# Supplementary Material

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### Introduction

SSD can determine the sample size for our experiment when we only have a few pilot data. Firstly, it can generate different synthetic training datasets and test datasets. Then it will calculate the corresponding different test classification error/ARI/AMI. The final step is to construct the plots in the terms of the sample size and the different metrics (test classification error/ARI/AMI). Users can determine the sample size by the above plots.

## **Preparations**

Before we dive into the main task, we need to load the package and an example dataset for our task. The dataset we use is the **pbmc\_68k** dataset from 10x Genomics.

We pre-processed the dataset: In this dataset, *phenoid* is the y label which has 10 classes. We sampled 15 observations from the original dataset for each class and assemble them as the pilot data. we normalize and scale the pilot data at first and then run principal component analysis (PCA) and keep 18 PCs according to JackStrawPlot and ElbowPlot mentioned in Seurat - Guided Clustering Tutorial. The JackStrawPlot and ElbowPlot are shown below:

For the whole dataset, we split the it into training data and test data first, then we ran principal component analysis (PCA). We pre-processed the training dataset using the same stratigies and keep 23 PCs according to JackStrawPlot and ElbowPlot, and then project the test set onto the reduced feature space obtained during the training.

We put the pro-processed data into our package and we can load them directly.

```
library(SSD)
# load data
pilot_data <- read.csv(system.file("extdata", "data_pbmc68k_pilot_18pc.csv",</pre>
                                      package = "SSD"),row.names=1)
print(table(pilot_data$phenoid))
#>
#>
                                                         CD19+_B
                  CD14+_Monocyte
#>
#>
                 CD4+/CD25_T_Req
#>
             CD4+/CD45RO+_Memory
#>
#>
                         CD56+ NK CD8+/CD45RA+ Naive Cytotoxic
#>
#>
                                                               15
#>
                CD8+_Cytotoxic_T
#>
```

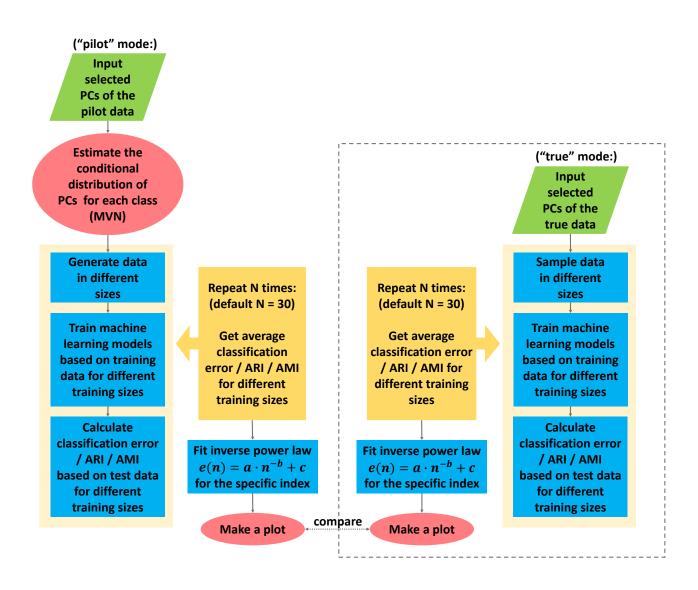


Figure 1: Workflow of the SSD package

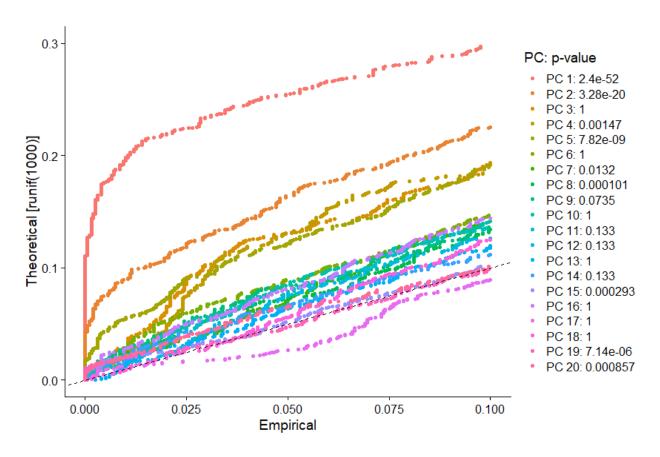


Figure 2: JackStrawPlot of the pilot data

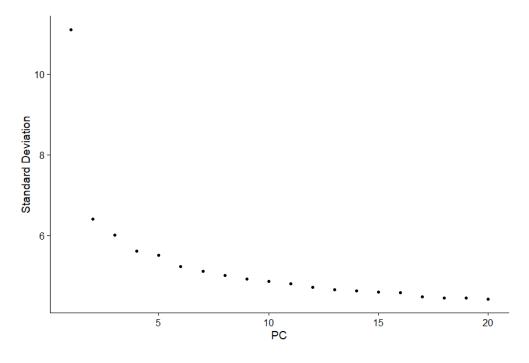


Figure 3: ElbowPlot of the pilot data

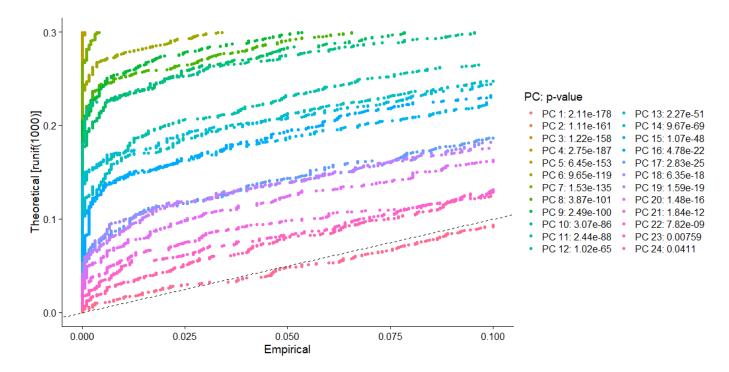


Figure 4: JackStrawPlot of the true training data

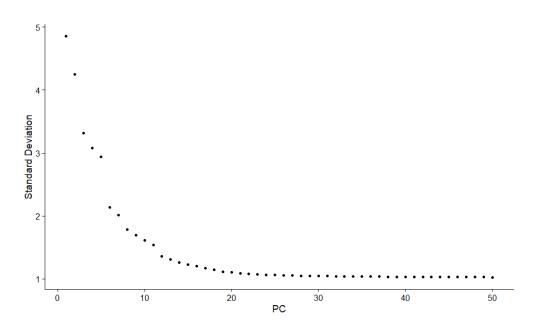


Figure 5: ElbowPlot of the true training data

```
train_data <- read.csv(system.file("extdata", "data_pbmc68k_train_23pc.csv",</pre>
                                      package = "SSD"),row.names=1)
test_data <- read.csv(system.file("extdata", "data_pbmc68k_test_23pc.csv",</pre>
                                    package = "SSD"),row.names=1)
print(table(train_data$phenoid))
#>
                  CD14+_Monocyte
                                                         CD19+_B
#>
                             3717
                                                             3206
#>
                 CD4+/CD25 T Req
                                    CD4+/CD45RA+/CD25- Naive T
#>
                             2712
                                                             3026
#>
             CD4+/CD45RO+_Memory
                                                  CD4+_T_Helper2
#>
                             5759
                                                            11345
                         CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
#>
                            14012
                                                           21875
#>
                CD8+_Cytotoxic_T
                                                       Dendritic
                                                              162
#>
                             1765
print(table(test_data$phenoid))
#>
#>
                  CD14+_Monocyte
                                                         CD19+_B
#>
                              100
                                                              100
#>
                 CD4+/CD25 T Req
                                     CD4+/CD45RA+/CD25- Naive T
#>
                              100
                                                              100
#>
             CD4+/CD45RO+ Memory
                                                  CD4+_T_Helper2
#>
                              100
                                                              100
                         CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
#>
                              100
                                                              100
#>
                                                       Dendritic
                CD8+_Cytotoxic_T
#>
                              100
                                                              100
```

### Task

### With pilot data, draw the plot and determine sample size using the built-in model

In the default setting, we use the built-in *random forest* to train the model. The **index** we use is Adjusted Rand Index (ARI). By default, the size of training data for each class is (30, 60, 90, 120, ..., 540, 570, 600) and the size of test data for each class is 300.

```
x_pilot = pilot_data[,-length(pilot_data)]
y_pilot = pilot_data[,length(pilot_data)]

result_pilot = ssd(x_pilot, y_pilot)
```

The plot is drawn only based on the pilot data and we could use the plot to determine the sample size if we don't have large enough true. We should focus on the trends of the plots because the results produced by synthetic data are usually better than true data, but the trends are pretty similar.

# With pilot data, draw the plot and determine sample size using the self-defined model

If you want to use the model defined by yourself. Then you need to write a "predict\_model" function including your model. The function should take  $train\_data\_x$  and  $train\_data\_y$  as the first two inputs to train the model and then take  $test\_data\_x$  as the third input and return the predicted result of  $test\_data\_x$ . Then you could set model to self and set func to  $predict\_model$ , and run the model using your self-defined

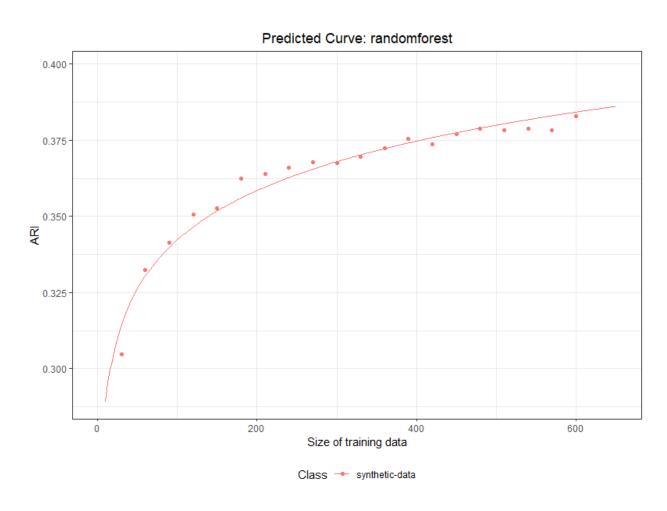


Figure 6: the result plot of the default setting

function.

```
library(e1071)
predict_model <- function(train_data_x, train_data_y, test_data_x){</pre>
    train_data = data.frame(train_data_x, as.factor(train_data_y))
    names(train_data)[length(train_data)] = "class"
    fit_svm<-svm(class~.,data=train_data,probability=TRUE)</pre>
    pred <- predict(fit_svm, test_data_x)</pre>
    return(pred)
}
```

result\_pilot\_self = ssd(x\_pilot, y\_pilot, model="self", func=predict\_model)

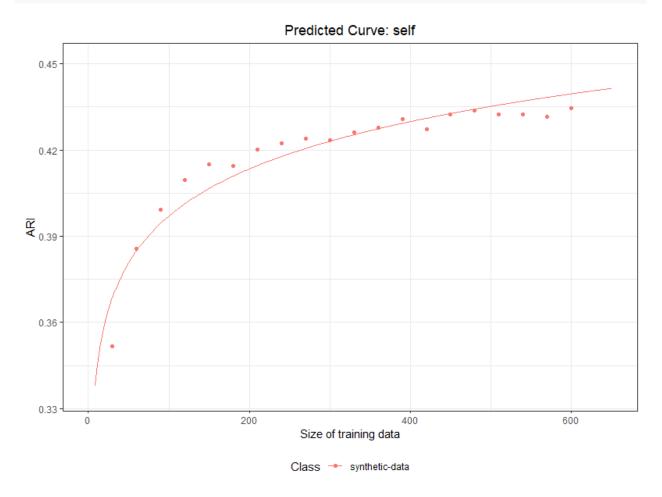


Figure 7: the result plot of the self-defined function setting

You could use the following code to check the function you defined. The result has to be the predicted value of  $test\_data\_x$ .

```
num_class=10
n_train=60
n_{test=100}
for(i in 1:num class){
  class_i_ids = which(train_data$phenoid == names(table(train_data$phenoid))[i])
```

```
train_test_i_ids = sample(class_i_ids, (n_train+n_test))
train_i_data = train_data[train_test_i_ids[1:n_train],]
test_i_data = train_data[train_test_i_ids[(n_train+1):(n_train+n_test)],]

if(i == 1){
    train_data_sample = train_i_data
    test_data_sample = test_i_data
}else{
    train_data_sample = rbind(train_data_sample, train_i_data)
    test_data_sample = rbind(test_data_sample, test_i_data)
}
}

train_data_x = train_data_sample[,-length(train_data_sample)]
train_data_y = train_data_sample$phenoid
test_data_x = test_data_sample[,-length(test_data_sample)]
result = predict_model(train_data_x, train_data_y, test_data_x)
```

### With pilot data and large true data, draw the plot and compare the result

If we have large enough true data and try to compare the plot drawn based on pilot data and the plot drawn based on true data, we could change mode to true and compare the results.

From the results above, even though the values of ARI are different, we can see that the two plots have almost the same trend, which could help us determine the sample size.

We could also try when index is classification error or AMI for this dataset.

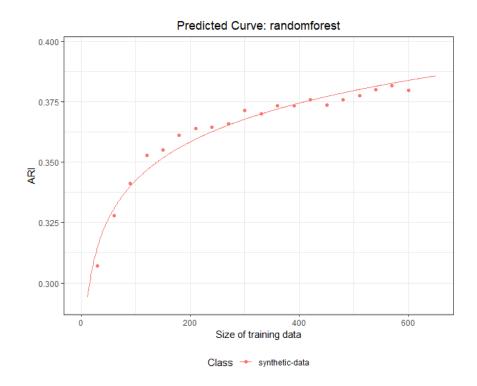


Figure 8: the result plots of comparison using 'ARI' as the index

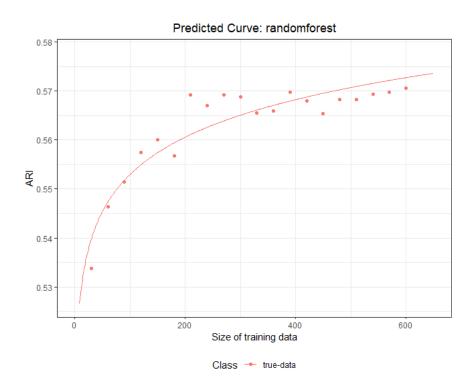


Figure 9: the result plots of comparison using 'ARI' as the index  $\,$ 

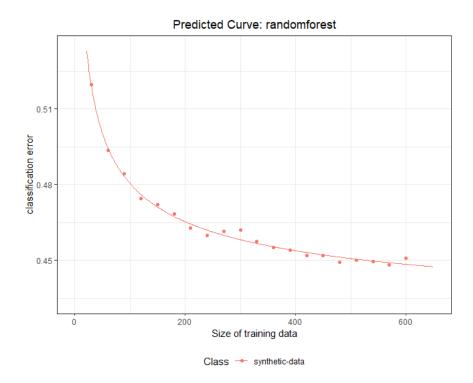


Figure 10: the result plots of comparison using 'classification error' as the index

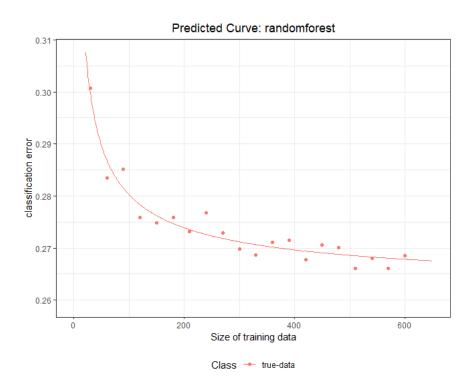


Figure 11: the result plots of comparison using 'classification error' as the index

# Predicted Curve: randomforest 0.46 0.42 0.42 0.38 0.38 0.36 0.36 Class synthetic-data

Figure 12: the result plots of comparison using 'AMI' as the index

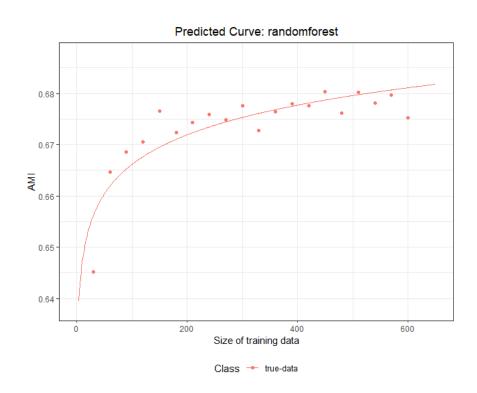


Figure 13: the result plots of comparison using 'AMI' as the index