DNA Sequencing and Data Analysis

Prof Noam Shomron

Hadas Volkov

Lecture 7, December 16, 2022

DNA Sequencing and Data Analysis

Friday 8:45 AM to 11:15 AM

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DNA Sequencing and Data Analysis

Sequence Mapping and Alignment

Class	Title	Content/assignments	Activity, location
1, 4.11	Introduction to Cells and DNA	Basic knowledge of biology	In the lecture hall, Noam
2, 11.11	DNA Sequencing past and present	Basic knowledge of molecular DNA	In the lecture hall, Noam
3, 18.11	Genomics technologies	DNA, RNA, technologies	In the lecture hall, Noam
4, 25.11	Introduction to Bioinformatics challenges in reading DNA	Focus on three methods: WES/WGS, RNA-seq, cell-free DNA	In the lecture hall, Noam
5, 2.12	Modern DNA Sequencing, 2nd wave File Formats, tools.	Analysis approaches for WES/WGS, RNA-seq, cell-free DNA	In the lecture hall, Hadas and Noam
6, 9.12	De novo Shotgun Assembly	The algorithms and methods behind the assembly problem	In computer class, Hadas and Noam
7, 16.12	**Sequence Mapping and Alignment**	**The algorithms behind mapping and alignment, fast and heuristics**	**In computer class, Hadas and Noam**
8, 23.12	Variant Calling and Somatic Variant Analysis	The bioinformatics behind discovery of novel mutations in cancer	In computer class, Hadas and Noam
9, 30.12	Nanopore data analysis introduction and class activity	The bioinformatics behind Nanopore analysis, activity on computers	In computer class, Hadas and Noam
10, 6.1	Practice molecular biology techniques	Pipetting, transferring small amounts of fluids, running a dry Nanopore experiment	In biology class, Meitar and Noam
11, 13.1	Nanopore DNA sequencing	Nanopore DNA sequencing, experimental run	In biology class, Meitar, Hadas, Assaf
12, 20.1	Nanopore data analysis	Nanopore DNA analysis, experimental run	In computer class, Hadas and Noam
13, 27.1	Nanopore data analysis and presentations	Groups present their results	In the lecture hall, Hadas and Noam

Why Do We Need Sequence Mapping?

Determine the origin of an unknown sequence

Find homologous sequences

Determine genomic position of a sequence

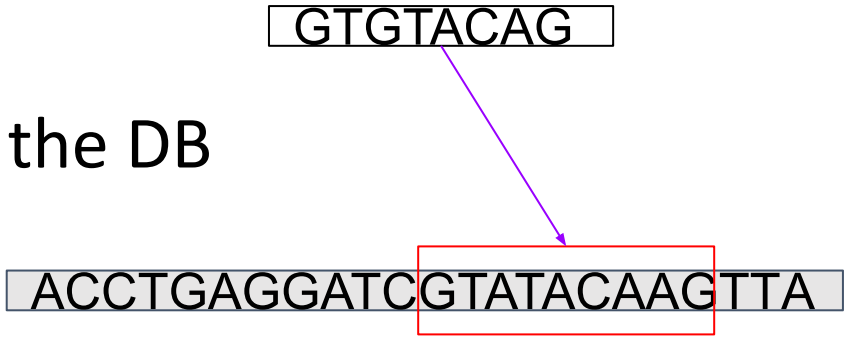
Identify genomic variants between samples (variant calling)

Determine the function of a sequence (annotation)

Two Stages of Sequence Mapping

1. **SEARCH** -

   Roughly find the position of the query in the DB



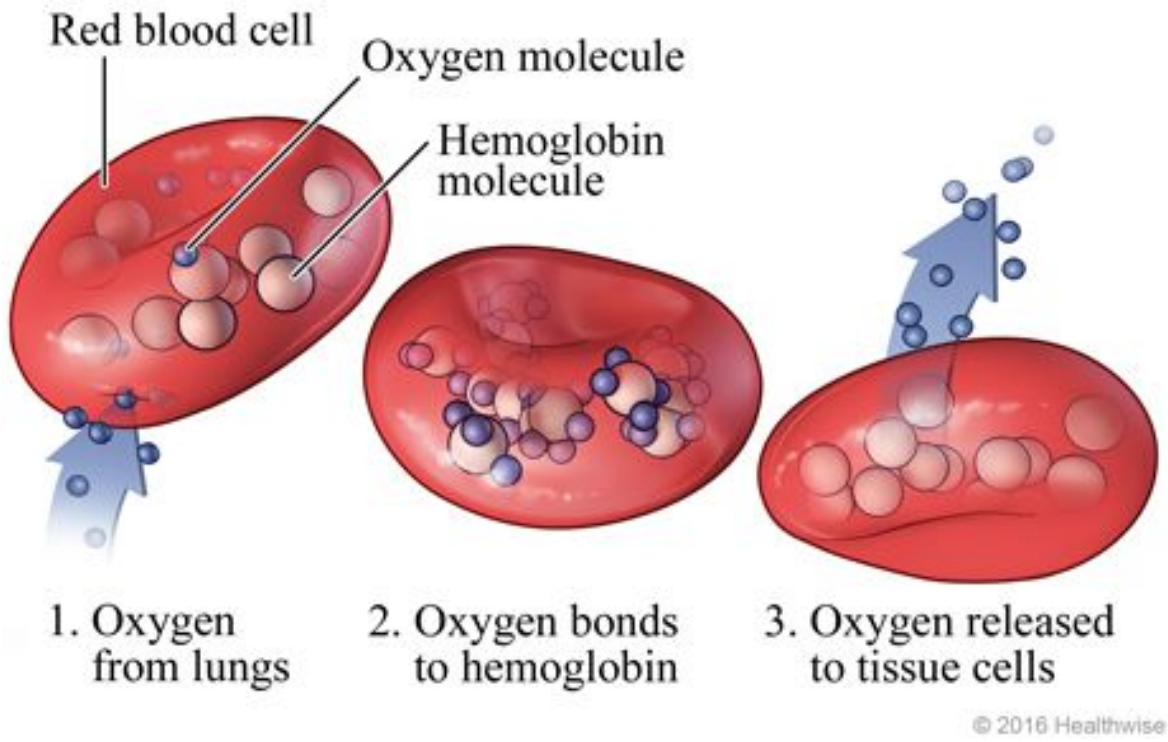
2. **ALIGN** -

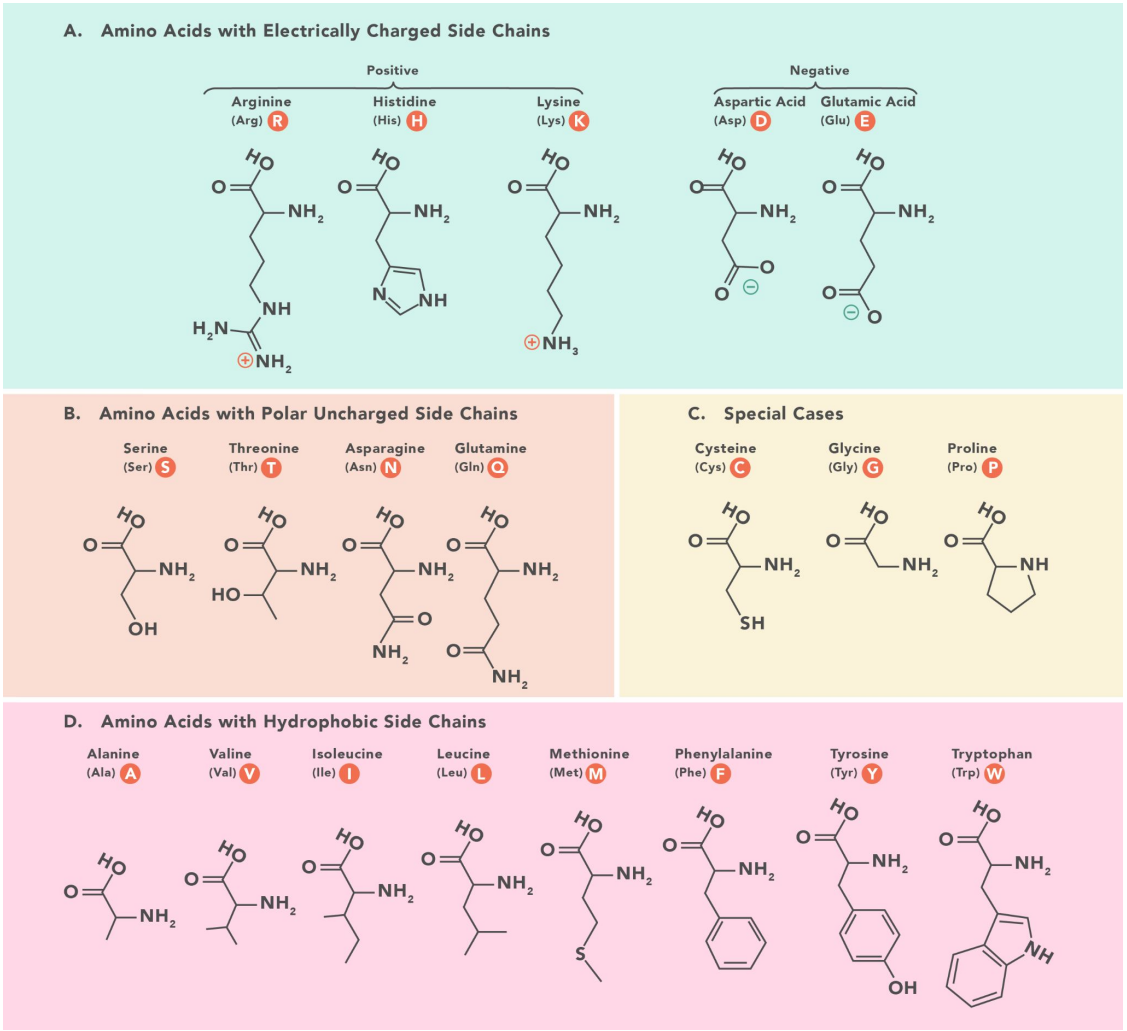
   Find the exact pairwise alignment of the query and the DB sequences

G	T	G	T	A	C	A	-	G
G	T	A	T	A	C	A	A	G

Pairwise Alignment

Hemoglobin Homologous





Pairwise Alignment

Hemoglobin Homologous

```
# NCBI Reference Sequence: NP_000508.1 (human hemoglobin subunit A)
r1 = skbio.Protein("MVLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHGKKVADAL\
            TNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR")

# NCBI Reference Sequence: NP_001004376.1 (chicken hemoglobin subunit A)
r2 = skbio.Protein("MVLSAADKNNVKGIFTKIAGHAEEYGAETLERMFTTYPPTKTYFPHFDLSHGSAQIKGHGKKVVAAL\
            IEAANHIDDIAGTLSKLSDLHAHKLRVDPVNFKLLGQCFLVVVAIHHPAALTPEVHASLDKFLCAVGTVLTAKYR")

# GenBank: QFF91579.1 (sei whale hemoglobin subunit A)
q1 = skbio.Protein("MVLFPADKSNVKATWAKIGNHGAEYGAEALERMFMNFPSTKTYFPHFDLGHDSAQVKGHGKKVADAL\
            TKAAGHMDNLLDALSDLSDLHAHKLRVDPVNFKLLSHCLLVTLALHLPAEFTPSVHASLDKFLASVSTVLTSKYR")
```

Pairwise Alignment

The Hamming Distance

Hamming distance is the number of symbols or positions of two strings at which their corresponding characters are different

```
def hamming_distance(string1, string2):
    if (len(string1) != len(string2)):
        raise Exception('Strings must be of equal length.')
    dist_counter = 0
    for n in range(len(string1)):
        if string1[n] != string2[n]:
            dist_counter += 1
    return dist_counter / len(string1)
```

The Hamming distance between r1 and q1 is: 0.1690
The Hamming distance between r2 and q1 is: 0.3169

Pairwise Alignment

The Hamming Distance

```
# NCBI Reference Sequence: XP_028905054.1 (platypus hemoglobin subunit A);
q2 = skbio.Protein("MLTDAEKKEVTALWGKAAGHGEEYGAEALERLFQAFPTTKTYFSHFDLSHGSAQIKAHGKKVADA\
LSTAAGHFDDMDSALSALSDLHAHKLRVDPVNFKLLAHCILVVLARHCPGEFTPSAHAAMDKFLSKVATVLTSKYR")
q2
```

```
Protein
--------------------------------------------------------------------------
Stats:
    length: 141
    has gaps: False
    has degenerates: False
    has definites: True
    has stops: False
--------------------------------------------------------------------------
0   MLTDAEKKEV TALWGKAAGH GEEYGAEALE RLFQAFPTTK TYFSHFDLSH GSAQIKAHGK
60  KVADALSTAA GHFDDMDSAL SALSDLHAHK LRVDPVNFKL LAHCILVVLA RHCPGEFTPS
120 AHAAMDKFLS KVATVLTSKY R
```

```
q2 = skbio.Protein("MLTDAEKKEVTALWGKAAGHGEEYGAEALERLFQAFPTTKTYFSHFDLSHGSAQIKAHGKKVADA\
LSTAAGHFDDMDSALSALSDLHAHKLRVDPVNFKLLAHCILVVLARHCPGEFTPSAHAAMDKFLSKVATVLTSKYR-")
```

Pairwise Alignment

The Hamming Distance

The Hamming distance between r1 and q2 is: 0.90845

The Hamming distance between r2 and q2 is: 0.92254

```
q2_aligned = skbio.Protein("M-LTDAEKKEVTALWGKAAGHGEEYGAEALERLFQAFPTTKTYFSHFDLSHGSAQIKAHGKKVADA\
            LSTAAGHFDDMDSALSALSDLHAHKLRVDPVNFKLLAHCILVVLARHCPGEFTPSAHAAMDKFLSKVATVLTSKYR")
print(r1)
print(q2_aligned)
```

```
MVLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHGKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPV
M-LTDAEKKEVTALWGKAAGHGEEYGAEALERLFQAFPTTKTYFSHFDLSHGSAQIKAHGKKVADALSTAAGHFDDMDSALSALSDLHAHKLRVDPV
```

The Hamming distance between r1 and q2_aligned is: 0.27465

The Hamming distance between r2 and q2_aligned is: 0.34507

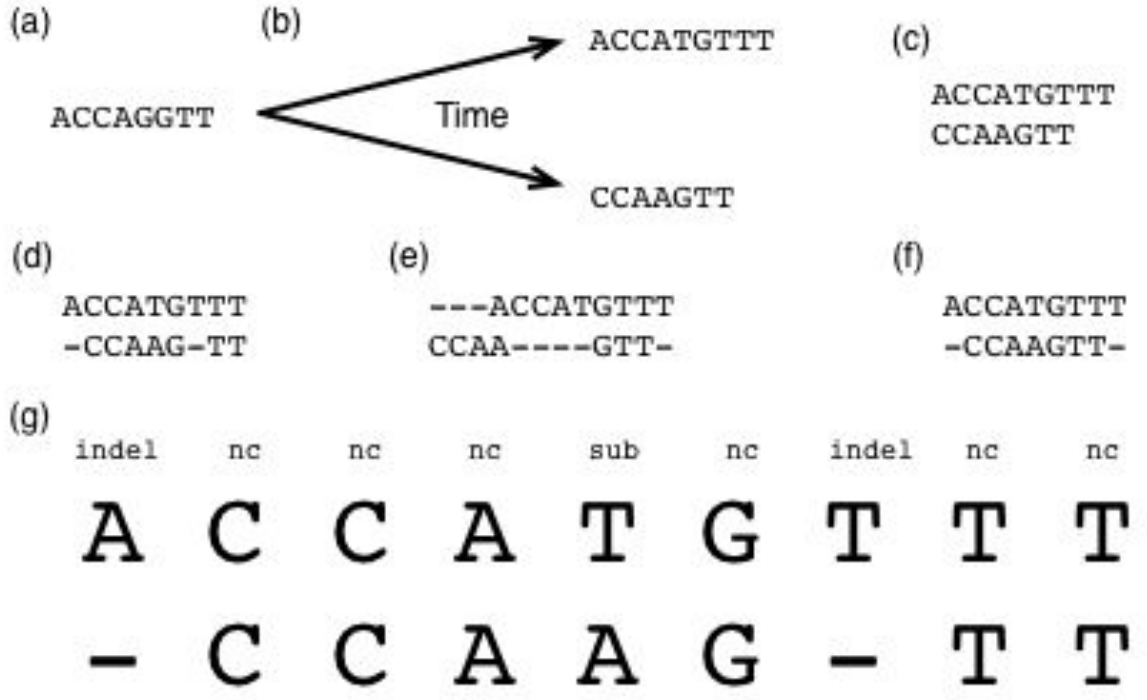
What Is Sequence Alignment?

Mutations:

Substitutions, where one DNA base is replaced with another

Insertions, where one or more contiguous DNA bases are inserted into a sequence

Deletions, where one or more contiguous DNA bases are deleted from a sequence.



Simple Align

ACCATGTTT

CCAAGTT

	A	C	C	A	T	G	T	T	T
C									
C									
A									
A									
G									
T									
T									

Simple Align

ACCATGTTT

CCAAGTT

	A	C	C	A	T	G	T	T	T
C		1	1						
C									
A									
A									
G									
T									
T									

Simple Align

ACCATGTTT

CCAAGTT

	A	C	C	A	T	G	T	T	T
C		1	1						
C		1	1						
A									
A									
G									
T									
T									

Simple Align

ACCATGTTT

CCAAGTT

	A	C	C	A	T	G	T	T	T
C		1	1						
C		1	1						
A	1			1					
A									
G									
T									
T									

Simple Align

ACCATGTTT

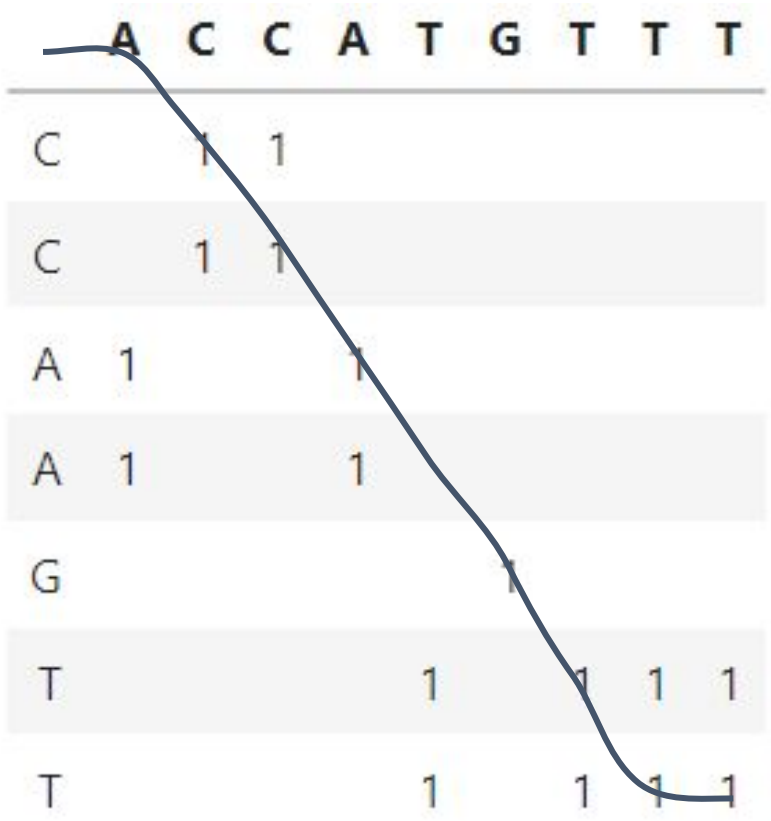
CCAAGTT

	A	C	C	A	T	G	T	T	T
C		1	1						
C		1	1						
A	1			1					
A	1			1					
G						1			
T					1		1	1	1
T					1		1	1	1

Simple Align

ACCATGTTT

CCAAGTT



		A	C	C	A	T	G	T	T	T
C			1	1						
C			1	1						
A	1				1					
A	1				1					
G							1			
T						1		1	1	1
T						1		1	1	1

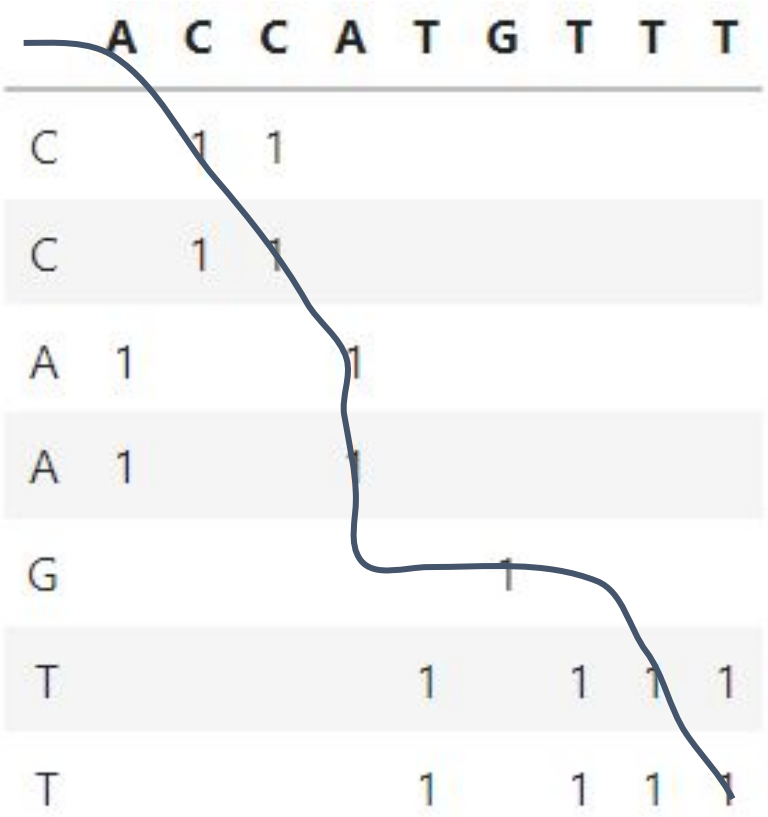
ACCATGTTT

-CCAAGTT -

Simple Align

ACCATGTTT

CCAAGTT



	A	C	C	A	T	G	T	T	T
C		1	1						
C		1	1						
A	1			1					
A	1			1					
G						1			
T					1		1	1	1
T					1		1	1	1

ACCA- -TGTTT

-CCAAG- - -TT

Simple Align

ACCATGTTT

CCAAGTT

ACCATGTTT

-CCAAGTT -

S = -1+1+1-1+1+1+1-1=4

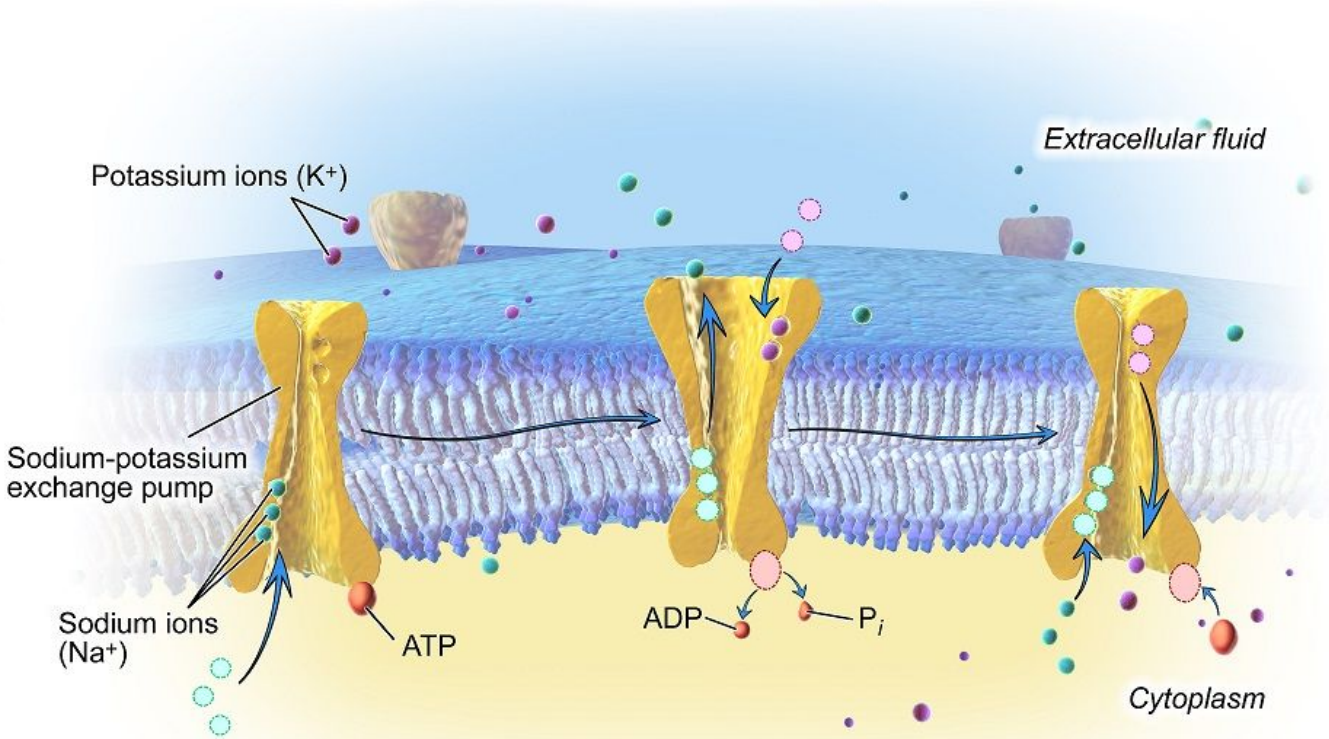
ACCA- -TGTTT

-CCAAG- - -TT

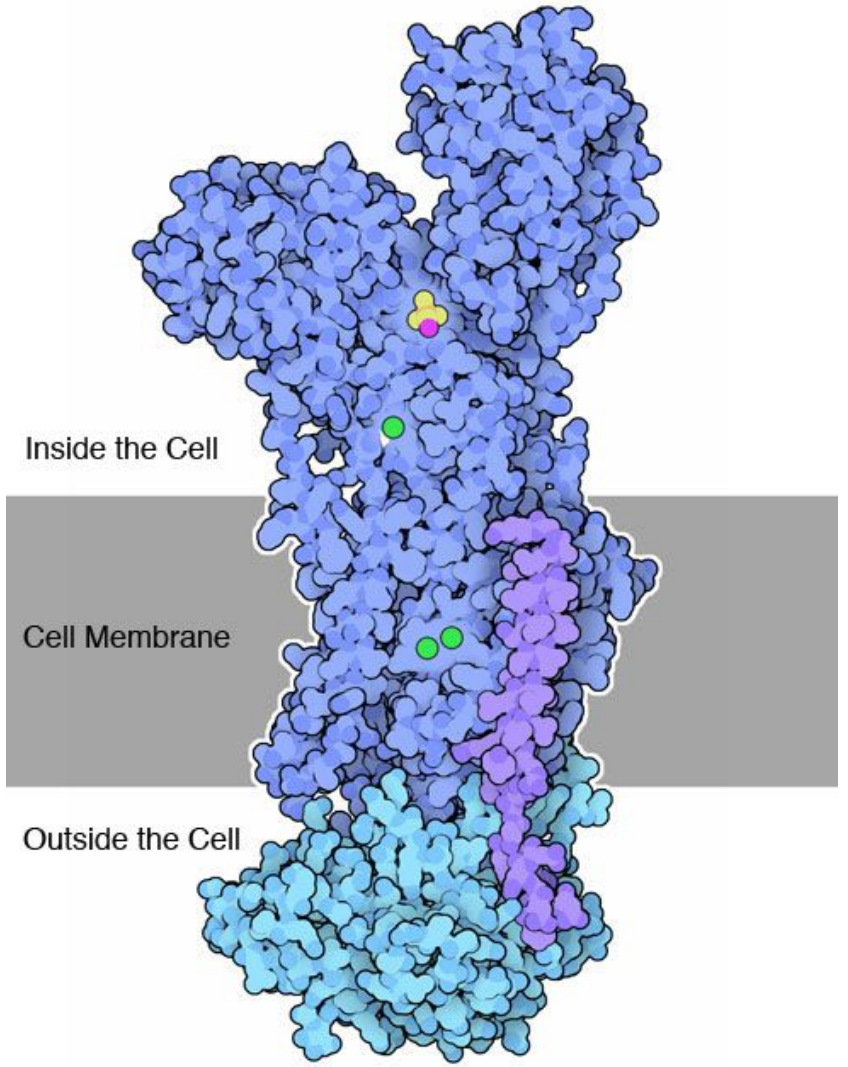
S = -1+1+1+1-1-1-1-1-1+1+1=-1

Simple Align

Too Simplistic



The Sodium-Potassium Exchange Pump



Simple Align

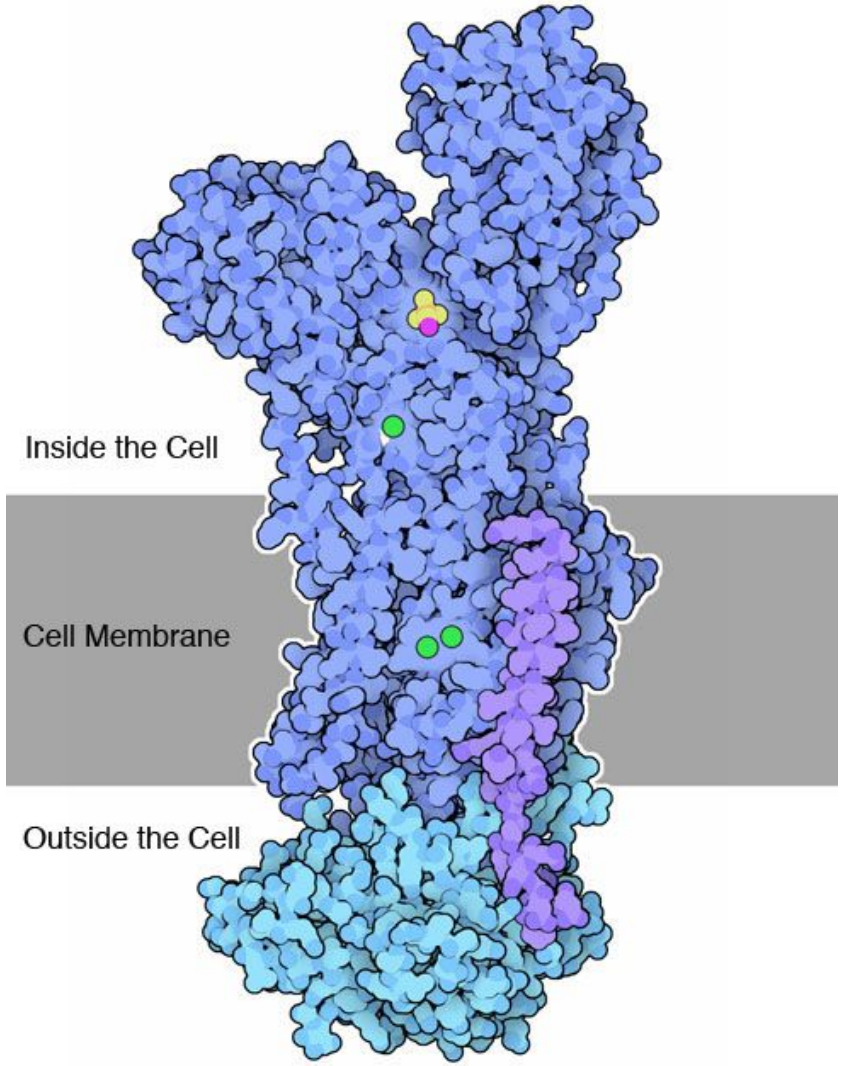
Too Simplistic

BLOSUM 62 scoring matrix

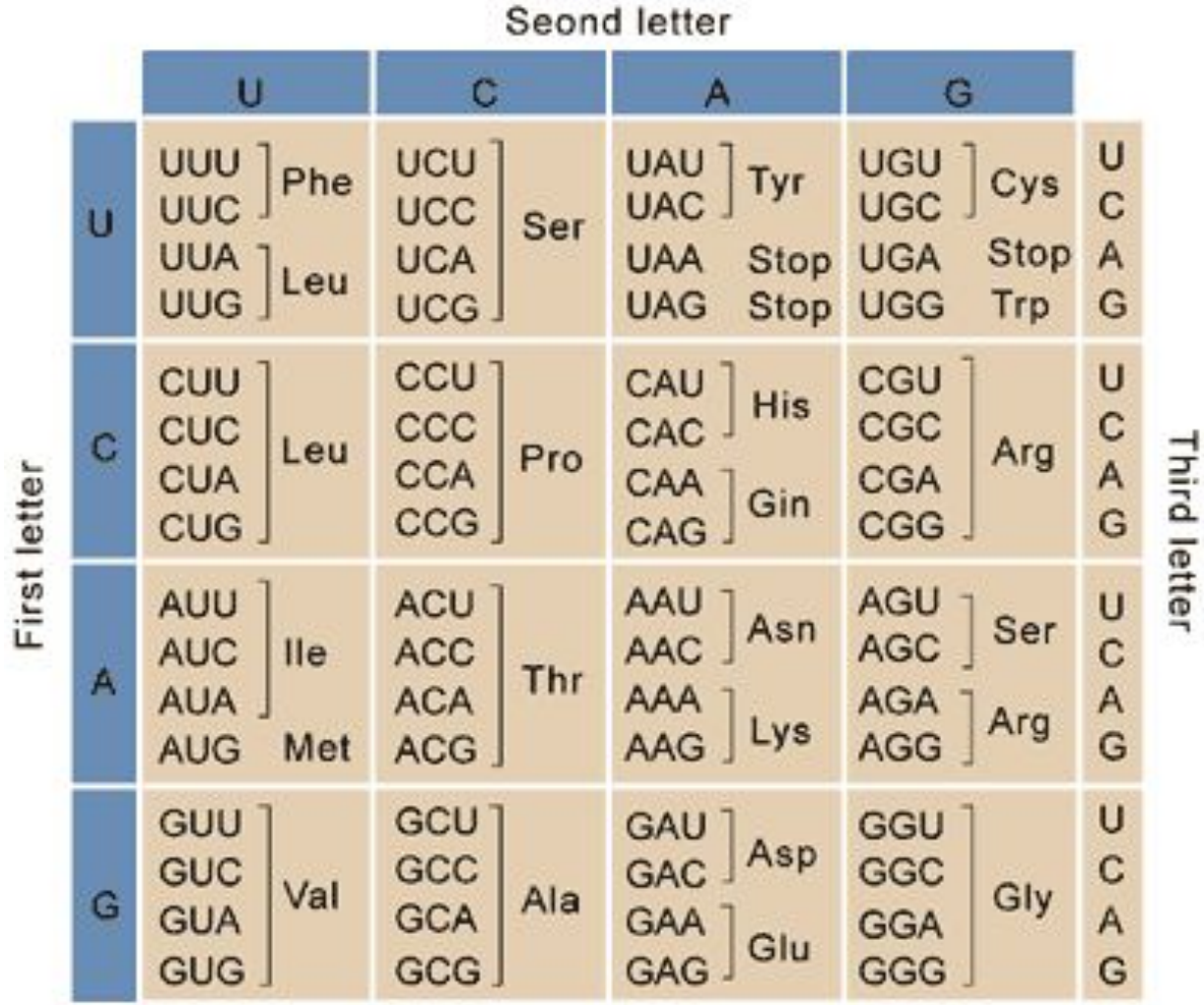
(positive values are shaded)

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	**4**																			
R	-1	**5**																		
N	-2	0	**6**																	
D	-2	-2	**1**	**6**																
C	0	-3	-3	-3	**9**															
Q	-1	**1**	0	0	-3	**5**														
E	-1	0	0	**2**	-4	**2**	**5**													
G	0	-2	0	-1	-3	-2	-2	**6**												
H	-2	0	**1**	-1	-3	0	0	-2	**8**											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	**4**										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	**2**	**4**									
K	-1	**2**	0	-1	-3	**1**	**1**	-2	-1	-3	-2	**5**								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	**1**	**2**	-1	**5**							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	**6**						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	**7**					
S	**1**	-1	**1**	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	**4**				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	**1**	**5**			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	**1**	-4	-3	-2	**11**		
Y	-2	-2	-2	-3	-2	-1	-2	-3	**2**	-1	-1	-2	-1	**3**	-3	-2	-2	**2**	**7**	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	**3**	**1**	-2	**1**	-1	-2	-2	0	-3	-1	**4**

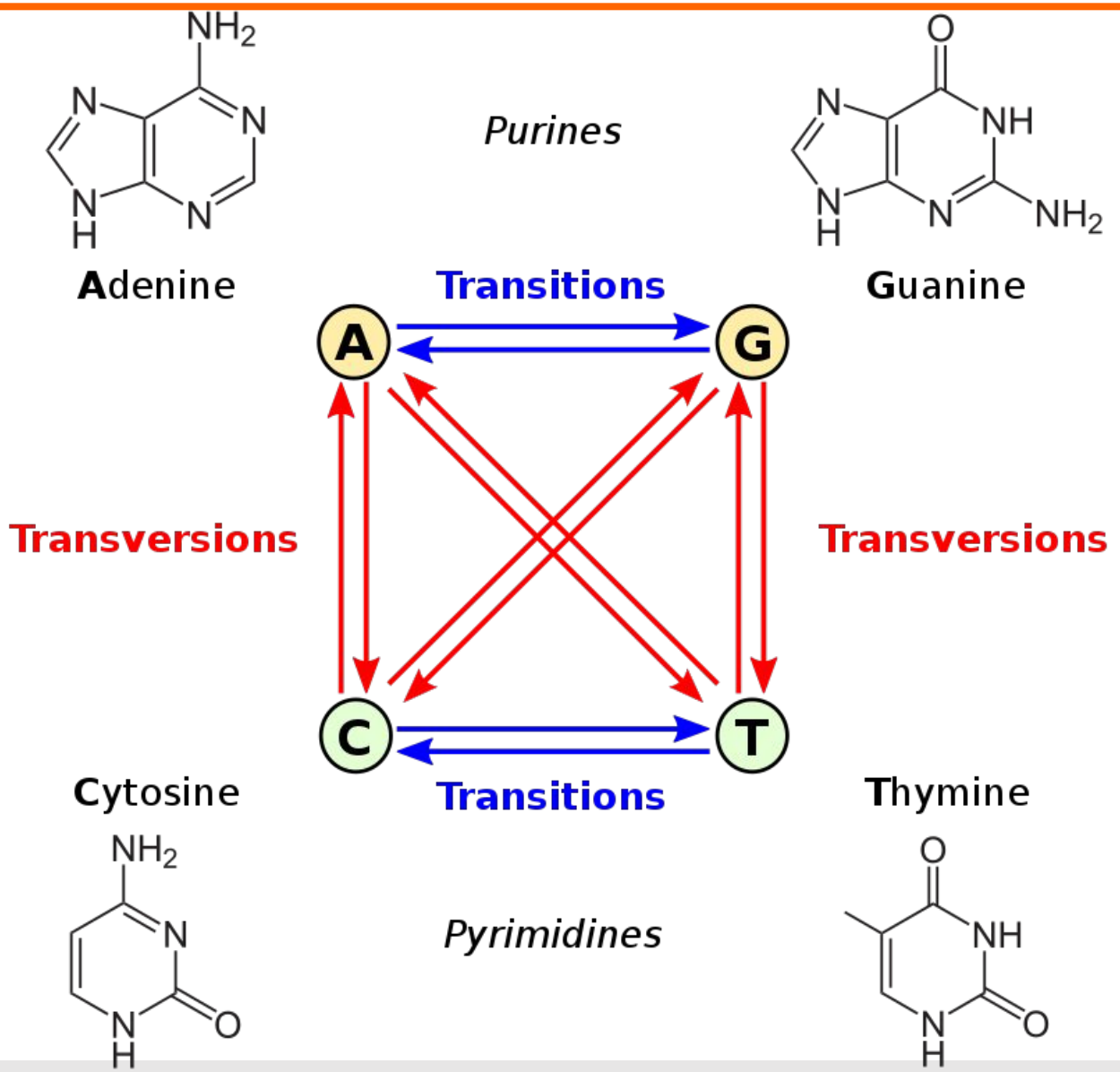
The values for amino acid substitutions were obtained from Henikoff S & Henikoff JG (1992) Amino acid substitutions matrices from protein blocks. *Proc. Natl. Acad. Sci.* **89**: 10915-10919.



Simple Align



Simple Align



Needleman-Wunsch Alignment

HEAGAWGHEE

PAWHEAE

F - The Dynamic Programming Matrix

T - The Traceback Matrix

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
W	0	0	0	0	0	0	0	0	0	0	0
H	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0

Needleman-Wunsch Alignment

max

$$F(0,0)=0$$
$$F(i,0)=F(i-1,0)-d$$
$$F(0,j)=F(0,j-1)-d$$

$$T(0,0)=.$$
$$T(i,0)=\leftarrow$$
$$T(0,j)=\uparrow$$

		H	E	A	G	A	W	G	H	E	E
	0	-8	-16	-24	-32	-40	-48	-56	-64	-72	-80
P	-8	0	0	0	0	0	0	0	0	0	0
A	-16	0	0	0	0	0	0	0	0	0	0
W	-24	0	0	0	0	0	0	0	0	0	0
H	-32	0	0	0	0	0	0	0	0	0	0
E	-40	0	0	0	0	0	0	0	0	0	0
A	-48	0	0	0	0	0	0	0	0	0	0
E	-56	0	0	0	0	0	0	0	0	0	0

		H	E	A	G	A	W	G	H	E	E
	•	←	←	←	←	←	←	←	←	←	←
P	↑										
A	↑										
W	↑										
H	↑										
E	↑										
A	↑										
E	↑										

Needleman-Wunsch Alignment

max

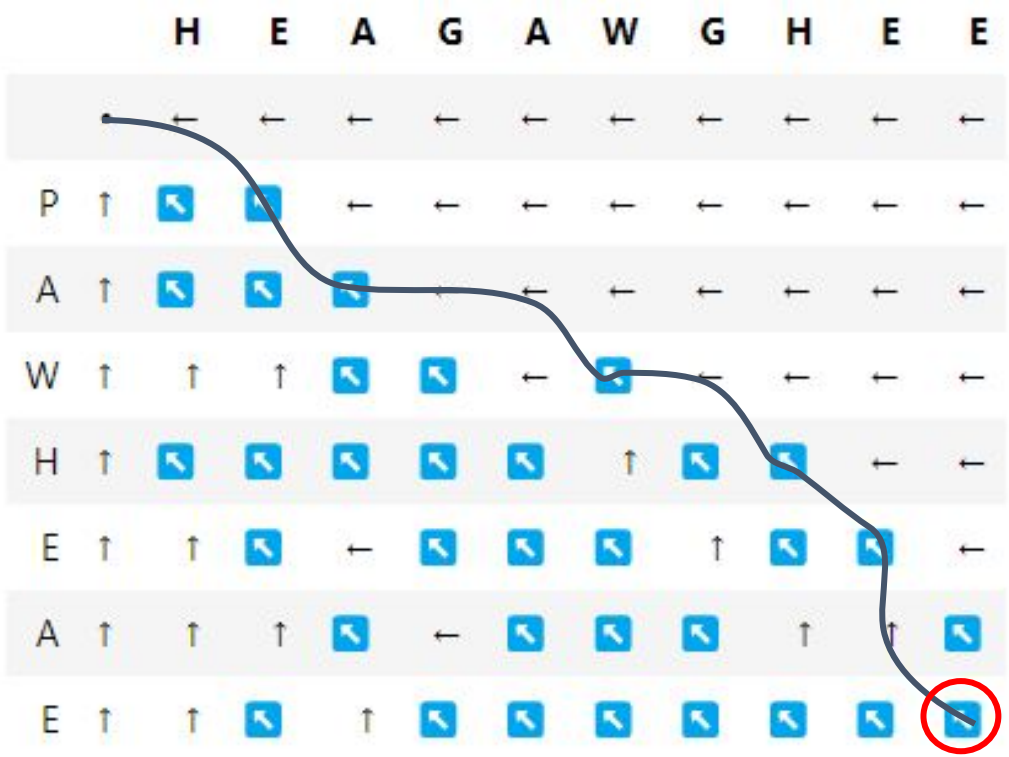
$$F(i-1,j-1)+s(ci,cj)$$
$$F(i-1,j)-d$$
$$F(i,j-1)-d$$

		H	E	A	G	A	W	G	H	E	E
	0	-8	-16	-24	-32	-40	-48	-56	-64	-72	-80
P	-8	-2	-9	-17	-25	-33	-41	-49	-57	-65	-73
A	-16	-10	-3	-4	-12	-20	-28	-36	-44	-52	-60
W	-24	-18	-11	-6	-7	-15	-5	-13	-21	-29	-37
H	-32	-14	-18	-13	-8	-9	-13	-7	-3	-11	-19
E	-40	-22	-8	-16	-16	-9	-12	-15	-7	3	-5
A	-48	-30	-16	-3	-11	-11	-12	-12	-15	-5	2
E	-56	-38	-24	-11	-6	-12	-14	-15	-12	-9	1

		H	E	A	G	A	W	G	H	E	E
	•	←	←	←	←	←	←	←	←	←	←
P	↑	↖	↖	←	←	←	←	←	←	←	←
A	↑	↖	↖	↖	←	←	←	←	←	←	←
W	↑	↑	↑	↖	↖	←	↖	←	←	←	←
H	↑	↖	↖	↖	↖	↖	↑	↖	↖	←	←
E	↑	↑	↖	←	↖	↖	↖	↑	↖	↖	←
A	↑	↑	↑	↖	←	↖	↖	↖	↑	↑	↖
E	↑	↑	↖	↑	↖	↖	↖	↖	↖	↖	↖

Needleman-Wunsch Alignment

		H	E	A	G	A	W	G	H	E	E
	0	-8	-16	-24	-32	-40	-48	-56	-64	-72	-80
P	-8	-2	-9	-17	-25	-33	-41	-49	-57	-65	-73
A	-16	-10	-3	-4	-12	-20	-28	-36	-44	-52	-60
W	-24	-18	-11	-6	-7	-15	-5	-13	-21	-29	-37
H	-32	-14	-18	-13	-8	-9	-13	-7	-3	-11	-19
E	-40	-22	-8	-16	-16	-9	-12	-15	-7	-3	-5
A	-48	-30	-16	-3	-11	-11	-12	-12	-15	-5	2
E	-56	-38	-24	-11	-6	-12	-14	-15	-12	-9	-1



HEAGAWGHE - E

- PA - -W - HEAE

Score = 1

Local vs. Global Alignment

Global alignment - try to match entire sequences

Useful for closely-related sequences of similar size

Local alignment - allow partial matching

Useful for sequences expected to contain some similarity regions

Smith-Waterman Local Alignment

HEAGAWGHEE

PAWHEAE

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
W	0	0	0	0	0	0	0	0	0	0	0
H	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0

Smith-Waterman Local Alignment

$$F(0,0)=0$$
$$F(i,0)=0$$
$$F(0,j)=0$$

$$T(0,0)=.$$
$$T(i,0)=.$$
$$T(0,j)=.$$

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
W	0	0	0	0	0	0	0	0	0	0	0
H	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	0	0	0	0	0	0	0	0

		H	E	A	G	A	W	G	H	E	E
	.	.	.	.	.	.	.	.	.	.	.
P	.										
A	.										
W	.										
H	.										
E	.										
A	.										
E	.										

Smith-Waterman Local Alignment

max

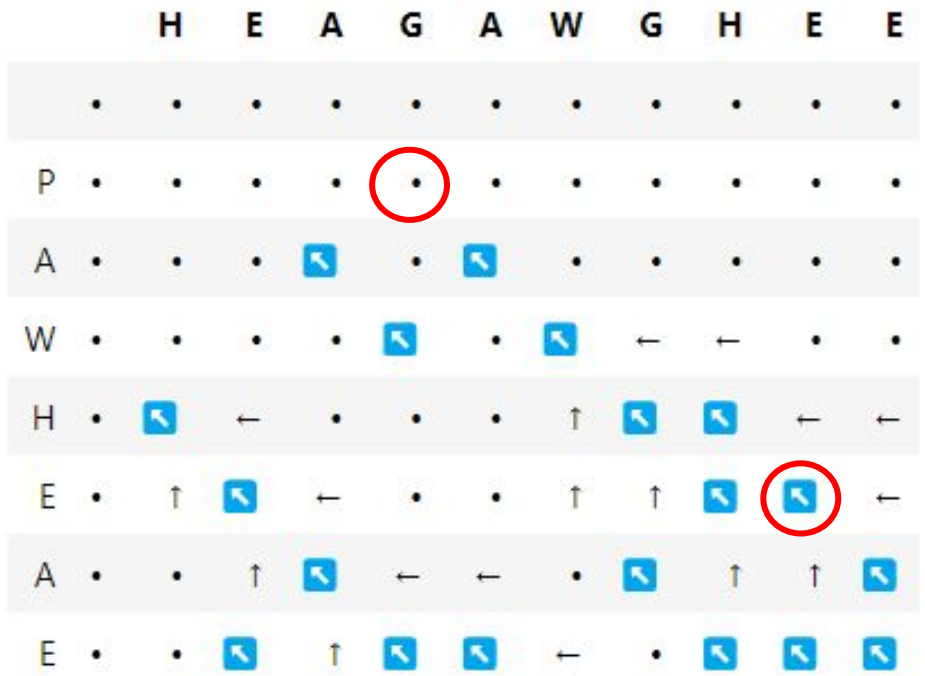
$$\begin{matrix} 0 \\ F(i-1,j-1)+s(ci,cj) \\ F(i-1,j)-d \\ F(i,j-1)-d \end{matrix}$$

AWGHE

AW-HE

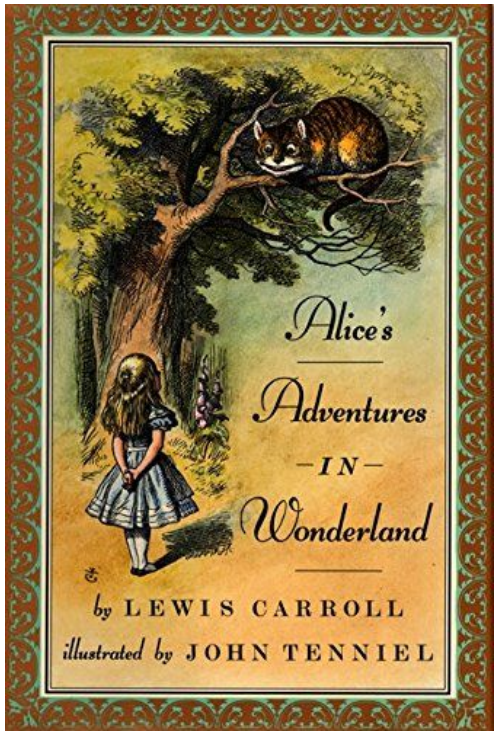
Score = 28

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	5	0	5	0	0	0	0	0
W	0	0	0	0	2	0	20	12	4	0	0
H	0	10	2	0	0	0	12	18	22	14	6
E	0	2	16	8	0	0	4	10	18	28	20
A	0	0	8	21	13	5	0	4	10	20	27
E	0	0	6	13	18	12	4	0	4	16	26



Search

Imagine we have a big book...



… and we want to search it for a specific sentence

It would be
“ so nice if
something
made sense
for a
change.

Lewis Carroll
Alice in Wonderland

Search

- How can we do it in a timely manner?
  - Brute force
  - Indexing
- Do we allow slight changes?

  e.g. : *"it **c**ould be so nice if something made sense"*
- Do we allow insertions and deletions?

  e.g. : *"it would be ~~**so**~~ nice if something made **a little** sense"*
- What if the sentence is repeated in several places in the book?

Sequence Mapping Challenges

Large DBs - millions to billions of nucleotides/AAs

Repetition - biological sequences tend to repeat

Noisy - sequencing errors and real biological variants

BLAST - Basic Local Alignment Search Tool

The most popular alignment tool

BLAST finds regions of similarity between biological sequences.

Compares nucleotide or protein sequences to sequence databases

Calculates the statistical significance of DB hits

Allows searching for **imperfect** sequence matches

Uses a **heuristic** algorithm to improve efficiency

BLAST - Algorithm

1. Index the DB
2. Generate query words
3. compute neighbour words
4. Search the DB for exact word matches - seeds
5. Elongate and combine seeds to get final alignment
6. Score alignment

BLAST - Indexing

Only needed the first time a DB is used

Mask repetitive and low-complexity regions - `ATATATTTATT` → `atatatttatt`

Break DB sequences into overlapping words of length *W*

- W=3 for amino acids
- W=11 for nucleotides

Create a lookup table of words with their positions

WTDFGYPAILKGGTAC

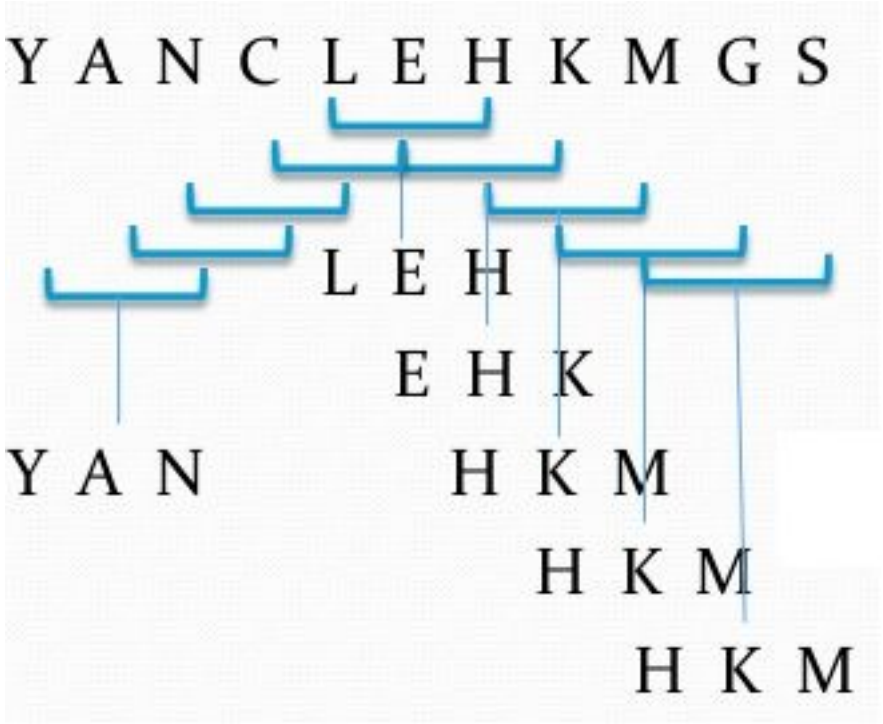
WTD	1
TDF	2
...	
TAC	14

BLAST - Breaking Query to Words

A query of length *L* produces *L-W+1* **overlapping** words of length *W*

L = 11

W = 3



BLAST - Finding Neighbour Words

1. For each word, find all neighbourhood words

   = words with one change

2. Use a scoring matrix to assign each neighbourhood word a score
3. Discard neighbourhood words with score < *T*

BLAST - Breaking Query to Words

Y A N C L E H K M G S

L E H

L K H
C E H
Q E H
L M H
L F H
L E R
D E H
…

BLOSUM 62 scoring matrix

(positive values are shaded)

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	4																			
R	-1	5																		
N	-2	0	6																	
D	-2	-2	1	6																
C	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
H	-2	0	1	-1	-3	0	0	-2	8											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4

The values for amino acid substitutions were obtained from Henikoff S & Henikoff JG (1992) Amino acid substitutions matrices from protein blocks. *Proc. Natl. Acad. Sci.* **89**: 10915-10919.

```
L  E  H
4+1+8 = 13
L  K  H
```

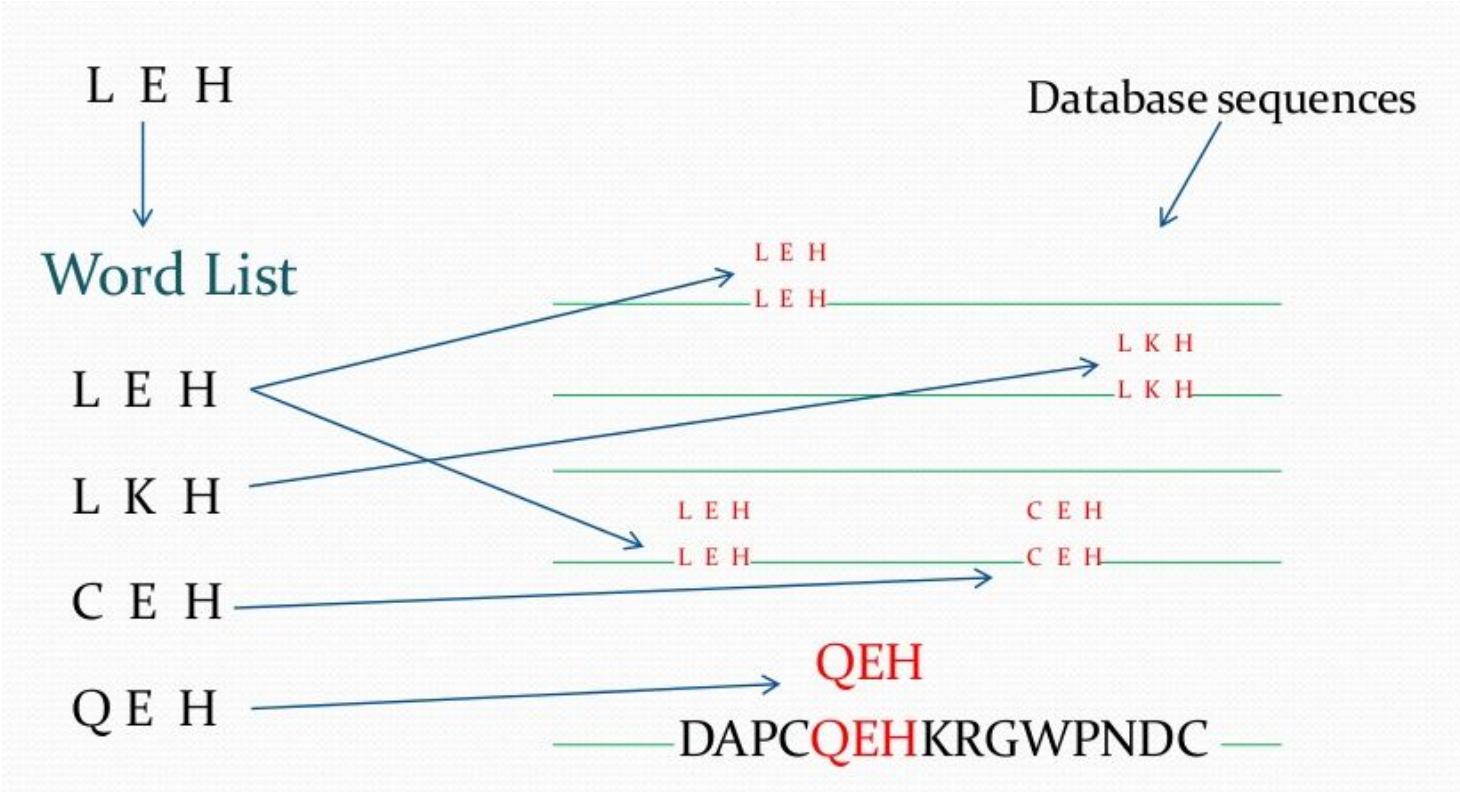
L K H	13
C E H	12
Q E H	11
L M H	10
L F H	9
L E R	9
D E H	9
…	

T=11

L K H	13
C E H	12
Q E H	11
L M H	10
L F H	9
L E R	9
D E H	9
…	

BLAST - Finding Alignment Seeds in DB

- Look for **exact** matches of query words with the DB words
- Masked regions are ignored

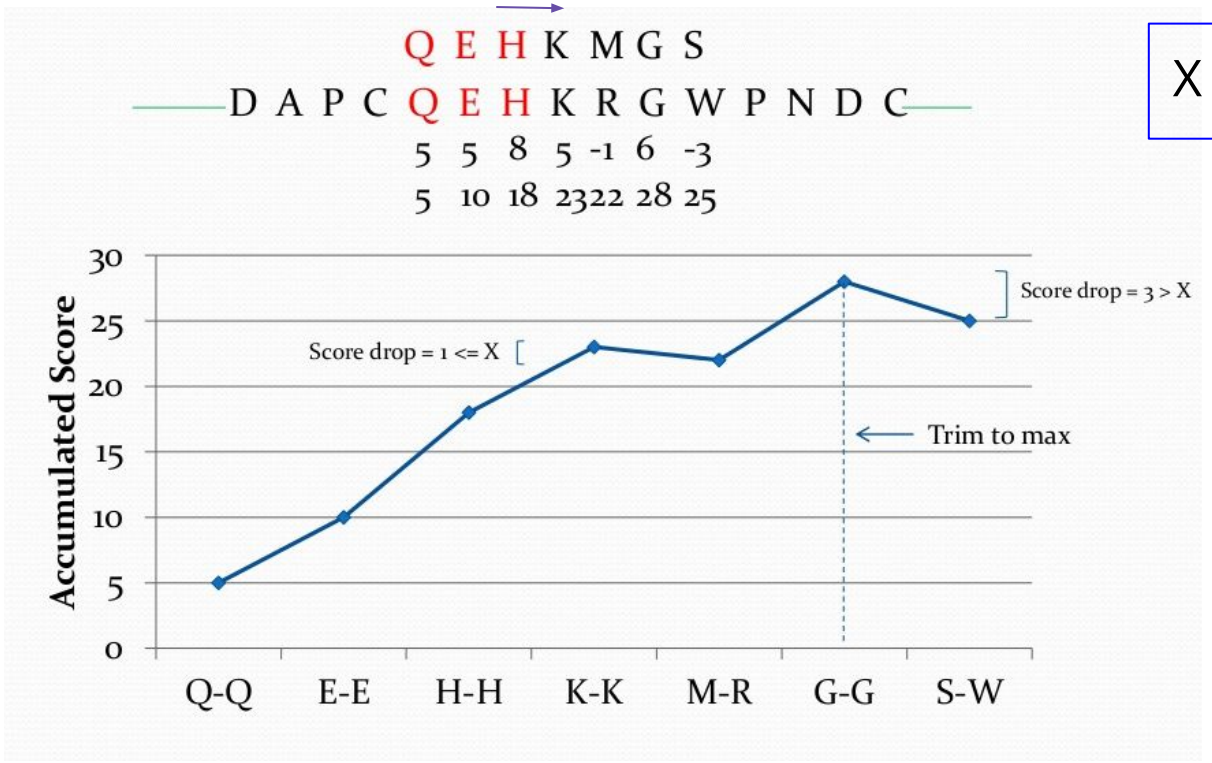


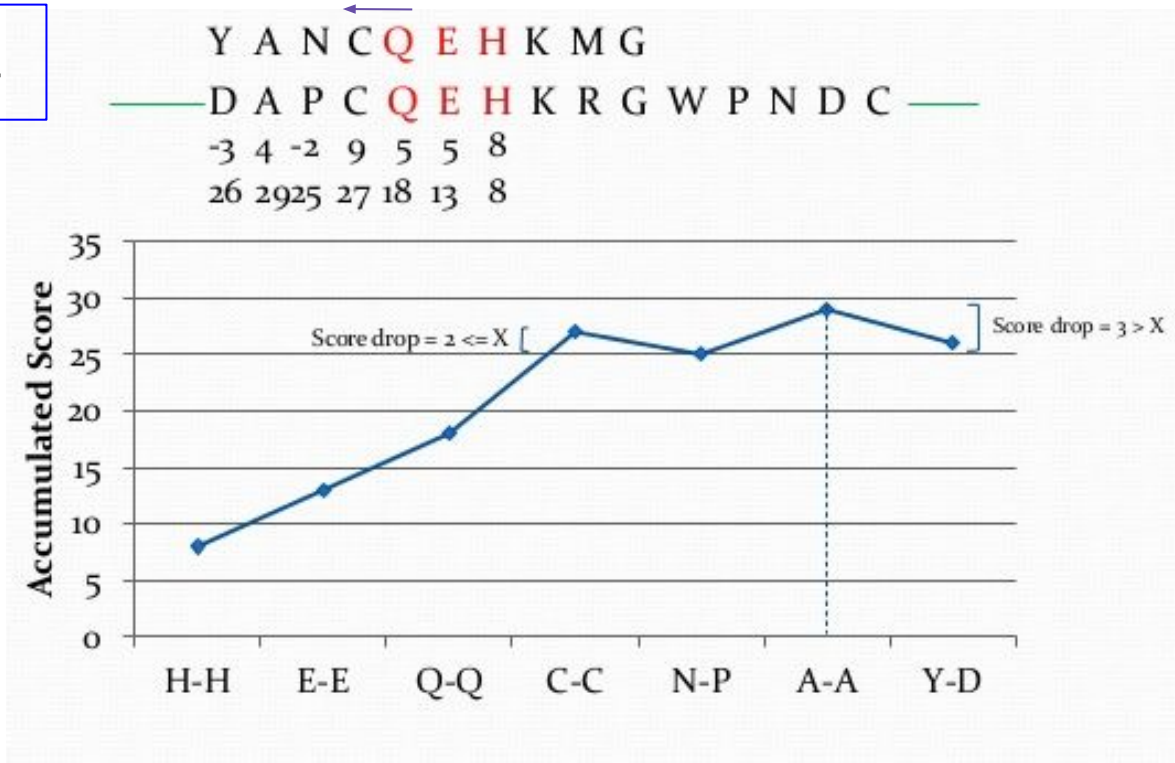
BLAST - Seed Elongation

Elongate each seed to both directions until a score drop > *X* is encountered

Query:
YANC**LEH**KMGS

X = 2





BLAST - Scoring the Alignment

Calculate total alignment score

Discard alignments with score < *S*

Remaining alignments are called High scoring Sequence Pairs - **HSP**s

BLAST - Scoring the Alignment

- Calculate alignment **bit score**
  - Independent of query length
  - Independent of DB size

$$S' = \frac{\lambda S - \ln(K)}{\ln(2)}$$

- Calculate **E-value** - the number of hits with score >= s that one can expect to find in DB by chance

$$E = \frac{L \times N}{2^{S'}}$$

L - query length , *N* - DB length , *S'* - bit score

Smaller E $\longrightarrow$ better hit

BLAST - Scoring the Alignment

W – word size (query and DB)

T – neighborhood words score cutoff

X – allowed score drop during seed elongation

S – HSP score cutoff

What would happen if we **increase** T?

Can We Use Blast in NGS?

Blastn - ~100 reads / sec

Human genome - ~ 3Gb

Assume 100bp reads

How long to map x10 data to the human genome?

Data required:
3 Gb x 10 = 30 Gb

Reads required:
30 Gb / 100 = 300 M reads

Time to map:
300 M reads / (100 reads/sec) = 3M sec = ~ **35 days**

Scale and Speed

We need to map millions to hundreds of millions of reads

Can we use Blast?

Blastn - ~100 reads / sec

Human genome - ~ 3Gb

Assume 100bp reads

How long to map x10 data to the human genome?

Hint: how many reads do we need?

BWA - Burrows-Wheeler Aligner

Specifically designed for mapping of short reads

Maps ~2,200 reads / sec (one CPU)

Allows parallel computing

Contains three algorithms - the most useful is **BWA-MEM**

BWA - Limitations

Only works for nucleotides (usually DNA, not RNA)

Less effective when:

- Queries are very long
- Reads are highly diverged from the reference
- Reads contain lots of sequencing errors

Usually offers a good accuracy-speed balance

BWA - Algorithm Overview

Step 1: Index the reference genome

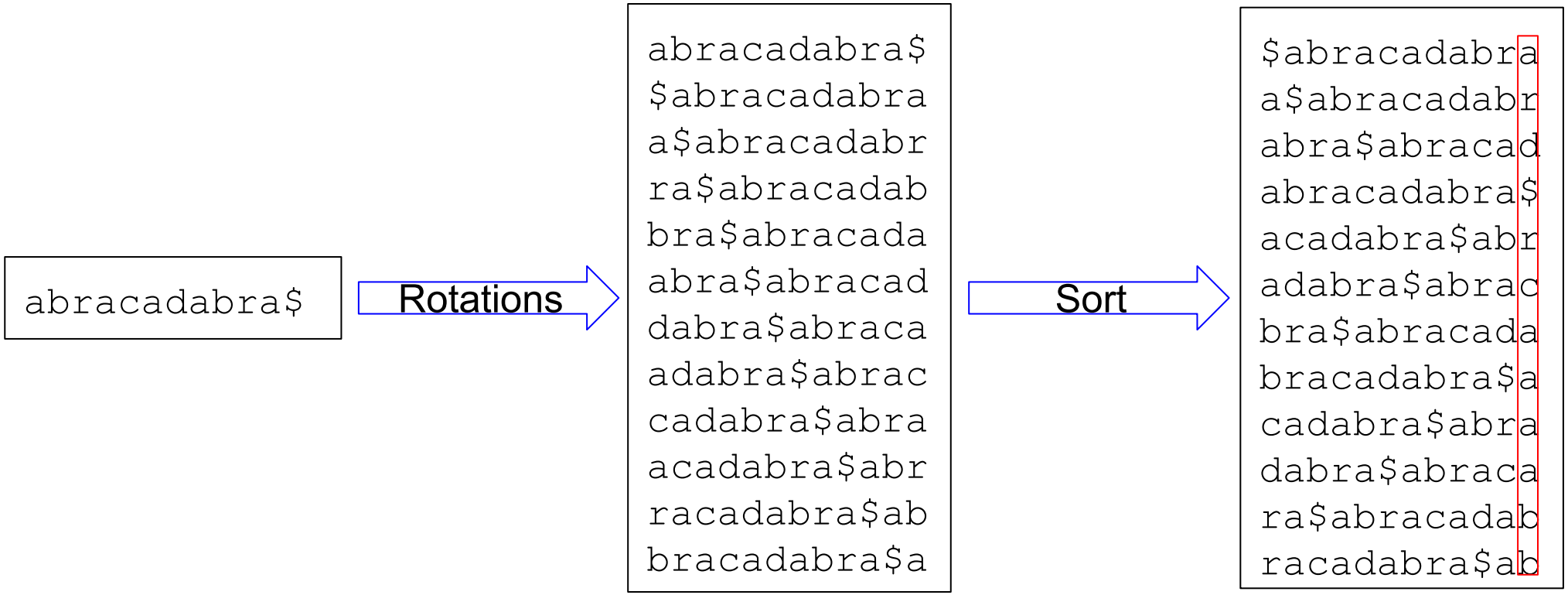
Step 2: Search for reads

Indexing is based on the **Burrows-Wheeler's transformation**

Index allows easy searching:

- Quick
- Memory efficient

BWA - The Burrows Wheeler Transform



BWT(abracadabra$) = ard$rcaaaabb

BWA - The Burrows Wheeler Transform

BWT is **reversible** - we can get back from BWT(G) to G

BWT(G) tends to cluster the same characters together - easy to compress

BWT(abracadabra$) = ard$rcaaaabb

Using some additional data structures, BWT(G) can be searched efficiently

BWA - The Burrows Wheeler Transform

1. Create index of reference genome:

   Input: reference in fasta format

```
$ bwa index genome.fasta
```

2. Map reads to reference:

   Input: reads file or pair (for PE data) in fastq format

```
$ bwa mem genome.fasta reads_R1.fq reads_R2.fq -o aln.sam
```