## Supplementary Material

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#### Introduction

SSD can determine the sample size for our experiment when we only have a few pilot data. Firstly, it can generate different synthetic training datasets and test datasets. Then it will calculate the corresponding different test classification error/ARI/AMI. The final step is to construct the plots in the terms of the sample size and the different metrics (test classification error/ARI/AMI). Users can determine the sample size by the above plots. In this supplementar materil, we use one real dataset to illustrate our algorithm's performance. The workflow can be seen in Figure 1.

### **Preparations**

Before we dive into the main task, we need to load the package and an example dataset for our task. The dataset is the **pbmc\_68k** dataset from 10x Genomics.

We pre-processed the dataset: in this dataset, *phenoid* is the response variable which has 10 classes. We sampled 15 observations from the original dataset for each class and assemble them as the pilot data. We normalize and scale the pilot data at first and then run principal component analysis (PCA) for the pilot data and keep 18 PCs according to *JackStrawPlot* and *ElbowPlot* mentioned in Seurat - Guided Clustering Tutorial. The *JackStrawPlot* and *ElbowPlot* are shown in Figures 2 and 3.

For the whole dataset, we split the it into training data and test data first, then we ran principal component analysis (PCA). We pre-processed the training dataset using the same stratigies and keep 23 PCs according to *JackStrawPlot* and *ElbowPlot* in Figures 4 and 5, and then project the test set onto the reduced feature space obtained during the training.

We put the pro-processed data into our package and we can load them directly.

```
library(SSD)
# load data
pilot_data <- read.csv(system.file("extdata", "data_pbmc68k_pilot_18pc.csv",</pre>
                                     package = "SSD"),row.names=1)
print(table(pilot_data$phenoid))
#>
#>
                  CD14+_Monocyte
                                                        CD19+_B
#>
#>
                 CD4+/CD25_T_Reg
                                    CD4+/CD45RA+/CD25-_Naive_T
#>
            CD4+/CD45RO+_Memory
#>
                                                 CD4+_T_Helper2
#>
                        CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
#>
#>
                CD8+_Cytotoxic_T
                                                      Dendritic
```

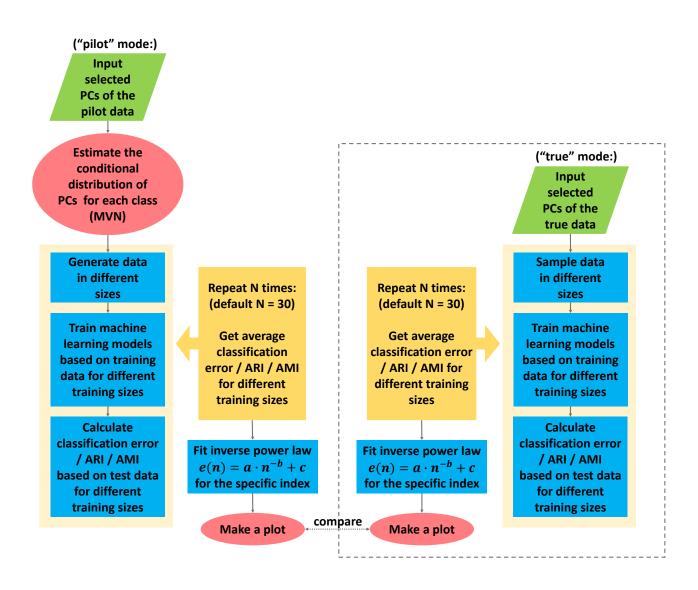


Figure 1: Workflow of the SSD package

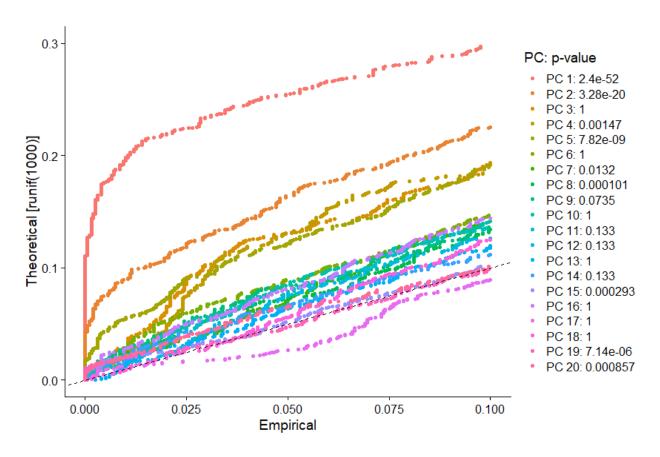


Figure 2: JackStrawPlot of the pilot data

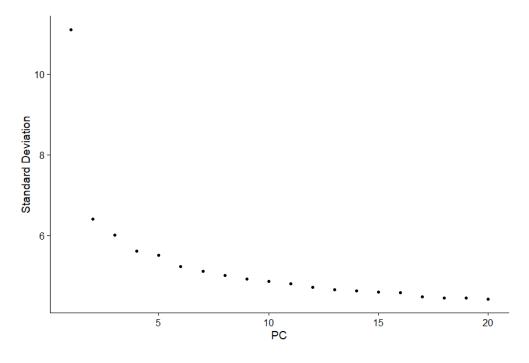


Figure 3: ElbowPlot of the pilot data

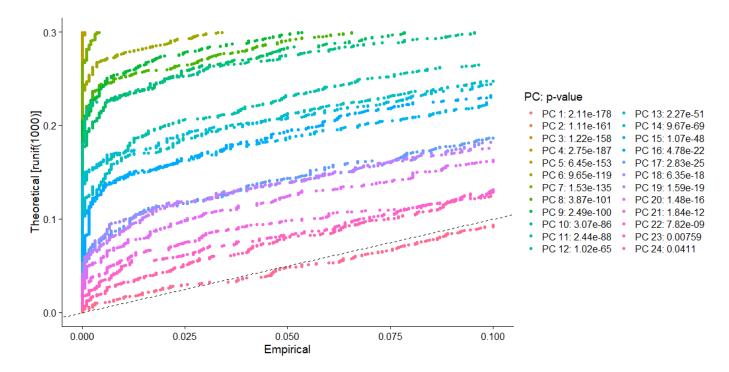


Figure 4: JackStrawPlot of the true training data

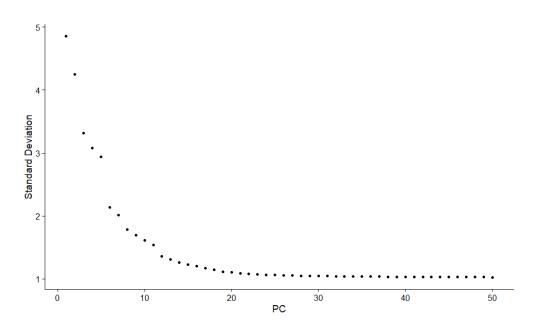


Figure 5: ElbowPlot of the true training data

```
train data <- read.csv(system.file("extdata", "data pbmc68k train 23pc.csv",
                                     package = "SSD"),row.names=1)
test data <- read.csv(system.file("extdata", "data pbmc68k test 23pc.csv",
                                    package = "SSD"),row.names=1)
print(table(train_data$phenoid))
#>
#>
                  CD14+_Monocyte
                                                        CD19+ B
#>
                            3717
                                                           3206
                 CD4+/CD25_T_Reg
#>
                                    CD4+/CD45RA+/CD25-_Naive_T
#>
                             2712
                                                           3026
#>
            CD4+/CD45RO+_Memory
                                                 CD4+_T_Helper2
#>
                            5759
                                                           11345
#>
                        CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
                           14012
                                                          21875
#>
                CD8+_Cytotoxic_T
                                                      Dendritic
                            1765
#>
                                                             162
print(table(test_data$phenoid))
#>
#>
                  CD14+ Monocyte
                                                        CD19+ B
#>
                             100
                                                             100
                                    CD4+/CD45RA+/CD25-Naive_T
#>
                 CD4+/CD25_T_Reg
#>
                             100
                                                             100
#>
             CD4+/CD45RO+_Memory
                                                 CD4+_T_Helper2
#>
                             100
                                                             100
#>
                        CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
                             100
                                                             100
#>
                CD8+_Cytotoxic_T
                                                      Dendritic
                                                             100
#>
```

#### Task

#### With pilot data, draw the plot and determine sample size using the built-in model

In the default setting, we use the built-in *random forest* to train the model. The **index** we use is Adjusted Rand Index (ARI). By default, the size of training data for each class is (30, 60, 90, 120, ..., 540, 570, 600) and the size of test data for each class is 300.

```
x_pilot = pilot_data[,-length(pilot_data)]
y_pilot = pilot_data[,length(pilot_data)]

result_pilot = ssd(x_pilot, y_pilot)
```

The learning curve in Figure 6 is drawn only based on the pilot data and we could use the plot to determine the sample size.

# With pilot data, draw the plot and determine sample size using the self-defined model

If you want to use the model defined by yourself. Then you need to write a "predict\_model" function including your model. The function should take  $train\_data\_x$  and  $train\_data\_y$  as the first two inputs to train the model and then take  $test\_data\_x$  as the third input and return the predicted result of  $test\_data\_x$ . Then you could set model to self and set func to  $predict\_model$ , and run the model using your self-defined

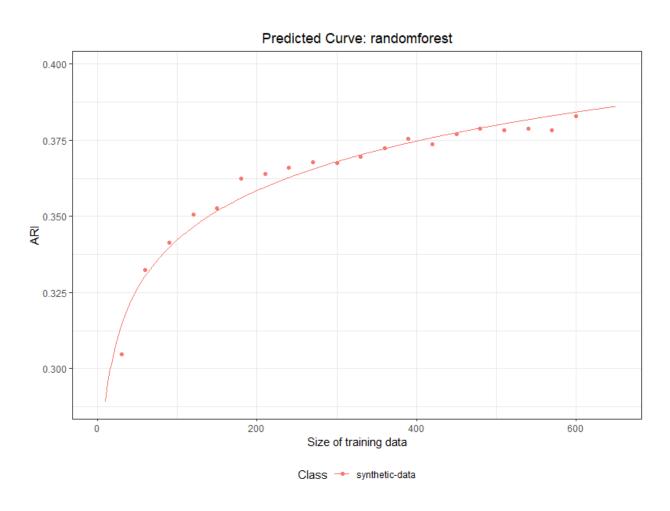


Figure 6: The learning curve of the default setting

function. The learning curve is shown in Figure 7.

```
library(e1071)

predict_model <- function(train_data_x, train_data_y, test_data_x){
    train_data = data.frame(train_data_x, as.factor(train_data_y))
    names(train_data)[length(train_data)] = "class"
    fit_svm<-svm(class~.,data=train_data,probability=TRUE)
    pred <- predict(fit_svm, test_data_x)
    return(pred)
}</pre>
```

result\_pilot\_self = ssd(x\_pilot, y\_pilot, model="self", func=predict\_model)

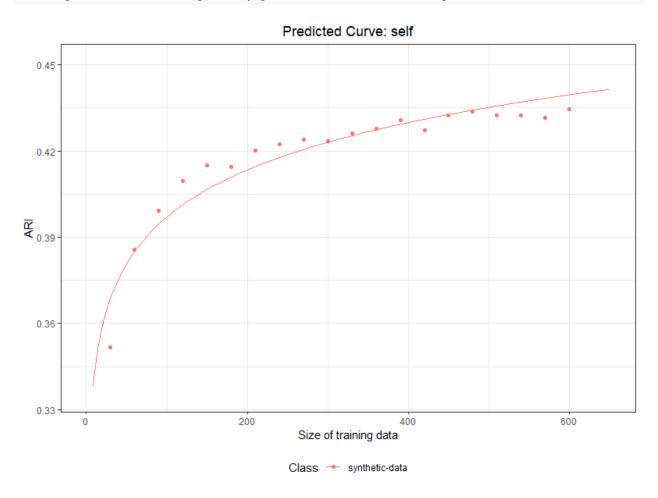


Figure 7: The learning curve of the self-defined function setting

You could use the following code to check the function you defined. The result has to be the predicted value of  $test\_data\_x$ .

```
num_class=10
n_train=60
n_test=100

for(i in 1:num_class){
   class_i_ids = which(train_data$phenoid == names(table(train_data$phenoid))[i])
```

```
train_test_i_ids = sample(class_i_ids, (n_train+n_test))
    train_i_data = train_data[train_test_i_ids[1:n_train],]
    test_i_data = train_data[train_test_i_ids[(n_train+1):(n_train+n_test)],]

if(i == 1){
        train_data_sample = train_i_data
        test_data_sample = test_i_data
    }else{
        train_data_sample = rbind(train_data_sample, train_i_data)
        test_data_sample = rbind(test_data_sample, test_i_data)
    }
}

train_data_x = train_data_sample[,-length(train_data_sample)]

train_data_y = train_data_sample[,-length(test_data_sample)]

result = predict_model(train_data_x, train_data_y, test_data_x)
```

#### With pilot data and large true data, draw the plot and compare the result

Here, we use the large ture data to compare the sythentic learning curve and true learning curve. If we have large enough true data and try to compare the plot drawn based on pilot data and the plot drawn based on true data, we could change mode to true and compare the results.

From the learning curves in Figures 8 and 9, even though the values of ARI are different, we can find that the two plots have almost the same trend, which could help us determine the sample size.

We could also try when **index** is *classification error* or *AMI* for this dataset. The results are shown in Figure 10-13. The graphs also show that the synthetic plots and the true plots have same trend.

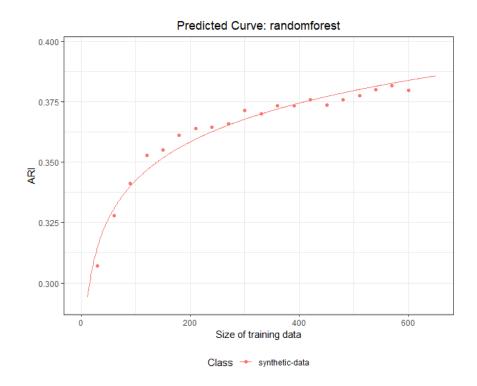


Figure 8: the result plots of comparison using 'ARI' as the index

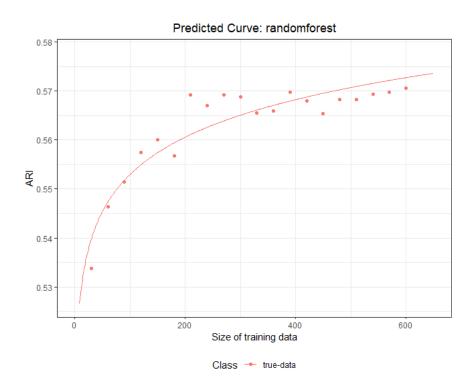


Figure 9: the result plots of comparison using 'ARI' as the index  $\,$ 

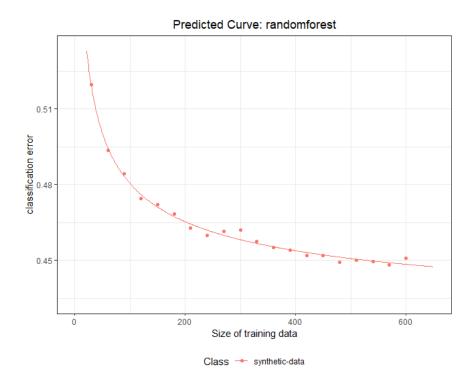


Figure 10: the result plots of comparison using 'classification error' as the index

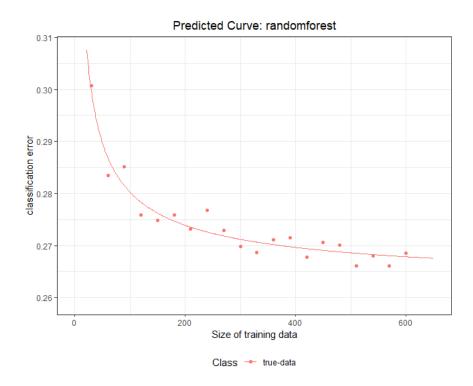


Figure 11: the result plots of comparison using 'classification error' as the index

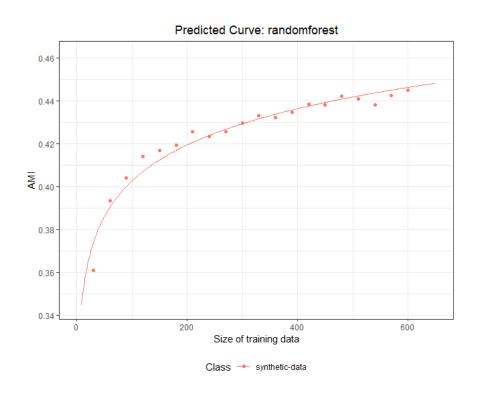


Figure 12: the result plots of comparison using 'AMI' as the index

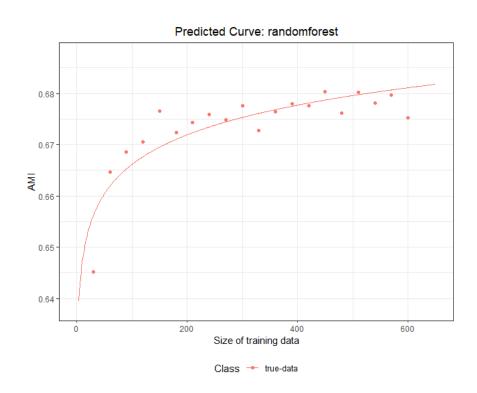


Figure 13: the result plots of comparison using 'AMI' as the index