id       = 'self')

res.blk <- remlf90(fixed   = phe_X ~ 1,
                   random  = ~ gg,
                   genetic = gen.globulus,
                   spatial = list(model = 'blocks',
                                  coord = globulus[, c('x', 'y')],
                                  id = 'bl'),
                   data    = globulus)
```

- The `blocks` spatial model is **equivalent** to `random = ~ bl`, but:
  - specifying `coord` is convenient for plotting (remember?)
  - `blocks` behaves as expected, even if `bl` is not a **factor**

Animal-spatial model: results

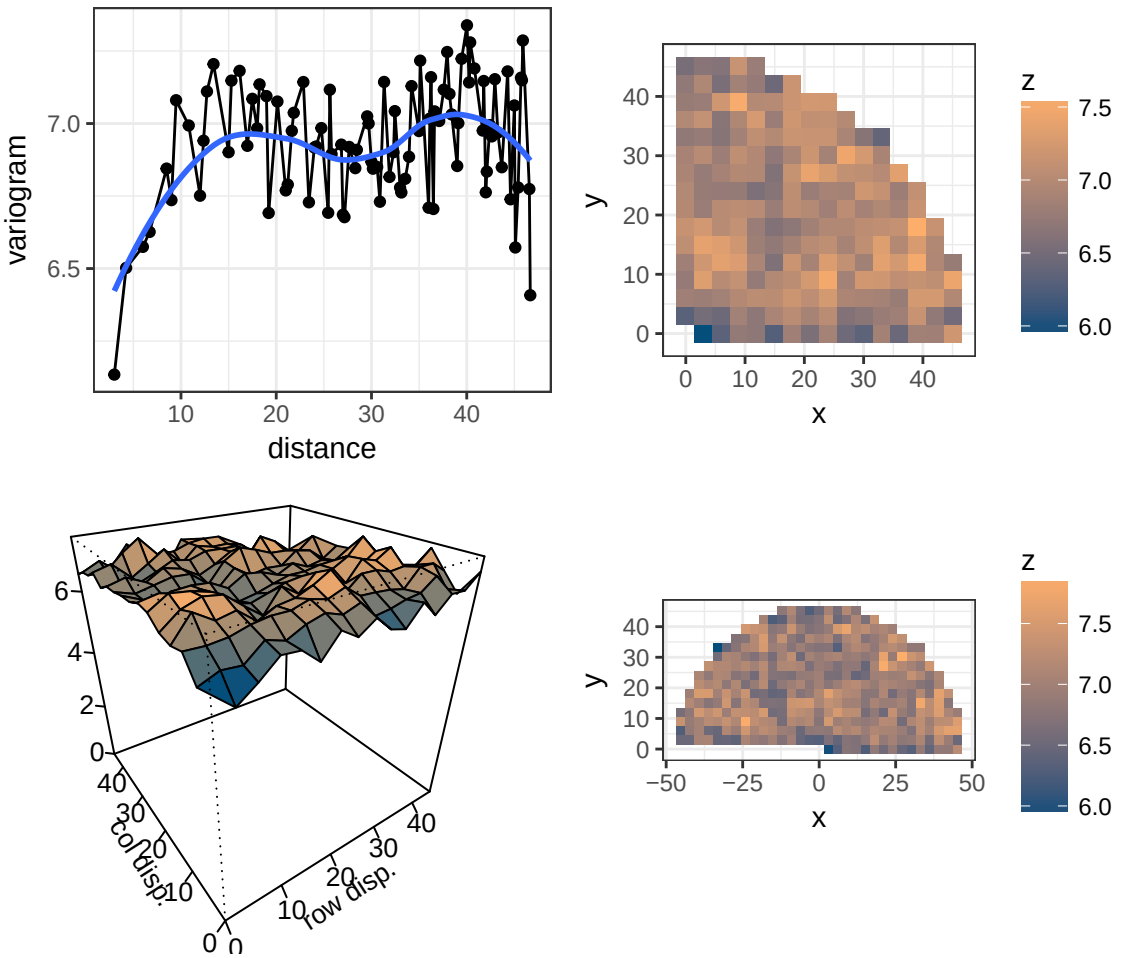
```
summary(res.blk)
```

```
## Formula: phe_X ~ 0 + Intercept + gg + pedigree + spatial
##    Data: globulus
##   AIC  BIC logLik
##  5734 5753  -2863
##
## Parameters of special components:
## spatial: n.blocks: 15
##
## Variance components:
##          Estimated variances  S.E.
## gg                     2.385 1.274
## genetic                5.275 1.836
## spatial                2.650 1.081
## Residual              10.279 1.601
##
##              Estimate    S.E.
## Heritability   0.2556 0.08989
```

```
##
## Fixed effects:
##             value   s.e.
## Intercept 14.762 0.6342
```

- Now the additive-genetic variance and the heritability have increased! (3.6 and 0.18 before)

Variogram of residuals



- There seems to remain some intra-block spatial autocorrelation

B-Splines model

- A continuous and smooth spatial surface built from a linear combination of **basis** functions
- The coefficients are modelled as a random effect

```
## Use the `em` method! `ai` does not like splines
res.spl  <- remlf90(fixed   = phe_X ~ 1,
                    random  = ~ gg,
                    genetic = gen.globulus,
                    spatial = list(model   = 'splines',
                                   coord   = globulus[, c('x','y')]),
                    data    = globulus, method  = 'em')
```

Autoregressive model

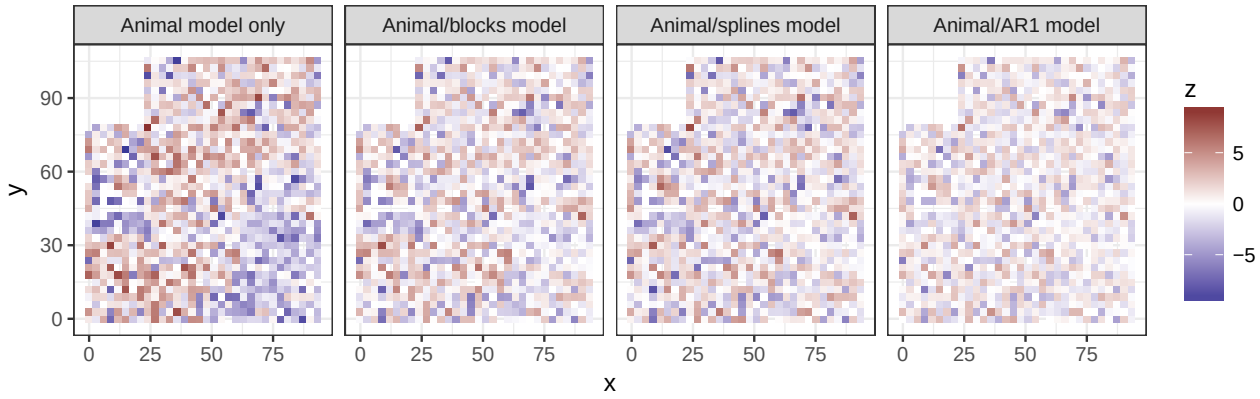
- A separable kronecker product of First order Autoregressive processes on the rows and the colums

```
res.ar1  <- remlf90(fixed   = phe_X ~ 1,
                    random  = ~ gg,
                    genetic = gen.globulus,
                    spatial = list(model = 'AR',
                                   coord = globulus[, c('x','y')]),
                    data    = globulus)
```

Change in model residuals

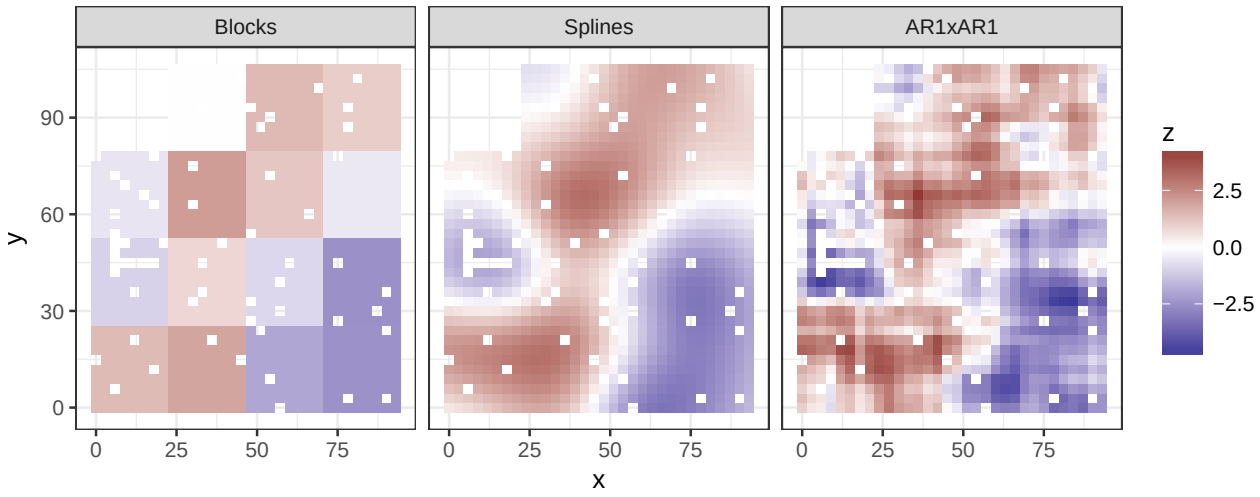
- We preserve the scale by using `compare.plots()`

```
compare.plots(
  list(`Animal model only` = plot(res.animal, 'residuals'),
       `Animal/blocks model` = plot(res.blk, 'residuals'),
       `Animal/splines model` = plot(res.spl, 'residuals'),
       `Animal/AR1 model` = plot(res.ar1, 'residuals')))
```



Comparison of spatial components

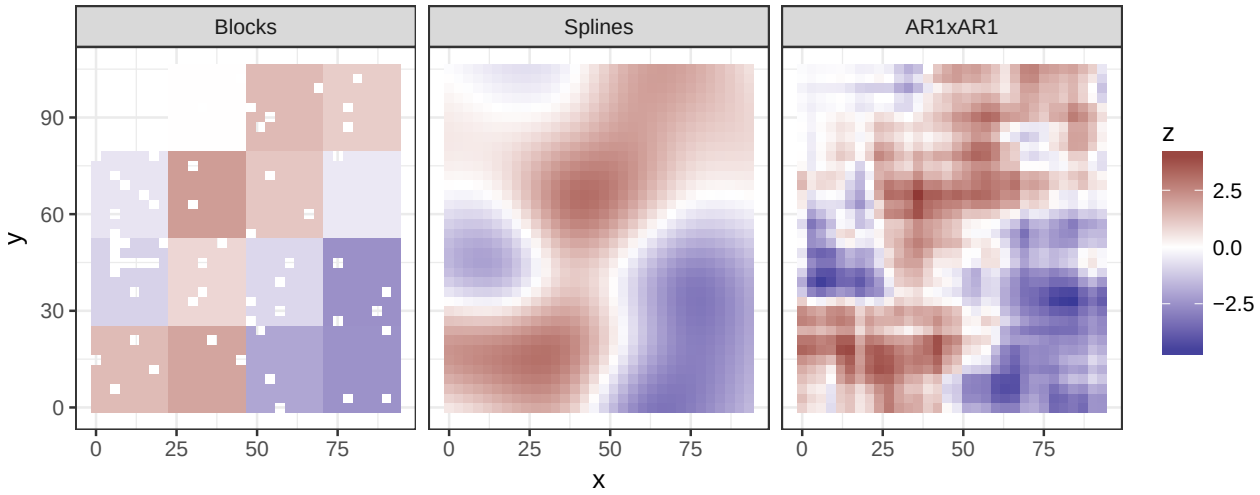
```
compare.plots(list(Blocks  = plot(res.blk, type = 'spatial'),
                   Splines = plot(res.spl, type = 'spatial'),
                   AR1xAR1 = plot(res.ar1, type = 'spatial')))
```



Prediction of the spatial effect in unobserved locations

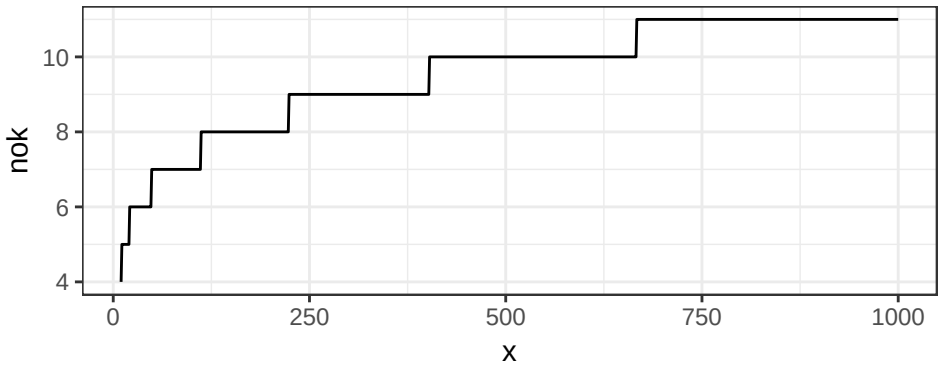
- The type `fullspatial` fills the holes (when possible)
- See `?plot.remlf90`

```
compare.plots(list(Blocks  = plot(res.blk, type = 'fullspatial'),
                   Splines = plot(res.spl, type = 'fullspatial'),
                   AR1xAR1 = plot(res.ar1, type = 'fullspatial')))
```



Spatial parameters | Number of knots of a `splines` model

- The smoothness of the spatial surface can be controlled modifying the number of base functions
- This is, directly determined by the **number of knots** (nok) in each dimension
- `n.knots` can be **used as an additional argument** in the `spatial` effect as a numeric vector of size 2.
- Otherwise, is determined by the function given in `breedR.getOption('splines.nok')`



Spatial parameters | Autoregressive parameters of a `AR` model

- Analogously, the *patchiness* of the `AR` effects can be controlled by the autoregressive parameter for each dimension
- `rho` can be **given as an additional argument** in the `spatial` effect as a numeric vector of size 2
- By default, **breedR** runs all the combinations in the grid produced by the values from `breedR.getOption('ar.eval')` and returns the one with largest likelihood
- It returns also the full table of combinations and likelihoods in `res$rho`

Exercise | Tuning spatial parameters

- Tuning parameters:
  - model `splines`: `n.knots`
  - model `AR`: `rho`

1. Increase the number of knots in the `splines` model and see if it improves the fit
2. Visualize the log-likelihood of the fitted `AR` models
3. Refine the grid around the most likely values, and refit using `rho = rho.grid`, where

```
rho.grid <- expand.grid(rho_r = seq(.7, .95, length = 4),
                        rho_c = seq(.7, .95, length = 4))
```

```
- What are now the most likely parameters?
```

Spatial #1 | B-splines model with increased `nok`

- `nok` were (6, 6) by default (see `summary()`)

```
res.spl99  <- remlf90(fixed  = phe_X ~ 1, random = ~ gg,
                      genetic = gen.globulus,
                      spatial = list(model   = 'splines',
                                     coord   = globulus[, c('x','y')],
                                     n.knots = c(9, 9)),
                      data = globulus, method = 'em')
```

```
summary(res.spl)
```

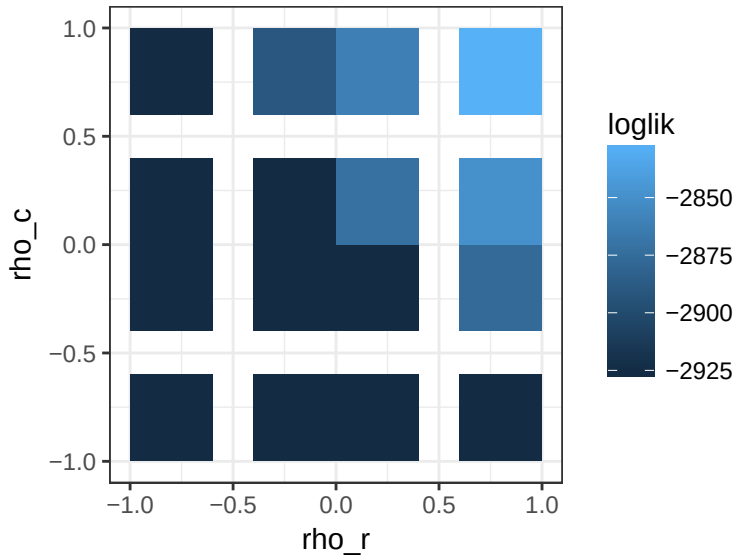
```
## Formula: phe_X ~ 0 + Intercept + gg + pedigree + spatial
##    Data: globulus
##   AIC     BIC logLik
##  5685 unknown  -2838
##
## Parameters of special components:
## spatial: n.knots: 12 12
##
## Variance components:
##          Estimated variances
## gg                     2.568
## genetic                4.498
## spatial                4.199
## Residual              10.070
##
## Fixed effects:
##            value   s.e.
## Intercept 14.479 0.9163
```

```
summary(res.spl99)
```

```
## Formula: phe_X ~ 0 + Intercept + gg + pedigree + spatial
##    Data: globulus
##   AIC     BIC logLik
##  5681 unknown  -2836
##
## Parameters of special components:
## spatial: n.knots: 15 15
##
## Variance components:
##          Estimated variances
## gg                     2.509
## genetic                4.651
## spatial                3.490
## Residual               9.552
##
## Fixed effects:
##            value   s.e.
## Intercept 14.611 0.6947
```

Spatial #2 | Visualize log-likelihoods

```
qplot(rho_r, rho_c,
      fill = loglik,
      geom = 'tile',
      data = res.ar1$rho)
```



rho_r	rho_c	loglik
-0.8	-0.8	-2925.648
-0.2	-0.8	-2925.647
0.2	-0.8	-2925.645
0.8	-0.8	-2925.636
-0.8	-0.2	-2925.647
-0.2	-0.2	-2925.645
0.2	-0.2	-2925.023
0.8	-0.2	-2876.893
-0.8	0.2	-2925.645
-0.2	0.2	-2925.645
0.2	0.2	-2871.691
0.8	0.2	-2849.814
-0.8	0.8	-2925.645
-0.2	0.8	-2890.606
0.2	0.8	-2860.981
0.8	0.8	-2828.017

Spatial #3 | Refine grid

```
rho.grid <- expand.grid(rho_r = seq(.7, .95, length = 4),
                        rho_c = seq(.7, .95, length = 4))
res.ar.grid  <- remlf90(fixed  = phe_X ~ gg,
                        genetic = list(model = 'add_animal',
                                       pedigree = globulus[,1:3],
                                       id = 'self'),
                        spatial = list(model = 'AR',
                                       coord = globulus[, c('x','y')],
                                       rho = rho.grid),
                        data = globulus)
summary(res.ar.grid)
```

```
## Formula: phe_X ~ 0 + gg + pedigree + spatial
```

```
##    Data: globulus
##   AIC  BIC logLik
##  5603 5617  -2798
##
## Parameters of special components:
## spatial: rho: 0.8666667 0.7833333
##
## Variance components:
##          Estimated variances  S.E.
## genetic                5.090 1.715
## spatial                4.984 1.053
## Residual               7.583 1.499
##
##              Estimate    S.E.
## Heritability   0.2878 0.09383
##
## Fixed effects:
##        value   s.e.
## gg.1  13.351 0.7195
## gg.2  14.331 0.9112
## gg.3  15.945 0.7698
## gg.4  11.585 0.9394
## gg.5  15.913 0.8200
## gg.6   9.593 1.6964
## gg.7  13.761 1.5681
## gg.8  15.521 0.7486
## gg.9  16.302 0.8260
## gg.10 12.684 1.1531
## gg.11 16.459 0.9849
## gg.12 16.801 1.1412
## gg.13 15.783 1.5665
## gg.14 14.211 0.6486
```

Competition

Theoretical model

- Each individual have **two** (unknown) Breeding Values (BV)
- The `direct` BV affects its **own** phenotype, while the `competition` BV affects its **neghbours'** (as the King moves)
- The effect of the neighbouring `competition` BVs is given by their sum **weighted by** $1/d^{\alpha}$, where α is a tuning parameter called `decay`
- Each set of BVs is modelled as a zero-mean **random effect** with structure matrix given by the **pedigree** and independent **variances** σ_a^2 and σ_c^2
- Both random effects are modelled jointly with **correlation** ρ

Permanent Environmental Effect (pec)

- **Optional** effect with **environmental** (rather than genetic) basis

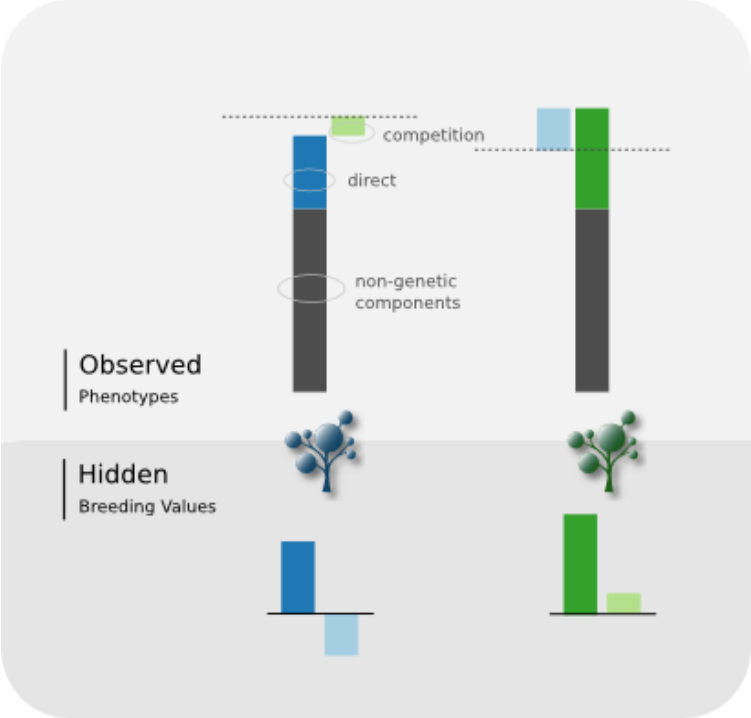


Figure 4: Competition model

- Modelled as an individual **independent** random effect that affects **neighbouring** trees in the same (weighted) way

Simulation of data

breedR implements a convenient dataset **simulator** which keeps a similar syntax.

- See `?simulation` for details on the syntax

```
# Simulation parameters
grid.size <- c(x=20, y=25) # cols/rows
coord <- expand.grid(sapply(grid.size,
                            seq))
Nobs <- prod(grid.size)
Nparents <- c(mum = 20, dad = 20)
sigma2_a <- 2   # direct add-gen var
sigma2_c <- 1   # compet add-gen var
rho      <- -.7 # gen corr dire-comp
sigma2_s <- 1   # spatial variance
sigma2_p <- .5  # pec variance
sigma2   <- .5  # residual variance

S <- matrix(c(sigma2_a,
              rho*sqrt(sigma2_a*sigma2_c),
              rho*sqrt(sigma2_a*sigma2_c),
              sigma2_c),
            2, 2)
```

```
set.seed(12345)
simdat <-
  breedR.sample.phenotype(
    fixed   = c(beta = 10),
    genetic = list(model = 'competition',
                   Nparents = Nparents,
                   sigma2_a = S,
                   check.factorial=FALSE,
                   pec = sigma2_p),
    spatial = list(model = 'AR',
                   grid.size = grid.size,
                   rho   = c(.3, .8),
                   sigma2_s = sigma2_s),
    residual.variance = sigma2
    )

## Remove founders
dat <- subset(simdat,
              !(is.na(simdat$sire)
                & is.na(simdat$dam)))
```

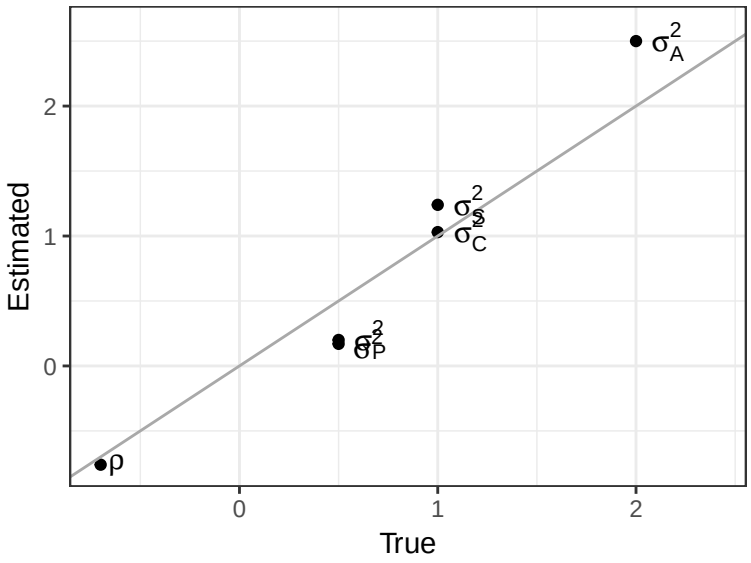
Fitting a competition model

```
system.time(
  res.comp <- remlf90(fixed   = phenotype ~ 1,
                      genetic = list(model = 'competition',
                                     pedigree = dat[, 1:3],
                                     id = 'self',
                                     coord = dat[, c('x', 'y')],
                                     competition_decay = 1,
                                     pec = list(present = TRUE)),
                      spatial = list(model = 'AR',
                                     coord = dat[, c('x', 'y')],
                                     rho   = c(.3, .8)),
                      data = dat,
                      method = 'em')  # AI diverges
  )
```

```
##    user  system elapsed
##  85.733   0.246  86.079
```

True vs. estimated parameters

	True	Estimated
direct	2.0	2.50
compet.	1.0	1.03
correl.	-0.7	-0.76
spatial	1.0	1.24
pec	0.5	0.20
residual	0.5	0.17



Exercise | Competition models

1. Plot the true vs predicted:
   - direct and competition Breeding Values
   - spatial effects
   - pec effects
2. Plot the residuals and their variogram
   - Do you think the residuals are independent?
   - How would you improve the analysis?

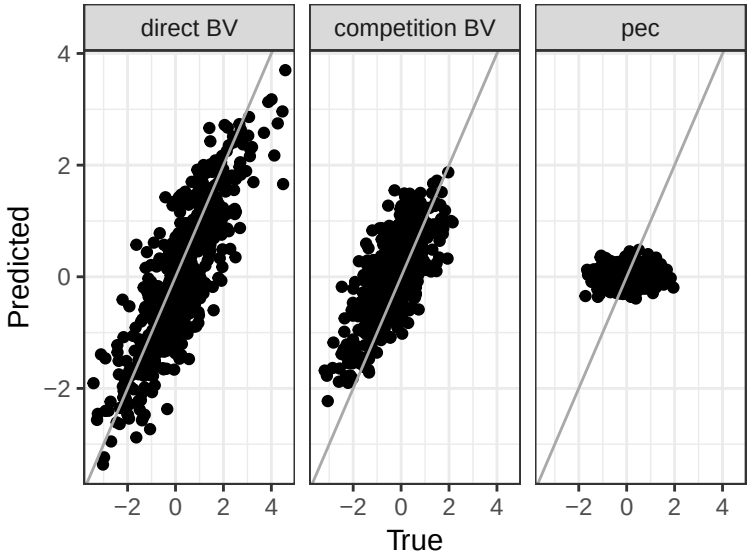
Competition #1 | True vs. predicted components

```
## compute the predicted effects for the observations
## by matrix multiplication of the incidence matrix and the BLUPs
pred <- list()
Zd <- model.matrix(res.comp)$'genetic_direct'
pred$direct <- Zd %*% ranef(res.comp)$'genetic_direct'

## Watch out! for the competition effects you need to use the incidence
## matrix of the direct genetic effect, to get their own value.
## Otherwise, you get the predicted effect of the neighbours on each
## individual.
pred$comp <- Zd %*% ranef(res.comp)$'genetic_competition'
pred$pec  <- model.matrix(res.comp)$pec %*% ranef(res.comp)$pec
```

Competition #1 | True vs. predicted components

```
comp.pred <-
  rbind(
    data.frame(
      Component = 'direct BV',
      True = dat$BV1,
```

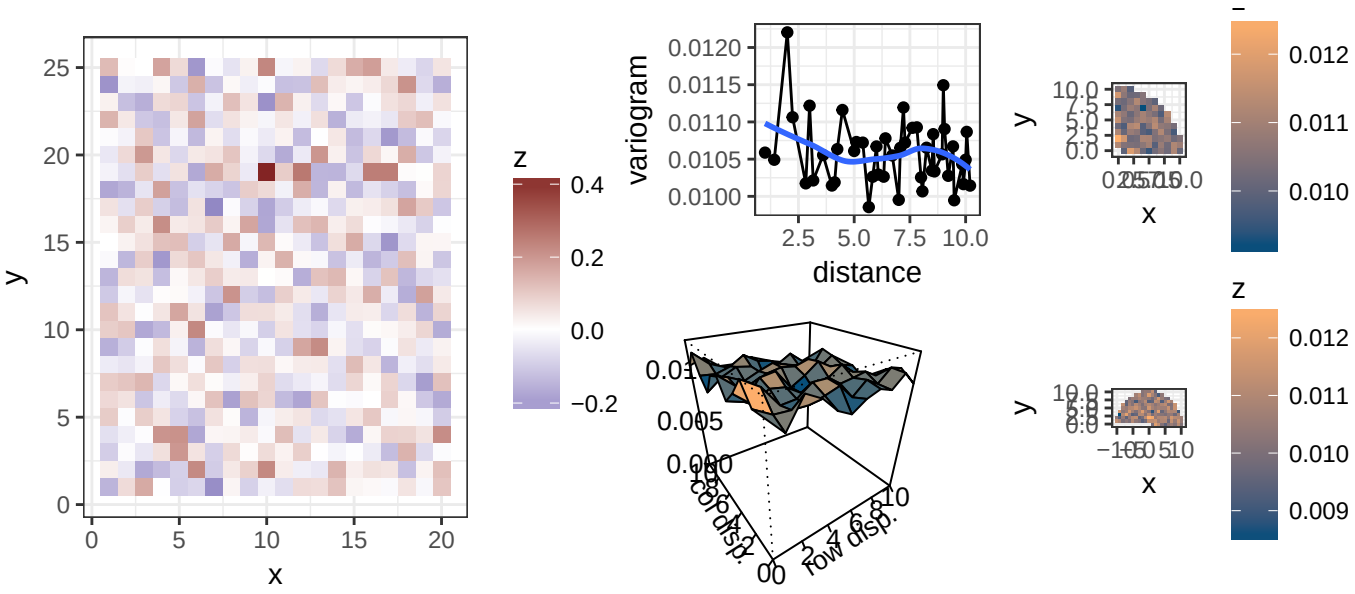
```
      Predicted = pred$direct),
    data.frame(
      Component = 'competition BV',
      True = dat$BV2,
      Predicted = pred$comp),
    data.frame(
      Component = 'pec',
      True      = dat$pec,
      Predicted = as.vector(pred$pec)))

ggplot(comp.pred,
       aes(True, Predicted)) +
  geom_point() +
  geom_abline(intercept = 0, slope = 1,
              col = 'darkgray') +
  facet_grid(~ Component)
```



The predition of the Permanent Environmental Competition effect is not precisely great...

Competition #2 | Map of residuals and their variogram

```
plot(res.comp, type = 'resid')
variogram(res.comp)
```



Generic component

The Generic model

This additional component allows to introduce a random effect ψ with **arbitrary** incidence and covariance matrices Z and Σ:

$$
\begin{aligned}
y =& \mu + X\beta + Z\psi + \varepsilon \\
\psi \sim& N(0, \sigma^2_\psi \Sigma_\psi) \\
\varepsilon \sim& N(0, \sigma^2_\varepsilon)
\end{aligned}
$$

Implementation of the generic component

```
## Fit a blocks effect using generic
inc.mat <- model.matrix(~ 0 + bl, globulus)
cov.mat <- diag(nlevels(globulus$bl))
res.blg <- remlf90(fixed  = phe_X ~ gg,
                   generic = list(block = list(inc.mat,
                                               cov.mat)),
                   data   = globulus)
```

Example of result

```
## Formula: phe_X ~ 0 + gg
##    Data: globulus
##   AIC  BIC logLik
##  5691 5701  -2844
##
## Parameters of special components:
##
```

```
##
## Variance components:
##          Estimated variances   S.E.
## block                  2.592 1.0640
## Residual              15.208 0.6825
##
## Fixed effects:
##        value   s.e.
## gg.1  13.534 0.6222
## gg.2  14.030 0.8464
## gg.3  16.106 0.5513
## gg.4  11.854 0.6824
## gg.5  15.883 0.5863
## gg.6  10.220 1.3041
## gg.7  13.995 1.0894
## gg.8  15.728 0.5410
## gg.9  16.478 0.5969
## gg.10 12.843 1.1225
## gg.11 16.744 0.6151
## gg.12 17.002 0.8464
## gg.13 16.297 1.0894
## gg.14 14.429 0.4730
```

Prediction

Predicting values for unobserved trees

- You can predict the Breeding Value of an **unmeasured tree**
- Or the expected phenotype of a death tree (or an hypothetical scenario)
- Information is gathered from the covariates, the spatial structure and the pedigree
- Simply **include the individual** in the dataset with the response set as `NA`

Leave-one-out cross-validation

- Re-fit the simulated competition data with one measurement removed
- Afterwards, compare the predicted values for the **unmeasured** individuals with their true simulated values

```
rm.idx <- 8
rm.exp <- with(dat[rm.idx, ],
               phenotype - resid)
dat.loo <- dat
dat.loo[rm.idx, 'phenotype'] <- NA
```

	True	Pred.loo
direct BV	-1.48	0.11
competition BV	0.46	0.36
exp. phenotype	6.80	9.90

Exercise | Cross validation

1. Extend the last table to include the predicted values with the full dataset
2. Remove 1/10th of the phenotypes randomly, and predict their expected phenotype
   - Have the parameter estimations changed too much?
3. Compute the Root Mean Square Error (RMSE) of Prediction with respect to the true values

Cross-validation #1 | Include prediction with full data

```
pred.BV.mat <- with(ranef(res.comp),
                    cbind(`genetic_direct`, `genetic_competition`))

valid.pred$Pred.full <- c(Zd[rm.idx, ] %*% pred.BV.mat,
                          fitted(res.comp)[rm.idx])
```

	True	Pred.full	Pred.loo
direct BV	-1.48	-1.30	0.11
competition BV	0.46	0.99	0.36
exp. phenotype	6.80	7.29	9.90

Cross-validation #2 | Perform cross-validation on 1/10th of the observations

```
rm.idx <- sample(nrow(dat), nrow(dat)/10)
dat.cv <- dat
dat.cv[rm.idx, 'phenotype'] <- NA
## Re-fit the model and build table
```

	Fully.estimated	CV.estimated
direct	2.50	2.69
compet.	1.03	0.86
correl.	-0.76	-0.80
spatial	1.24	1.03
pec	0.20	0.51
residual	0.17	0.14

Cross-validation #3 | MSE of Prediction

```
true.exp.cv <- with(dat[rm.idx, ], phenotype - resid)
round(sqrt(mean((fitted(res.comp.cv)[rm.idx] - true.exp.cv)^2)), 2)
```

```
## [1] 1.5
```

Multiple traits

breedR provides a basic interface for multi-trait models which only requires specifying the different traits in the main formula using `cbind()`.

```
## Filter site and select relevant variables
dat <-
  droplevels(
    douglas[douglas$site == "s3",
            names(douglas)[!grepl("H0[^4]|AN|BR|site", names(douglas))]]
  )

res <-
  remlf90(
    fixed = cbind(H04, C13) ~ orig,
    # random = ~ block,
    genetic = list(
      model = 'add_animal',
      pedigree = dat[, 1:3],
      id = 'self'),
    data = dat
  )
```

```
## Warning in build_pedigree(1:3, data = ped.df): The pedigree has been
## recoded. Check attr(ped, 'map').
```

```
## Using default initial variances given by default_initial_variance()
## See ?breedR.getOption.
```

A full covariance matrix across traits is estimated for each random effect, and all results, including heritabilities, are expressed effect-wise:

```
## Formula: cbind(H04, C13) ~ 0 + orig + pedigree
##    Data: dat
##    AIC   BIC logLik
##  30968 31010 -15476
##
## Parameters of special components:
##
##
## Variance components:
##                                        Estimated variances   S.E.
## genetic.direct.H04                                   918.1  438.6
## genetic.direct.H04_genetic.direct.C13               1872.4  824.0
## genetic.direct.C13                                  5827.6 1829.6
## Residual.H04                                        8373.7  461.7
## Residual.H04_Residual.C13                          10922.0  755.3
## Residual.C13                                       18439.0 1484.2
##
##                   Estimate    S.E.
## Heritability:H04    0.0990 0.04589
## Heritability:C13    0.2391 0.07036
##
## Fixed effects:
##              value    s.e.
## orig.H04.pA 352.00  6.2389
```

```
## orig.H04.pB 370.90 10.7947
## orig.H04.pC 346.93 13.0788
## orig.H04.pF 339.66  6.2268
## orig.H04.pG 313.00 24.0430
## orig.H04.pH 305.39 19.9334
## orig.H04.pI 323.29 20.0946
## orig.H04.pJ 343.87 19.8567
## orig.H04.pK 335.48 19.6409
## orig.C13.pA 460.01 13.6444
## orig.C13.pB 494.58 19.8635
## orig.C13.pC 430.86 25.5477
## orig.C13.pF 429.48 12.5501
## orig.C13.pG 376.42 48.3133
## orig.C13.pH 376.98 43.4266
## orig.C13.pI 404.62 43.6194
## orig.C13.pJ 418.91 43.2856
## orig.C13.pK 441.99 43.0567
```

Although the results are summarized in tabular form, the covariance matrices can be recovered directly:

```
res$var[["genetic", "Estimated variances"]]
```

```
##            direct.H04 direct.C13
## direct.H04     918.08     1872.4
## direct.C13    1872.40     5827.6
```

```
## Use cov2cor() to compute correlations
cov2cor(res$var[["genetic", "Estimated variances"]])
```

```
##            direct.H04 direct.C13
## direct.H04  1.0000000  0.8094938
## direct.C13  0.8094938  1.0000000
```

Estimates of fixed effects and BLUPs of random effects can be recovered with `fixef()` and `ranef()` as usual. The only difference is that they will return a list of matrices rather than vectors, with one column per trait.

The standard errors are given as attributes, and are displayed in tabular form whenever the object is printed.

```
fixef(res)       ## printed in tabular form, but...
```

```
## $orig
##    value.H04 value.C13  s.e..H04 s.e..C13
## pA  352.0025  460.0097  6.238914 13.64437
## pB  370.8997  494.5846 10.794693 19.86351
## pC  346.9318  430.8644 13.078774 25.54773
## pF  339.6614  429.4795  6.226796 12.55013
## pG  313.0000  376.4231 24.043034 48.31334
## pH  305.3889  376.9779 19.933367 43.42664
## pI  323.2885  404.6216 20.094619 43.61939
## pJ  343.8727  418.9064 19.856683 43.28562
## pK  335.4828  441.9861 19.640911 43.05671
```

```
unclass(fixef(res))  ## actually a matrix of estimates with attribute "se"
```

```
## $orig
##         H04      C13
## pA 352.0025 460.0097
## pB 370.8997 494.5846
```

```
## pC 346.9318 430.8644
## pF 339.6614 429.4795
## pG 313.0000 376.4231
## pH 305.3889 376.9779
## pI 323.2885 404.6216
## pJ 343.8727 418.9064
## pK 335.4828 441.9861
## attr(,"se")
##          H04      C13
## pA  6.238914 13.64437
## pB 10.794693 19.86351
## pC 13.078774 25.54773
## pF  6.226796 12.55013
## pG 24.043034 48.31334
## pH 19.933367 43.42664
## pI 20.094619 43.61939
## pJ 19.856683 43.28562
## pK 19.640911 43.05671
```

```
str(ranef(res))
```

```
## List of 1
##  $ genetic: num [1:1525, 1:2] -6.02 -12.93 -10.16 33.51 6.77 ...
##   ..- attr(*, "dimnames")=List of 2
##   .. ..$ : chr [1:1525] "19" "21" "23" "25" ...
##   .. ..$ : chr [1:2] "H04" "C13"
##   ..- attr(*, "se")= num [1:1525, 1:2] 23 22.8 23.6 23 23.6 ...
##   .. ..- attr(*, "dimnames")=List of 2
##   .. .. ..$ : chr [1:1525] "19" "21" "23" "25" ...
##   .. .. ..$ : chr [1:2] "H04" "C13"
##   ..- attr(*, "names")= chr [1:3050] "1" "2" "3" "4" ...
##  - attr(*, "class")= chr [1:2] "ranef.breedR" "breedR_estimates"
```

```
head(ranef(res)$genetic)
```

```
##           H04         C13
## 19  -6.016271   40.093547
## 21 -12.925035 -108.673107
## 23 -10.164449   23.276658
## 25  33.507715   80.855347
## 27   6.768289   -5.018311
## 29  22.201575   32.078520
```

Recovering the breeding values for each observation in the original dataset follows the same procedure as for one trait: multiply the incidence matrix by the BLUP matrix. The result, however, will be a matrix with one column per trait.

```
head(model.matrix(res)$genetic %*% ranef(res)$genetic)
```

```
##           H04        C13
## 151  5.923689  -6.612036
## 153  7.760706  22.486000
## 155 -7.414378 -38.978615
## 157  7.894009   3.756494
## 159  3.536361 -10.654445
## 161 12.431919  12.736590
```

Some more features

Metagene interface

- We have used simulated data from the `metagene` software
- If you simulate data, import the results with `read.metagene()`
- Use several common methods with a `metagene` object:
  - `summary()`, `plot()`, `as.data.frame()`
- Plus some more specific `metagene` functions:
  - `b.values()`, `get.ntraits()`, `ngenerations()`, `nindividuals()`, `get.pedigree()`
- And specific functions about spatial arrangement:
  - `coordinates()` extract coordinates
  - `sim.spatial()` simulates some spatial autocorrelation

Simulation framework

- The function `breedR.sample.phenotype()` simulates datasets from all the model structures available in **breedR**
- Limitation: only one generation, with random matings of founders
- See `?simulation` for details

Remote computation

If you have access to a **Linux** server through **SSH**, you can perform computations remotely

- Take advantage of more **memory** or **faster** processors
- **Parallelize** jobs
- Free **local resources** while fitting models
- See `?remote` for details

Package options

- **breedR** features a list of configurable options
- Use `breedR.setOption(...)` for changing an option during the current sesion
- Set options permanently in the file `$HOME/.breedRrc`
- see `?breedR.option` for details

```
breedR.getOption()
```

```
## $ar.eval
## [1] -0.8 -0.2  0.2  0.8
##
## $breedR.bin
## [1] "/home/facu/Work/Proyectos/2013.T4F/bin/PROGSF90/linux/32bit"
##
```

```
## $splines.nok
## determine.n.knots
##
## $default.initial.variance
## default_initial_variance
##
## $col.seq
## [1] "#034E7B" "#FDAE6B"
##
## $col.div
## [1] "#3A3A98FF" "#832424FF"
##
## $cygwin
## [1] "C:/cygwin"
##
## $cygwin.home
## [1] "/home/facu"
##
## $ssh.auth.sock
## [1] "/tmp/ssh-auth-sock-facu"
##
## $remote.host
## [1] "elnuevo"
##
## $remote.user
## [1] "fmunoz"
##
## $remote.port
## [1] 22
##
## $remote.bin
## [1] "/usr/local/lib/R/site-library/breedR/bin/linux"
##
## $ssh.options
## [1] "-x -o BatchMode=yes -o TCPKeepAlive=yes -e none"
```