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## **Assignment 2**

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```

library(class)

# Read CSV from working directory into R

MyData <- read.csv(file="redwine.csv", header=TRUE, sep=",")

### Question 2

## a.

# Numerical summary of data

str(MyData)

## 'data.frame':    1599 obs. of  12 variables:
## $ fixed.acidity      : num  7.4 7.8 7.8 11.2 7.4 7.4 7.9 7.3 7.8 7.5 ...
## $ volatile.acidity   : num  0.7 0.88 0.76 0.28 0.7 0.66 0.6 0.65 0.58 0.5 ...
## $ citric.acid        : num  0 0 0.04 0.56 0 0 0.06 0 0.02 0.36 ...
## $ residual.sugar     : num  1.9 2.6 2.3 1.9 1.9 1.8 1.6 1.2 2 6.1 ...
## $ chlorides          : num  0.076 0.098 0.092 0.075 0.076 0.075 0.069 0.065 0.073 0.
071 ...
## $ free.sulfur.dioxide : num  11 25 15 17 11 13 15 15 9 17 ...
## $ total.sulfur.dioxide: num  34 67 54 60 34 40 59 21 18 102 ...
## $ density            : num  0.998 0.997 0.997 0.998 0.998 ...
## $ pH                 : num  3.51 3.2 3.26 3.16 3.51 3.51 3.3 3.39 3.36 3.35 ...
## $ sulphates          : num  0.56 0.68 0.65 0.58 0.56 0.56 0.46 0.47 0.57 0.8 ...
## $ alcohol            : num  9.4 9.8 9.8 9.8 9.4 9.4 9.4 10 9.5 10.5 ...
## $ quality            : int  5 5 5 6 5 5 5 7 7 5 ...

library(psych)
describe(MyData)

##               vars      n mean    sd median trimmed   mad  min
## fixed.acidity      1 1599  8.32  1.74   7.90    8.15  1.48 4.60
## volatile.acidity    2 1599  0.53  0.18   0.52    0.52  0.18 0.12
## citric.acid         3 1599  0.27  0.19   0.26    0.26  0.25 0.00
## residual.sugar      4 1599  2.54  1.41   2.20    2.26  0.44 0.90
## chlorides           5 1599  0.09  0.05   0.08    0.08  0.01 0.01
## free.sulfur.dioxide  6 1599 15.87 10.46  14.00   14.58 10.38 1.00
## total.sulfur.dioxide 7 1599 46.47 32.90  38.00   41.84 26.69 6.00
## density             8 1599  1.00  0.00   1.00    1.00  0.00 0.99
## pH                 9 1599  3.31  0.15   3.31    3.31  0.15 2.74
## sulphates          10 1599  0.66  0.17   0.62    0.64  0.12 0.33
## alcohol            11 1599 10.42  1.07  10.20   10.31  1.04 8.40
## quality            12 1599  5.64  0.81   6.00    5.59  1.48 3.00
##               max range skew kurtosis   se
## fixed.acidity   15.90 11.30 0.98    1.12 0.04
## volatile.acidity  1.58  1.46 0.67    1.21 0.00
## citric.acid      1.00  1.00 0.32   -0.79 0.00
## residual.sugar   15.50 14.60 4.53   28.49 0.04
## chlorides        0.61  0.60 5.67   41.53 0.00
## free.sulfur.dioxide 72.00 71.00 1.25    2.01 0.26
## total.sulfur.dioxide 289.00 283.00 1.51    3.79 0.82
## density          1.00  0.01 0.07    0.92 0.00

```

## pH	4.01	1.27	0.19	0.80	0.00
## sulphates	2.00	1.67	2.42	11.66	0.00
## alcohol	14.90	6.50	0.86	0.19	0.03
## quality	8.00	5.00	0.22	0.29	0.02

#### summary(MyData)

## fixed.acidity	volatile.acidity	citric.acid	residual.sugar
## Min. : 4.60	Min. :0.1200	Min. :0.000	Min. : 0.900
## 1st Qu.: 7.10	1st Qu.:0.3900	1st Qu.:0.090	1st Qu.: 1.900
## Median : 7.90	Median :0.5200	Median :0.260	Median : 2.200
## Mean : 8.32	Mean :0.5278	Mean :0.271	Mean : 2.539
## 3rd Qu.: 9.20	3rd Qu.:0.6400	3rd Qu.:0.420	3rd Qu.: 2.600
## Max. :15.90	Max. :1.5800	Max. :1.000	Max. :15.500
## chlorides	free.sulfur.dioxide	total.sulfur.dioxide	
## Min. :0.01200	Min. : 1.00	Min. : 6.00	
## 1st Qu.:0.07000	1st Qu.: 7.00	1st Qu.: 22.00	
## Median :0.07900	Median :14.00	Median : 38.00	
## Mean :0.08747	Mean :15.87	Mean : 46.47	
## 3rd Qu.:0.09000	3rd Qu.:21.00	3rd Qu.: 62.00	
## Max. :0.61100	Max. :72.00	Max. :289.00	
## density	pH	sulphates	alcohol
## Min. :0.9901	Min. :2.740	Min. :0.3300	Min. : 8.40
## 1st Qu.:0.9956	1st Qu.:3.210	1st Qu.:0.5500	1st Qu.: 9.50
## Median :0.9968	Median :3.310	Median :0.6200	Median :10.20
## Mean :0.9967	Mean :3.311	Mean :0.6581	Mean :10.42
## 3rd Qu.:0.9978	3rd Qu.:3.400	3rd Qu.:0.7300	3rd Qu.:11.10
## Max. :1.0037	Max. :4.010	Max. :2.0000	Max. :14.90
## quality			
## Min. :3.000			
## 1st Qu.:5.000			
## Median :6.000			
## Mean :5.636			
## 3rd Qu.:6.000			
## Max. :8.000			

```
# Graphical summary of data
```

```
pairs(MyData, main="Scatter Plot of Variables")
```

## Scatter Plot of Variables



### # Correlation of attributes

```
z <- cor(MyData)
z[lower.tri(z,diag=TRUE)]=NA
z=as.data.frame(as.table(z))
z=na.omit(z)
z=z[order(-abs(z$Freq)),]
head(z, n=10)
```

##	Var1	Var2	Freq
## 97	fixed.acidity	pH	-0.6829782
## 25	fixed.acidity	citric.acid	0.6717034
## 85	fixed.acidity	density	0.6680473
## 78	free.sulfur.dioxide	total.sulfur.dioxide	0.6676665
## 26	volatile.acidity	citric.acid	-0.5524957
## 99	citric.acid	pH	-0.5419041
## 128	density	alcohol	-0.4961798
## 143	alcohol	quality	0.4761663
## 134	volatile.acidity	quality	-0.3905578
## 113	chlorides	sulphates	0.3712605

*# After exploring the dataset, I generated a correlation matrix to understand how the attributes are related to each other. I created a list of these relationships, sorted them, and printed out the ones with the largest absolute values. Obviously, a correlation of 1 would be the highest signify a perfect, positive correlation whereas a -1 would signify a perfect, negative correlation.*

*# The highest correlation patterns in the data seem to be between fixed.acidity & pH, fixed.acidity % citric.acid, fixed.acidity & density, and freesulfur.dioxide & total.sulfur.dioxide. All of them have roughly a 0.68 correlation - only fixed.acidity and pH have an inverse relationship. Although these correlations are not extremely high, they indicate that there is some dependency of variables on each other which is potentially harmful to us when building a model.*

## b.

*# Create binary variable final\_quality using mean*

```
MyData$final_quality <- with(ifelse(quality>mean(quality), 1, 0), data=MyData)
```

*# Creating dataset without original quality attribute*

```
myvars <- names(MyData) %in% c("quality")
fulldataset <- MyData[!myvars]
head(fulldataset, n=1)
```

##	fixed.acidity	volatile.acidity	citric.acid	residual.sugar	chlorides	
## 1	7.4	0.7	0	1.9	0.076	
##	free.sulfur.dioxide	total.sulfur.dioxide	density	pH	sulphates	alcohol
## 1	11	34	0.9978	3.51	0.56	9.4
##	final_quality					
## 1	0					

```
# Splitting data into train and test
```

```
set.seed(1)
rows <- sample(x=nrow(fulldataset), size=.80*nrow(fulldataset))
trainset <- fulldataset[rows, ]
testset <- fulldataset[-rows, ]
```

```
# Logistic Regression
```

```
glm.fit <- glm(final_quality ~ fixed.acidity+volatile.acidity+citric.acid+residual.sugar+
chlorides+free.sulfur.dioxide+total.sulfur.dioxide+density+pH+sulphates+alcohol, data=trainset, family=binomial)
summary(glm.fit)
```

```
##
## Call:
## glm(formula = final_quality ~ fixed.acidity + volatile.acidity +
##      citric.acid + residual.sugar + chlorides + free.sulfur.dioxide +
##      total.sulfur.dioxide + density + pH + sulphates + alcohol,
##      family = binomial, data = trainset)
##
```

```
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## -3.3285  -0.8640   0.3094   0.8477   2.2338
##
```

```
## Coefficients:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)    19.150545   85.590617   0.224  0.82296
## fixed.acidity     0.111649   0.105894   1.054  0.29173
## volatile.acidity  -2.844256   0.534364  -5.323 1.02e-07 ***
## citric.acid      -0.988874   0.630212  -1.569  0.11662
## residual.sugar    0.006617   0.061182   0.108  0.91388
## chlorides        -4.933251   1.738512  -2.838  0.00454 **
## free.sulfur.dioxide  0.016142   0.009139   1.766  0.07734 .
## total.sulfur.dioxide -0.014461   0.003119  -4.636 3.55e-06 ***
## density          -25.095517  87.419908  -0.287  0.77406
## pH               -0.921976   0.793994  -1.161  0.24557
## sulphates         2.567119   0.499801   5.136 2.80e-07 ***
## alcohol           0.872373   0.113699   7.673 1.68e-14 ***
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
##
## (Dispersion parameter for binomial family taken to be 1)
```

```
##      Null deviance: 1770.7  on 1278  degrees of freedom
## Residual deviance: 1352.0  on 1267  degrees of freedom
## AIC: 1376
```

```
##
## Number of Fisher Scoring iterations: 4
```

```
# Atleast 4 of the predictors appear to be statistically significant. alcohol,
# sulphates, total.sulfur.dioxide, and volative.acidity all have very small p
# values meaning they have the largest impact on the response variable of final
# quality.
```

```
## c.
```

### *# Confusion Matrix*

```
glm.probs <- predict(glm.fit, testset, type="response")
glm.preds <- ifelse(glm.probs>0.5, 1, 0)
confusion_matrix_glm <- table(testset$final_quality,glm.preds)
print(confusion_matrix_glm)
```

```
##      glm.preds
##      0      1
## 0 105    27
## 1   45   143
```

### *# Fraction of Correct Predictions*

```
Correct_Predictions_fraction = (confusion_matrix_glm[1,1]+confusion_matrix_glm[2,2])/sum(
confusion_matrix_glm)
sprintf("Overall Fraction of Correct Predictions are: %f",Correct_Predictions_fraction)
```

```
## [1] "Overall Fraction of Correct Predictions are: 0.775000"
```

*# The confusion tells us about the performance of the model. There were True  
# Negatives (105) and True Positives (143) and, as can be seen above, constitute  
# about 77.5% of results. The rest were misclassified - so about 22.5%. 27  
# points were False Positives and 45 were False Negatives. This tells us our  
# logistic regression model is wrong about one fourth of the time and tends to  
# be too conservative. It is wrongly classifying good wine (in class 1 that have  
# quality above the mean) as bad wine (in class 0 with quality below mean) more  
# than it is classifying bad wine as good wine (although that is happening a  
# fair bit as well).*

```
## d.
```

```
variables <- which(names(fulldataset)%in%c("fixed.acidity","volatile.acidity","citric.aci  
d","residual.sugar","chlorides","free.sulfur.dioxide","total.sulfur.dioxide","density","p  
H","sulphates","alcohol"))
```

```
test_error <- data.frame("k"=1:11)
```

```
set.seed(1)
for(k in 1:11)
{
  knn.pred <- knn(train=trainset[, variables], test=testset[, variables], cl=trainset$fi  
nal_quality, k=k)
  test_error$error[k]= round(sum(knn.pred!=testset$final_quality)/nrow(testset)*100,2)
}
```

```
print(test_error)
```

```
##      k error
## 1    1 25.62
## 2    2 35.00
## 3    3 34.06
## 4    4 37.19
```

```
## 5    5 36.25
## 6    6 37.50
## 7    7 32.81
## 8    8 32.50
## 9    9 34.06
## 10   10 34.69
## 11   11 31.87
```

*# As seen above, the model with the lowest test error is when  $k=1$  with an error of approximately 25 and therefore can be concluded to be performing the best on this dataset.*

```
### Question 3
```

```
## a.
```

*# Split data into training and test - 80/20*

```
set.seed(1)
```

```
rows <- sample(x=nrow(fulldataset), size=0.8*nrow(fulldataset))
trainset <- fulldataset[rows, ]
testset <- fulldataset[-rows, ]
```

```
## b.
```

*# LDA*

```
library (MASS)
```

```
## Warning: package 'MASS' was built under R version 3.4.3
```

```
lda.fit <- lda(final_quality ~ fixed.acidity+volatile.acidity+citric.acid+residual.sugar+
chlorides+free.sulfur.dioxide+total.sulfur.dioxide+density+pH+sulphates+alcohol, data=trainset)
lda.pred <- predict(lda.fit, testset)
confusion_matrix_lda <- table(testset$final_quality, lda.pred$class)
print(confusion_matrix_lda)
```

```
##
##      0    1
## 0 105  27
## 1  47 141
```

```
test_error_lda <- sum(lda.pred$class!=testset$final_quality)/nrow(testset)
sprintf("The test error for LDA is: %f", test_error_lda)
```

```
## [1] "The test error for LDA is: 0.231250"
```



```
## c.

# QDA

qda.fit <- qda(final_quality ~ fixed.acidity+volatile.acidity+citric.acid+residual.sugar+
chlorides+free.sulfur.dioxide+total.sulfur.dioxide+density+pH+sulphates+alcohol, data=trainset)
qda.pred <- predict(qda.fit, testset)
confusion_matrix_qda <- table(testset$final_quality, qda.pred$class)
print(confusion_matrix_qda)

##
##      0    1
##  0  90  42
##  1  36 152

test_error_qda <- sum(qda.pred$class!=testset$final_quality)/nrow(testset)
sprintf("The test error for QDA is: %f", test_error_qda)

## [1] "The test error for QDA is: 0.243750"
```

## SUMMARY OF SECTION 4.5 – COMPARISON OF CLASSIFICATION METHODS

Logistic regression, LDA, QDA, and kNN are all different classification methods that each have their strengths and weaknesses – understanding the scenarios in which they are most useful can help us build more accurate models. Logistic regression and LDA both create linear decision boundaries (only difference between the two is their fitting procedures) and therefore when the true decision boundary is linear in form, these methods perform well. kNN, on the other hand, is completely non-parametric (does not assume shape), and hence performs better when the true boundary is highly-nonlinear. The disadvantage with kNN is that we cannot infer anything about the individual predictors and their impact. QDA falls somewhere in between the two: able to fit a wider range of shapes than linear methods but not as flexible as kNN. By making assumptions about the data, it is able to perform better at lower number of training examples than kNN. There are several in between states as well – such as transformations of the predictors – that can be performed to move between these four main types of classification methods. It is important to realize the benefits and shortcomings of each method so that we can apply the correct one when the problem needs it.