How do cis-regulatory elements evolve in plants?

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Outline

1 Introduction

2 Methods

3 Progress

What is the project?

Introduction 000

Main question: How do *cis*-regulatory elements evolve in plants?

- Cis meaning same
 - in this case elements on the same DNA strand
- i.e. how do non-coding elements on the same DNA strand affect gene expression?

What are *cis*-regulatory elements?

Cis-regulatory elements

Introduction 000

> They are usually enhancers and promoters that control development and physiology by regulating gene expression. 1

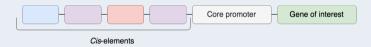


Figure 1: Example promoter containing cis-regulatory elements. Cis-elements can be repeated more than once (purple) and can recruit different transcription factors (blue/red), all driving the same gene (green)

¹Wittkopp and Kalay (2012)

Introduction

Gene expression / phenotypic variation

- For most of the twentieth century, evolution of protein-coding sequences was commonly thought to be primarily (if not solely) responsible for phenotypic evolution
- Whereas more recent studies show that mutations which affect the function of these sequences contribute to phenotypic diversity within and between species
 - Many studies imply divergent *cis*-regulatory activity in phenotypic evolution ²

²Carroll (2008)

How can you study the evolution of *cis*-regulatory elements, in plants?

Materials

- Start with a little known plant: Arabidopsis thaliana
 - Has an extremely well covered genome, as well as transcription factor database
- Will later move towards *Nicotiana* benthamiana, to evaluate how transferable work on model systems are to others'

Plants Nicotiona tohacum Arabidopsis thaliana

Figure 2: Example of Arabidopsis and Nicotiana

Promoter study

Procedure to evaluate?

First choose two types of promoters to study:

The most constitutively expressed and Nitrogen-responsive

For each promoter type:

- Identify motifs where TFs bind, check if any TFs are commonly used
- Determine whether any patterns or similarities are present in groups/types of TFs
- 3 Incorporate nucleosome occupancy data (whether DNA is wrapped around a nucleosome or is open)
- Verify hypothesis in lab

Identifying TF binding motifs of interest

Using MEME and FIMO to predict TF binding motifs 0 to 1 458 (1 458 bp) 1 to 1 458 (1 458 bp) Figure 3: Example FIMO output for promoter of AT1G71820

Lab work

Progress towards testing selected regions of interest

- Designed primers in Benchling and ordered them for each of the promoters/promoter parts if mutating RE cut site
- Ran PCR with primers and At template DNA
 - didn't work, changed conditions
 - tried different DNA template worked
- Made own At CTAB DNA template
 - PCR didnt work
- Using higher concentration of template DNA worked

References I

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- Sean B. Carroll. Evo-Devo and an Expanding Evolutionary Synthesis: A Genetic Theory of Morphological Evolution. Cell, 134(1):25–36, July 2008. ISSN 0092-8674. doi: 10.1016/j.cell.2008.06.030.