

Snakemake tutorial

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Introduction













What is snakemake?

• Workflow manager: builds reproducible, scalable data analysis pipelines.

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- Language: a Python-based DSL (Domain-Specific Language) with readable rule syntax.
- Scaling: runs on local, server, cluster, and cloud—no workflow changes needed.
- Software management: declare required envs/containers/conda, auto-deployed anywhere.
- Reporting: turns runs into portable, browser-based reports for easy sharing.
- Dependency/retries: tracks input → output dependencies using a DAG and supports selective re-runs.
- **Bioconda**: bioinformatics package channel.
- **PyPI**: installable via pip (Python); shows supported Python versions.
- Containers: official container support (e.g., Docker images/builds).





Useful website

Workflow catalog

- https://snakemake.github.io/snakemake-workflow-catalog/
- https://github.com/snakemake-workflows

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Plugin

- https://snakemake.github.io/snakemake-plugin-catalog/

















Pre-requirements

Git

https://git-scm.com/install/

Docker

- * Install Docker Desktop or Docker Engine (choose one).
- Docker desktop (for GUI OS. such as Windows, Mac, Linux Ubuntu GUI)
 https://www.docker.com/products/docker-desktop/
- Docker engine (for CLI OS. such as Linux OS)
 https://docs.docker.com/engine/install/



Use Conda

Download

wget https://repo.anaconda.com/miniconda/Miniconda3-latest-Linux-x86_64.sh

Run the command below and press Enter when prompted

Type "yes" when asked to set up conda init

/bin/bash Miniconda3-latest-Linux-x86_64.sh

Activate Conda

source ~/.bashrc

Create a snakemake environment for the snakemake tutorial

conda create -n snakemake_9.11.1 -c bioconda -c conda-forge snakemake=9.11.1

conda activate

conda activate snakemake_9.11.1

Install tutorial dependencis

conda install -c bioconda -c conda-forge bwa

conda install -c bioconda samtools





Download

The --privileged option is required for Singularity.

docker run -it -v [local_volume]:[docker_volume] --privileged shinejh0528/snakemake:9.11.1_for_tutorial /bin/bash

* For more information, please visit my docker hub https://hub.docker.com/r/shinejh0528/snakemake





Tutorial repository download

Repository download

git clone https://github.com/Shin-jongwhan/snakemake_tutorial.git

* For more information, please visit my git repository https://github.com/Shin-jongwhan/snakemake_tutorial















Genome download (hg38)

UCSC - download link

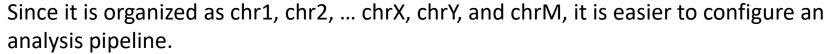
In this tutorial, we use this FASTA file.



<u>Name</u>	<u>Last modified</u>	<u>Size</u>	<u>Descript</u>
Parent Directory		_	
analysisSet/	2023-01-06 17:06	-	
est.fa.gz	2019-10-14 13:54	1.5G	
est.fa.gz.md5	2019-10-14 13:54	44	
genes/	2024-12-23 12:50	-	
<u>hg38.2bit</u>	2015-04-30 16:16	797M	
<u>hg38.agp.gz</u>	2014-01-15 20:55	842K	
<u>hg38.chrom.sizes</u>	2013-12-24 21:06	11K	
<u>hg38.chromAlias.bb</u>	2024-04-08 20:13	243K	
<u>hg38.chromAlias.txt</u>	2021-10-06 13:44	27K	
<u>hg38.chromFa.tar.gz</u>	2014-01-23 17:18	938M	
<u>hg38.chromFaMasked.tar.gz</u>	2014-01-23 17:10	487M	
<u>hg38.fa.align.gz</u>	2014-01-08 23:43	2.4G	
<u>hg38.fa.gz</u>	2014-01-15 21:14	938M	
<u>hg38.fa.masked.gz</u>	2014-01-15 21:24	487M	
<u>hg38.fa.out.gz</u>	2014-01-15 20:56	172M	
LION IIFNIII L	0010 10 04 01-00	1 00	



Genome download (hg38)





```
cat hg38.fa | grep ">"
chrl
chr10
chrl1
chrll KI270721vl random
chr12
chr13
chr14
chr14 GL000009v2 random
chrl4 GL000225vl random
chrl4 KI270722vl random
chrl4 GL000194vl random
chrl4 KI270723vl random
chrl4 KI270724vl random
chrl4 KI270725vl random
chrl4 KI270726vl random
chr15
chrl5 KI270727vl random
chr16
chrl6 KI270728vl random
```





Leave only chr1 sequence to test and bwa indexing

```
# Using seqtk
echo "chr1" > list.txt
seqtk subseq hg38.fa list.txt | seqtk seq -l 50 > hg38_chr1.fa

# fasta index
bwa index hg38_chr1.fa
```





Download Human example data (quick start)

hg38_chr1.fa

https://drive.google.com/file/d/1-oIZNcatSH9gT5iHFhxm0Yy4fP34CnC7/view?usp=sharing

hg38_chr.fa.idx.tar.gz (in the same directory as hg38_chr1.fa)

https://drive.google.com/file/d/1quRANcCjdxHGoKAfsPB3pkZkrOlozrw /view?usp=sharing

* Use this index data after decompressing with the command tar -xzf hg38_chr1.fa.idx.tar.gz

test fastq 0.25 M reads

R1: https://drive.google.com/file/d/1AqONWh9GJZKQvs0ls6ltny4fLqapv0eE/view?usp=sharing

R2: https://drive.google.com/file/d/1J4P5IZA8RpRK8LGATbquv52HUGQLHFf6/view?usp=sharing















file name: 'Snakefile' (default)





file name: 'Snakefile' (default)

You can execute snakemake in Snakefile folder by default Or you can specify Snakefile path by -s option

Option

```
-n: dry-run
```

-p: print command

--jobs [int] : number of parallel jobs (or use --cores)

--cores [int] : number of threads (or use --jobs)

-s [Snakefile_path] : specify Snakefile path

Last arguments [path]: Specify the output path as an absolute path. This ensures that Snakemake constructs the dependency graph (DAG) correctly and executes all preceding rules in order to generate the specified output.

```
# Activate conda
conda activate snakemake_9.11.1

# Or using docker
docker run -it -v [local_volume]:[docker_volume] shinejh0528/snakemake:9.11.1 /bin/bash

# Run pipeline In Snakefile path
snakemake --jobs 10 -np /biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.bam

# Or specify Snakefile path
snakemake --jobs 10 \
-s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/1_mapping/Snakefile -p \
/ biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.bam
```





Running Pipeline

```
(snakemake 9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake totorial/test data
snakemake -s /TBI/People/tbi/jhshin/pipeline/snakemake tutorial/basic/l mapping/Snakefile -np /biarchive/project/jh
shin/test/snakemake_totorial/test_data/test.bam
ost: husky
Building DAG of jobs...
ob stats:
Wed Sep 17 16:28:06 2025]
 snakemake totorial/test data/test 2.fastq.gz
hell command:
               bwa mem /biarchive/project/jhshin/test/snakemake totorial/test data/test l.fastg.gz /biarchive/proje
 jhshin/test/snakemake totorial/test data/test 2.fastq.gz
                                                                        | samtools view -Sb -
                                                                                                        > /biarchive
 roject/jhshin/test/snakemake_totorial/test_data/test.bam
   (check individual jobs above for details)
   output files have to be generated:
 is was a dry-run (flag -n). The order of jobs does not reflect the order of execution.
```

```
(snakemake 9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake totorial/test data
snakemake --jobs 10 -s /TBI/People/tbi/jhshin/pipeline/snakemake tutorial/basic/l mapping/Snakefile -p /biarchive/
oject/jhshin/test/snakemake_totorial/test_data/test.bam
 ssuming unrestricted shared filesystem usage.
ost: husky
uilding DAG of jobs...
sing shell: /bin/bash
 ales claiming more threads will be scaled down.
 b stats:
Execute 1 jobs...
[Fri Sep 26 13:40:27 2025]
 calrule bwa map:
 ell command:
               bwa mem -t 8 /biarchive/project/jhshin/reference/hg38/ucsc/hg38 chrl.fa /biarchive/project/jhshin/te
 snakemake totorial/test data/test l.fastq.gz/biarchive/project/jhshin/test/snakemake totorial/test data/test 2.fa/
q.gz
               | samtools view -Sb -
                                               > /biarchive/project/jhshin/test/snakemake totorial/test data/test.b
 Fri Sep 26 13:41:24 2025]
 mplete log(s): /biarchive/project/jhshin/test/snakemake totorial/test data/.snakemake/log/2025-09-26T134027.015432
 nakemake.log
```

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Dry-run Real run



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Output





2. Mapping with wildcard

Snakefile

The **{sample}** wildcard can be used in other arguments.





2. Mapping with wildcard

Running Pipeline

```
snakemake --jobs 10 \
-s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/2_mapping_with_wildcard/Snakefile \
-p \
/biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.bam
```

The output is same as '1. Create single rule - mapping'





3. Sorting read alignments

Snakemakefile

```
rule samtools_sort:
input:
bam = "/biarchive/project/jhshin/test/snakemake_tutorial/test_data/{sample}.bam"
output:
bam = "/biarchive/project/jhshin/test/snakemake_tutorial/test_data/{sample}.sorted.bam"
params:
thread = 8
shell:
"samtools sort -@ {params.thread} -T {wildcards.sample} "
"-O bam {input.bam} > {output.bam}"
```

Shell cmd Options

- T [str]

When sorting, Samtools splits the input BAM into **temporary files and then merges** them into the final output BAM. The –T option specifies the prefix for these temporary files.

Example: -T test \rightarrow creates temporary files like test.0000.bam, test.0001.bam during sorting.

-@ [int]
specify thread





3. Sorting read alignments

Running Pipeline

```
snakemake --jobs 10 \
-s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/3_sorting_read_alignments/Snakefile \
-p \
/biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.sorted.bam
```

```
snakemake 9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake totorial/test data
 snakemake --jobs 10 -s /TBI/People/tbi/jhshin/pipeline/snakemake tutorial/basic/3 sorting read alignments/Snakefile
-p /biarchive/project/jhshin/test/snakemake totorial/test data/test.sorted.bam
ssuming unrestricted shared filesystem usage.
ost: husky
Building DAG of jobs...
Jsing shell: /bin/bash
ules claiming more threads will be scaled down.
ob stats:
                count
amtools sort
select jobs to execute...
Execute 1 jobs...
 ocalrule samtools sort:
 nell command: samtools sort -@ 8 -T test -O bam /biarchive/project/jhshin/test/snakemake totorial/test data/test.bam
 > /biarchive/project/jhshin/test/snakemake totorial/test data/test.sorted.bam
 omplete log(s): /biarchive/project/jhshin/test/snakemake totorial/test data/.snakemake/log/2025-09-26T144510.465119
```



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3. Sorting read alignments

Output





4. Indexing bam

Snakefile

```
rule samtools_index:
input:
bam = "/biarchive/project/jhshin/test/snakemake_tutorial/test_data/{sample}.sorted.bam"
output:
"/biarchive/project/jhshin/test/snakemake_tutorial/test_data/{sample}.sorted.bam.bai"
shell:
"samtools index {input.bam}"
```

Running pipeline

```
snakemake --jobs 10 \
-s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/4_indexing_bam/Snakefile \
-p \
/biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.sorted.bam.bai
```





5. Let's combine everything

Snakefile

You can use Python syntax!

```
sAnalysis_dir = "/biarchive/project/jhshin/test/snakemake_tutorial/test_data/"

rule bwa_map:
    input:
        fastq_1 = sAnalysis_dir + "{sample}_1.fastq.gz",
        fastq_2 = sAnalysis_dir + "{sample}_2.fastq.gz"

    output:
        bam = sAnalysis_dir + "result/mapping/{sample}/{sample}.bam"
    params:
        genome_fa = "/biarchive/project/jhshin/reference/hg38/ucsc/hg38_chr1.fa",
        thread = 8

shell:
    """

bwa mem -t {params.thread} {params.genome_fa} {input.fastq_1} {input.fastq_2} \
        | samtools view -Sb - \
        > {output.bam}
    """
```

```
rule samtools sort:
 input:
   bam = sAnalysis_dir + "result/mapping/{sample}/{sample}.bam"
 output:
   bam = sAnalysis dir + "result/mapping/{sample}/{sample}.sorted.bam"
 params:
   thread = 8
 shell:
    "samtools sort -@ {params.thread} -T {wildcards.sample} "
   "-O bam {input.bam} > {output.bam}"
rule samtools index:
 input:
   bam = sAnalysis dir + "result/mapping/{sample}/{sample}.sorted.bam"
 output:
   bam_bai = sAnalysis_dir + "result/mapping/{sample}/{sample}.sorted.bam.bai"
 shell:
    "samtools index {input.bam}"
```



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5. Let's combine everything

Running pipeline

```
snakemake --jobs 10 \
-s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/5_combine_1-4_rules/Snakefile \
-p \
/biarchive/project/jhshin/test/snakemake_tutorial/test_data/result/mapping/test/test.sorted.bam.bai
```

Output

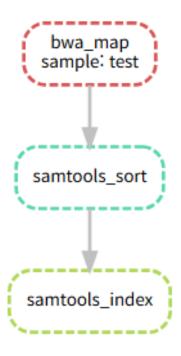




6. Generate DAG graph

Command

snakemake -s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/5_combine_1-4_rules/Snakefile \ /biarchive/project/jhshin/test/snakemake_tutorial/test_data/result/mapping/test/test.sorted.bam.bai \ --dag | dot -Tsvg > dag.svg





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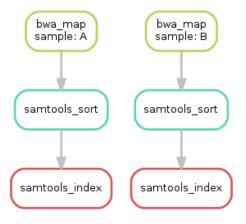


6. Generate DAG graph

You can generate a DAG graph for each sample using {sample_1,sample_2,...}.



we create a **visualization of the DAG** using the dot command provided by <u>Graphviz</u>. For the given target files, Snakemake specifies the DAG in the dot language and pipes it into the dot command, which renders the definition into <u>SVG format</u>. The rendered DAG is piped into the file dag.svg and will look similar to this:















In this tutorial, we see

- 1. How to record benchmark
- 2. How to record log
- 3. How to use container





The Snakefile from "Basic tutorial – 2. Mapping with wildcard" section, with additional benchmark, log, and container directives.

```
rule bwa map:
 input:
   genome_fa = "/biarchive/project/jhshin/reference/hg38/ucsc/hg38_chr1.fa",
   fastq 1 = "/biarchive/project/jhshin/test/snakemake tutorial/test data/{sample} 1.fastq.gz",
   fastq 2 = "/biarchive/project/jhshin/test/snakemake tutorial/test data/{sample} 2.fastq.gz"
  output:
   bam = "/biarchive/project/jhshin/test/snakemake tutorial/test data/{sample}.bam"
  params:
   thread = 8
  log:
   stdout = "/biarchive/project/jhshin/test/snakemake tutorial/test data/log/bwa map/{sample}.stdout.txt",
   stderr = "/biarchive/project/jhshin/test/snakemake tutorial/test data/log/bwa map/{sample}.stderr.txt"
  benchmark:
    "/biarchive/project/jhshin/test/snakemake tutorial/test data/benchmark/bwa map/{sample}.txt"
  container:
    "docker://shinejh0528/bwa-mem2:1.2.0-samtools"
  shell:
    bwa mem -t {params.thread} {input.genome fa} {input.fastq 1} {input.fastq 2} 2> {log.stderr} \
    | samtools view -Sb - \
   > {output.bam} \
   2> {log.stderr}
```





Running pipeline

To run Snakemake with a container, use the options --use-singularity and --singularity-args.

* --singularity-args "--bind /biarchive": **specifies a mount point** in Singularity so that the /biarchive path is accessible inside the container.

```
snakemake --jobs 10 \
    --use-singularity \
    --singularity-args "--bind /biarchive" \
    -s /TBI/People/tbi/jhshin/pipeline/snakemake_tutorial/basic/6_mapping_with_wildcard_with_other_opt/Snakefile \
    -p \
    /biarchive/project/jhshin/test/snakemake_tutorial/test_data/test.bam
```





Running pipeline

- 1. First, pull the container image from Docker Hub as a Singularity image.
- 2. Then, the rules run inside the container image.

```
(snakemake 9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake totorial/test data
 snakemake --jobs 10 --use-singularity --singularity-args "--bind /biarchive" -s /TBI/People/tbi/jhshin/pipeline/sna
kemake tutorial/basic/6 mapping with wildcard with other opt/Snakefile /biarchive/project/jhshin/test/snakemake totor
ial/test data/test.bam -p
Assuming unrestricted shared filesystem usage.
host: husky
Building DAG of jobs...
Pulling singularity image docker://shinejh0528/bwa-mem2:1.2.0-samtools.
Using shell: /bin/bash
Provided cores: 10
Rules claiming more threads will be scaled down.
Job stats:
           count
owa map
total
Select jobs to execute...
Execute 1 jobs...
```





Log

You can view the log files of each rule and sample.

```
snakemake 9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake totorial/test data/log/bwa map
 cat test.stderr.txt
[M::bwa idx load from disk] read 0 ALT contigs
[M::process] read 500000 sequences (75500000 bp)...
[M::mem pestat] # candidate unique pairs for (FF, FR, RF, RR): (1, 18746, 16, 2)
[M::mem pestat] skip orientation FF as there are not enough pairs
[M::mem pestat] analyzing insert size distribution for orientation FR...
[M::mem pestat] (25, 50, 75) percentile: (318, 367, 418)
[M::mem pestat] low and high boundaries for computing mean and std.dev: (118, 618)
[M::mem pestat] mean and std.dev: (370.05, 76.57)
[M::mem pestat] low and high boundaries for proper pairs: (18, 718)
[M::mem pestat] analyzing insert size distribution for orientation RF...
[M::mem pestat] (25, 50, 75) percentile: (1153, 1920, 9747)
[M::mem pestat] low and high boundaries for computing mean and std.dev: (1, 26935)
[M::mem pestat] mean and std.dev: (4812.56, 4316.80)
[M::mem pestat] low and high boundaries for proper pairs: (1, 35529)
[M::mem pestat] skip orientation RR as there are not enough pairs
[M::mem pestat] skip orientation RF
[M::mem process seqs] Processed 500000 reads in 391.234 CPU sec, 49.555 real sec
[main] Version: 0.7.18-r1243-dirty
[main] CMD: bwa mem -t 8 /biarchive/project/jhshin/reference/hg38/ucsc/hg38 chrl.fa /biarchive/project/jhshin/test/sn
akemake totorial/test data/test l.fastq.gz /biarchive/project/jhshin/test/snakemake totorial/test data/test 2.fastq.g
[main] Real time: 53.727 sec; CPU: 392.737 sec
```





Benchmark

In the benchmark data, you can check CPU, memory, and disk usage.

```
(snakemake_9.11.1) jhshin@husky:/biarchive/project/jhshin/test/snakemake_totorial/test_data/benchmark/bwa_map

$ cat test.txt

s h:m:s max_rss max_vms max_uss max_pss io_in io_out mean_load cpu_time

54.0623 0:00:54 1852.50 2355.79 1847.91 1849.98 74.80 0.00 641.76 347.16
```



Introducing my template









Template

Template URL

https://github.com/Shin-jongwhan/snakemake_template/tree/main/impertoire/1.0.0

Features

- 1. Define configuration (config).
- 2. Organize Docker containers.
- 3. List samples in samples.tsv.
- 4. In workflow/rules/common.smk, define configuration, outputs, and options, then import each sub_rule.smk.













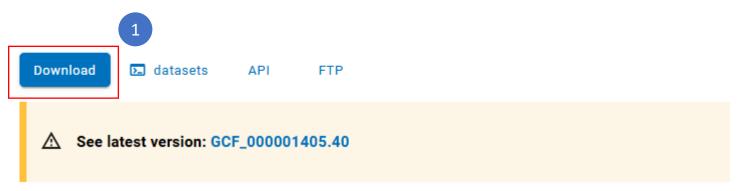


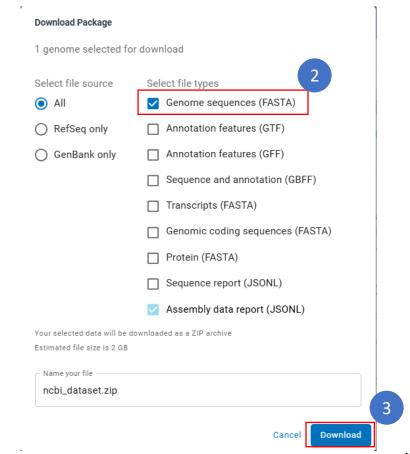
Genome download (hg38)

NCBI - download link

GCF_000001405.26 is GRCh38.p13 (Genome Reference Consortium Human Build 38, patch 13), maintained by NCBI RefSeq.

Genome assembly GRCh38





National Library of Medicine

National Center for Biotechnology Information



Genome download (hg38)

Since it is provided as an assembly, it may be difficult to set up an analysis pipeline.



```
$ cat GCF_000001405.26_GRCh38_genomic.fna | grep ">"

NC_000001.11 Homo sapiens chromosome 1, GRCh38 Primary Assembly

NT_187361.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG1_UNLOCALIZED

NT_187362.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG2_UNLOCALIZED

NT_187363.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG3_UNLOCALIZED

NT_187364.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG4_UNLOCALIZED

NT_187365.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG5_UNLOCALIZED

NT_187366.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG6_UNLOCALIZED

NT_187367.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG7_UNLOCALIZED

NT_187368.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG8_UNLOCALIZED

NT_187369.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG9_UNLOCALIZED

NT_187369.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG9_UNLOCALIZED

NT_187369.1 Homo sapiens chromosome 1 unlocalized genomic scaffold, GRCh38 Primary Assembly HSCHR1_CTG9_UNLOCALIZED

NC_000002.12 Homo sapiens chromosome 2, GRCh38 Primary Assembly
```





Thank you!:)

jonghwan.shin@theragenbio.com







