

PhD Diary

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1 Tasks [3/9]

1.1 DONE Redo Kitagawa et al. [2] data figures

1.1.1 DONE Write optimising procedure for q

```

1  from node_diffusion import do_internode_diffusion
2  from diffusion_functions import D_eff
3  import numpy as np
4  import matplotlib.pyplot as plt
5  from scipy.optimize import leastsq
6
7  cells = 151 # need to take two extra measurements to get sample of 5 without slicing
8  cell_um = 100
9  points_per_cell = 10 # Needs to be divisible by 2
10 Xs = cells*points_per_cell
11 dx = cell_um//points_per_cell
12 dx2 = dx**2
13
14 b = 0.0
15 dt = 0.01
16 # convert to seconds
17 t = 1/dt
18 num_seconds = 60*60*14
19 ts = int(t * num_seconds)
20
21 moss_values = [0.08, 0.15, 0.3, 0.2, 0.1]
22
23 err = 1
24 q = 0.0015
25
26
27 def optimize_q(q):
28     u = np.zeros((Xs, 1))
29     u[(Xs//2)-(points_per_cell//2):(Xs//2)+(points_per_cell//2)] = 1
30     D = D_eff(83, q, cell_um)
31     nds, u = do_internode_diffusion(
32         u, dx2, D, dt, b, cell_um, points_per_cell, ts)
33     fig = plt.figure(0, figsize=(10, 10))
34     fig.clf()
35     fig, ax = plt.subplots(1, 1, figsize=(5, 5), num=0)
36
37     ax.plot(np.arange(-(cells*cell_um)//2, +(cells*cell_um)//2, step=cell_um)+50,
38            nds, marker='o', label='Model')
39
40     ax.set_xlim(-250, 250)
41     ax.set_ylim(-0.02, 1)
42
43     ax.set_xlabel(r'$\mu$ m$^2$')
44     ax.plot(np.linspace(-200, 200, num=5), moss_values,
45            label='Kitawga et al.', marker='o')
46
47     err = np.square((nds[(len(nds)//2)-2:(len(nds)//2)+3] - moss_values)).sum()
48
49     fig.suptitle('Q={0} \ t err^2 = {1}'.format(q, err))
50     fig.tight_layout()
51     fig.savefig('./images/{0}.png'.format(str(q).replace('.', '_')))
52

```

```

53     # Minimise sum of squared errors
54     return err
55
56 result = leastsq(optimize_q, [q])
57 print(result)

```

1.1.2 **DONE** minimise difference of model to data

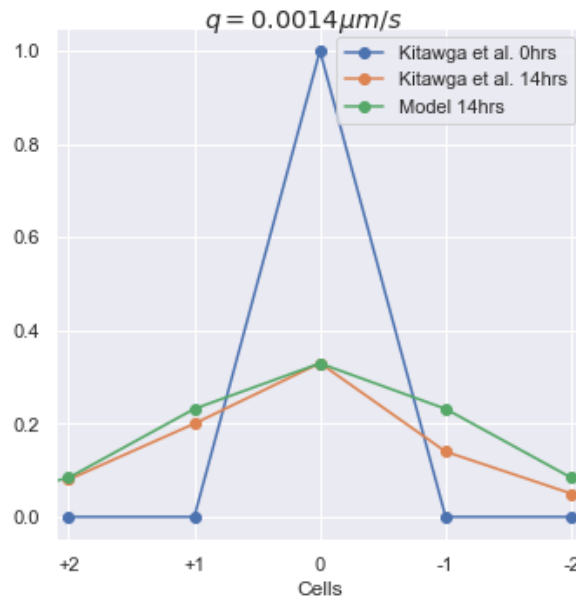


Figure 1: Testing against Kitagawa data

1.1.2.1 Mock treatment

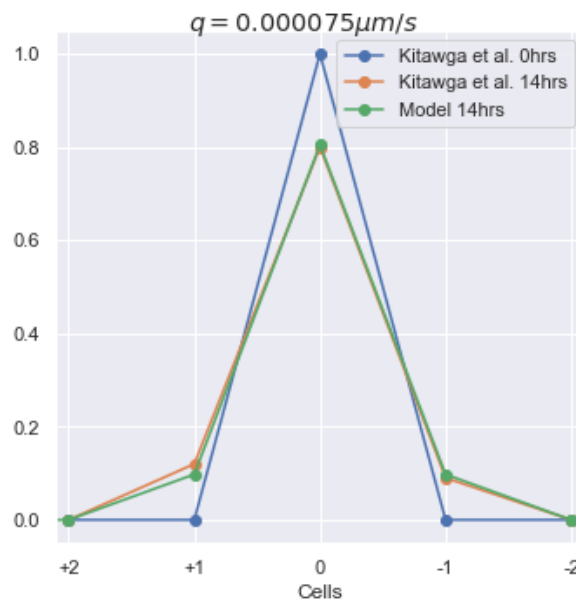


Figure 2: Testing against Kitagawa data with aba treatment

1.1.2.2 ABA treatment

1.2 **TODO** Does it make sense to use D_{eff} even for intra-cellular diffusion1.2.1 **TODO** Test effect of using D vs D_{eff} 1.3 **TODO** Move towards fully node based graph model with proof of results

Still use a low Δx but just use $\text{dist} = \text{cell}$ to approx

- Cell length / 10 works consistently

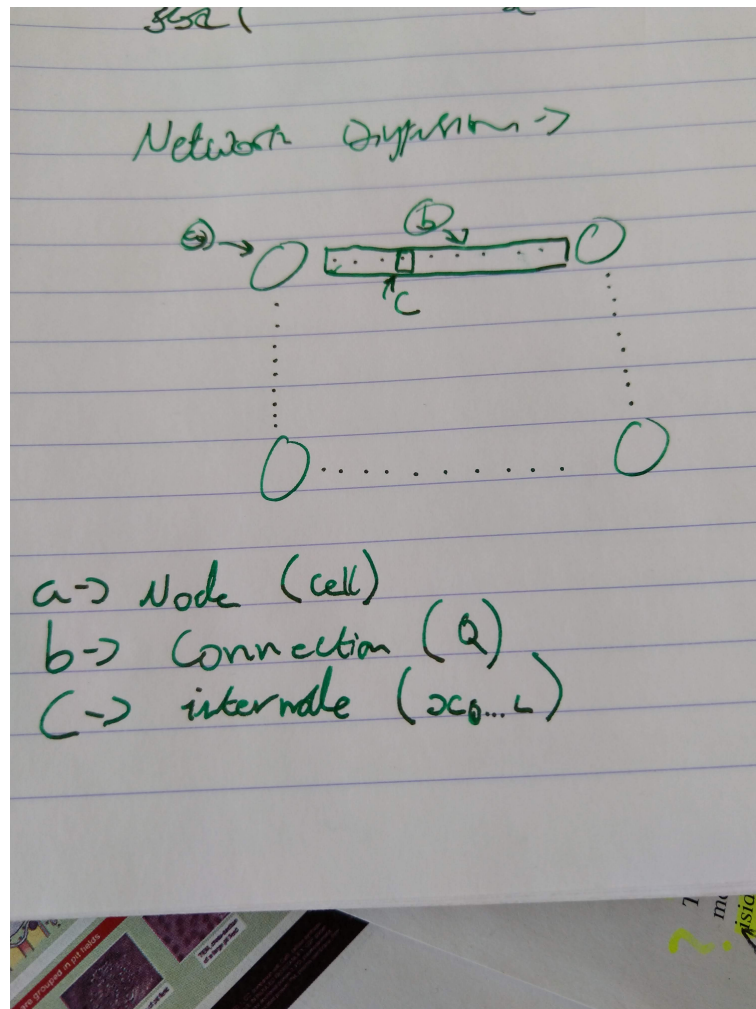


Figure 3: Sketch of idea

1.4 **TODO** Test D_{eff} vector idea

```

1 def diffuse_vectorise(un, g, b, dt, dx2, dy2, a):
2     """
3     Takes a state, rate of decay, production, delta time, delta space and
4     flux of molecule. Uses these data to compute next time state
5     """
6     return (un[1:-1, 1:-1] + a *
7             (((un[2:, 1:-1] - 2 * un[1:-1, 1:-1] + un[:-2, 1:-1]))/dx2 +
8              ((un[1:-1, 2:] - 2 * un[1:-1, 1:-1] + un[1:-1, :-2])/dy2))) * \
9             g + b

```

1.5 DONE Finish presentation**1.6 TODO** Finish chapter 1.5 of report**1.7 TODO** Give practice presentation to Morris group**1.8 TODO** Follow up on images of bombardment assays with Christine**1.9 TODO** Use Deinum [1] A.69 with numerical solution**2 Questions****2.1 How best to make use of D_{eff}**

Not sure how best to use, as it will give a linear curve for diffusion.

- Could probably do a D_{eff} with a k , for cell number, term

$$\alpha = \frac{Dql}{D + ql} \quad (1)$$

References

- [1] E E Deinum. *Simple Models for Complex Questions on Plant Development*. PhD thesis, s.n., S.l., 2013. 00011.
- [2] Munenori Kitagawa, Takumi Tomoi, Tomoki Fukushima, Yoichi Sakata, Mayuko Sato, Kiminori Toyooka, Tomomichi Fujita, and Hitoshi Sakakibara. Absciscic Acid Acts as a Regulator of Molecular Trafficking through Plasmodesmata in the Moss *Physcomitrella patens*/i. *Plant and Cell Physiology*, December 2018. ISSN 0032-0781. doi: 10.1093/pcp/pcy249.