

Review article

Gastrointestinal microbiota-directed nutritional and therapeutic interventions for inflammatory bowel disease: opportunities and challenges

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Abstract

Evidence-based research has confirmed the role of gastrointestinal microbiota in regulating intestinal inflammation. These data have generated interest in developing microbiota-based therapies for the prevention and management of inflammatory bowel disease (IBD). Despite in-depth understanding of the etiology of IBD, it currently lacks a cure and requires ongoing management. Accumulating data suggest that an aberrant gastrointestinal microbiome, often referred to as dysbiosis, is a significant environmental instigator of IBD. Novel microbiome-targeted interventions including prebiotics, probiotics, fecal microbiota transplant, and small molecule microbiome modulators are being evaluated as therapeutic interventions to attenuate intestinal inflammation by restoring a healthy microbiota composition and function. In this review, the effectiveness and challenges of microbiome-centered interventions that have the potential to alleviate intestinal inflammation and improve clinical outcomes of IBD are explored.

Keywords: dietary fiber; dysbiosis; fecal microbiota transplant; microbiota restoration therapy; intestinal inflammation; microbial metabolism

Introduction

Inflammatory bowel disease (IBD) is a chronic inflammatory condition of the gastrointestinal (GI) tract primarily resulting from a dysregulated host immune response to environmental factors, including atypical GI microbiota [1, 2]. Crohn's disease (CD) and ulcerative colitis (UC) are the two most common types of IBD. Patients with CD exhibit discontinuous (patchy), transmural inflammation, affecting multiple layers of the intestinal wall. In contrast, UC is characterized by continuous inflammation, typically limited to the large intestine (colon and rectum). Additionally, inflammation in UC is confined to the inner mucosal lining. Both CD and UC patients tend to have a GI microbiota that is less diverse [3, 4] and exhibit reduced metabolic capacity [5, 6]. A decrease in the number of *Faecalibacterium* is considered to be the "hallmark" of IBD [3], while an increase in *Proteobacteria* and *Actinobacteria* is a characteristic of dysbiosis [7, 8]. Such alterations in the gut microbiome composition can impact microbial metabolome leading to the disruption of intestinal immune function and barrier integrity. However, whether such dysbiosis is an outcome of existing inflammation or has a causative role in IBD development is largely unknown [9] (Figure 1).

The interplay among commensal microbes, the intestinal epithelium, and resident immune cells shapes the composition and function of the gut microbiota and the intestinal immune

response [2]. A diet consistently high in ultra-processed, low-fiber foods may disrupt the delicate communication among gut bacteria, the intestinal lining, and immune cells, potentially leading to chronic inflammation, including IBD [10]. Dietary fibers present in fruits and vegetables play a constructive role in preserving the diversity of microbes within the intestines of individuals in good health [11]. Gut microbiota uses these dietary fibers as metabolic substrate and produces short-chain fatty acids (SCFAs) such as acetate, propionate, and butyrate through fermentation. These SCFAs play a pivotal role in governing the intestinal immune response through their interactions with immune pathways [12, 13]. Besides fiber fermentation metabolites, host molecules including bile acids also play a role in host immunity and microbiota composition. Dysregulation of microbial bile acid metabolism is known to contribute to IBD development [14] and the growth of *Clostridioides difficile*—an opportunistic pathogen that infects the large intestine [15]. The role of the SCFAs and bile acids in regulating intestinal immune response through alterations to the gut microbiota will be discussed further within this review.

The role of the dysbiotic gut microbiome in contributing to the adverse immune response found in IBD has led to interest in strategies that either favorably alter the microbiome composition and/or restore the balanced gut microbiome. Prebiotics,

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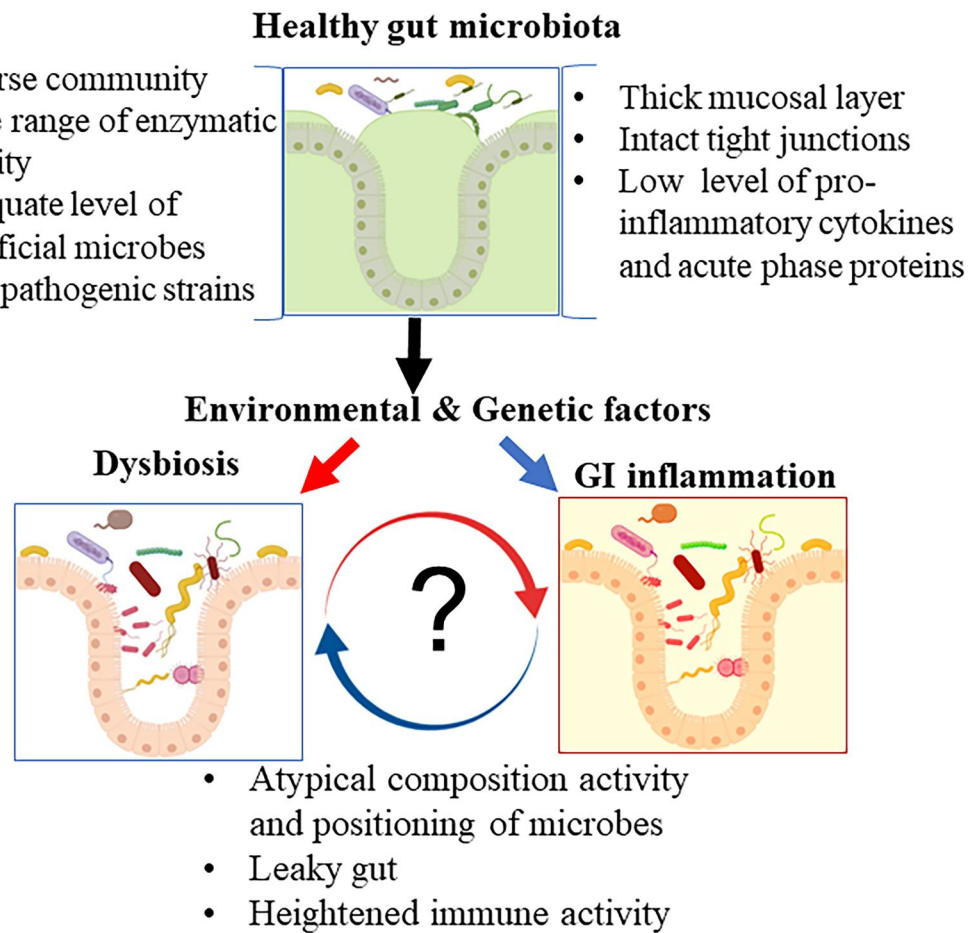


Figure 1. A schematic diagram showing that atypical gut microbiota (i.e. dysbiosis—a relative difference in both composition and metabolic activity of gut microbiota) could be a cause or consequence—or both—of GI inflammation. The right arrow represents GI inflammation that induces dysbiosis. The left arrow represents dysbiosis that elicits GI inflammation. The figure was illustrated with the help of BioRender software (accessed in 2021). GI = gastrointestinal.

Box 1. Glossary

Prebiotics: gut microbiota-accessible dietary fibers that nourish and promote the growth of beneficial bacteria.

Probiotics: beneficial live microorganisms found in fermented foods and supplements.

Synbiotics: a combination of prebiotics and probiotics that attempt to maximize the potential benefits of both prebiotics and probiotics.

Paraprobiotics: inactive (dead) bacterial cells or bacterial fragments that exhibit therapeutic effects.

Postbiotics: metabolites produced by the bacteria that elicit beneficial properties.

probiotics, paraprobiotics, postbiotics, and synbiotics (Box 1) in dietary components or supplements have been studied for their potential benefit to individuals with IBD and will be discussed in the later parts of this article.

The present therapeutic strategies for the treatment of IBD are focused primarily on managing symptoms rather than a curative or preventive approach [16, 17]. IBD patients are typically treated with 5-aminosalicylate, corticosteroids, antibiotics, immunomodulators, and biologics [16]. Novel biological therapies, including monoclonal antibodies for tumor necrosis factor (TNF), are promising; however, a subgroup of IBD patients who are non-responsive, develop adverse reactions, and/or are at risk of developing secondary infections is further complicating the management of IBD [18]. Herein, there is a need to advance innovative treatment modalities characterized by diminished adverse

effects and enhanced clinical efficacy, with the goal of more effectively managing IBD. Preclinical and clinical studies suggest that an imbalanced GI microbiota disrupts the intestinal immune balance, contributing to the development of IBD over time [19, 20]. Therefore, ameliorating the pre-existing dysbiosis by dietary modification or restoring microbiota homeostasis could represent potential approaches. To understand the opportunities and limitations of using nutritional and therapeutic interventions to target microbiota for managing and potentially treating IBD, we conducted a review of relevant research published online. We searched databases such as PubMed, Web of Science, and Google Scholar, focusing on research articles, systematic reviews, and meta-analyses published after 2000. Our search included keywords such as IBD, UC, CD, nutrition, dietary fibers, gut microbiome, and microbiome-based therapeutic approaches for IBD.

Precisely, this review delves into previous research that considers the relative contribution of gut microbiota dysbiosis to the onset and exacerbation of IBD. It then navigates through various nutritional interventions, including prebiotics and probiotics, aimed at restoring microbial homeostasis and ameliorating inflammation in IBD patients. Furthermore, the review explores new therapeutic approaches, such as microbiota-restoration therapy and microbiota-targeted small molecules that beneficially modify host-microbe interactions, holding promise for reshaping IBD treatment approaches. Altogether, this review discusses ongoing microbiome-focused investigational therapies, their limitations, emerging opportunities, and challenges associated with developing microbiome-informed interventions for improving IBD.

Microbiota dysbiosis elicits the disease or disease induces dysbiosis: the causation conundrum in IBD

Several studies suggest that a dysbiotic GI microbiome plays a role in the development of IBD [21, 22]. However, whether an alteration in the composition and metabolic activity of the GI microbiota in IBD patients constitutes a cause or an outcome of the inflamed environment within the intestines remains to be established. The high variability between studies and the lack of a common microbe identified across all studies limit our ability to pinpoint a specific group of microbes that drives or dampens IBD pathogenesis. In a healthy host, the diverse microbial composition plays a symbiotic role in the digestion of dietary fibers, synthesis of specific vitamins, and a favorable modulation of intestinal immune responses [23]. Moreover, a diverse microbiome prevents the colonization of pathogenic bacteria. Environmental influences, such as prolonged usage of broad-spectrum antibiotics or a deficiency in fiber within the diet, disrupt symbiotic rapport between the host and the microbiota. Although it remains to be determined whether dysbiosis directly causes inflammation in IBD patients, preclinical studies using germ-free mice colonized with specific bacteria have identified strains capable of triggering intestinal inflammation. For instance, germ-free mice lacking interleukin-10 (IL-10 KO) developed mild inflammation in the cecum, while *Enterococcus faecalis* colonization led to distal colitis [24]. Interestingly, colonization with *Candida albicans*, *Lactobacillus casei*, *L. reuteri*, *L. acidophilus*, a *Bifidobacterium* sp., *Lactococcus lactis*, or a *Bacillus* sp. did not induce inflammation in any part of the GI tract of germ-free IL-10 KO mice [24]. These findings in mice suggest that IBD could result from the presence of specific bacterial species, although this remains to be verified in patients with IBD. A strong correlation exists between dysbiosis and IBD, though a causal relationship between specific bacterial species and inflammation remains unclear (Figure 1). Additional research is required to understand the complex relationship between genetics, GI microbial composition, and inflammation in the pathogenesis of IBD.

Limitations of present IBD therapies

Therapeutic strategies for IBD chiefly aim to manage the disease by reducing ongoing intestinal inflammation and preventing sustained inflammation-induced tissue damage and fibrosis. Principal anti-inflammatory and immunosuppressive drugs currently in use to manage IBD are 5-aminosalicylate (5-ASA), corticosteroids (such as glucocorticoids [GCs]), immunomodulators (e.g. methotrexate), and biologics (e.g. TNF antagonists) [25]. The main goals of these drugs are to achieve and maintain a state of

remission, prevent hospitalization, and improve the quality of life (QOL) to the best possible extent. For a subset of IBD patients with reduced response to potent agents such as biologics, dose escalation and increased frequency become necessary.

Corticosteroid therapy is an effective treatment in reducing disease activity and induction of remission in patients with IBD. However, the chronic use of GCs can produce central nervous system, musculoskeletal, metabolic, dermatologic, cardiovascular, and immunological adverse effects [26]. In addition, GCs have adverse GI effects such as indigestion and bleeding in the upper abdomen upon eating, which can exacerbate IBD [27]. Second-generation GCs, such as budesonide and beclomethasone dipropionate, primarily applied topically, represent a better alternative for systemic GCs. 5-ASA, a standard therapy used for mild to moderate UC, can produce acute and chronic adverse effects such as nausea, abdominal pain, and diarrhea [27]. Biological therapies that include TNF- α inhibitors, such as infliximab, adalimumab, golimumab, and certolizumab pegol, are being effectively used to suppress excessive intestinal inflammation for inducing and maintaining remission in patients with IBD [28]. However, treatment failures associated with anti-TNF therapy, including primary non-responsiveness to treatment or secondary loss of response to these drugs, can occur due to immune-mediated neutralizing antibodies [28–30]. The use of these biological therapies is associated with a higher incidence of specific types of opportunistic infections [8, 31], which underscores the crucial need for a new line of therapeutic agents and adjuvant therapies that deliver sustained remission and improve clinical outcomes of IBD. Beyond conventional treatments, alternative therapeutic approaches such as microbiome-targeted therapy hold promise in managing IBD symptoms and improving the QOL for IBD patients.

The effect of prebiotics, probiotics, and synbiotics in IBD

Microbiota-accessible carbohydrates such as fermentable dietary fibers (FDFs) play pivotal role in maintaining a symbiotic relationship between host and microbiota in the GI tract. Prebiotics are types of FDFs that selectively nourish beneficial gut bacteria. A reduced intake of naturally occurring FDFs favors the expansion of colonic mucus-degrading bacteria that compromises the GI barrier function and elicits colonic inflammation [32–34]. Bioactive products, such as amines, phenols, and sulfur compounds, derived from microbial protein fermentation, also increase the likelihood of intestinal inflammation [35]. In a seminal study [33], the authors observed that fiber deprivation significantly increases the growth of mucus-eroding microbiota and susceptibility to the intestinal pathogen *Citrobacter rodentium*. Supporting the notion that FDFs are beneficial for GI health, an intervention with psyllium fiber successfully attenuated dextran sulfate sodium (DSS)-induced experimental colitis [36]. Recently, Bretin et al. demonstrated that psyllium fiber elevates bile acids and engages the farnesoid X receptor to provide protection against colitis [37]. These empirical observations underscore the critical role of dietary fiber in regulating intestinal inflammation. While naturally occurring FDFs are known to benefit gut microbiota and intestinal health, supplementation with highly processed fibers has been shown to exacerbate colitis in preclinical models [38–41]. These detrimental effects, particularly of inulin-type fructans, have also been observed in a subgroup of IBD patients [42]. Along the same lines, research studies are exploring the efficacy of probiotics, live beneficial bacteria, in attenuating

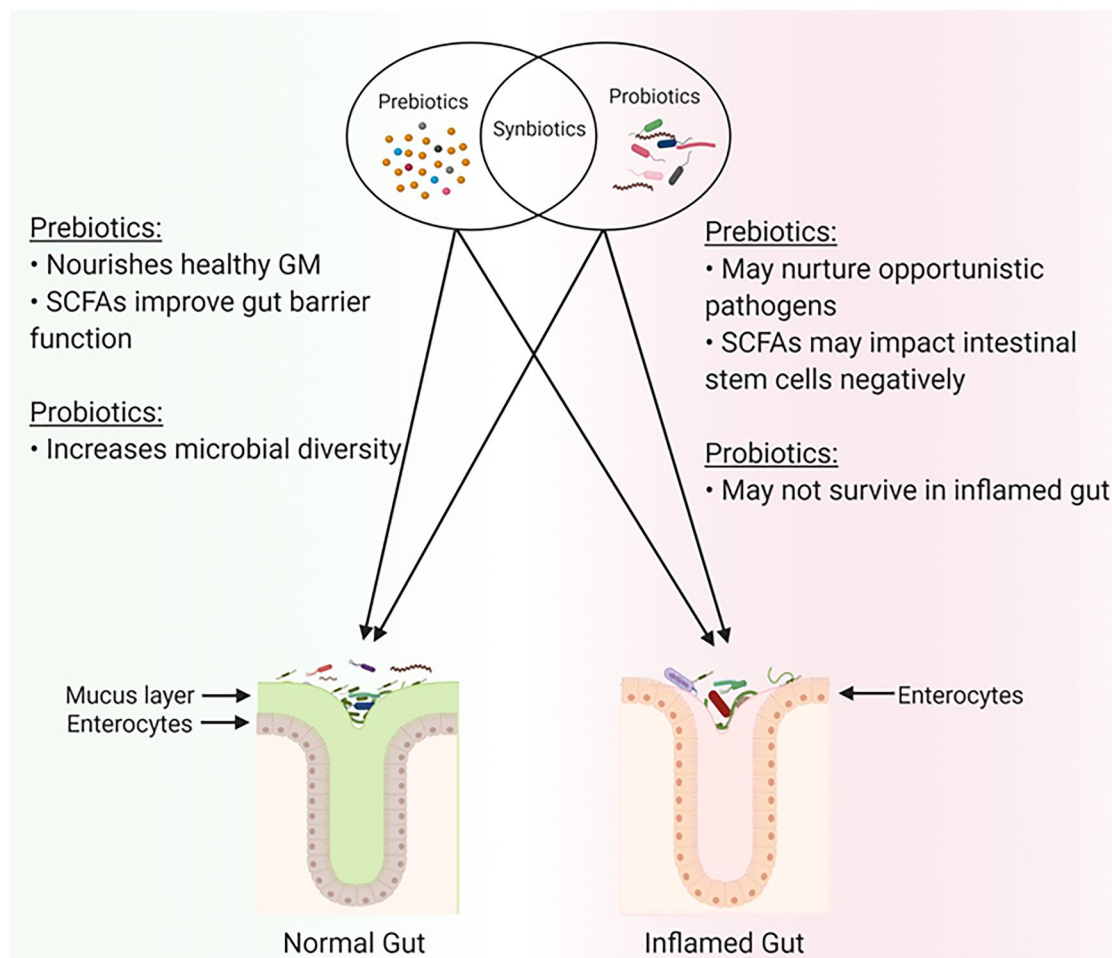


Figure 2. A schematic diagram showing the effect of prebiotics, probiotics, and synbiotics. In healthy individuals, the consumption of prebiotics, probiotics, and synbiotics has been reported to be beneficial in decreasing inflammation and protecting the GI tract from colonization by opportunistic pathogens. However, in IBD, the effect of consuming prebiotics, probiotics, and synbiotics may be detrimental and could further exacerbate inflammation, nurture opportunistic pathogens, and negatively impact intestinal progenitor cells. The figure was illustrated with the help of BioRender software (accessed in 2021). IBD = inflammatory bowel disease, GI = gastrointestinal, SCFAs = short-chain fatty acids.

intestinal inflammation; alleviating common IBD symptoms such as diarrhea, abdominal pain, and bloating; and ultimately improving the QOL of IBD patients. Synbiotics, which consist of a mixture of both prebiotics and probiotics, are becoming more popular to aid in the survival and growth of beneficial bacteria in the lower GI tract [43, 44]. Overall, dietary intervention(s) that can support (i.e. prebiotics), restore (ie probiotics), or both (i.e. synbiotics) may represent a viable strategy to maintain a healthy GI tract and mitigate intestinal inflammation [44] (Figure 2).

Prebiotics in IBD prevention: potentials and limitations

According to the International Scientific Association for Probiotics and Prebiotics (ISAPP), a prebiotic is “a substrate that is selectively utilized by host microorganisms conferring a health benefit” [45]. Specifically, prebiotics are classified as a substrate that must be resistant to the conditions of the GI tract, fermentable by the GI microbiota, and cannot be hydrolysed by the enzymes of the small intestine. Prebiotic consumption has been considered safe for healthy adults, demonstrating the ability to enhance microbial diversity and promote health. Nonetheless, it remained elusive whether prebiotics yield similar benefits for individuals with IBD [42].

The effects of prebiotics on colonic inflammation have been evaluated in experimental models of IBD. Fructan (inulin and fructooligosaccharides [FOS]), which is a linear chain of fructose with a β (2→1) linkage, and galacto-oligosaccharides (GOS), which is a chain of lactose molecules connected by β (1→6), β (1→3), and β (1→4) linkages [46], are among the most-evaluated prebiotics regarding their effects on intestinal inflammation. The daily co-administration of inulin and FOS increases the proliferation of *Lactobacillus* and *Bifidobacteria*, decreases mucosal pro-inflammatory cytokines IL-1 β and IFN- γ , and improves crypt damage in dextran DSS-fed HLA-B27 transgenic rats [47]. In a mouse model of DSS-induced UC, administration of low- and high-molecular-weight oat-derived beta-glucan decreased myeloperoxidase activity, malondialdehyde levels, and nitric oxide levels [48]. Similarly, intervention with purified pectin with various degrees of esterification significantly attenuated intestinal epithelial injury, inflammation, and oxidative stress in an experimental model of colitis [49]. Moreover, studies using germinated barley [50], psyllium fiber [37], and xylooligosaccharides [51], which are not primarily considered as prebiotic fibers, exhibited a protective response to intestinal inflammation.

Although significant evidence suggests that dietary fibers and their bacterial fermentation products benefit the management of IBD, it is crucial to consider these findings in the context of both

healthy and diseased (ongoing intestinal inflammation) states. Emerging data suggest that FDFs such as inulin may worsen symptoms in some patients with IBD [38, 39, 42]. This highlights the need for further research on the specific benefits of different fiber types in IBD. Our own work has revealed that the dietary fiber inulin exacerbates intestinal inflammation [38] and colitis-associated colon tumorigenesis [39], while structurally distinct fiber pectin offers protection against colitis, emphasizing the potential risks and benefits associated with distinct fiber types [38]. This observation finds support in other studies as well [40, 41]. The use of FDFs, such as arabinoxylans, β -glucans, β -fructans, and pectin shows promise in promoting beneficial gut microbes and metabolites in the distal gut. However, extensive research is needed to fully understand their effects in isolation or in combination, especially in the inflamed intestinal environment of IBD patients.

Probiotics: do they improve IBD?

The ISAPP defines probiotics as “live microorganisms that, when administered in adequate amounts, confer a health benefit on the host.” This definition encompasses only those microbial species proven to provide health advantages through rigorously controlled studies. Any live cultures of traditional fermented foods lacking evidence of efficacy are excluded [52]. Certain strains of *Lactobacillus* and *Bifidobacteria* are the most-used probiotic microbes [53]. A long-term study examining the effects of probiotics in patients with UC reported that patients receiving mesalazine (5-ASA) and a probiotic blend of *L. salivarius*, *L. acidophilus*, and *Bifidobacterium bifidus* had an improvement in their clinical symptoms, based on the Modified Mayo Disease Activity Index and Physician’s Global Assessment, compared with patients receiving mesalazine alone. The combined therapy group had a decrease in stool frequency and clinical improvement over the 24-month study [54]. Similarly, probiotic yogurt containing *L. rhamnosus* and *Limosilactobacillus reuteri* increased the T-reg cell count in IBD patients after 30 days of consumption [55]. In a double-blind, placebo-controlled trial in UC and CD patients, inflammation was decreased in UC but not in CD patients after consuming a multi-strain (*L. rhamnosus*, *L. plantarum*, *L. acidophilus*, *E. faecium*) probiotic [56]. Overall, the available data on the therapeutic efficacy of probiotics for improving clinical symptoms in IBD patients remains limited. It is conceivable that specific factors such as the extent of ongoing intestinal inflammation in IBD patients may influence treatment response. Recently, the utilization of genetically engineered bacteria and postbiotics has introduced a novel avenue for addressing intestinal inflammatory conditions [57–60]. These genetically modified probiotics hold the potential to exert a dual impact: they can help regulate intestinal dysbiosis while simultaneously releasing therapeutic compounds directly into the intestine. This approach circumvents the need for systemic drug administration and mitigates associated systemic side effects. Probiotics show promise in animal studies but their effectiveness in humans may be limited by stomach acids, bile salts, and active inflammation in patients with IBD [61].

Synbiotics: the promise of combined modulation of the GI microbiota in IBD

Synbiotics are designed to maximize the potential of both probiotics (live bacteria) and FDFs (nourishment for gut bacteria) [62]. A randomized-controlled trial [63] compared the effects of synbiotics, prebiotics, and probiotics on the QOL in patients with UC where participants received either *B. longum* (probiotic), psyllium fiber (an FDF), or a combination of both (synbiotic). The study

found the greatest improvement in QOL scores in the synbiotic group [63]. A systematic review and meta-analysis of the use of probiotics, prebiotics, and synbiotics in IBD found that synbiotics were associated with a significant improvement in remission rates of UC [64]. Still, identifying synergistic combinations of prebiotics and probiotics, where prebiotics enhance the benefits of added probiotics, remains a complex challenge. The future of synbiotics relies on a methodical formulation and design process. This involves the careful selection of synbiotics that harmonize and enhance each other’s effects, guided by their specific mechanisms of action and the intended health benefits [65].

Despite enormous promise and interest, driven primarily by preclinical studies, there are only a few limited, well-designed, randomized-controlled trials to support the concept that microbiome-targeted nutritional and probiotic approaches may mitigate clinical symptoms and cure IBD. In preclinical studies, it is evident that prebiotics, probiotics, and synbiotics can maintain a diverse and healthy GI microbiota and beneficially modulate intestinal immune activity [37, 66–68]. The efficacy of these treatments suggests that consuming fruits and vegetables containing fermentable fibers increases the QOL in IBD patients. However, large-scale randomized-controlled trials are needed to validate the potential protective efficacy of microbiome-centered dietary interventions.

Microbiome-targeted investigational therapies for IBD

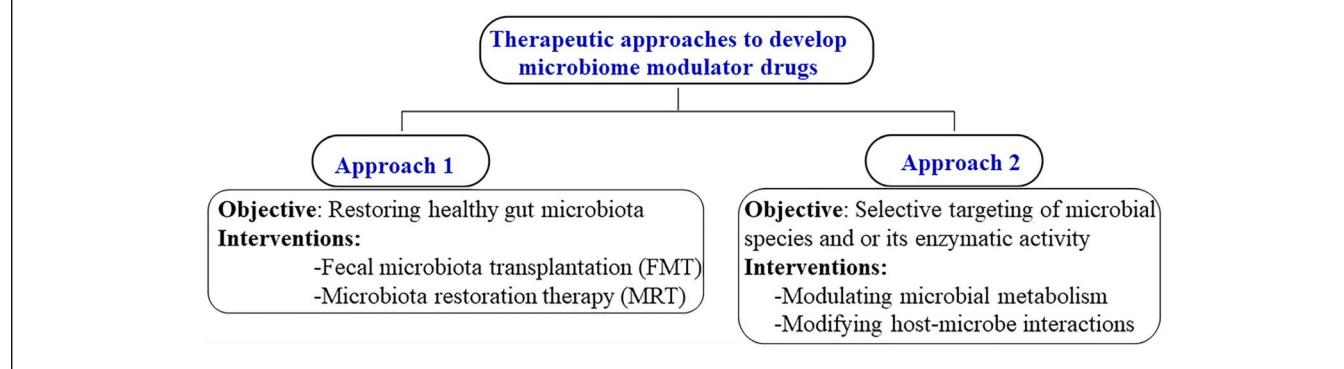
Microbiome-based therapeutic approaches

Commensal and symbiotic microbes co-evolved with the host and synthesize certain vitamins (K and B vitamins); biodegrade glycosaminoglycans; and produce SCFAs, secondary bile acids, and essential amino acids [69, 70]. The potential for gut microbiota and its metabolites to regulate intestinal immune homeostasis has fueled interest in therapeutic approaches targeting the restoration of healthy microbial composition and their metabolic activity (Box 2).

Restoring healthy microbiota

Fecal microbiota transplantation (FMT) involves the therapeutic infusion or engraftment of homogenized fecal suspension from healthy donors into the GI tract of patients suffering from IBD [71]. This approach has gained significant attention because of the successful implementation of FMT in treating *Clostridium difficile* infection [72, 73]. Despite the huge promise of FMT in limiting *C. difficile* infection, standardization among various studies, particularly regarding the route of administration, dose, volume, frequency, and method of preparation (i.e. fresh, frozen, single-dose, or pooled sample) of the fecal transplant, is required to compare the efficacy of various FMT trials appropriately [74, 75]. Analysing the effectiveness of FMT interventions becomes intricate when pooled samples are involved, as attributing outcomes to specific donors within the pool becomes difficult. Moreover, the risk of transferring opportunistic pathogens such as *Escherichia coli* increases with the number of donors in the pool.

Because there are many unknown factors associated with the efficacy and safety of FMT, more defined and targeted microbial populations are being investigated as therapeutic strategies to treat IBD. Microbiota restoration therapy is targeted to ameliorate the dysbiosis due to IBD by using defined microflora [76]. Data suggest that the gut bacteria of individuals with IBD differ in composition and metabolic capacity compared with those of healthy individuals. Relatively lower levels of *Faecalibacterium prausnitzii*, *Blautia faecis*, *Roseburia inulinivorans*, *Ruminococcus*

Box 2. Flow chart depicting the common approaches to developing GI microbiota-based therapeutic interventions**Table 1.** Microbiota restoration therapy studies

Therapeutic candidate	Conditions	Composition	Current phase	Mode of action	Clinical trial ID
SER-287 (Seres Therapeutics)	UC	Consists of a complex and diverse bacterial spore ecology	Phase 2b	Decreases immune activation	NCT03759041
VE202 (Vedanta Biosciences)	IBD	A defined consortium of live bacteria designed to modulate the activity of regulatory T cells	Phase 2	Designed to modulate the activity of regulatory T cells	NCT05370885
FIN-524 (Finch Therapeutics)	UC	Rationally selected microbiota product, lyophilized bacterial strains grown in pure culture	Preclinical	Immuno-modulatory properties	Not applicable
FIN-525 (Finch Therapeutics)	CD	Rationally selected microbiota product, lyophilized bacterial strains grown in pure culture	Preclinical	Immuno-modulatory properties	Not applicable
RBX2660 (Rebiotix)	Pediatric UC	Live microbes from screened human donors	Phase 3	Immuno-modulatory properties	NCT03931941
MET-2 (Mount Sinai Hospital, Ontario)	UC	Defined consortium of human commensal bacteria derived from a healthy donor	Phase 1	Gut microbiome restoration and mucosal healing	NCT02865616

IBD = inflammatory bowel disease, UC = ulcerative colitis, CD = Crohn's disease.

torques, and *C. lavalense* were shown in CD and relapsed CD patients [77–79]. These bacterial genera can produce SCFAs, secrete antimicrobial peptides, produce anti-inflammatory effects, and enhance certain immune responses that could restore perturbed intestinal homeostasis [9, 52, 80]. Currently, several microbiome companies are conducting research trials to evaluate potential of defined microbial populations (SER-287, SER-301, VE202, FIN-524, FIN-525, RBX2660, and MET-2) for the treatment of IBD (Table 1). Recent approval of Vowst by the U.S. Food and Drug Administration (FDA)—the first orally administered microbiota-based therapeutic for preventing recurrent *C. difficile* infection—underscores the potential and promise of microbiota-replacement therapy for improving outcomes in IBD patients.

Modifying host-microbe interactions: future therapeutics

Therapeutic strategies can be developed by modifying microbial metabolism or interfering with the host-microbe interactions. These promising therapeutic approaches are elegantly discussed [81] and are tabularized in Table 2. Microbially derived metabolites have been extensively evaluated for their potential use as medicinal drugs, although most of these compounds have not been advanced to clinical trials (Table 2). The compound EB8018

is a FimH inhibitor that specifically blocks the binding of adherent-invasive *E. coli* to the intestinal epithelium [82] and is undergoing evaluation in a clinical trial in CD patients. Peptide SG-2-0776 is a compound purported to bind to the interface of the cell membrane and the extracellular matrix to promote mucosal healing (Second Genome pipeline) [81]. SG-2-0776 is currently being evaluated in a preclinical model for IBD. The compounds SYMB-104 and SYMB-202 have been shown to decrease intestinal inflammation in mice by increasing the levels of T-reg cells [81]. Collectively, microbial metabolites, which maintain immune homeostasis, mucosal integrity, and immune maturation, are being evaluated as potential therapeutics to cure IBD.

Future perspectives

Our understanding of the interactions between dietary macronutrients, GI microbiota, and their effect on inflammation and intestinal pathology is rapidly evolving. Considering the complex nature of IBD and the heterogeneity in pathology and symptoms in patients, we are still in the nascent stage of determining the combinations of microbiota-targeted therapy that can attenuate the clinical complications in IBD patients. Emerging data from

Table 2. Microbiome-targeted small molecules for IBD

Therapeutic candidate	Conditions	Phase	Mode of action	Company
EB8018	CD	Phase Ib	FimH inhibitor that blocks the adherence of invasive <i>Escherichia coli</i> to the intestinal epithelium protein, FimH	Enterome Biosciences
SG-2-0776	IBD	Preclinical	Mechanism of action yet to be established. Hypothesized to bind to the proteins at the interface of the cell membrane and the extracellular matrix, facilitates mucosal healing	Second Genome
SYMB-104 (polysaccharide A)	IBD	Preclinical	The compound is naturally produced by GI commensal organisms and stimulates the production of regulatory T cells to suppress inflammation	Symbiotix Biotherapies
SYMB-202 (outer membrane vesicles)	IBD	Preclinical	Stimulates regulatory T cells, suppresses inflammation	Symbiotix Biotherapies
PEM compounds	UC	Preclinical	Modulates the enteric signaling network (composed of GI immune cells, the enteric nervous system, and the GI microbiome) in a GI region-specific manner to elicit positive outcomes	Kintai Therapeutics

IBD: inflammatory bowel disease; UC: ulcerative colitis; CD: Crohn's disease. Table 2 is adopted from reference [81].

gut microbiome-targeted therapies, focusing on preventive strategies for IBD, are encouraging. These early studies indicate the potential of microbiome manipulation in managing and potentially preventing this chronic inflammatory condition. However, their translation into clinical use requires a detailed mechanistic understanding of the enzymatic activities of microbial population and their virulence, particularly in the inflammatory environment. Moreover, the effectiveness of these treatments hinges on establishing whether dysbiosis triggers and fuels intestinal inflammation in this specific subset of IBD patients. Eventually, large-scale randomized-controlled trials are needed to optimize the response of microbiome-directed treatments in IBD patients to alleviate intestinal inflammation and manage IBD effectively.

Authors' contributions

D.P. and V.S. conceived and conceptualize the review. D.P., D.V.T.N., G.J., and S.V. prepared the original draft. D.P. and D.V.T.N. completed graphics and visualization. D.P., D.V.T.N., G.J., R.C., A. K.T., and V.S. reviewed and finalized the draft. All authors read and approved the final manuscript.

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Conflicts of interest

The authors declare no conflict of interest. The authors also clarify that they do not have any direct affiliations with the companies whose therapeutic candidates are discussed in the manuscript.

References

1. Franzosa EA, Sirota-Madi A, Avila-Pacheco J et al. Gut microbiome structure and metabolic activity in inflammatory bowel disease. *Nat Microbiol* 2019;**4**:293–305.
2. Ahlawat S, Kumar P, Mohan H et al. Inflammatory bowel disease: tri-directional relationship between microbiota, immune system and intestinal epithelium. *Crit Rev Microbiol* 2021;**47**:254–73.
3. Pascal V, Pozuelo M, Borrueal N et al. A microbial signature for Crohn's disease. *Gut* 2017;**66**:813–22.
4. Halfvarson J, Brislawn CJ, Lamendella R et al. Dynamics of the human gut microbiome in inflammatory bowel disease. *Nat Microbiol* 2017;**2**:17004.
5. Laserna-Mendieta EJ, Clooney AG, Carretero-Gomez JF et al. Determinants of reduced genetic capacity for butyrate synthesis by the gut microbiome in crohn's disease and ulcerative colitis. *J Crohns Colitis* 2018;**12**:204–16.
6. Heinken A, Hertel J, Thiele I. Metabolic modelling reveals broad changes in gut microbial metabolism in inflammatory bowel disease patients with dysbiosis. *NPJ Syst Biol Appl* 2021;**7**:19.
7. Zhou Y, Xu ZZ, He Y et al. Gut Microbiota Offers Universal Biomarkers across Ethnicity in Inflammatory Bowel Disease Diagnosis and Infliximab Response Prediction. *mSystems* 2018;**3**:e00188–17.
8. Dahal RH, Kim S, Kim YK et al. Insight into gut dysbiosis of patients with inflammatory bowel disease and ischemic colitis. *Front Microbiol* 2023;**14**:1174832.
9. Ni J, Wu GD, Albenberg L et al. Gut microbiota and IBD: causation or correlation? *Nat Rev Gastroenterol Hepatol* 2017;**14**:573–84.
10. Lewis JD, Abreu MT. Diet as a Trigger or Therapy for Inflammatory Bowel Diseases. *Gastroenterology* 2017;**152**:398–414 e6.
11. Holscher HD. Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes* 2017;**8**:172–84.
12. Bilotta AJ, Cong Y. Gut microbiota metabolite regulation of host defenses at mucosal surfaces: implication in precision medicine. *Precis Clin Med* 2019;**2**:110–9.
13. Sun M, Wu W, Liu Z et al. Microbiota metabolite short chain fatty acids, GPCR, and inflammatory bowel diseases. *J Gastroenterol* 2017;**52**:1–8.
14. Sinha SR, Haileselassie Y, Nguyen LP et al. Dysbiosis-Induced Secondary Bile Acid Deficiency Promotes Intestinal Inflammation. *Cell Host Microbe* 2020;**27**:659–70 e5.
15. Foley MH, Walker ME, Stewart AK et al. Bile salt hydrolases shape the bile acid landscape and restrict *Clostridioides difficile* growth in the murine gut. *Nat Microbiol* 2023;**8**:611–28.

16. Pithadia AB, Jain S. Treatment of inflammatory bowel disease (IBD). *Pharmacol Rep* 2011;**63**:629–42.
17. Ko JK, Auyeung KK. Inflammatory bowel disease: etiology, pathogenesis and current therapy. *Curr Pharm Des* 2014;**20**:1082–96.
18. Kirchgesner J, Lemaitre M, Carrat F et al. Risk of serious and opportunistic infections associated with treatment of inflammatory bowel diseases. *Gastroenterology* 2018;**155**:337–46 e10.
19. Zhang M, Sun K, Wu Y et al. Interactions between intestinal microbiota and host immune response in inflammatory bowel disease. *Front Immunol* 2017;**8**:942.
20. Zuo T, Kamm MA, Colombel JF et al. Urbanization and the gut microbiota in health and inflammatory bowel disease. *Nat Rev Gastroenterol Hepatol* 2018;**15**:440–52.
21. Nishida A, Inoue R, Inatomi O et al. Gut microbiota in the pathogenesis of inflammatory bowel disease. *Clin J Gastroenterol* 2018;**11**:1–10.
22. Matsuoka K, Kanai T. The gut microbiota and inflammatory bowel disease. *Semin Immunopathol* 2015;**37**:47–55.
23. Khan I, Ullah N, Zha L et al. Alteration of Gut Microbiota in Inflammatory Bowel Disease (IBD): Cause or consequence? IBD treatment targeting the gut microbiome. *Pathogens* 2019;**8**:126.
24. Balish E, Warner T. Enterococcus faecalis induces inflammatory bowel disease in interleukin-10 knockout mice. *Am J Pathol* 2002;**160**:2253–7.
25. Neurath M. Current and emerging therapeutic targets for IBD. *Nat Rev Gastroenterol Hepatol* 2017;**14**:688.
26. Lewis JD, Scott FI, Brensinger CM et al. Increased mortality rates with prolonged corticosteroid therapy when compared with antitumor necrosis factor- α -directed therapy for inflammatory bowel disease. *Am J Gastroenterol* 2018;**113**:405–17.
27. Rutgeerts PJ. Review article: the limitations of corticosteroid therapy in Crohn's disease. *Aliment Pharmacol Ther* 2001;**15**:1515–25.
28. Danese S, Vuitton L, Peyrin-Biroulet L. Biologic agents for IBD: practical insights. *Nat Rev Gastroenterol Hepatol* 2015;**12**:537–45.
29. Ben-Horin S, Chowers Y. Review article: loss of response to anti-TNF treatments in Crohn's disease. *Aliment Pharmacol Ther* 2011;**33**:987–95.
30. Vermeire S, Gils A, Accossato P et al. Immunogenicity of biologics in inflammatory bowel disease. *Therap Adv Gastroenterol* 2018;**11**:1756283X17750355.
31. Ford AC, Peyrin-Biroulet L. Opportunistic infections with anti-tumor necrosis factor- α therapy in inflammatory bowel disease: meta-analysis of randomized controlled trials. *Am J Gastroenterol* 2013;**108**:1268–76.
32. Schroeder BO, Birchenough GMH, Stahlman M et al. Bifidobacteria or fiber protects against diet-induced microbiota-mediated colonic mucus deterioration. *Cell Host Microbe* 2018;**23**:27–40 e7.
33. Desai MS, Seekatz AM, Koropatkin NM et al. A dietary fiber-deprived gut microbiota degrades the colonic mucus barrier and enhances pathogen susceptibility. *Cell* 2016;**167**:1339–53 e21.
34. Riva A, Kuzyk O, Forsberg E et al. A fiber-deprived diet disturbs the fine-scale spatial architecture of the murine colon microbiome. *Nat Commun* 2019;**10**:4366.
35. Diether NE, Willing BP. Microbial fermentation of dietary protein: an important factor in diet(-)microbe(-)host interaction. *Microorganisms* 2019;**7**:19.
36. Llewellyn SR, Britton GJ, Contijoch EJ et al. Interactions between diet and the intestinal microbiota alter intestinal permeability and colitis severity in mice. *Gastroenterology* 2018;**154**:1037–46/e2.
37. Bretin A, Zou J, San Yeoh B et al. Psyllium fiber protects against colitis via activation of bile acid sensor farnesoid X receptor. *Cell Mol Gastroenterol Hepatol* 2023;**15**:1421–42.
38. Singh V, Yeoh BS, Walker RE et al. Microbiota fermentation-NLRP3 axis shapes the impact of dietary fibres on intestinal inflammation. *Gut* 2019;**68**:1801–12.
39. Tian S, Paudel D, Hao F et al. Refined fiber inulin promotes inflammation-associated colon tumorigenesis by modulating microbial succinate production. *Cancer Rep (Hoboken)* 2023;**6**:e1863.
40. He Y, Peng X, Liu Y et al. Long-term maternal intake of inulin exacerbated the intestinal damage and inflammation of offspring rats in a DSS-induced colitis model. *Food Funct* 2022;**13**:4047–60.
41. Miles JP, Zou J, Kumar MV et al. Supplementation of low- and high-fat diets with fermentable fiber exacerbates severity of DSS-induced acute colitis. *Inflamm Bowel Dis* 2017;**23**:1133–43.
42. Armstrong HK, Bording-Jorgensen M, Santer DM et al. Unfermented beta-fructan fibers fuel inflammation in select inflammatory bowel disease patients. *Gastroenterology* 2023;**164**:228–40.
43. Ghouri YA, Richards DM, Rahimi EF et al. Systematic review of randomized controlled trials of probiotics, prebiotics, and synbiotics in inflammatory bowel disease. *Clin Exp Gastroenterol* 2014;**7**:473–87.
44. Geier MS, Butler RN, Howarth GS. Inflammatory bowel disease: current insights into pathogenesis and new therapeutic options; probiotics, prebiotics and synbiotics. *Int J Food Microbiol* 2007;**115**:1–11.
45. Gibson GR, Hutkins R, Sanders ME et al. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol* 2017;**14**:491–502.
46. Davani-Davari D, Negahdaripour M, Karimzadeh I et al. Prebiotics: definition, types, sources, mechanisms, and clinical applications. *Foods* 2019;**8**:92.
47. Hoentjen F, Welling GW, Harmsen HJ et al. Reduction of colitis by prebiotics in HLA-B27 transgenic rats is associated with microflora changes and immunomodulation. *Inflamm Bowel Dis* 2005;**11**:977–85.
48. Liu B, Lin Q, Yang T et al. Oat beta-glucan ameliorates dextran sulfate sodium (DSS)-induced ulcerative colitis in mice. *Food Funct* 2015;**6**:3454–63.
49. Fan L, Zuo S, Tan H et al. Preventive effects of pectin with various degrees of esterification on ulcerative colitis in mice. *Food Funct* 2020;**11**:2886–97.
50. Faghfoori Z, Shakerhosseini R, Navai L et al. Effects of an oral supplementation of germinated barley foodstuff on serum CRP level and clinical signs in patients with ulcerative colitis. *Health Promot Perspect* 2014;**4**:116–21.
51. Jana UK, Kango N, Pletschke B. Hemicellulose-derived oligosaccharides: emerging prebiotics in disease alleviation. *Front Nutr* 2021;**8**:670817.
52. Hill C, Guarner F, Reid G et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol* 2014;**11**:506–14.
53. Holzapfel WH, Haberer P, Geisen R et al. Taxonomy and important features of probiotic microorganisms in food and nutrition. *Am J Clin Nutr* 2001;**73**:365S–73S.
54. Palumbo VD, Romeo M, Marino Gammazza A et al. The long-term effects of probiotics in the therapy of ulcerative colitis: a clinical study. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2016;**160**:372–7.

55. Lorea Baroja M, Kirjavainen PV, Hekmat S et al. Anti-inflammatory effects of probiotic yogurt in inflammatory bowel disease patients. *Clin Exp Immunol* 2007;**149**:470–9.
56. Bjarnason I, Sission G, Hayee B. A randomised, double-blind, placebo-controlled trial of a multi-strain probiotic in patients with asymptomatic ulcerative colitis and Crohn's disease. *Inflammopharmacology* 2019;**27**:465–73.
57. Esposito G, Pesce M, Seguella L et al. Engineered *Lactobacillus paracasei* Producing Palmitoylethanolamide (PEA) prevents colitis in mice. *Int J Mol Sci* 2021;**22**:2945.
58. Hamady ZZ, Scott N, Farrar MD et al. Xylan-regulated delivery of human keratinocyte growth factor-2 to the inflamed colon by the human anaerobic commensal bacterium *Bacteroides ovatus*. *Gut* 2010;**59**:461–9.
59. Zhang T, Zhang W, Feng C et al. Stronger gut microbiome modulatory effects by postbiotics than probiotics in a mouse colitis model. *NPJ Sci Food* 2022;**6**:53.
60. Praveschotinunt P, Duraj-Thatte AM, Gelfat I et al. Engineered *E. coli* Nissle 1917 for the delivery of matrix-tethered therapeutic domains to the gut. *Nat Commun* 2019;**10**:5580.
61. Pesce M, Seguella L, Del Re A et al. Next-generation probiotics for inflammatory bowel disease. *Int J Mol Sci* 2022;**23**:5466.
62. Swanson KS, Gibson GR, Hutkins R et al. The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of synbiotics. *Nat Rev Gastroenterol Hepatol* 2020;**17**:687–701.
63. Fujimori S, Gudis K, Mitsui K et al. A randomized controlled trial on the efficacy of synbiotic versus probiotic or prebiotic treatment to improve the quality of life in patients with ulcerative colitis. *Nutrition* 2009;**25**:520–5.
64. Zhang XF, Guan XX, Tang YJ et al. Clinical effects and gut microbiota changes of using probiotics, prebiotics or synbiotics in inflammatory bowel disease: a systematic review and meta-analysis. *Eur J Nutr* 2021;**60**:2855–75.
65. Gomez Quintero DF, Kok CR, Hutkins R. The Future of Synbiotics: Rational Formulation and Design. *Front Microbiol* 2022;**13**:919725.
66. Yilmaz I, Dolar ME, Ozpinar H. Effect of administering kefir on the changes in fecal microbiota and symptoms of inflammatory bowel disease: A randomized controlled trial. *Turk J Gastroenterol* 2019;**30**:242–53.
67. Zhu Y, Xu Y, Wang X et al. Probiotic cocktail alleviates intestinal inflammation through improving gut microbiota and metabolites in colitis mice. *Front Cell Infect Microbiol* 2022;**12**:886061.
68. Lunken GR, Tsai K, Schick A et al. Prebiotic enriched exclusive enteral nutrition suppresses colitis via gut microbiome modulation and expansion of anti-inflammatory T cells in a mouse model of colitis. *Cell Mol Gastroenterol Hepatol* 2021;**12**:1251–66.
69. Lloyd-Price J, Abu-Ali G, Huttenhower C. The healthy human microbiome. *Genome Med* 2016;**8**:51.
70. Jandhyala SM, Talukdar R, Subramanyam C et al. Role of the normal gut microbiota. *World J Gastroenterol* 2015;**21**:8787–803.
71. Wang ZK, Yang YS, Chen Y et al. Intestinal microbiota pathogenesis and fecal microbiota transplantation for inflammatory bowel disease. *World J Gastroenterol* 2014;**20**:14805–20.
72. Reinisch W. Fecal microbiota transplantation in inflammatory bowel disease. *Dig Dis* 2017;**35**:123–6.
73. Sartor RB, Wu GD. Roles for intestinal bacteria, viruses, and fungi in pathogenesis of inflammatory bowel diseases and therapeutic approaches. *Gastroenterology* 2017;**152**:327–39 e4.
74. Knox NC, Forbes JD, Van Domselaar G et al. The gut microbiome as a target for IBD treatment: are we there yet? *Curr Treat Options Gastroenterol* 2019;**17**:115–26.
75. Yalchin M, Segal JP, Mullish BH et al. Gaps in knowledge and future directions for the use of faecal microbiota transplant in the treatment of inflammatory bowel disease. *Therap Adv Gastroenterol* 2019;**12**:1756284819891038.
76. Blount KF, Shannon WD, Deych E et al. Restoration of bacterial microbiome composition and diversity among treatment responders in a phase 2 trial of RBX2660: an investigational microbiome restoration therapeutic. *Open Forum Infect Dis* 2019;**6**:ofz095.
77. Fujimoto T, Imaeda H, Takahashi K et al. Decreased abundance of *Faecalibacterium prausnitzii* in the gut microbiota of Crohn's disease. *J Gastroenterol Hepatol* 2013;**28**:613–9.
78. Takahashi K, Nishida A, Fujimoto T et al. Reduced abundance of butyrate-producing bacteria species in the fecal microbial community in Crohn's disease. *Digestion* 2016;**93**:59–65.
79. Sokol H, Pigneur B, Watterlot L et al. *Faecalibacterium prausnitzii* is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc Natl Acad Sci USA* 2008;**105**:16731–6.
80. Canani RB, Costanzo MD, Leone L et al. Potential beneficial effects of butyrate in intestinal and extraintestinal diseases. *World J Gastroenterol* 2011;**17**:1519–28.
81. Cully M. Microbiome therapeutics go small molecule. *Nat Rev Drug Discov* 2019;**18**:569–72.
82. Chevalier G, Laveissiere A, Desachy G et al.; MOBIDIC Study Investigators. Blockage of bacterial FimH prevents mucosal inflammation associated with Crohn's disease. *Microbiome* 2021;**9**:176.