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REVIEW PAPER

Identifying target traits and molecular mechanisms for wheat breeding under a changing climate

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Abstract

Global warming is causing changes in temperature at a rate unmatched by any temperature change over the last 50 million years. Crop cultivars have been selected for optimal performance under the current climatic conditions. With global warming, characterized by shifts in weather patterns and increases in frequency and magnitude of extreme weather events, new ideotypes will be required with a different set of physiological traits. Severe pressure has been placed on breeders to produce new crop cultivars for a future, rapidly-changing environment that can only be predicted with a great degree of uncertainty and is not available in the present day for direct experiments or field trials. Mathematical modelling, therefore, in conjunction with crop genetics, represents a powerful tool to assist in the breeding process. In this review, drought and high temperature are considered as key stress factors with a high potential impact on crop yield that are associated with global warming, focusing on their effects on wheat. Modelling techniques are described which can help to quantify future threats to wheat growth under climate change and simple component traits that are amenable to genetic analysis are identified. This approach could be used to support breeding programmes for new wheat cultivars suitable for future environments brought about by the changing climate.

Key words: Crop modelling, deconvoluting complex traits, drought and heat stress, G×E interactions, wheat genetics.

Introduction

The largest temperature changes of the past million years happened as a result of the glacial cycles, during which global mean temperature changed by 4–7 °C between the ice ages and the warm interglacial periods (Solomon *et al.*, 2007). The changes in temperature during the last 450 000 years at Vostok station in east Antarctica, as derived from deuterium isotopic measurements on ice cores, are shown in Fig. 1 (Jouzel *et al.*, 1987; Jouzel *et al.*, 1993; Jouzel *et al.*, 1996; Petit *et al.*, 1999). Although changes in temperature were large in the past, global warming at the end of each ice age was a relatively gradual process taking approximately 5000 years (Solomon *et al.*, 2007). The current rate of global climate change is much more rapid. If, as predicted, an increase of 5 °C in global mean temperature is achieved by the end of the century (this is the upper end of the range

predicted by Global Climate Models in the IPCC 4th Assessment Report), then the Earth will have experienced a rate of change which could not be matched by any global mean temperature increase of the last 50 million years (Solomon *et al.*, 2007).

Breeders select new cultivars of agricultural crops that are better suited to a specific environment utilizing available resources in the most optimal way. However, cultivars that are recommended for use at present might not be suitable if the climate changes. Breeding for a new cultivar usually takes 10–12 years, if the target traits are known and the environment in which to test new lines is available. Faced with the prospect of a rapidly changing climate, breeders do not have access to the climatic conditions of even the near future in which to carry out field trials, and they do not

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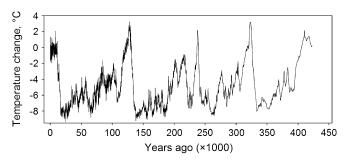


Fig. 1. Temperature changes during the last several glacial cycles from the Vostok ice core in Antarctica as derived from deuterium isotopic measurements on ice cores (figure created from a dataset published in Petit *et al.*, 1999).

know which wheat traits might be important in 15–25 years time. For example, breeding for drought tolerance in wheat could be justified for the current climate in the UK. However, in the future, heat stress at flowering, resulting in substantial yield losses, could be a greater threat to wheat production.

Most Global Climate Models predict increases in summer dryness and winter wetness in most parts of the northern middle and high latitudes during this century (Fig. 2). They also predict a substantial increase in temperature and in the frequency and magnitude of extreme weather events (Solomon et al., 2007). Using climate projections for the UK, based on the UKCIP02 climate modelling experiments completed by the Hadley Centre (Hulme et al., 2002), it has been shown that heat waves will increase substantially in frequency (by an order of magnitude), length, severity, and peak temperature by the end of the century (Semenov, 2007). Even isolated incidents of extreme high temperature during sensitive stages of crop development, flowering for example, could reduce grain yield significantly, while a continuous period of extreme high temperature could ruin a harvest altogether.

With global warming, changes in climate and extreme weather events are likely to impact agricultural crops, but it is not yet clear what the extent of yield losses will be and whether breeding for new, stress-tolerant cultivars will be required. In this review, drought and high temperature are considered; two key stress factors associated with global warming that have a high potential impact on crop yield, focusing on their effects on wheat. Modelling techniques are described that can help to quantify future threats to wheat growth under climate change and to identify the component wheat traits and genes that can be used for wheat genetic improvement. These techniques will support breeding programmes in a rapidly changing climate.

Predicting the impacts of drought and heat stress under climate change

Crop simulation models are used increasingly in basic and applied research in plant sciences (Passioura, 1996; Sinclair and Muchow, 2001; Debaeke and Aboudrare, 2004; Porter and Semenov, 2005). Simulation models provide the best-

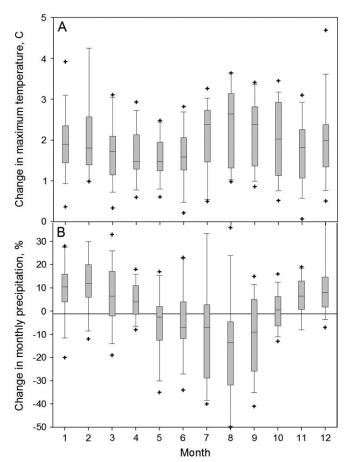


Fig. 2. Absolute changes in monthly mean maximum temperature (A) and relative changes in monthly mean precipitation (B) as predicted by global climate models used in the latest IPCC 4th Assessment Report for the SRES-A1B emission scenario for 2050s compared with the baseline scenario (1960–1990) at Rothamsted, UK. The boundaries of the box indicate the 25th and 75th percentiles, the line within the box marks the median, whiskers below and above the box indicate the 10th and 90th percentiles and outliers show minimum and maximum values.

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known approach for integrating our understanding of complex plant processes that are influenced by weather and other environmental factors. They are useful in guiding the direction of fundamental research by providing quantitative predictions and highlighting gaps in our knowledge (Tardieu, 2003; Hansen, 2005; Semenov *et al.*, 2007). Combined with high-resolution climate scenarios based on Global Climate Models, crop simulation models are extensively used to assess the impact of climate change and to identify potential future threats (Semenov *et al.*, 1993, 1996; Jamieson *et al.*, 2000; Ewert *et al.*, 2002; Carbone *et al.*, 2003; Richter and Semenov, 2005; Olesen *et al.*, 2007).

In the following example, the relative importance of drought and high temperature stress on wheat yields under climate change has been quantified using a crop simulation model combined with high resolution climate change scenarios (Semenov, 2009). According to the UKCIP02 projections, the reduction of precipitation during summer and the substantial increase in maximum temperature

during spring and summer could be major stress factors for wheat production in the UK in the future (Hulme et al., 2002).

In an experiment on the effects of CO₂ and temperature on grain yield, Mitchell and co-workers (1993) observed that a temperature of 27 °C or higher half-way through anthesis, when 50% of the ears in a population had flowered, resulted in a high number of sterile grains. Although the effect of reduced grain numbers on final yield could be compensated for during grain-filling by the production of larger grains, the yield losses could still be high. In another study, Wheeler et al. (1996) used temperature-gradient tunnel systems to demonstrate that a temperature of 31 °C or higher prior to anthesis could reduce the number of grains per ear considerably, with a concomitant effect on grain yield. A similar result was obtained by Ferris et al. (1998). These studies provide convincing evidence that a short episode of high temperature around the time of flowering can reduce wheat grain yield substantially (Wheeler et al., 2000). In our example study (Semenov, 2009), heat stress around flowering was quantified by the probability P_{ant}^{T27} of the maximum temperature exceeding 27 °C at least once during the 10 d after the beginning of anthesis.

Considerable yield losses can also be the outcome of prolonged drought over the growing season. The reduction in grain yield due to water stress can be quantified by the drought stress index (DSI), which is calculated as $DSI=1 - Y_{WL}/Y_{Pot}$, where Y_{WL} and Y_{Pot} are simulated water-limited grain yields and potential grain yields, respectively. The potential yield is calculated for a crop for which daily water demands are satisfied. The 95-percentile for DSI, $Q_{\rm DSI}^{95}$, represents the level of yield losses due to water stress that can be expected, on average, once every 20 years.

Daily site-specific climate scenarios were constructed by using the LARS-WG stochastic weather generator (Semenov, 2007), which downscaled UKCIP02 predictions (Hulme et al., 2002) to the resolution required by crop simulation models. Weather generators have been adopted in climate change studies as a computationally inexpensive tool to generate climate scenarios with high temporal and spatial resolutions, based on the output from Global Climate Models (Wilks, 1992; Barrow and Semenov, 1995). Two winter wheat cultivars, cv. Avalon and cv. Mercia, were analysed; both are obligate winter wheat cultivars with moderate to weak day-length responses that have been calibrated previously using field experiments in the UK (Wolf et al., 1996; Ewert et al., 2002; Lawless et al., 2005). Mercia is a late-flowering cultivar that matures, on average, 2 weeks later than cv. Avalon.

The baseline scenario, representing 1960-1990, was compared with the climate predictions for the 2050s based on the high emission scenario (2050HI) (in other words the climate predicted if greenhouse gas emissions remain 'business as usual'; Nakicenovic and Swart, 2000). The predicted increases in maximum temperature for the 2050HI scenario were between 2 °C and 4 °C and the probability of the maximum temperature exceeding 27 °C around flowering would be affected significantly by such large increases in

temperature mean. However, this would be offset somewhat because wheat development is driven by thermal time, so in a warmer climate wheat flowering would occur earlier in the season, before the summer temperature peak. Table 1 shows the average day (day of the year) of flowering and average maximum temperature \bar{T}_{max} for this day for cvs Avalon and Mercia for the baseline and 2050HI scenarios. The increase of \bar{T}_{max} at flowering in the 2050HI scenario compared with the baseline was only 0.35 for cv. Avalon and 1.06 °C for cv. Mercia, because the wheat was predicted to flower almost 2 weeks earlier. The relative changes between the baseline and 2050HI scenarios in probability P_{ant}^{T27} for earlyflowering cv. Avalon were significantly smaller in magnitude (Fig. 3c) than changes for late-flowering cv. Mercia, which increased more than 2-fold for most of England and Wales (Fig. 3d).

To calculate changes in 95-percentiles of the DSI distribution, $Q_{\rm DSI}^{95}$, 150 years of daily weather for the baseline and 2050HI climate scenarios were used (Fig. 3a, b). For both cultivars, $Q_{\rm DSI}^{95}$ was predicted to decrease with a significantly larger decrease for early-flowering cv. Avalon compared with late-flowering cv. Mercia. For future scenarios, despite the fact that monthly precipitation was predicted to be lower for the summer months, $Q_{\rm DSI}^{95}$ was predicted to decrease in England and Wales. Two factors contributed to this decrease. The first was wheat phenology: both cultivars were predicted to mature almost 3 weeks earlier for the 2050HI scenarios compared with the baseline scenario (Table 1). Because soil water deficit increases towards the end of crop growth, a crop can avoid the most severe drought stress by maturing earlier. The second factor was related to changes in the precipitation pattern. Although summer was predicted to be drier for the UKCIP02 projections, winter was predicted to be wetter. Additional precipitation during winter and early spring would be stored in the soil (the exact amount of stored water would depend on soil available water capacity) and made available to the crop during late spring and early summer.

This study demonstrates that the impacts of changing climate on wheat can be counter-intuitive and that the severity of the impact depends strongly on cultivar characteristics and spatial and temporal patterns of climate changes. Drought is the most significant environmental stress in agriculture worldwide and improving yield in water-limited environments is a major goal of plant breeding (Tambussi et al., 2007; Cattivelli et al., 2008). Some researchers suggest that the impact of drought will increase with climate change (Foulkes et al., 2007; Witcombe et al., 2008) and emphasize the importance of breeding for drought-tolerant crops. The analyses presented here, however, predict that the impact of drought stress on two existing wheat cultivars in the UK will decrease with climate change. Therefore, drier and warmer summers, which are expected in the UK, will not necessarily mean additional yield losses due to water stress. An increase in the frequency of heat stress around flowering as a result of global warming represents a greater risk for sustainable wheat production in the UK (Barnabás et al., 2008).

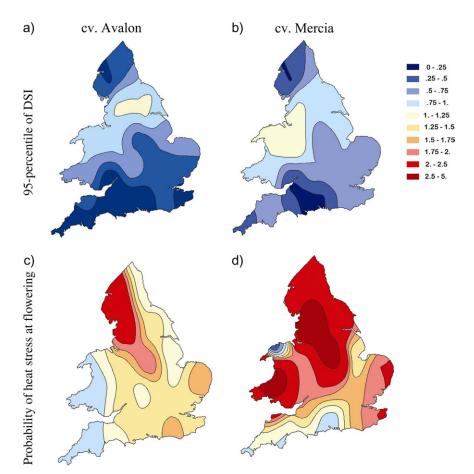


Fig. 3. Relative changes in 95-percentile for drought stress index and the probability of maximum temperature exceeding 27 °C around flowering for early developing cv. Avalon and late developing cv. Mercia for the 2050s high emission scenario compared with the 1960-1990s for England and Wales, UK.

Table 1. The average date of flowering and maturity and the average maximum temperature $\overline{T}_{\rm max}$ at flowering calculated for cv. Avalon and cv. Mercia and for baseline and 2050HI climate scenarios at Rothamsted

Cultivar		Baseline	2050HI
Avalon	Flowering	9 June	24 May
	Maturity	8 Aug	18 July
	$\bar{T}_{ m max}$ at flowering (°C)	18.50	18.85
Mercia	Flowering	19 June	5 June
	Maturity	23 Aug	2 Aug
	\bar{T}_{max} at flowering (°C)	19.36	20.42

Deconvoluting complex wheat traits

Even when abiotic threats are known, it may be difficult to progress with crop breeding if the target for improvement is a complex trait, such as water-use efficiency or thermotolerance. It may be difficult to establish relationships between such traits and simple component traits, which can be assigned to quantitative trait loci (QTL), markers or single genes and targeted by breeders and geneticists. Crop simulation models may be useful here to deconvolute a complex trait into a ranked list of simpler traits suitable

for further genetic analysis and breeding. As an example, there follows a demonstration of how modelling works for water-use efficiency in wheat (Semenov *et al.*, 2009).

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A considerable amount of work has been done to develop ways of improving crop yield under limited water supply (Collins et al., 2008), in particular for wheat (Richards et al., 2002; Condon et al., 2004). The term 'water use efficiency' (WUE) has many definitions, depending on the range of biological processes studied and the temporal scale. At the leaf level, WUE can be defined as the ratio of instantaneous carbon dioxide assimilation over transpiration (Jones, 1992), whereas at the crop level, WUE is usually defined as the ratio of final crop yield over total seasonal evapotranspiration. It has been shown experimentally that crop WUE is sensitive to environmental conditions, with humidity being particularly important because it directly affects stomatal conductance (Day et al., 1978; Lawlor et al., 1981). However, improving WUE in itself is of limited interest if the crop produces a very low grain yield and it may be more sensible to explore the potential of modifying other, less complex physiological traits to improve wheat yield in water-limited environments (Sinclair and Muchow, 2001).

Plants exhibit different strategies for survival and growth under conditions of limited water supply or high

evaporative demands (Jones, 2004; Tambussi et al., 2007). Plants may avoid the effects of severe drought by developing early and shortening the growing season, by conserving available water through reducing leaf size and regulating stomatal closure, or by extracting water more efficiently with an improved root system. By conserving water or extracting it more efficiently they avoid becoming water-stressed despite the drought conditions. Plants may also develop improved tolerance to water deficit; in other words they become water-stressed but are able to maintain productivity anyway. The Sirius wheat simulation model was used to quantify the effects of variations in both groups of traits on wheat performance under waterlimited conditions at two contrasting European sites representative of major wheat-growing regions (Semenov et al., 2009).

Water stress avoidance traits: phenology

Modifying the duration of crop photosynthesis and its timing in relation to seasonal variations of solar radiation may have significant effects on yield (Akkaya et al., 2006; Richards, 2006). The phyllochron (P) is the thermal time required for the appearance of successive leaves, and is a major driver of phenological development (Jamieson et al., 1995, 1998a, 2007). By modifying P, the rate of crop development is altered and, therefore, the date of flowering and maturity. The rate at which the leaves appear, determined by P, will influence the rate at which the canopy develops and water is depleted; increasing this rate shortens the duration of the vegetative development phase. Genetic variations of P of up to 20% have been reported for wheat (Mossad et al., 1995; Ishag et al., 1998).

Increasing the duration of the grain-filling period (GFD) has also been suggested as a possible trait for increasing grain yield in wheat (Evans and Fischer, 1999). The rate of grain growth depends on intercepted radiation during grainfilling, and the recycling of water-soluble carbohydrates that are stored in the stem or produced by the breakdown of leaf tissue during senescence. In the Sirius model, the labile carbohydrate pool is calculated as a fixed 25% of biomass at anthesis, and is translocated to the grain in thermal time at a rate proportional to GFD^{-1} . Increasing GFD has the potential to increase the amount of radiation intercepted by the crop during grain-filling and, therefore, grain yield. However, if the end of grain-filling were to occur earlier as a result of leaves senescing, grain yield would decrease, both through the loss of intercepted radiation and the fact that there would be insufficient time for all of the labile carbohydrate pool to be translocated to the grain. Genetic variations of GFD for wheat of up to 40% have been observed (Robert et al., 2001; Charmet et al., 2005; Akkaya et al., 2006).

Water stress avoidance traits: root growth and distribution

In the Sirius model, it is assumed that only a proportion of available soil water can be extracted from the soil layer by

the plant on any day. A model parameter λ describes the efficiency of the root system to extract water through the vertical soil profile. This simple approach to soil water extraction by wheat crops has been proven to be robust for contrasting environments and soil conditions (Meinke et al., 1997; Jamieson et al., 1998b), as well as for several other crop species (Robertson et al., 1993; Dardanelli et al., 2004). A faster water uptake would reduce the stress experienced by the plant in anticipation of additional water coming in the form of precipitation or irrigation later in the season. In environments with a low probability of additional water, however, an alternative strategy that reduced water uptake would be less risky and might achieve a higher yield (Manschadi et al., 2006).

Root vertical extension rate (RVER) is constant in Sirius (Jamieson et al., 1998b). In the model, roots can potentially grow up to a maximum depth and stop growing at anthesis. Allowing roots to extend at a higher rate would enable plants to capture water more efficiently and prevent water leaching to deeper, unavailable soil layers.

Water stress avoidance traits: canopy expansion

The Sirius model describes the canopy as a series of layers associated with individual, main-stem leaves, and simulates the effect of tillering only through the potential size of any layer (Lawless et al., 2005). Individual leaf layers are described according to their lifetime, growth and senescence periods, and their potential maximum surface areas. Sirius uses one cultivar parameter, α_{max} , the maximum potential surface area of the largest culm leaf layer, to describe potential growth of all culm leaf layers. By varying $\alpha_{\text{max}},$ the rate of canopy expansion and maximum achievable canopy size is changed. This, in turn, changes the pattern of light interception and transpiration and therefore affects grain yield. The reported range of genetic variations for flag leaf area under unlimited water and nitrogen supplies is up to 40% (Fischer et al., 1998; Shearman et al., 2005).

Water stress tolerance traits

In Sirius, water stress is predicted to affect the growth of the crop in two ways: by limiting production of new biomass and by accelerating leaf senescence. Both of these limitations depend on a stress factor SF, that is calculated daily as a ratio of actual over potential evapotranspiration. Daily production of new biomass decreases proportionally to the drought biomass reduction factor DBF defined as DBF= SF^{β} , where β is currently a cultivar-independent constant.

In the current version of Sirius, the ontogenic rate of leaf senescence is constant in thermal time. This rate could be accelerated by low nitrogen availability (inadequate to sustain either leaf expansion or grain-filling) or water stress. Maintaining green leaf area longer after anthesis results in increases in yield and crop nitrogen (N), if N uptake is also maintained (Austin, 1999; Triboi and Triboi-Blondel, 2002). In the presence of water stress, however, the rate of leaf senescence increases because the daily increment of thermal time is modified by the drought leaf senescence factor, which is a function of *DSF*, a maximum possible acceleration of senescence. Reducing *DSF* should make leaves stay green longer under water deficit.

Effects of trait variations on grain yield

Sirius was run with the perturbed values of parameters described above. Changes in mean yield are presented in Table 2. The two parameters that had the largest impact on grain yield under limited water availability were the response of the plant with respect to leaf senescence (DSF) and biomass accumulation (β) (Fig. 4). Both of these parameters are linked with drought tolerance and are calculated using SF, the water stress factor. The model predicted that, under severe water limitation (DSI>0.6) at a site in Seville, Spain, grain yield of modified cv. Cartaya in which DSF is decreased by 20% could be up to 70% higher than that of the unmodified control, while a 20% reduction of DBF, the drought biomass reduction factor, could increase yield by up to 12% in dry years. There is some experimental data to support this: it has been shown for wheat and sorghum that cultivars with a stay-green trait that allow the plant to retain more green leaves when drought stress is applied post-anthesis have a higher grain yield and biomass than normal cultivars (Richards, 2006; Foulkes et al., 2007).

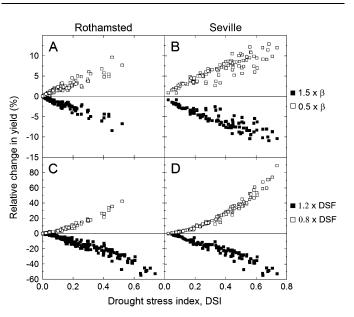
Changes in the maximum potential surface area of culm leaves (α_{max}) were predicted to have a relatively small, but consistent effect on grain yield (Table 2). A 20% increase in α_{max} resulted in mean yield increases of nearly 4% at Rothamsted in the UK and 3.5% at Seville.

An increase of the root vertical extension rate (RVER)was predicted to have a very small positive effect on grain yield at both sites for wheat grown on shallow, sandy soil. Increasing the rate of relative crop water uptake (k) was beneficial at both sites, but changes in mean yield were small (0.6% at Rothamsted and 1.1% at Seville) and would be difficult to measure in practice. With the higher value of k, the plant would take up soil water more quickly and therefore require precipitation later in the growing season. This would seem to be a better strategy at both sites. The vertical distribution of k can be associated with root architecture (Dardanelli et al., 2004), which has been shown to have significant implications for water uptake and crop productivity in water-scarce environments (Manschadi et al., 2006). In experimental and simulation studies, Manschadi and co-workers have shown that differences in root architecture between drought-tolerant wheat (SeriM82) and barley (Mackay) and the standard wheat cv. Hartog arise from two contrasting adaptation strategies. The compact, uniform, and deep root architecture of SeriM82 reduces water use early in the season, but increases access to water during grain-filling. On the other hand, barley cv. Mackay develops a large and shallow root system with greater potential for water extraction early in the season, thereby optimizing the timing of soil water extraction. A simulation analysis using APSIM has shown that modifica-

Table 2. Relative changes (%) in mean yields simulated by Sirius for cv. Avalon at Rothamsted, UK and cv. Cartaya at Seville, Spain for 100 years of the baseline scenario in response to changes in model and cultivar parameters

The * symbol indicates that the differences in means are significant at the 0.05 level calculated using paired t test.

Symbol	Description	Perturbed values	Relative changes in mean yield,%	
			Rothamsted	Seville
Р	Phyllochron	1.2× <i>P</i>	2.1	-0.1
		0.8× <i>P</i>	-14.3*	-1.9
GFD	Grain-filling duration	1.2×GFD	0.6	-3.6*
		0.8×GFD	-6.6*	0.2
RVER	Root vertical extension rate	2×RVER	0.0	0.2*
		0.5×RVER	-0.6*	-4.3*
λ	Efficiency of root to extract	1.4	-2.6*	-5.6*
	water	0.7	0.6*	1.1*
α_{max}	Maximum potential surface	$1.2 \times \alpha_{max}$	3.9*	3.4*
	area of the largest culm leaf layer	0.8×α <i>max</i>	-5.8*	-5.1*
DSF	Maximum acceleration of	1.2×DSF	-5.6*	-17.6*
	leaf senescence in response to water deficit	0.8×DSF	5.8*	19.7*
β	Parameters modifying	$1.5 \times \beta$	-1.8*	-5.1*
	the sensitivity of crops response to water deficit	0.5×β	1.9*	5.7*



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Fig. 4. Relative changes in grain yield versus drought stress index, DSI, calculated for 100 years in response to an increase (closed symbols) or a decrease (open symbols) of scaling exponent of the drought stress factor β (A, B), and of maximum acceleration of leaf senescence DSF (C, D) for cv. Avalon at Rothamsted, UK, and cv. Cartaya at Seville, Spain, simulated by Sirius for sandy soil with 90 mm AWC at each site (Semenov $et\ al.$, 2009).

tion of the rooting pattern could lead to a long-term average gain of 14.5% of yield in dry seasons for a wide range of environments in southern Queensland, Australia (Manschadi *et al.*, 2006).

Another simulation study performed in low-rainfall areas of Southern Australia predicted that yields could be enhanced by an increased depth of rooting (Dreccer et al., 2002). Moreover, this analysis showed that limiting water uptake from deeper soil layers early in the season through greater root-length distribution in the surface soil layers would result in increased availability of water for postanthesis growth, and therefore would improve grain yields. Richards and Passioura (1989) investigated experimentally the effect of root xylem vessel diameter on the ability of roots to extract subsoil water during drought in field trials in eastern Australia. They found that a wheat cultivar with approximately 20% narrower vessels yielded 8% more than the control cultivar in dry environments, whereas yield differences in wetter environments were not significant (Richards and Passioura, 1981; Richards and Passioura, 1989).

Surprisingly, changes in the parameters controlling crop development (P and GFD) were not predicted to improve grain yields consistently either at Rothamsted or Seville (Fig. 5). Indeed, changes in P produced an almost random scatter of yield responses for all levels of DSI. A possible explanation for this is that the cultivars used in the study, cvs Avalon and Cartaya, have an optimal rate of development for climates in the UK and Spain, respectively. According to the results presented here, 20% changes in P would not improve crop performance under water-limited conditions. In an experimental study of the effect of early flowering on drought resistance in two mapping populations, Beaver×Soissons and Rialto×Spark, phenotypic

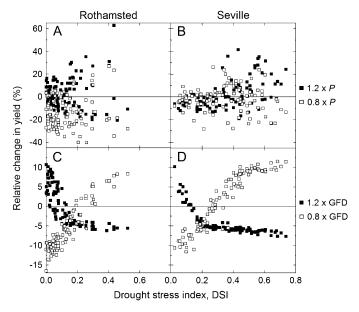


Fig. 5. Relative changes in grain yield versus drought stress index, DSI, calculated for 100 years in response to a 20% increase (closed symbols) or 20% decrease (open symbols) in phyllochron P (A, B) and grain-filling duration GFD (C, D) for cv. Avalon at Rothamsted, UK, and cv. Cartaya at Seville, Spain, simulated by Sirius for sandy soil with 90 mm AWC at each site (Semenov et al., 2009).

correlations amongst the lines between flowering date and yield or yield loss under drought were all non-significant (Foulkes et al., 2007).

Changes in GFD were predicted to produce positive or negative changes in yield, depending on the value of DSI, if the soil were shallow with 90 mm available water capacity (AWC). For wet years with low DSI, an increase in GFD would improve yield.

This analysis illustrates how crop simulation models can be used to explore the relationships between simple and complex traits. By modelling crop growth in variable environments, the stability of these relationships can be tested and a better understanding of the behaviour of a complex trait in response to changes in simple, underlying component traits can be achieved. Attempts to identify QTL for better yields under limited water supplies that are based solely on a limited number of experiments may lead to incorrect conclusions for those traits that demonstrate highly variable responses in different environments. For example, as shown in Fig. 5, for many 'trait-individual year' combinations, trait effects on yield associated with an individual year can change sign (negative to positive) and magnitude.

Linking model parameters with QTL and genes

Crop simulation models integrate the current understanding of crops derived from physiological studies, but very few, if any, incorporate knowledge derived from genetic studies (White and Hoogenboom, 2003). Crop models include cultivar parameters that have been derived by inference from phenotypic characteristics measured in different environments, using calibration procedures with subsequent validation. The criteria for success in using crop models as an integral aspect of crop genetics research and breeding is the ability to associate model cultivar parameters with measurable traits and resolvable QTL. If this link is established, then genetics research will benefit from crop modelling by focusing the search for genes and QTL for those component traits that are identified by modelling analysis as having the best potential (Tardieu, 2003; Quilot et al., 2005; Tardieu et al., 2005).

The next step in utilizing crop models for genetic research would be the development of a theoretical framework that allows reliable predictions of the phenotypic consequences when making alterations to the genetic make-up of a plant (Hammer et al., 2005, 2006). Progress in the design of molecular breeding strategies for complex traits would be greatly accelerated as a result (Cooper et al., 2005). One of the possible approaches is to link phenotypic consequence to changes in genetics via stable associations with crop model parameters. This approach was used to study sorghum breeding scenarios for a broad range of waterlimited environments in Australia using APSIM (Hammer et al., 2005). Fifteen QTL for four component traits of grain yield were defined and the phenotypic variation associated

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with these QTL was modelled in terms of the influence of each QTL on control processes in the APSIM model. There were significant differences in rate of increase in average yield in the target population of environments (TPE) with cycles of selection among three marker-assisted selection scenarios. The first two scenarios require physiological understanding and integrative modelling; in the third scenario markers have been assigned to physiological traits and marker weights have been derived from the simulated value of that trait in the TPE. When physiologically weighted marker selection was used, the rate of yield improvement was enhanced and the average yield reached a higher level.

In a different approach, a gene-to-phenotype $(G \rightarrow P)$ modelling framework for quantitative genetics was proposed that explicitly deals with the context-dependent gene effects that are attributed to genes functioning within networks, i.e. epistasis, $G \times E$ interactions and pleiotropy (Podlich and Cooper, 1999; Cooper and Podlich, 2002). Using $(G \rightarrow P)$ models that are based on the E(NK) genenetwork model and the QU-GENE simulation platform for quantitative analysis of genetic models (Podlich and Cooper, 1998), it has been demonstrated that it is possible to design molecular breeding strategies for complex traits that, on average, will outperform phenotypic selection (Cooper et al., 2005).

Elucidating drought and heat stress response mechanisms at the molecular level and their manipulation in plant biotechnology

Crop simulation models may also guide the use of genetic manipulation to produce crop varieties that can tolerate the effects of climate change. Drought tolerance is already a target for wheat in the UK, but as we have described above, crop simulation models indicate that heat tolerance may be of equal if not greater importance. As we have discussed above, both are complex traits in which the effects of single genes may be difficult to discern. Nevertheless, genes that play an important role in imparting these traits will have to be identified if biotechnology is going to play a useful part in producing crops for future climates.

The classic molecular biology approach to stress tolerance is to impose the stress and look for genes that are expressed in response. A group of candidate genes are then selected based on likely function. Proteins involved in signalling (such as protein kinases) or the control of gene expression (such as transcription factors) are often favoured on the basis that they have the potential for wide-ranging effects on plant physiology. In the case of drought and heat tolerance, proteins that act as protectants under stress conditions have also attracted much attention. The candidate genes are then tested by modifying their expression in transgenic plants, or by analysing mutants that lack their activity. Typically, the stress that is applied is extreme and this attracts scepticism from some plant physiologists. Nevertheless, there have been notable successes in elucidating the systems that control responses to both drought and heat stress and some of the genes that are involved do increase tolerance when over-expressed in transgenic plants, at least under experimental conditions.

Water stress brings about a wide range of effects on plants that differ from species to species and between different varieties, developmental stages, organs, and tissue types (see Barnabás et al., 2008, for a comprehensive review). The hormone abscisic acid (ABA) plays a key role, initiating a network of signalling pathways involving multiple protein kinases. These include ATH1, a transmembrane histidine kinase receptor (Wohlbach et al., 2008), mitogen-activated protein kinases (MAPKs) (Colcombet and Hirt, 2008), calcium-dependent protein kinases (CDPKs) (Hrabak et al., 2003) and sucrose non-fermenting-1-related protein kinases (SnRKs). SnRKs are divided into three subfamilies: SnRK1, 2, and 3 (Halford et al., 2003; Halford and Hey, 2009). Of these, SnRK2 and SnRK3 have been known to be involved in stress responses for several years. For example, some Arabidopsis SnRK2s have been shown to be activated by hyperosmotic and saline stresses (Boudsocq et al., 2004; Kobayashi et al., 2004) and over-expression of one has been shown to improve drought tolerance (Umezawa et al., 2004). The Arabidopsis SnRK3 family includes SOS2, which is involved in conferring salt tolerance (Liu et al., 2000). Transcription factors of the ABA response element binding protein class (AREBPs) are substrates for members of all three SnRK families (Kobayashi et al., 2005; Furihata et al., 2006; Zhang et al., 2008).

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SnRK1 is better known as a central regulator of metabolism (Halford, 2006), and while the nature of its relationship with ABA requires further study it is a possible conduit for cross-talk between metabolic and stress signalling. Plants manipulate carbohydrate metabolism, for example, by interconverting insoluble starch with soluble sugars, to mitigate the effects of osmotic stress brought about by drought (Wingler et al., 1999; Reddy, 2000; Hoekstra et al., 2001; Villadsen et al., 2005). Regulation of gene expression is not the only mechanism by which such changes can be brought about. For example, ADP-glucose pyrophosphorylase, a key enzyme in the starch biosynthetic pathway, is redox-activated in a process that requires SnRK1 (Tiessen et al., 2003), while trehalose phosphate synthase is a substrate for SnRK1 (Harthill et al., 2006). Trehalose 6-phosphate is a key signalling metabolite in plants while trehalose itself is present at relatively high levels in resurrection plants. Furthermore, transgenic rice plants containing high levels of trehalose have been shown to have increased drought tolerance (Garg et al., 2002).

Another substrate for SnRK1 is fructose-6-phosphate,2kinase/fructose-2,6-bisphosphatase (F2KP) (Kulma et al., 2004) and this enzyme has been implicated in bringing about drought-induced changes in carbohydrate metabolism in barley; these changes include the accumulation of hexoses at the expense of sucrose and starch (Villadsen

et al., 2005). Regulation of F2KP by SnRK1 also involves a 14-3-3 protein (Kulma et al., 2004).

Calcium-dependent protein kinases (CDPKs) are also involved in transducing stress signals, including signals for water and heat stress. There are too many studies on this subject to describe here, but they have been reviewed extensively by others (Hrabak et al., 2003; Ludwig et al., 2004; Wan et al., 2007). A theme that emerges from these reviews is the interrelationship of different stress responses, particularly those of cold, salt, water, and osmotic stress. Indeed, cold, salt, and water stress will all induce osmotic stress. It is also notable that over-expression of a CDPK has been shown to increase drought tolerance in rice, at least under experimental conditions (Saijo et al., 2000).

CDPKs are closely linked with ABA signalling. For example, Arabidopsis CDPKs AtCPK10 and AtCPK30 have been shown to be required for the activation of an ABA-inducible promoter (Sheen, 1996). Many CDPK genes themselves contain an ABA response element in their promoter and some have been shown to be induced at the transcriptional level by ABA (Wan et al., 2007).

Transcription factors that are known to be involved in water stress responses include dehydration-responsive element binding protein (DREB)-1 and -2 (Sakuma et al., 2006; Seki et al., 2001), ABA response element binding proteins (AREBPs) (Choi et al., 2000), transcription factors of the zinc finger homeodomain (ZFHD)-1 type (Tran et al., 2007), members of the myeloblastosis (MYB) and myelocytomatosis (MYC) families (Abe et al., 1997), and of the no apical meristem (NAM), ATAF1 and 2, and cup-shaped cotyledon (CUC) family (NAC) (Olsen et al., 2005). A comprehensive review of their roles was provided by Shinozaki and Yamaguchi-Shinozaki (2007). The action of AREBPs, DREB1, MYC, and MYB requires ABA, while that of DREB2, ZFHD1, and NAC is ABA-independent. Overexpression of another transcription factor, plant nuclear factor-Y (NF-Y) has been shown to confer increased drought tolerance in maize in the field (Nelson et al., 2007).

Many of the factors that are expressed during water stress are also expressed when heat stress is applied. In some cases this may be misleading because elevated temperatures will cause water stress unless ambient humidity is adjusted to prevent it. The effects of different abiotic stresses, including drought and heat, do overlap, as we have discussed above, but each different stress also presents the plant with some specific problems and heat stress is no exception. As we have described above, high temperatures cause wheat and other cereals to develop and mature more quickly; they also bring about an increase in respiration and an inhibition of photosynthesis. The latter is caused by a reduction in the activity of ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) and the efficiency of photosystem II (Al-Khatib and Paulsen, 1999). The reduction in Rubisco activity occurs because the enzyme responsible for maintaining its activity, Rubisco activase, is labile at even moderately high temperatures. Genetic manipulation of Rubisco activase to improve its stability at high temperatures is a potentially important target (Kurek et al., 2007).

Another effect of high temperature is oxidative damage and high activities of superoxide dismutase and catalase have been shown to be associated with good thermotolerance in wheat (Sairam et al., 2000). Temperatures greater than 35 °C during wheat grain development have also been shown to cause changes in the expression of different groups of seed storage proteins, with consequent effects on dough quality (Blumenthal et al., 1993; Irmak et al., 2008). Heat stress is also likely to increase the levels of free asparagine (Lea et al., 2007), one of the precursors of acrylamide, a toxic contaminant that is formed during hightemperature processing of wheat and other crops. On the other hand, moderate increases in summer temperature in the UK may actually have a beneficial effect in reducing the levels of free asparagine (Curtis et al., 2009).

Transcription factors that are specifically associated with heat stress include heat shock factors (HSFs). The HSFs are quite a large family with at least 21 members, and their interaction appears to be complex (Baniwal et al., 2004; Kotak et al., 2007). Nevertheless, simple over-expression experiments with HSFs have resulted in increased thermotolerance in transgenic plants (Ogawa et al., 2007; Yokotani et al., 2008). Heat stress can cause proteins, RNA and other molecules to fold incorrectly, affecting their assembly, translocation, turnover, and activity. Heat-shock proteins (HSPs) and other chaperones that keep proteins and RNA in their correct conformation are expressed to mitigate this problem and these have therefore also attracted much attention (Wang et al., 2004). The expression of HSPs is under the control of HSFs (Schoffl et al., 1998).

The key question for plant biotechnology is: will the manipulation of any of these genes provide a consistent improvement in drought and/or heat tolerance under field as opposed to laboratory conditions? At present there are no transgenic crop varieties being marketed on the basis of improved drought or heat tolerance that we are aware of, but all of the major plant biotechnology companies claim to have such varieties in development. Monsanto appears to be leading the way with plans to market drought-tolerant corn possibly as early as 2012. The exact nature of the traits involved is not yet in the public domain and there may be more than one; Monsanto has experimented with overexpression of transcription factors (Nelson et al., 2007) and RNA chaperones (Castiglioni et al., 2008). The yield increases that are claimed are relatively modest, from 6.7% to 13.4% under drought conditions (http://www.monsanto. com/pdf/droughttolerantcorn/drought slides.pdf), so these varieties are not going to green the desert. Nevertheless, Monsanto claims that the yield increases are significant and, of course, these varieties would hopefully represent the start of a process of long-term improvement.

Conclusions

A step-by-step procedure is described here for a rational design of wheat ideotypes for future environments under a changing climate using mathematical modelling and indepth knowledge of molecular regulatory mechanisms and wheat genetics. This approach can be used to underpin the breeding process for new wheat cultivars that are suitable for the rapidly changing environments brought about by global warming. The aim is to enable breeders to focus their efforts on those traits that promise an enhanced performance of the wheat crop under climate change.

By running crop simulation models for future scenarios brought about by climate change, the abiotic stresses that will have major impacts on wheat in the future can be identified. Those wheat physiological traits that need improvement to adapt to changing environments can then be shortlisted. The outcome depends on predicted changes in weather patterns, which have a high degree of uncertainty. However, important conclusions can be drawn. For example, wheat is particularly sensitivity to heat stress during flowering, with potentially severe consequences for grain yield. Severe heat stress is therefore likely to have an increasing impact on wheat yield as the climate warms. On the other hand, global warming will accelerate wheat development, reducing the impact of summer drought, at least in the UK.

Some caution is required when using process-based crop simulation models to analyse complex traits, since the ability of the model to predict complicated trait×environment interactions depends on the assumptions made in the model. Nevertheless, such an approach allows us to investigate these complex interactions in a way that is not possible in field experiments. Crop simulation modelling can be used to deconvolute the complex traits that are selected for improvement, breaking down the responses of complex traits to variation in simpler component traits. By modelling crop growth in variable environments, a better understanding of the relationships between the complex and component traits can be achieved and the simple traits can be ranked in order of their influence on the complex traits. The simple traits may then be amenable to mapping to genetic loci and used to prioritize research on QTL or single marker genes and in the assessment of the available genetic variability in wheat populations.

If traits can be assigned to single genes or small numbers of genes they may also be amenable to genetic modification. However, much of the work at the molecular level on elucidating the systems that enable plants to respond to drought and heat stress has been done in *Arabidopsis* and transferring the technology to wheat with its relatively complex genetics will be difficult. Furthermore, while the market for biotech corn, soybean, and cotton is well established in the Americas and Asia, the market for biotech wheat is not and wheat is attracting much less investment from plant biotech companies as a result. Wheat is the major crop in Europe, of course, but with the development of plant biotechnology stalled in Europe under the weight of legislation and lack of public confidence, there is no prospect of investment in biotech varieties for the European market.

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References

Abe H, Yamaguchi-Shinozaki K, Urao T, Iwasaki T, Hosokawa D, Shinozaki K. 1997. Role of Arabidopsis MYC and MYB homologs in drought- and abscisic acid-regulated gene expression. *The Plant Cell* **9,** 1859–1868.

Akkaya A, Dokuyucu T, Kara R, Akçura M. 2006. Harmonization ratio of post- to pre-anthesis durations by thermal times for durum wheat cultivars in a Mediterranean environment. *European Journal of Agronomy* **24,** 404–408.

Al-Khatib K, Paulsen GM. 1999. High-temperature effects on photosynthetic processes in temperate and tropical cereals. *Crop Science* **39**, 119–125.

Austin RB. 1999. Yield of wheat in the United Kingdom: recent advances and prospects. *Crop Science* **39**, 1604–1610.

Baniwal SK, Bharti K, Chan KY, et al. 2004. Heat stress response in plants: a complex game with chaperones and more than twenty heat stress transcription factors. *Journal of Bioscience* **29,** 471–487.

Barnabás B, Jäger K, Fehér A. 2008. The effect of drought and heat stress on reproductive processes in cereals. *Plant, Cell and Environment* **31,** 11–38.

Downloaded from http://jxb.oxfordjournals.org at BBSRC on July 20, 2010

Barrow EM, Semenov MA. 1995. Climate change scenarios with high spatial and temporal resolution for agricultural applications. *Forestry* **68,** 349–360.

Blumenthal CS, Barlow EWR, Wrigley CW. 1993. Growth environment and wheat quality: the effect of heat stress on dough properties and gluten proteins. *Journal of Cereal Science* **18**, 3–21.

Boudsocq M, Barbier-Brygoo H, Lauriere C. 2004. Identification of nine sucrose nonfermenting 1-related protein kinases 2 activated by hyperosmotic and saline stresses in *Arabidopsis thaliana*. *Journal of Biological Chemistry* **279**, 41758–41766.

Carbone GJ, Kiechle W, Locke C, Mearns LO, McDaniel L, Downton MW. 2003. Response of soybean and sorghum to varying spatial scales of climate change scenarios in the southeastern United States. *Climatic Change* **60**, 73–98.

Castiglioni P, Warner D, Bensen RJ, et al. 2008. Bacterial RNA chaperones confer abiotic stress tolerance in plants and improved grain yield in maize under water-limited conditions. *Plant Physiology* **147,** 446–455.

Cattivelli L, Rizza F, Badeck FW, Mazzucotelli E, Mastrangelo AM, Francia E, Mare C, Tondelli A, Stanca AM. 2008. Drought tolerance improvement in crop plants: an integrated view from breeding to genomics. *Field Crops Research* **105**, 1–14.

Charmet G, Robert N, Branlard G, Linossier L, Martre P, Triboï E. 2005. Genetic analysis of dry matter and nitrogen accumulation and protein composition in wheat kernels. *Theoretical and Applied Genetics* **111**, 540–550.

X

Choi HI, Hong JH, Ha JO, Kang JY, Kim SY. 2000. ABFs, a family of ABA-responsive element binding factors. Journal of Biological Chemistry 275, 1723-1730.

Colcombet J, Hirt H. 2008. Arabidopsis MAPKs: a complex signalling network involved in multiple biological processes. Biochemical Journal 413, 217-226.

Collins NC, Tardieu F, Tuberosa R. 2008. Quantitative trait loci and crop performance under abiotic stress: where do we stand? Plant Physiology 147, 469-486.

Condon AG, Richards RA, Rebetzke GJ, Farquhar GD. 2004. Breeding for high water-use efficiency. Journal of Experimental Botany **55,** 2447-2460.

Cooper M, Podlich DW. 2002. The E(NK) model: extending the NK model to incorporate gene-by-environment interactions and epistasis for diploid genomes. Complexity 7, 31-47.

Cooper M, Podlich DW, Smith OS. 2005. Gene-to-phenotype models and complex trait genetics. Australian Journal of Agricultural Research 56, 895-918.

Curtis TY, Muttucumaru N, Shewry PR, Parry MA, Powers SJ, Elmore JS, Mottram DS, Hook S, Halford NG. 2009. Evidence for genetic and environmental effects on free amino acid levels in wheat grain: implications for acrylamide formation during processing. Journal of Agricultural and Food Chemistry 57, 1013-1021.

Dardanelli JL, Ritchie JT, Calmon M, Andriani JM, Collino DJ. 2004. An empirical model for root water uptake. Field Crops Research **87.** 59–71.

Day W, Legg BJ, French BK, Johnston AE, Lawlor DW, Jeffers WDC. 1978. Drought experiment using mobile shelters: effect of drought on barley yield, water-use and nutrient-uptake. Journal of Agricultural Science 91, 599-623.

Debaeke P, Aboudrare A. 2004. Adaptation of crop management to water-limited environments. European Journal of Agronomy 21, 433-446.

Dreccer MF, Rodriguez D, Ogbonnaya F. 2002. Tailoring wheat for marginal environments: a crop modelling study. In: 12th Australasian plant breeding conference. Perth: The Australasian Plant Breeding Association, Inc. 457-462.

Evans LT, Fischer RA. 1999. Yield potential: its definition, measurement and significance. Crop Science 39, 1544-1551.

Ewert F, Rodriguez D, Jamieson P, et al. 2002. Effects of elevated CO₂ and drought on wheat: testing crop simulation models for different experimental and climatic conditions. Agriculture, Ecosystems and Environment 93, 249-266.

Ferris R, Ellis RH, Wheeler TR, Hadley P. 1998. Effect of high temperature stress at anthesis on grain yield and biomass of fieldgrown crops of wheat. Annals of Botany 82, 631-639.

Fischer RA, Rees D, Sayre KD, Lu ZM, Condon AG, Saavedra AL. 1998. Wheat yield progress associated with higher stomatal conductance and photosynthetic rate, and cooler canopies. Crop Science 38, 1467-1475.

Foulkes MJ, Sylvester-Bradley R, Weightman R, Snape JW. 2007. Identifying physiological traits associated with improved drought resistance in winter wheat. Field Crops Research **103,** 11–24.

Furihata T, Maruyama K, Fujita Y, Umezawa T, Yoshida R, Shinozaki K, Yamaguchi-Shinozaki K. 2006. Abscisic aciddependent multisite phosphorylation regulates the activity of a transcription activator AREB1. Proceedings of the National Academy of Sciences, USA 103, 1988-1993.

Garg AK, Kim JK, Owens TG, Ranwala AP, Do Choi Y, Kochian LV, Wu RJ. 2002. Trehalose accumulation in rice plants confers high tolerance levels to different abiotic stresses. Proceedings of the National Academy of Sciences, USA 99, 15898-15903.

Halford NG. 2006. Regulation of carbon and amino acid metabolism: roles of sucrose nonfermenting-1-related protein kinase-1 and general control nonderepressible-2-related protein kinase. Advances in Botanical Research Incorporating Advances in Plant Pathology **43,** 93-142.

Halford NG, Hey S. 2009. SNF1-related protein kinases (SnRKs) act within an intricate network that links metabolic and stress signalling in plants. Biochemical Journal 419, 247-259.

Halford NG, Hey S, Jhurreea D, Laurie S, McKibbin RS, Paul MJ, Zhang Y. 2003. Metabolic signalling and carbon partitioning: role of Snf1-related (SnRK1) protein kinase. Journal of Experimental Botany **54,** 467–475.

Hammer G, Cooper M, Tardieu F, Welch S, Walsh B, van Eeuwijk F, Chapman S, Podlich D. 2006. Models for navigating biological complexity in breeding improved crop plants. Trends in Plant Science 11, 587-593.

Hammer GL, Chapman S, van Oosterom E, Podlich DW. 2005. Trait physiology and crop modelling as a framework to link phenotypic complexity to underlying genetic systems. Australian Journal of Agricultural Research 56, 947-960.

Hansen JW. 2005. Integrating seasonal climate prediction and agricultural models for insights into agricultural practice. Philosophical Transactions of the Royal Society Series B, Biological Sciences 360, 2037-2047.

Harthill JE, Meek SEM, Morrice N, Peggie MW, Borch J, Wong BHC, MacKintosh C. 2006. Phosphorylation and 14-3-3 binding of Arabidopsis trehalose-phosphate synthase 5 in response to 2-deoxyglucose. The Plant Journal 47, 211-223.

Hoekstra FA, Golovina EA, Buitink J. 2001. Mechanisms of plant desiccation tolerance. Trends in Plant Science 6, 431-438.

Hrabak EM, Chan CWM, Gribskov M, et al. 2003. The Arabidopsis CDPK-SnRK superfamily of protein kinases. Plant Physiology 132,

Hulme M, Jenkins GJ, Lu X, et al. 2002. Climate change scenarios for the United Kingdom: the UKCIP02 scientific report. Norwich: Tyndall Centre for Climate Change Research and School of Environmental Sciences, University of East Anglia.

Irmak S, Naeem HA, Lookhart GL, MacRitchle F. 2008. Effect of heat stress on wheat proteins during kernel development in wheat near-isogenic lines differing at Glu-D1. Journal of Cereal Science 48, 513-516.

Ishag HM, Mohamed BA, Ishag KHM. 1998. Leaf development of spring wheat cultivars in an irrigated heat-stressed environment. Field Crops Research 58, 167-175.

Jamieson PD, Berntsen J, Ewert F, Kimball BA, Olesen JE, Pinter PJJ, Porter JR, Semenov MA. 2000. Modelling CO₂ effects on wheat with varying nitrogen supplies. *Agriculture, Ecosystems and Environment* **82**, 27–37.

Jamieson PD, Brooking IR, Porter JR, Wilson DR. 1995. Prediction of leaf appearance in wheat: a question of temperature. *Field Crops Research* **41,** 35–44.

Jamieson PD, Brooking IR, Semenov MA, MeMaster GS, White JW, Porter JR. 2007. Reconciling alternative models of phenological development in winter wheat. *Field Crops Research* **103**, 36–41.

Jamieson PD, Brooking IR, Semenov MA, Porter JR. 1998a. Making sense of wheat development: a critique of methodology. *Field Crops Research* **55,** 117–127.

Jamieson PD, Semenov MA, Brooking IR, Francis GS. 1998b. Sirius: a mechanistic model of wheat response to environmental variation. *European Journal of Agronomy* **8,** 161–179.

Jones H. 2004. What is water use efficiency? In: Bacon MA, ed. *Water use efficiency inplant biology*. Oxford: Blackwell Publishing, 27–41.

Jones HG. 1992. Plants and microclimate. A quantitative approach to environmental plant physiology. Cambridge: Cambridge University Press.

Jouzel J, Lorius C, Petit JR, Genthon C, Barkov NI, Kotlyakov VM, Petrov VM. 1987. Vostok ice core: a continuous isotope temperature record over the last climatic cycle (160,000 years). *Nature* **329**, 403–408.

Jouzel J, Barkov NI, Barnola JM, Bender M, Chappellaz J, Genthon C, Kotlyakov VM, Lipenkov V, Lorius C, Petit JR, Raynaud D, Raisbeck G, Ritz C, Sowers T, Stievenard M, Yiou F, Yiou P. 1993. Extending the Vostok ice-core record of palaeoclimate to the penultimate glacial period. *Nature* **364**, 407–412.

Jouzel J, Waelbroeck C, Malaize B, Bender M, Petit JR, Stievenard M, Barkov NI, Barnola JM, King T, Kotlyakov VM, Lipenkov V, Lorius C, Raynaud D, Ritz C, Sowers T. 1996. Climatic interpretation of the recently extended Vostok ice records. *Climate Dynamics* 12, 513–521.

Kobayashi Y, Yamamoto S, Minami H, Kagaya Y, Hattori T. 2004. Differential activation of the rice sucrose nonfermenting1-related protein kinase2 family by hyperosmotic stress and abscisic acid. *The Plant Cell* **16**, 1163–1177.

Kobayashi Y, Murata M, Minami H, Yamamoto S, Kagaya Y, Hobo T, Yamamoto A, Hattori T. 2005. Abscisic acid-activated SNRK2 protein kinases function in the gene-regulation pathway of ABA signal transduction by phosphorylating ABA response element-binding factors. *The Plant Journal* 44, 939–949.

Kotak S, Larkindale J, Lee U, von Koskull-Doring P, Vierling E, Scharf KD. 2007. Complexity of the heat stress response in plants. *Current Opinion in Plant Biology* **10**, 310–316.

Kulma A, Villadsen D, Campbell DG, Meek SEM, Harthill JE, Nielsen TH, MacKintosh C. 2004. Phosphorylation and 14-3-3 binding of Arabidopsis 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase. *The Plant Journal* **37**, 654–667.

Kurek I, Chang TK, Bertain SM, Madrigal A, Liu L, Lassner MW, Zhu GH. 2007. Enhanced thermostability of Arabidopsis Rubisco

activase improves photosynthesis and growth rates under moderate heat stress. *The Plant Cell* **19,** 3230–3241.

Lawless C, Semenov MA, Jamieson PD. 2005. A wheat canopy model linking leaf area and phenology. *European Journal of Agronomy* **22,** 19–32.

Lawlor DW, Day W, Johnston AE, Legg BJ, Parkinson KJ. 1981. Growth of spring barley under drought: crop development, photosynthesis, dry-matter accumulation and nutrient content. *Journal of Agricultural Science* **96,** 167–186.

Lea PJ, Sodek L, Parry MAJ, Shewry R, Halford NG. 2007. Asparagine in plants. *Annals of Applied Biology* **150,** 1–26.

Liu YB, Tabashnik BE, Masson L, Escriche B, Ferre J. 2000. Binding and toxicity of *Bacillus thuringiensis* protein Cry1C to susceptible and resistant diamondback moth (Lepidoptera: Plutellidae). *Journal of Economic Entomology* **93,** 1–6.

Ludwig AA, Romeis T, Jones JDG. 2004. CDPK-mediated signal-ling pathways: specificity and cross-talk. *Journal of Experimental Botany* **55,** 181–188.

Manschadi AM, Christopher J, Devoil P, Hammer GL. 2006. The role of root architectural traits in adaptation of wheat to water-limited environments. *Functional Plant Biology* **33**, 823–837.

Meinke H, Hammer G, van Keulen H, Rabbinge R, Keating BA. 1997. Improving wheat simulation capabilities in Australia from a cropping systems perspective: water and nitrogen effects on spring wheat in a semi-arid environment. *European Journal of Agronomy* **7,** 75–88.

Mitchell RAC, Mitchell VJ, Driscoll SP, Franklin J, Lawlor DW. 1993. Effects of increased CO₂ concentration and temperature on growth and yield of winter-wheat at two levels of nitrogen application. *Plant, Cell and Environment* **16,** 521–529.

Downloaded from http://jxb.oxfordjournals.org at BBSRC on July 20, 2010

Mossad MG, Ortiz-Ferrara G, Mahalakshmi V, Fischer RA. 1995. Phyllochron response to vernalization and photoperiod in sprind wheat. *Crop Science* **35**, 168–171.

Nakicenovic N, Swart R. 2000. *Emissions scenarios*. 2000. Special Report of the Intergovernmental Panel on Climate Change. Cambridge: Cambridge University Press, 570.

Nelson DE, Repetti PP, Adams TR, et al. 2007. Plant nuclear factor Y (NF-Y) B subunits confer drought tolerance and lead to improved corn yields on water-limited acres. *Proceedings of the National Academy of Sciences, USA* **104,** 16450–16455.

Ogawa D, Yamaguchi K, Nishiuchi T. 2007. High-level over-expression of the Arabidopsis *HsfA2* gene confers not only increased themotolerance but also salt/osmotic stress tolerance and enhanced callus growth. *Journal of Experimental Botany* **58**, 3373–3383.

Olesen JE, Carter TR, Diaz-Ambrona CH, et al. 2007. Uncertainties in projected impacts of climate change on European agriculture and terrestrial ecosystems based on scenarios from regional climate models. *Climatic Change* **81,** 123–143.

Olsen AN, Ernst HA, Lo Leggio L, Skriver K. 2005. NAC transcription factors: structurally distinct, functionally diverse. *Trends in Plant Science* **10,** 79–87.

Passioura JB. 1996. Simulation models: science, snake oil, education or engineering. *Agronomy Journal* **88**, 690–694.

Petit JR, Jouzel J, Raynaud D, et al. 1999. Climate and atmospheric history of the past 420 000 years from the Vostok ice core, Antarctica. Nature 399, 429-436.

Podlich DW, Cooper M. 1998. QU-GENE: a simulation platform for quantitative analysis of genetic models. Bioinformatics 14, 632-653.

Podlich DW, Cooper M. 1999. Modelling plant breeding programs as search strategies on a complex response surface. Lecture Notes in Computer Science 1585, 171-178.

Porter JR, Semenov MA. 2005. Crop responses to climatic variability. Philosophical Transactions of the Royal Society, Series B, Biological Sciences 360, 2021-2035.

Quilot B, Kervella J, Genard M, Lescourret F. 2005. Analysing the genetic control of peach fruit quality through an ecophysiological model combined with a QTL approach. Journal of Experimental Botany 56, 3083-3092.

Reddy AR. 2000. Photosynthesis and fructose 2.6-bisphosphate content in water stressed wheat leaves. Cereal Research Communications 28. 131-137.

Richards RA. 2006. Physiological traits used in the breeding of new cultivars for water-scarce environments. Agricultural Water Management 80, 197-211.

Richards RA, Passioura JB. 1981. Seminal root morphology and water-use of wheat.1. Environmental effects. Crop Science 21, 249-252.

Richards RA, Passioura JB. 1989. A breeding program to reduce the diameter of the major xylem vessel in the seminal roots of wheat and its effect on grain-yield in rain-fed environments. Australian Journal of Agricultural Research 40, 943-950.

Richards RA, Rebetzke GJ, Condon AG, van Herwaarden AF. 2002. Breeding opportunities for increasing the efficiency of water use and crop yield in temperate cereals. Crop Science 42, 111-121.

Richter GM, Semenov MA. 2005. Modelling impacts of climate change on wheat yields in England and Wales: assessing drought risks. Agricultural Systems 84, 77-97.

Robert N, Hennequet C, Bérard P. 2001. Dry matter and nitrogen accumulation in wheat kernel: genetic variation in rate and duration of grain filling. Journal of Genetics and Breeding 55, 297-305.

Robertson MJ, Fukai S, Ludlow MM, Hammer GL. 1993. Water extraction by grain sorghum in a sub-humid environment. I. Analysis of the water extraction pattern. Field Crops Research 33, 81-97.

Saijo Y, Hata S, Kyozuka J, Shimamoto K, Izui K. 2000. Overexpression of a single Ca²⁺-dependent protein kinase confers both cold and salt/drought tolerance on rice plants. The Plant Journal 23, 319-327.

Sairam RK, Srivastava GC, Saxena DC. 2000. Increased antioxidant activity under elevated temperatures: a mechanism of heat stress tolerance in wheat genotypes. Biologia Plantarum 43, 245-251.

Sakuma Y, Maruyama K, Osakabe Y, Qin F, Seki M, Shinozaki K, Yamaguchi-Shinozaki K. 2006. Functional analysis of an Arabidopsis transcription factor, DREB2A, involved in droughtresponsive gene expression. The Plant Cell 18, 1292-1309.

Schoffl F, Prandl R, Reindl A. 1998. Regulation of the heat-shock response. Plant Physiology 117, 1135-1141.

Seki M, Narusaka M, Abe H, Kasuga M, Yamaguchi-Shinozaki K, Carninci P, Hayashizaki Y, Shinozaki K. 2001. Monitoring the expression pattern of 1300 Arabidopsis genes under drought and cold stresses by using a full-length cDNA microarray. The Plant Cell 13, 61-72.

Semenov MA. 2007. Development of high resolution UKCIP02-based climate change scenarios in the UK. Agricultural and Forest Meteorology 144, 127-138.

Semenov MA. 2009. Impacts of climate change on wheat in England and Wales. Journal of the Royal Society Interface 6, 343-350.

Semenov MA, Jamieson PD, Martre P. 2007. Deconvoluting nitrogen use efficiency in wheat: a simulation study. European Journal of Agronomy 26, 283-294.

Semenov MA, Martre P, Jamieson PD. 2009. Quantifying effects of simple wheat traits on yield in water-limited environments using a modelling approach. Agricultural and Forest Meteorology 149, 1095-1104.

Semenov MA, Porter JR, Delecolle R. 1993. Climatic change and the growth and development of wheat in the UK and France. European Journal of Agronomy 2, 293-304.

Semenov MA, Wolf J, Evans LG, Eckersten H, Iglesias A. 1996. Comparison of wheat simulation models under climate change.2. Application of climate change scenarios. Climate Research 7, 271–281.

Shearman VJ, Sylvester-Bradley R, Scott RK, Foulkes MJ. 2005. Physiological processes associated with wheat yield progress in the UK. Crop Science 45, 175-185.

Sheen J. 1996. Ca²⁺-dependent protein kinases and stress signal transduction in plants. Science 274, 1900-1902.

Shinozaki K, Yamaguchi-Shinozaki K. 2007. Gene networks involved in drought stress response and tolerance. Journal of Experimental Botany 58, 221-227.

Sinclair TR, Muchow RC. 2001. System analysis of plant traits to increase grain yield on limited water supplies. Agronomy Journal 93, 263-270

Solomon S, Qin D, Manning M, Marquis M, Averyt K, Tignor MMB, LeRoy Miller H, Chen Z. 2007. Climate change 2007: the physical science basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovermental Panel on Climate Change. New York: Cambridge University Press.

Tambussi EA, Bort J, Araus JL. 2007. Water use efficiency in C₃ cereals under Mediterranean conditions: a review of physiological aspects. Annals of Applied Biology 150, 307-321.

Tardieu F. 2003. Virtual plants: modelling as a tool for genomics of tolerance to water deficit. Trends in Plant Science 8, 9-14.

Tardieu F, Reymond M, Muller B, Granier C, Simonneau T, Sadok W, Welcker C. 2005. Linking physiological and genetic analyses of the control of leaf growth under changing environmental conditions. Australian Journal of Agricultural Research 56, 937-946.

Tiessen A, Prescha K, Branscheid A, Palacios N, McKibbin R, Halford NG, Geigenberger P. 2003. Evidence that SNF1-related kinase and hexokinase are involved in separate sugar-signalling pathways modulating post-translational redox activation of ADP-glucose pyrophosphorylase in potato tubers. The Plant Journal 35, 490-500.

Tran LSP, Nakashima K, Sakuma Y, Osakabe Y, Qin F, Simpson SD, Maruyama K, Fujita Y, Shinozaki K, Yamaguchi-Shinozaki K. 2007. Co-expression of the

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stress-inducible zinc finger homeodomain ZFHD1 and NAC transcription factors enhances expression of the ERD1 gene in Arabidopsis. The Plant Journal 49, 46-63.

Triboi E, Triboi-Blondel AM. 2002. Productivity and grain or seed composition: a new approach to an old problem-invited paper. European Journal of Agronomy 16, 163-186.

Umezawa T, Yoshida R, Maruyama K, Yamaguchi-Shinozaki K, Shinozaki K. 2004. SRK2C, a SNF1-related protein kinase 2, improves drought tolerance by controlling stress-responsive gene expression in Arabidopsis thaliana. Proceedings of the National Academy of Sciences, USA 101, 17306-17311.

Villadsen D, Rung JH, Nielsen TH. 2005. Osmotic stress changes carbohydrate partitioning and fructose-2,6-bisphosphate metabolism in barley leaves. Functional Plant Biology 32, 1033-1043.

Wan B, Lin Y, Mou T. 2007. Expression of rice Ca²⁺-dependent protein kinases (CDPKs) genes under different environmental stresses. FEBS Letters 581, 1179-1189.

Wang ML, Zhao Y, Chen F, Yin XC. 2004. Inheritance and potentials of a mutated dwarfing gene ndf1 in Brassica napus. Plant Breeding 123, 449-453.

Wheeler TR, Batts GR, Ellis RH, Hadley P, Morison JlL. 1996. Growth and yield of winter wheat (Triticum aestivum) crops in response to CO₂ and temperature. Journal of Agricultural Science 127, 37-48.

Wheeler TR, Craufurd PQ, Ellis RH, Porter JR, Prasad PVV. 2000. Temperature variability and the yield of annual crops. Agriculture, Ecosystems and Environment 82, 159-167.

White JW, Hoogenboom G. 2003. Gene-based approaches to crop simulation: past experiences and future opportunities. Agronomy Journal 95, 52-64.

Wilks DS. 1992. Adapting stochastic weather generation algorithms for climate changes studies. Climatic Change 22, 67-84.

Wingler A, Quick WP, Bungard RA, Bailey KJ, Lea PJ, **Leegood RC.** 1999. The role of photorespiration during drought stress: an analysis utilizing barley mutants with reduced activities of photorespiratory enzymes. Plant, Cell and Environment 22,

Witcombe JR, Hollington PA, Howarth CJ, Reader S, Steele KA. 2008. Breeding for abiotic stresses for sustainable agriculture. Philosophical Transactions of the Royal Society, Series B, Biological Sciences 363, 703-716.

Wohlbach DJ, Quirino BF, Sussman MR. 2008. Analysis of the Arabidopsis histidine kinase ATHK1 reveals a connection between vegetative osmotic stress sensing and seed maturation. The Plant Cell **20,** 1101–1117.

Wolf J, Evans LG, Semenov MA, Eckersten H, Iglesias A. 1996. Comparison of wheat simulation models under climate change. 1. Model calibration and sensitivity analyses. Climate Research 7, 253-270.

Yokotani N, Ichikawa T, Kondou Y, Matsui M, Hirochika H, Iwabuchi M, Oda K. 2008. Expression of rice heat stress transcription factor OsHsfA2e enhances tolerance to environmental stresses in transgenic Arabidopsis. Planta 227, 957-967.

Zhang Y, Andralojc PJ, Hey SJ, Primavesi LF, Specht M, Koehler J, Parry MAJ, Halford NG. 2008. Arabidopsis sucrose non-fermenting-1-related protein kinase-1 and calcium-dependent protein kinase phosphorylate conserved target sites in ABA response element binding proteins. Annals of Applied Biology 153, 401-409.