SSD

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Introduction

SSD can determine the sample size for our experiment when we only have a few pilot data. Firstly, it can generate different synthetic training datasets and test datasets. Then it will calculate the corresponding different test classification error/ARI/AMI. The final step is to construct the plots in the terms of the sample size and the different metrics (test classification error/ARI/AMI). Users can determine the sample size by the above plots.

Preparations

Before we dive into the main task, we need to load the package and an example dataset for our task. The dataset we use is the **pbmc_68k** dataset from 10x Genomics.

We pre-processed the dataset: In this dataset, *phenoid* is the y label which has 10 classes. We sampled 15 observations from the original dataset for each class and assemble them as the pilot data, we normalize and scale the pilot data at first and then run principal component analysis (PCA) and keep 18 PCs according to JackStrawPlot and ElbowPlot mentioned in Seurat - Guided Clustering Tutorial. The JackStrawPlot and ElbowPlot are shown below:

For the whole dataset, we pre-processed it using the same stratigies and keep 23 PCs according to *JackStrawPlot* and *ElbowPlot*.

We put the pro-processed data into our package and we can load them directly.

```
library(SSD)
pilot_data <- read.csv(system.file("extdata", "data_pbmc68k_pilot_18pc.csv",</pre>
                                     package = "SSD"),row.names=1)
print(table(pilot_data$phenoid))
#>
                  CD14+_Monocyte
                                                         CD19+_B
#>
                                                              15
                 CD4+/CD25_T_Req
                                    CD4+/CD45RA+/CD25-_Naive_T
#>
#>
#>
             CD4+/CD45RO+_Memory
                                                 CD4+_T_Helper2
#>
                        CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
#>
#>
                               15
#>
                CD8+ Cytotoxic T
                                                      Dendritic
#>
                                                              15
train_data <- read.csv(system.file("extdata", "data_pbmc68k_train_23pc.csv",</pre>
```

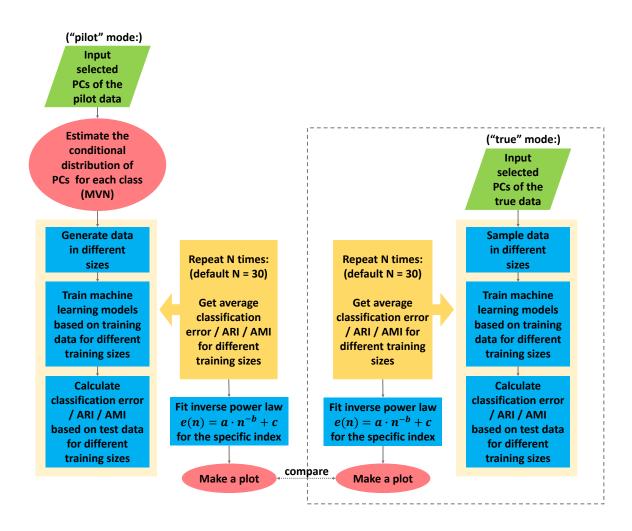


Figure 1: Workflow of the SSD package

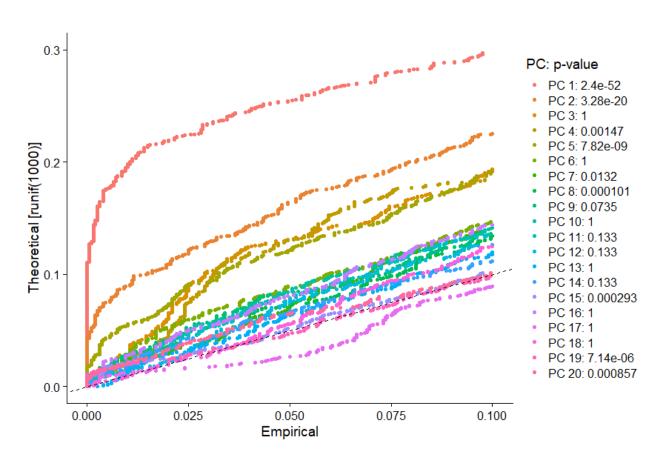


Figure 2: JackStrawPlot of the pilot data

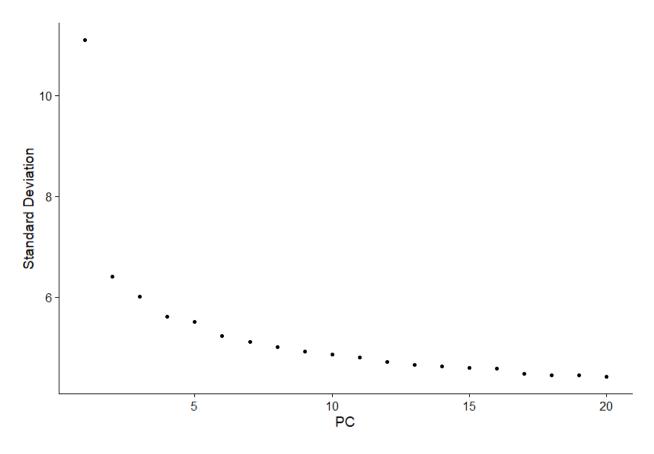


Figure 3: ElbowPlot of the pilot data

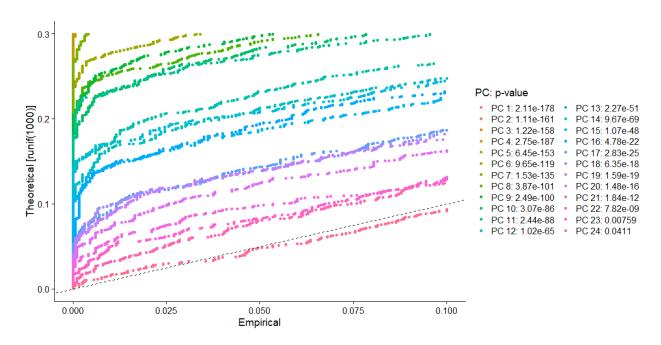


Figure 4: JackStrawPlot of the whole dataset

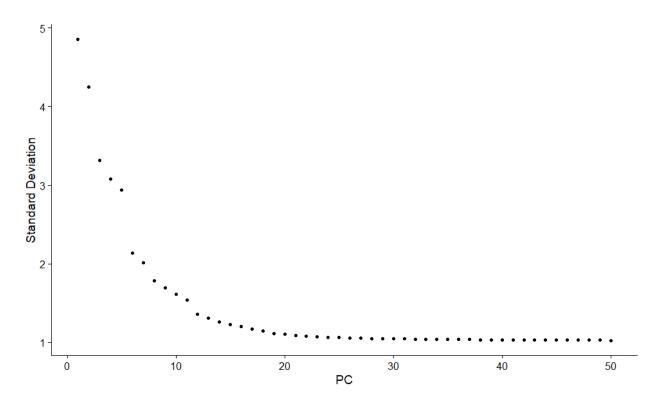


Figure 5: ElbowPlot of the whole dataset

```
package = "SSD"),row.names=1)
test_data <- read.csv(system.file("extdata", "data_pbmc68k_test_23pc.csv",</pre>
                                    package = "SSD"),row.names=1)
print(table(train_data$phenoid))
#>
                  CD14+_Monocyte
                                                         CD19+_B
#>
                             3717
                                                            3206
                 CD4+/CD25_T_Reg
#>
                                    CD4+/CD45RA+/CD25-_Naive_T
#>
                             2712
                                                            3026
#>
             CD4+/CD45RO+_Memory
                                                 CD4+_T_Helper2
#>
                             5759
                                                           11345
#>
                         CD56+_NK CD8+/CD45RA+_Naive_Cytotoxic
                            14012
#>
                                                           21875
#>
                CD8+_Cytotoxic_T
                                                       Dendritic
#>
                             1765
                                                             162
print(table(test_data$phenoid))
#>
#>
                  CD14+_Monocyte
                                                         CD19+_B
#>
                                                             100
                              100
#>
                 CD4+/CD25_T_Reg
                                    CD4+/CD45RA+/CD25-Naive_T
#>
                              100
                                                             100
#>
             CD4+/CD45RO+_Memory
                                                 CD4+_THelper2
#>
#>
                        {\it CD56+\_NK~CD8+/CD45RA+\_Naive\_Cytotoxic}
#>
                              100
                                                             100
#>
                CD8+_Cytotoxic_T
                                                       Dendritic
```

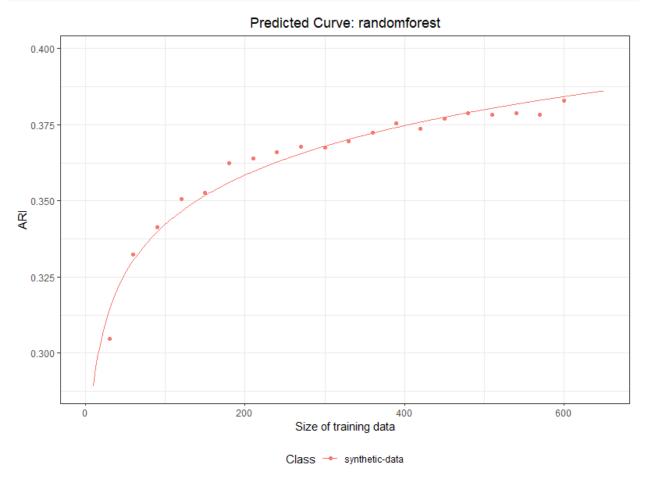
#> 100 100

Task

With pilot data, draw the plot and determine sample size using the built-in model

In the default setting, we use the built-in *random forest* to train the model. The **index** we use is Adjusted Rand Index (ARI). By default, the size of training data for each class is (30, 60, 90, 120, ..., 540, 570, 600) and the size of test data for each class is 300.

```
x_pilot = pilot_data[,-length(pilot_data)]
y_pilot = pilot_data[,length(pilot_data)]
result_pilot = ssd(x_pilot, y_pilot)
```



The plot is drawn only based on the pilot data and we could use the plot to determine the sample size if we don't have large enough true. We should focus on the trends of the plots because the results produced by synthetic data are usually better than true data, but the trends are pretty similar.

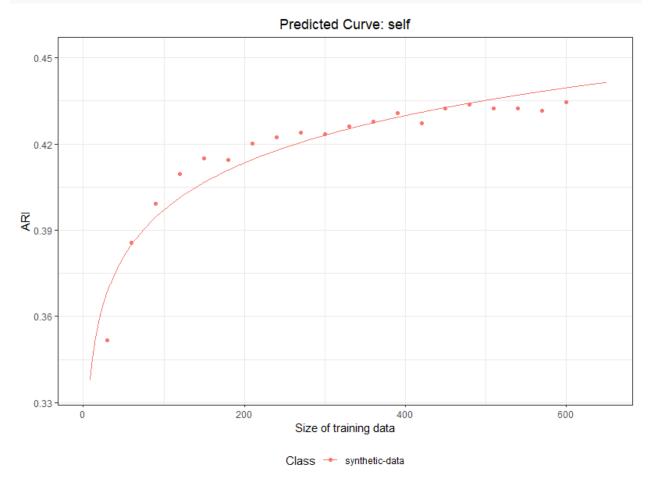
With pilot data, draw the plot and determine sample size using the self-defined model

If you want to use the model defined by yourself. Then you need to write a "predict_model" function including your model. The function should take $train_data_x$ and $train_data_y$ as the first two inputs to

train the model and then take test data x as the third input and return the predicted result of test data x. Then you could set model to self and set func to predict_model, and run the model using your self-defined function.

```
library(e1071)
predict_model <- function(train_data_x, train_data_y, test_data_x){</pre>
    train_data = data.frame(train_data_x, as.factor(train_data_y))
    names(train_data)[length(train_data)] = "class"
    fit_svm<-svm(class~.,data=train_data,probability=TRUE)</pre>
    pred <- predict(fit_svm, test_data_x)</pre>
    return(pred)
}
```

result_pilot_self = ssd(x_pilot, y_pilot, model="self", func=predict_model)



You could use the following code to check the function you defined. The result has to be the predicted value of $test_data_x$.

```
num_class=10
n_train=60
n_{test=100}
for(i in 1:num_class){
  class_i_ids = which(train_data$phenoid == names(table(train_data$phenoid))[i])
```

```
train_test_i_ids = sample(class_i_ids, (n_train+n_test))
train_i_data = train_data[train_test_i_ids[1:n_train],]
test_i_data = train_data[train_test_i_ids[(n_train+1):(n_train+n_test)],]

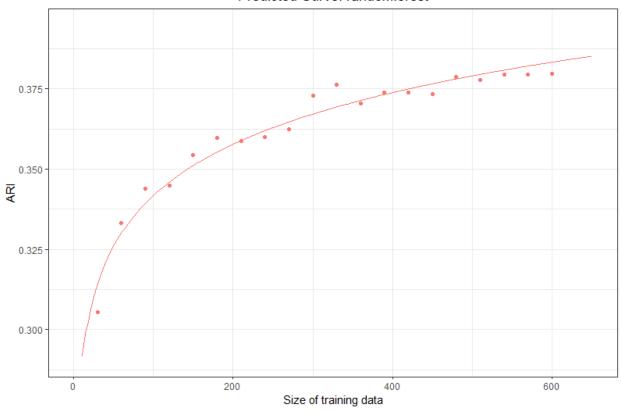
if(i == 1){
    train_data_sample = train_i_data
    test_data_sample = test_i_data
}else{
    train_data_sample = rbind(train_data_sample, train_i_data)
    test_data_sample = rbind(test_data_sample, test_i_data)
}

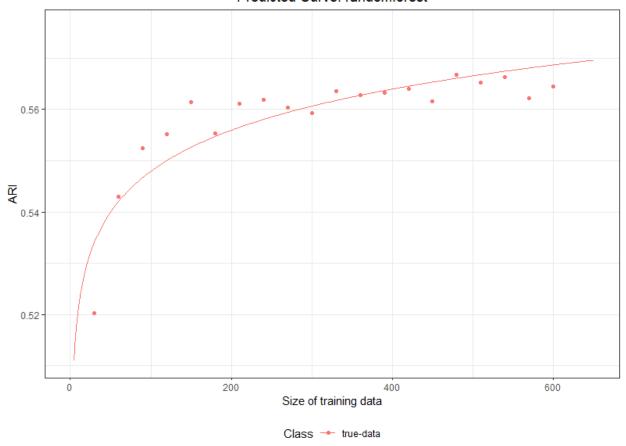
train_data_x = train_data_sample[,-length(train_data_sample)]
train_data_y = train_data_sample[,-length(test_data_sample)]

result = predict_model(train_data_x, train_data_y, test_data_x)
```

With pilot data and large true data, draw the plot and compare the result

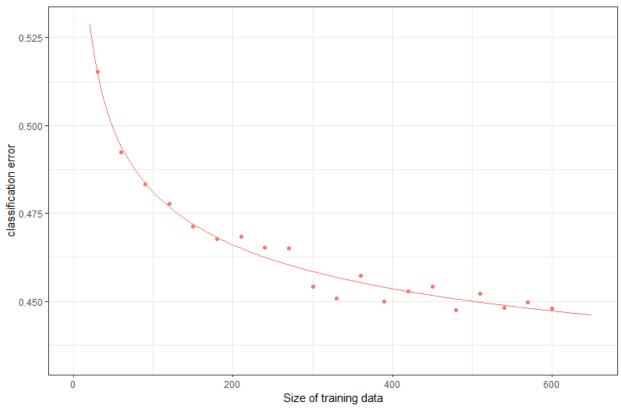
If we have large enough true data and try to compare the plot drawn based on pilot data and the plot drawn based on true data, we could change mode to true and compare the results.

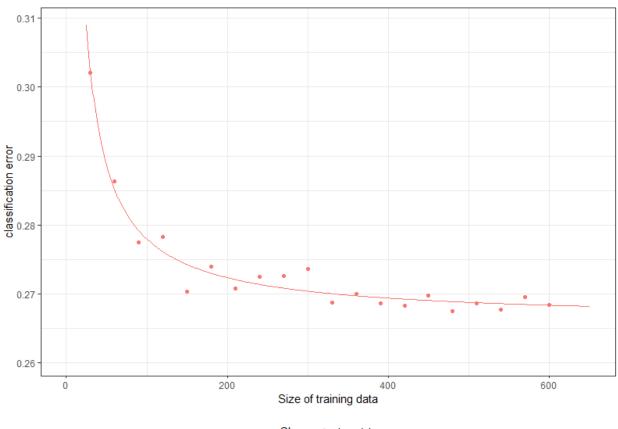




From the results above, even though the values of ARI are different, we can see that the two plots have almost the same trend, which could help us determine the sample size.

We could also try when index is classification error or AMI for this dataset.





Class - true-data

