

1 **Running head:** CRANIAL MORPHOLOGICAL DIVERSITY IN
2 TENRECS

3 Morphological diversity of tenrec
4 (Afrosoricida, Tenrecidae) skulls compared
5 to their closest relatives, the golden moles
6 (Afrosoricida, Chrysochloridae)

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14 diversity, tenrecs

15 Abstract

16 Morphologically diverse groups have long attracted the interest of
17 biologists. Many studies now recognise the importance of quantifying
18 patterns of morphological diversity to gain new insights into evolutionary
19 patterns. Tenrecs (Afrosoricida, Tenrecidae) are a family of small
20 mammals which is often cited as an example of an exceptionally
21 morphologically diverse group. However, this assumption has not been
22 tested. Here we use geometric morphometric analyses of skull shape to
23 test whether tenrecs are more morphologically diverse than their closest
24 relatives, the golden moles (Afrosoricida, Chrysochloridae). Contrary to
25 our expectations, we find that tenrec skulls are only more morphologically
26 diverse than golden moles when measured in lateral view. Furthermore,
27 the similarities among the species-rich *Microgale* tenrec Genus appear to
28 mask higher morphological diversity in the rest of the Family. Our results
29 reveal new insights into the morphological diversity of tenrecs and
30 highlight the importance of using quantitative methods to test qualitative
31 assumptions about patterns of morphological diversity.

32 Introduction

33 Morphological diversity has long attracted the attention of biologists.
34 There are many famous examples of exceptional morphological diversity
35 including in the beaks of Darwin's finches, the body and limbs of
36 Caribbean *Anolis* lizards, and the pharyngeal jaws of cichlid fish (Gavrilets
37 & Losos, 2009). Morphological diversity is important because it has
38 implications for studies of adaptive radiations - where close relatives
39 exhibit a range of divergent morphologies - (Losos, 2010), convergent
40 evolution - where distant relatives exhibit similar morphologies - (e.g.
41 Muschick et al., 2012; Harmon et al., 2005) and our understanding of
42 biodiversity (Roy & Foote, 1997). However, apart from a few examples
43 (e.g. Goswami et al., 2011; Ruta et al., 2013; Brusatte et al., 2008), it is still
44 common to study morphological diversity from a qualitative rather than
45 quantitative perspective.

46 Morphological diversity is rarely studied from a quantitative
47 perspective because it is difficult to quantify. Studies are inevitably
48 constrained to measure the diversity of specific traits rather than overall
49 morphologies (Roy & Foote, 1997). Different traits (such as cranial
50 compared to limb morphologies) may yield different patterns of
51 morphological diversity (Foth et al., 2012). Furthermore, linear
52 measurements of morphological traits can restrict our understanding of
53 overall morphological variation (Rohlf & Marcus, 1993) . Some of these
54 problems can be solved by using geometric morphometric approaches
55 (Rohlf & Marcus, 1993; Adams et al., 2013) that provide more detailed
56 insights into morphological variation. Yet few studies have used these
57 techniques to specifically address questions about morphological diversity

58 in mammals.

59 Tenrecs (Afrosoricida: Tenrecidae) are a morphologically diverse
60 mammalian group (Soarimalala & Goodman, 2011; Olson & Goodman,
61 2003). The Family contains 34 species, 31 of which are endemic to
62 Madagascar (Olson, 2013). Body sizes of tenrecs span three orders of
63 magnitude (2.5 to > 2,000g); a greater range than all other Families, and
64 most Orders, of living mammals (Olson & Goodman, 2003). Within this
65 vast size range there are tenrecs which convergently resemble shrews
66 (*Microgale* tenrecs), moles (*Oryzorictes* tenrecs) and hedgehogs (*Echinops*
67 and *Setifer* tenrecs) (Eisenberg & Gould, 1969) even though they are not
68 closely related to these species (Stanhope et al., 1998). Despite these
69 interesting features, the morphological diversity of tenrecs has never been
70 properly quantified.

71 Here we present the first quantitative investigation of morphological
72 diversity in tenrecs. We use geometric morphometric approaches to
73 compare cranial morphological diversity in tenrecs to their sister taxa, the
74 golden moles (Afrosoricida, Chrysochloridae). Tenrecs inhabit a wider
75 variety of ecological niches than golden moles (Soarimalala & Goodman,
76 2011; Bronner, 1995) so we expect tenrecs to be more morphologically
77 diverse. However, we only find a significant difference in the
78 morphological diversity of skulls in lateral view, not dorsal or ventral. In
79 contrast, when we restricted our data to include a subsample of the
80 morphologically similar *Microgale* tenrecs, we found that tenrecs were
81 more morphologically diverse than golden moles in all three analyses.
82 Our results highlight the importance of using quantitative methods to test
83 assumptions about patterns of morphological diversity.

84 **Materials and Methods**

85 Our methods for measuring cranial morphological diversity involved
86 several steps of data collection, processing and analysis. For clarity, figure
87 1 summarises all of these steps which are described in detail below.

88 **Morphological data collection**

89 One of us (SF) photographed crania of tenrecs and golden moles at the
90 Natural History Museum London (BMNH), the Smithsonian Institute
91 Natural History Museum (SI), the American Museum of Natural History
92 (AMNH), Harvard's Museum of Comparative Zoology (MCZ) and the
93 Field Museum of Natural History, Chicago (FMNH). We photographed
94 the specimens with a Canon EOS 650D camera fitted with an EF 100mm
95 f/2.8 Macro USM lens using a standardised procedure to minimise
96 potential error (see supplementary material for details).

97 We collected pictures of the skulls in dorsal, ventral and lateral views
98 (right side of the skull). A full list of museum accession numbers and
99 details on how to access the images can be found in the supplementary
100 material.

101 In total we collected pictures from 182 skulls in dorsal view (148
102 tenrecs and 34 golden moles), 173 skulls in ventral view (141 tenrecs and
103 32 golden moles) and 171 skulls in lateral view (140 tenrecs and 31 golden
104 moles) representing 31 species of tenrec (out of the total 34 in the family
105 (Olson, 2013)) and 12 species of golden moles (out of a total of 21 in the
106 family (Asher et al., 2010)). We used the taxonomy of Wilson and Reeder
107 (2005) supplemented with more recent sources (Olson, 2013) to define our

108 species.

109 We used a combination of landmarks (type 2 and type 3, (Zelditch
110 et al., 2012)) and semilandmarks to characterise the shapes of our
111 specimens. Figure 2 shows our landmarks (points) and semilandmarks
112 (outline curves) for the skulls in each of the three views. Corresponding
113 definitions of each of the landmarks can be found in the supplementary
114 material.

115 We used the TPS software series (Rohlf, 2009) to process and landmark
116 the pictures (Fig. 1). We digitised all landmarks and semilandmarks in
117 tpsDIG, version 2.17 (Rohlf, 2013). We re-sampled the outlines to the
118 minimum number of evenly spaced semilandmark points required to
119 represent each outline accurately (MacLeod, 2013, details in
120 supplementary material). We used TPSUtil (Rohlf, 2012) to create
121 “sliders” files that defined which points in our TPS files should be treated
122 as semilandmarks (Zelditch et al., 2012). We conducted all subsequent
123 analyses in R version 3.0.2 (R Core Team, 2014, Fig. 1).

124 We used the `gpagen` function in the `geomorph` package (Adams et al.,
125 2013) to run a general Procrustes alignment (Rohlf & Marcus, 1993) of the
126 landmark coordinates while sliding the semilandmarks by minimising
127 Procrustes distance (Bookstein, 1997). We used these Procrustes-aligned
128 coordinates of all species to calculate average shape values for each
129 species ($n = 43$) which we then used for a principal components analysis
130 (PCA) with the `plotTangentSpace` function (Adams et al., 2013).

Calculating morphological diversity

We calculated morphological diversity using the results of our principal components analyses. We selected the principal components (PC) axes which accounted for 95% of the cumulative variation for each of our three skull analyses. These axes represent the dimensions of our full cranial morphospace (Polly et al., 2013).

We used the scores from the PC axes to compare cranial morphologies in two ways (Fig. 1). First, we used non parametric MANOVAs (Anderson, 2001) to test whether tenrecs and golden moles occupied significantly different positions within our full cranial morphospaces (e.g. Serb et al., 2011; Ruta et al., 2013). We then compared morphological diversity within tenrecs to the diversity within golden moles, defining morphological diversity as the mean Euclidean distance between each species and its Family centroid. If tenrecs are more morphologically diverse than golden moles, then they should be more dispersed within our full cranial morphospaces.

Our groups have unequal sample sizes (31 tenrec species compared to 12 golden mole species). Morphological diversity is usually decoupled from taxonomic diversity (e.g. Ruta et al., 2013; Hopkins, 2013) so larger groups are not necessarily more morphologically diverse. However, comparing morphological diversity in tenrecs to the diversity of a smaller Family could still bias our results. To account for this, we used pairwise permutation tests as follows.

We assigned each species to either “tenrecs” or “golden moles” at random and then calculated the difference in morphological diversity for the new groupings as described above. We repeated this procedure 1000

157 times to generate a null distribution of the expected differences in
158 morphological diversity between a group with 31 members (“tenrecs”)
159 compared to one with 12 members (“golden moles”). If there is no
160 difference between the morphological diversity of tenrecs and golden
161 moles, then the group identity (“tenrec” or “golden mole”) of each species
162 is arbitrary,

163 Finally, we compared our observed measures of the differences in
164 morphological diversity between the two Families to our null
165 distributions to determine whether there were significant differences after
166 taking sample size into account.

167 The majority of tenrec species (19 out of 31 in our dataset) belong to
168 the *Microgale* (shrew-like) Genus that has relatively low morphological
169 diversity (Soarimalala & Goodman, 2011; Jenkins, 2003). This may mask
170 signals of higher morphological diversity among other tenrecs. To test
171 this, we created a subset of our tenrec data that included just five of the
172 *Microgale* species, each representing one of the five sub-divisions of
173 *Microgale* outlined by Soarimalala and Goodman (2011), i.e. small,
174 small-medium, medium, large and long-tailed species. We compared the
175 morphological diversity of this subset of tenrecs (n=19: five *Microgale* and
176 12 non *Microgale* species) to that of golden moles using the methods
177 described above (Fig. 1).

178 Results

179 Figure 3 depicts the morphospace plot derived from our principal
180 components analysis of average Procrustes-superimposed shape

181 coordinates for skulls in lateral view. Similar plots for our analyses of
182 skulls in dorsal and ventral views can be found in the supplementary
183 material. To compare morphological diversity in the two families, we used
184 the principal components axes which accounted for 95% of the cumulative
185 variation in each of our skull analyses: dorsal (n=6 axes), ventral (n=7
186 axes) and lateral (n=7 axes).

187 First, we compared the position of each Family within the
188 morphospace plots. Tenrecs and golden moles occupy significantly
189 different positions in the dorsal (npMANOVA, $F_{1,42} = 68.13$, $R^2 = 0.62$,
190 $p=0.001$), ventral (npMANOVA, $F_{1,42} = 103.33$, $R^2 = 0.72$, $p=0.001$) and
191 lateral (npMANOVA, $F_{1,42} = 76.7$, $R^2=0.652$, $p=0.001$) skull
192 morphospaces, indicating that the Families have very different,
193 non-overlapping cranial morphologies.

194 Secondly, we compared the morphological diversity within each
195 Family. Based on our measures of mean Euclidean distance to the Family's
196 centroid, tenrec skulls are more morphologically diverse than golden mole
197 skulls when they are measured in lateral view but not in dorsal or ventral
198 view (table 1). In contrast, when we compared morphological diversity
199 within the sub-sample of 19 tenrecs (including just five *Microgale* species)
200 to the 12 golden mole species, we found that tenrecs had significantly
201 higher morphological diversity than golden moles in all analyses (table 1).

202 Our pairwise permutation tests for each analysis confirmed that
203 differences in morphological diversity were not artefacts of differences in
204 sample size (see supplementary material).

Discussion

Tenrecs (Tenrecidae) are often cited as an example of a mammalian group with high morphological diversity (Olson, 2013; Soarimalala & Goodman, 2011; Eisenberg & Gould, 1969) and we expected them to be more morphologically diverse than their closest relatives the golden moles (Chrysochloridae). However, tenrecs were only more morphologically diverse than golden moles in one of our three skull analyses (lateral view; table 1). Furthermore, the morphologically similar *Microgale* tenrecs seem to mask high morphological diversity in the rest of the tenrec Family; reducing our data to include a sub-sample of this *Microgale* species revealed that the remaining tenrecs were significantly more morphologically diverse than golden moles across all three skull analyses (table 1). These results highlight the importance of using quantitative methods to test qualitative assumptions about patterns of morphological diversity.

In our full analyses, tenrecs only had higher morphological diversity than golden moles when the skulls were measured in lateral view. This is most likely due to our choice of landmarks. The two outline curves in lateral view (Fig. 2) emphasise morphological variation in the back and top of the skulls, indicating that tenrecs are more morphologically diverse than golden moles in their three dimensional height. These lateral aspects of the skull morphology could not be included in the dorsal and ventral analyses. In contrast, our landmarks in the dorsal, and particularly ventral, views focus on morphological variation in the overall outline shape of the skull and palate (Fig. 2). The result that tenrecs are no more diverse than golden moles in these areas makes intuitive sense: most

231 tenrecs have broad, non-specialised diets (Olson, 2013) so there is no
232 obvious functional reason why they should have significantly diverse
233 palate morphologies. Therefore, comparing the morphologies in three
234 separate views allowed us to identify the more morphologically variable
235 skull regions.

236 Measures of morphological variation are sensitive to the sampling
237 used. If a particular morphotype is over-represented then the similarities
238 among those species will reduce the overall morphological variation
239 within the group (Foote, 1991). This appears to be the case for our data: it
240 is only when we included a sub-sample of *Microgale* tenrecs that we found
241 higher morphological diversity in tenrecs compared to golden moles
242 across all three skull analyses (table 1). These results indicate that the
243 overall morphological diversity within tenrecs is not as large as is often
244 assumed (e.g. Eisenberg & Gould, 1969; Olson, 2013) because the majority
245 of the Family are members of a single, morphologically similar Genus.

246 Of course our results are based on a single morphological axis; the
247 diversity of skull shape. It is difficult to quantify overall morphological
248 diversity because any study is inevitably constrained by its choice of
249 specific traits (Roy & Foote, 1997). Many other studies have also used
250 skulls to study morphological variation within species (Blagojević &
251 Milošević-Zlatanović, 2011; Bornholdt et al., 2008), to delineate species
252 boundaries within a clade (e.g. Panchetti et al., 2008) or for
253 cross-taxonomic comparative studies of morphological (dis)similarities
254 (e.g. Ruta et al., 2013; Goswami et al., 2011; Wroe & Milne, 2007).
255 However, variation in skull shape is only one aspect of overall
256 morphology. Quantifying variation in other morphological traits could
257 yield different patterns. Therefore future work should extend our

258 approach beyond just skulls to gain a more complete understanding of the
259 overall morphological diversity of tenrecs and golden moles.

260 We have presented the first quantitative investigation of morphological
261 diversity in tenrecs. We found that tenrec skulls are more morphologically
262 diverse than their closest relatives the golden moles, but only in some
263 aspects of their morphology. Furthermore, our results indicate that the
264 similarities among the species rich *Microgale* tenrecs mask signals of
265 higher morphological diversity among the rest of the Family. Of course
266 our results are restricted to just one axis of morphological variation and
267 further analysis of other traits is required. However, our results represent
268 a significant step towards a more quantitative understanding of patterns
269 of morphological diversity in tenrecs.

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References

- Adams, D., Otárola-Castillo, E. & Paradis, E. 2013. geomorph: an r package for the collection and analysis of geometric morphometric shape data. *Methods in Ecology and Evolution* **4**: 393–399.
- Anderson, M. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecology* **26**: 32–46.
- Asher, R.J., Maree, S., Bronner, G., Bennett, N., Bloomer, P., Czechowski, P., Meyer, M. & Hofreiter, M. 2010. A phylogenetic estimate for golden moles (Mammalia, Afrotheria, Chrysochloridae). *BMC Evolutionary Biology* **10**: 1–13.
- Blagojević, M. & Milošević-Zlatanović, S. 2011. Sexual shape dimorphism in Serbian roe deer (*Capreolus capreolus* L.). *Mammalian Biology - Zeitschrift für Säugetierkunde* **76**: 735–740.
- Bookstein, F. 1997. Landmark methods for forms without landmarks: morphometrics of group differences in outline shape. *Medical image analysis* **1**: 225–243.
- Bornholdt, R., Oliveira, L.R. & Fabián, M.E. 2008. Size and shape variability in the skull of *Myotis nigricans* (schinz, 1821) (chiroptera: Vespertilionidae) from two geographic areas in brazil. *Brazilian Journal of Biology* **68**: 623–631.
- Bronner, G. 1995. *Systematic revision of the golden mole genera Amblysomus, Chlorotalpa and Calcochloris (Insectivora: Chrysochloromorpha; Chrysochloridae)*. Ph.D. thesis.

- 301 Brusatte, S., Benton, M., Ruta, M. & Lloyd, G. 2008. Superiority,
302 competition and opportunism in the evolutionary radiation of
303 dinosaurs. *Science* **321**: 1485–1488.
- 304 Eisenberg, J.F. & Gould, E. 1969. The Tenrecs: A Study in Mammalian
305 Behaviour and Evolution. *Smithsonian Contributions to Zoology* **27**: 1–152.
- 306 Foote, M. 1991. Morphological and taxonomic diversity in a clade's
307 history: the blastoid record and stochastic simulations. *Museum of*
308 *Paleontology, The University of Michigan* **28**: 101–140.
- 309 Foth, C., Brusatte, S. & Butler, R. 2012. Do different disparity proxies
310 converge on a common signal? Insights from the cranial morphometrics
311 and evolutionary history of *Pterosauria* (Diapsida: Archosauria). *Journal*
312 *of Evolutionary Biology* **25**: 904–915.
- 313 Gavrillets, S. & Losos, J. 2009. Adaptive radiation: contrasting theory with
314 data. *Science* **323**: 732–736.
- 315 Goswami, A., Milne, N. & Wroe, S. 2011. Biting through constraints:
316 cranial morphology, disparity and convergence across living and fossil
317 carnivorous mammals. *Proceedings of the Royal Society B: Biological*
318 *Sciences* **278**: 1831–1839.
- 319 Harmon, L., Kolbe, J., Cheverud, J. & Losos, J. 2005. Convergence and the
320 multidimensional niche. *Evolution* **59**: 409–421.
- 321 Hopkins, M. 2013. Decoupling of taxonomic diversity and morphological
322 disparity during decline of the Cambrian trilobite family
323 *Pterocephaliidae*. *Journal of Evolutionary Biology* **26**: 1665–1676.

324 Jenkins, P. 2003. *Microgale, shrew tenrecs*, pp. 1273–1278. The University of
325 Chicago Press, Chicago.

326 Losos, J. 2010. Adaptive radiation, ecological opportunity, and
327 evolutionary determinism. American Society of Naturalists E. O. Wilson
328 Award Address. *The American Naturalist* **175**: 623–639. 10.1086/652433.

329 MacLeod, N. 2013. Landmarks and semilandmarks: Difference without
330 meaning and meaning without difference.

331 Muschick, M., Indermaur, A. & Salzburger, W. 2012. Convergent evolution
332 within an adaptive radiation of cichlid fishes. *Current Biology* **22**: 1–7.

333 Olson, L. & Goodman, S. 2003. *Phylogeny and biogeography of tenrecs*, pp.
334 1235–1242. The University of Chicago Press, Chicago.

335 Olson, L.E. 2013. Tenrecs. *Current Biology* **23**: R5–R8.

336 Panchetti, F., Scalici, M., Carpaneto, G. & Gibertini, G. 2008. Shape and
337 size variations in the cranium of elephant-shrews: a morphometric
338 contribution to a phylogenetic debate. *Zoomorphology* **127**: 69–82.

339 Polly, P.D., Lawing, A.M., Fabre, A.C. & Goswami, A. 2013. Phylogenetic
340 principal components analysis and geometric morphometrics. *Hystrix,*
341 *the Italian Journal of Mammalogy* **24**: 1–9.

342 R Core Team 2014. *R: A Language and Environment for Statistical Computing*.
343 R Foundation for Statistical Computing, Vienna, Austria. URL
344 <http://www.R-project.org/>.

345 Rohlf, F. 2009. Morphometrics at SUNY Stony Brook. URL
346 <http://life.bio.sunysb.edu/morph/index.html>.

- 347 Rohlf, F. 2012. *TPSUtil ver 1.53*. Morphometrics at SUNY Stony Brook.
348 URL <http://life.bio.sunysb.edu/morph/>.
- 349 Rohlf, F. 2013. *TPSDig2 ver 2.17*. Morphometrics at SUNY Stony Brook.
350 URL <http://life.bio.sunysb.edu/morph/>.
- 351 Rohlf, J. & Marcus, L. 1993. A revolution in morphometrics. *Trends in*
352 *Ecology & Evolution* **8**: 129–132.
- 353 Roy, K. & Foote, M. 1997. Morphological approaches to measuring
354 biodiversity. *Trends in Ecology & Evolution* **12**: 277–281.
- 355 Ruta, M., Angielczyk, K., Fröbisch, J. & Benton, M. 2013. Decoupling of
356 morphological disparity and taxic diversity during the adaptive
357 radiation of anomodont therapsids. *Proceedings of the Royal Society B:*
358 *Biological Sciences* **280**: 20131071.
- 359 Serb, J., Alejandrino, A., Otárola-Castillo, E. & Adams, D. 2011.
360 Morphological convergence of shell shape in distantly related scallop
361 species (mollusca: Pectinidae). *Zoological Journal of the Linnean Society*
362 **163**: 571–584.
- 363 Soarimalala, V. & Goodman, S. 2011. *Les petits mammifères de Madagascar*.
364 Guides sur la diversité biologique de Madagascar. Association Vahatra,
365 Antananarivo, Madagascar.
- 366 Stanhope, M., Waddell, V., Madsen, O., de Jong, W., Hedges, S., Cleven,
367 G., Kao, D. & Springer, M. 1998. Molecular evidence for multiple
368 origins of insectivora and for a new order of endemic african insectivore
369 mammals. *Proceedings of the National Academy of Sciences* **95**: 9967–9972.

- 370 Wilson, D. & Reeder, D. 2005. *Mammal species of the world. A taxonomic and*
371 *geographic reference (3rd ed)*. Johns Hopkins University Press.
- 372 Wroe, S. & Milne, N. 2007. Convergence and remarkably consistent
373 constraint in the evolution of carnivore skull shape. *Evolution* **61**:
374 1251–1260.
- 375 Zelditch, M., Swiderski, D. & Sheets, D. 2012. *Geometric Morphometrics for*
376 *Biologists, second edition*. Academic Press, Elsevier, United States of
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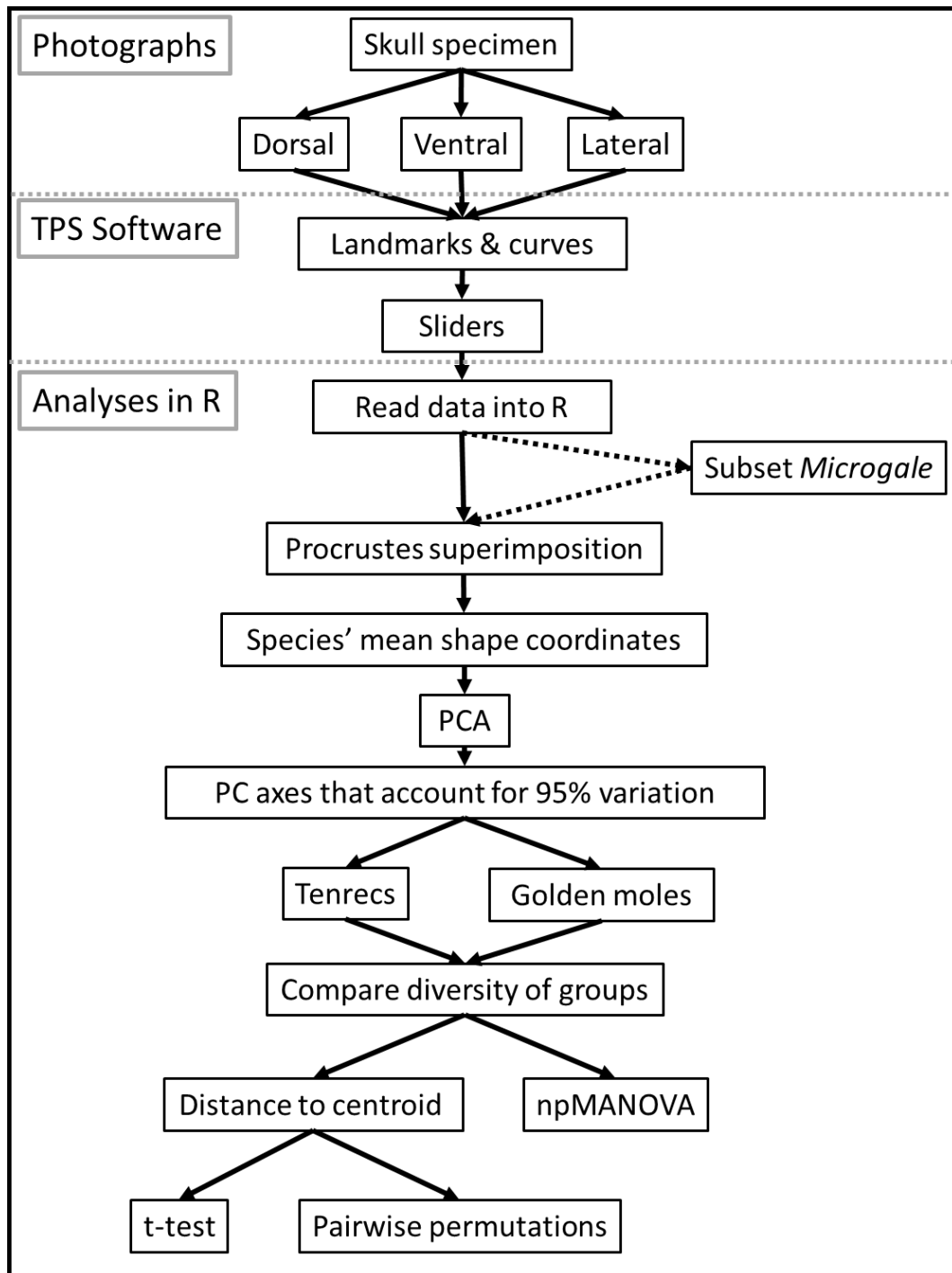


Figure 1: Summary of the main steps in our data collection, processing and analysis protocol. Note that skulls were photographed in three views and the analyses were repeated separately for each view. The dashed arrows refer to the analyses we repeated including only a subset of *Microgale* tenrecs.

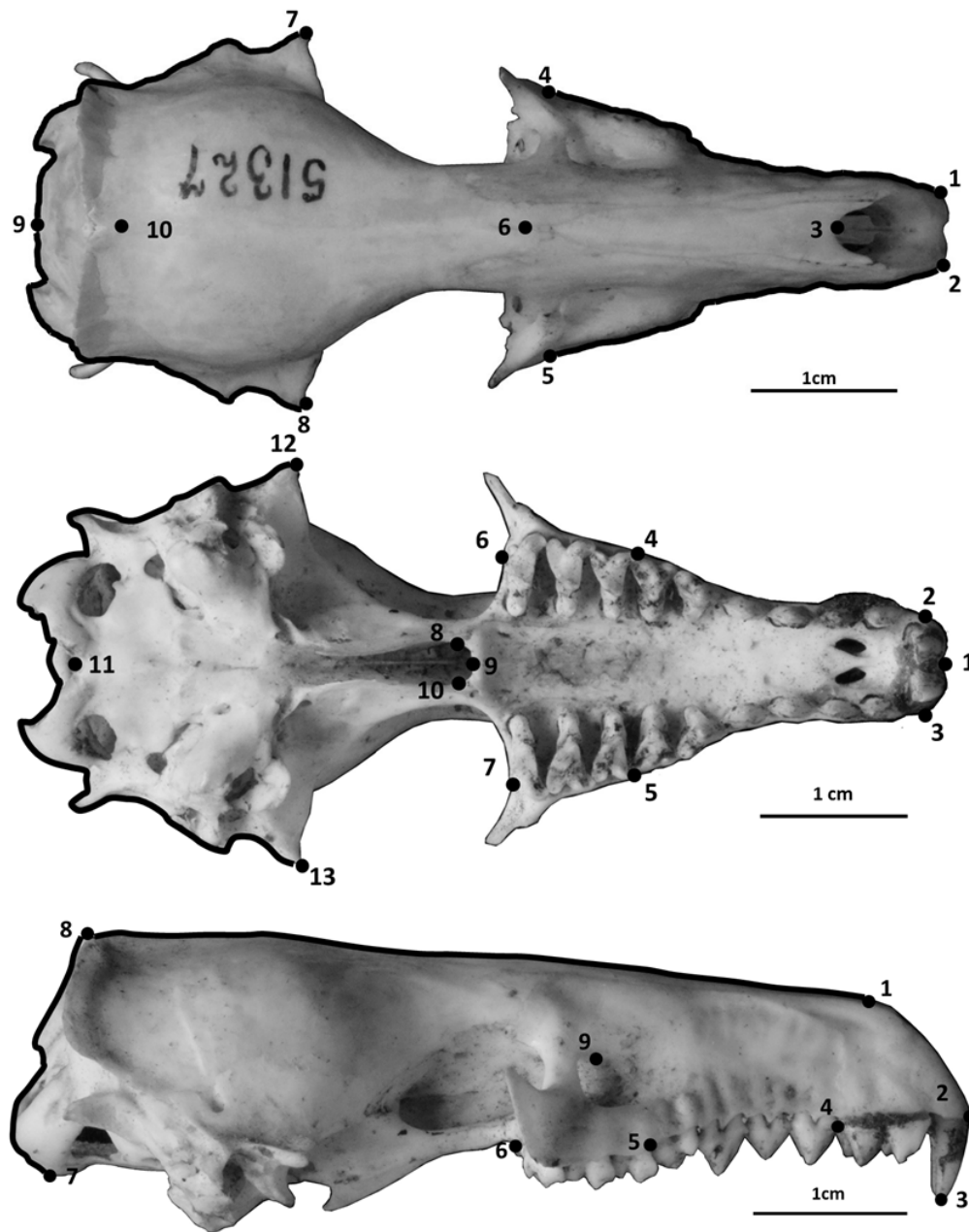


Figure 2: Landmarks (numbered points) and curves (black lines) used to capture the shape of skulls in dorsal, ventral and lateral views respectively. Curves were re-sampled to the same number of evenly-spaced points. See supplementary material for descriptions of the curves and landmarks. The specimens belong to two different *Potamogale velox* (Tenrecidae) skulls: accession number AMNH 51327 (dorsal) and BMNH 1934.6.16.2 (ventral and lateral)

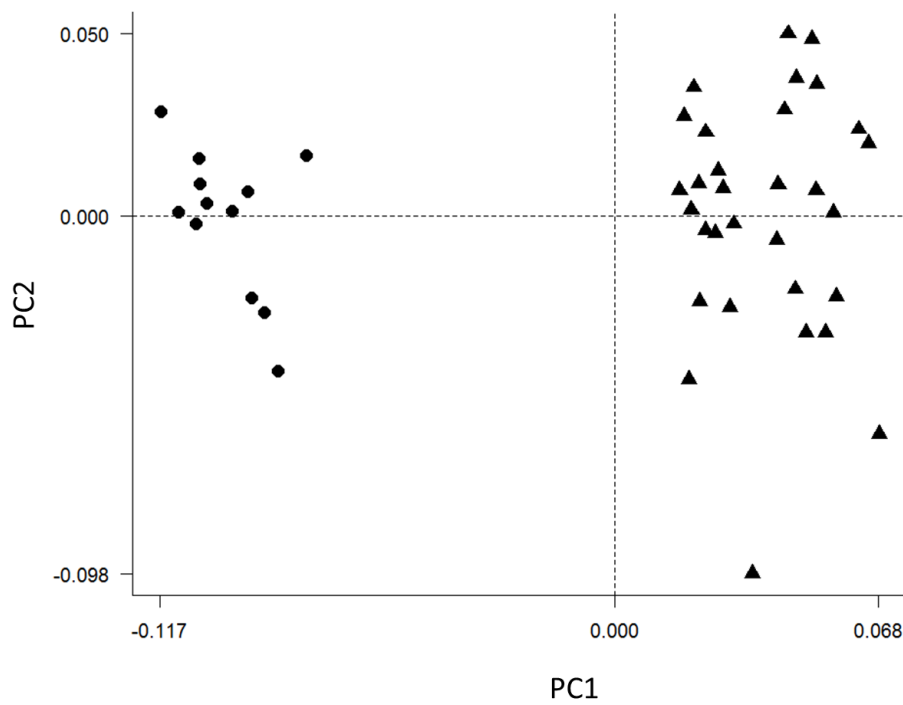


Figure 3: Principal components plot of the morphospace occupied by tenrecs (triangles, $n = 31$ species) and golden moles (circles, $n = 12$) for the skulls in lateral view. Each point represents the average skull shape of an individual species. Axes are PC1 and PC2 of the average scores from a PCA analysis of mean Procrustes shape coordinates for each species.

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Table 1:

Morphological diversity in tenrecs and golden moles for each of the three analyses (skulls in dorsal, ventral and lateral view). Results are shown for all 31 species of tenrec (left) and 19 species of tenrec (right) including just five *Microgale* species. Significant differences ($p < 0.05$) are highlighted in bold.

Tenrec species	Analysis	Tenrecs (mean \pm s.e)	Golden moles (mean \pm s.e)	t	p
31	Skulls dorsal	0.036 \pm 0.0029	0.029 \pm 0.0032	-1.63	0.11
	Skulls ventral	0.048 \pm 0.0034	0.044 \pm 0.0041	-0.676	0.51
	Skulls lateral	0.044 \pm 0.0041	0.032 \pm 0.0037	-2.16	0.04
	Mandibles	0.049 \pm 0.0044	0.067 \pm 0.0054	2.62	0.014
17	Skulls dorsal	0.044 \pm 0.0025	0.029 \pm 0.003	-3.62	0.001
	Skulls ventral	0.054 \pm 0.004	0.042 \pm 0.004	-2.23	0.04
	Skulls lateral	0.054 \pm 0.005	0.031 \pm 0.0037	-3.47	0.002
	Mandibles	0.055 \pm 0.0049	0.062 \pm 0.005	1.003	0.325