

# **PYMOL NOTES**

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# 1. General Manipulation

## category

```
create protein, polymer  
create ligand, organic  
create solvent, solvent  
create ions, resn NA+K+CL+CA+MG+ZN+MN+FE+CU+CO+NI
```

## highlight binding site

```
select binding_site, protein within 5 of ligand  
show sticks, binding_site  
color orange, binding_site
```

## define docking box

```
change wd  
create protein, polymer  
create ligand, organic  
create solvent, solvent  
create ions, resn NA+K+CL+CA+MG+ZN+MN+FE+CU+CO+NI  
select ligand_selection, resn UNK  
run shaded_box.py  
draw_shaded_box()
```

## Show surface of binding pocket

```
show surface, binding_site  
set transparency, 0.5, binding_site
```

## 2. Ligand Observation

### waters near the ligand

```
create waters_near_ligand_obj, solvent within 4 of ligand  
show lines, waters_near_ligand_obj  
color blue, waters_near_ligand_obj
```

### Ions in Binding Site

```
select ions_near_ligand, ions within 5 of ligand  
show spheres, ions_near_ligand  
color yellow, ions_near_ligand
```

### Metal Coordination (if present)

Elements: Zn, Mg, Ca, etc.

```
select zn, resn ZN  
create zn_obj, zn  
distance metal_coord, zn_obj, ligand_obj within 2.5  
set dash_color, white, metal_coord  
set dash_width, 3
```

### Hydrogen Bonds

```
distance hbonds_lig_prot, ligand, protein, 3.2, mode=2  
distance hbonds_lig_solvent, ligand, solvent, 3.2, mode=2
```

### Salt Bridges / Ionic Interactions

Detect charged groups in protein near charged groups in ligand (e.g., acidic/basic residues near ligand ions):

```
select charged_prot_near_lig, (protein and (resn ARG+LYS+HIS+ASP+GLU)) within 4 of ligand  
show sticks, charged_prot_near_lig
```

### PI / PI

```
select arom_prot_near_lig, (protein and resn PHE+TYR+TRP+HIS) within 5 of ligand  
show sticks, arom_prot_near_lig
```

## Label interacting residues

```
label binding_site and name CA, "%s`%s" % (resn, resi)
```

## Orient the view:

```
orient ligand
```

```
zoom ligand, 10
```

### 3. Protein Observation

#### Dynamic Factors

Blue: low B-factor (rigid)

White: intermediate

Red: high B-factor (flexible)

color blue, polymer and b < 20

color white, polymer and b >= 20 and b < 40

color red, polymer and b >= 40

#### Protein / Protein Rx

fetch 1H4G, async=0

select interfaceA, byres (chain A within 4 of chain B)

select interfaceB, byres (chain B within 4 of chain A)

show cartoon, chain A+B

show sticks, interfaceA

show sticks, interfaceB

color red, interfaceA

color blue, interfaceB

distance contacts, chain A within 4 of chain B, chain B within 4 of chain A

label interfaceA, resn + resi

label interfaceB, resn + resi

#### Superimpose Proteins

super ptn\_1, ptn\_2        # ptn\_1 will be pushed to the position of ptn\_2 and align on it (ptn\_2 is the ref.)

#### Fast sequence

print(cmd.get\_fastastr("protein"))

#### hiding H

hide (hydro)

## 4. Images and Videos

ray tracing for final images

ray 1600,1200

png binding\_site\_view.png, dpi=300

**Scenes**

scene 001, store

**video**

mset 1 x 600 # 20 sec (30 frame/sec)

mview store, 1

mview store, 200

mview store, 300

mview store, 400

mplay

mstop

frame 1

**label size**

set label\_size, 14

## 5. Dashes

### dash color

```
set dash_color, red      # Strong visual, good for highlighting  
set dash_color, blue  
set dash_color, green  
set dash_color, yellow  
set dash_color, orange  
set dash_color, cyan  
set dash_color, magenta  
set dash_color, white  
set dash_color, black  
set dash_color, grey50  # Soft neutral tone
```

### dash width

```
set dash_width, 4        # Default is 2; 4 is bolder, good for visibility
```

### dash gab

```
set dash_gap, 0.4        # Default is 0.3; 0.4–0.5 for better separation
```

### dash length

```
set dash_length, 0.3     # Default is 0.2; 0.3–0.4 for better visibility
```

## 6. Labels

### label color

```
set label_color, white      # Common options: white, black, red, yellow, etc.
```

```
set label_color, black
```

```
set label_color, red
```

```
set label_color, yellow
```

```
set label_color, cyan
```

```
set label_color, green
```

### Label size

```
set label_size, 18      # (default = 14) Good for presentations; try 12–24 based on need
```

### Label font

```
set label_font_id, 7      # Common fonts:
```

```
# 5: Sans, 6: Sans Bold, 7: Serif, 8: Serif Bold, 9: Fixed Width (monospace)
```

### Label transparency

```
set label_transparency, 0.3 # 0 = opaque, 1 = fully transparent
```

## 7. colorblind-friendly Commands

```
bg_color white
set antialias=1
set orthoscopic=1
set gamma=1.15
set cartoon_fancy_helices, 1
set cartoon_fancy_sheets, 1
set_color wred, [0.788,0.000,0.140]
set_color wblue, [0.31,0.506,0.686]
set_color wgold, [0.855,0.647,0.125]
set_color wgreen, [0.134,0.545,0.134]
set_color wgray, [0.800,0.800,0.800]
set_color wrose, [0.65,0.47,0.55]
set_color wpurple, [0.37,0.31,0.62]
set_color mpurple, [0.75,0.57,0.80]
set_color mpgrey, [0.73,0.68,0.82]
set ray_shadows, 0
set ray_trace_fog, 1
```

## 8. Reference Values

INTERACTION TYPE	DISTANCE RANGE (Å)	ANGLE CRITERIA (°)	NOTES
<b>HYDROGEN BOND</b>	2.5 – 3.5 Å (ideal ~2.8)	>120° (donor-H-acceptor)	H-bond is directional; angle close to 180° is ideal
<b>SALT BRIDGE (IONIC)</b>	≤ 4.0 Å	—	Between oppositely charged groups (e.g., Lys–Asp)
<b>Π–Π STACKING</b>	3.3 – 4.0 Å	Parallel or T-shaped	Between aromatic rings (e.g., Phe–Phe or ligand ring)
<b>CATION–Π</b>	3.5 – 6.0 Å	—	Between a positive charge (e.g., Lys <sup>+</sup> ) and aromatic ring
<b>VAN DER WAALS</b>	3.0 – 4.0 Å	—	Close but not overlapping atoms; weak individually
<b>HYDROPHOBIC CONTACT</b>	≤ 4.0 Å	—	Nonpolar groups (e.g., Val–Leu, methyl–phenyl)
<b>METAL COORDINATION</b>	1.8 – 2.5 Å (strong)	~90°–180°	For Zn, Mg, Fe, Ca, etc. Usually involves O or N
<b>HALOGEN BOND</b>	3.0 – 3.5 Å	~160°–180°	Between halogen (Br, Cl, I) and O/N; highly directional
<b>WATER BRIDGE</b>	H-bonds through water	Same as H-bonds	Bridging ligand and protein via water molecule
<b>CH–Π INTERACTION</b>	~2.5 – 3.0 Å	—	Between C-H and aromatic system; often overlooked
FEATURE	TYPICAL RANGE	NOTES	
<b>CA–CA (ADJACENT RESIDUES)</b>	~3.8 Å	Peptide bond spacing	
<b>C=O TO N (H-BOND IN HELIX/SHEET)</b>	~2.8 Å	Backbone H-bonds	
<b>H-BOND IN A-HELIX</b>	i → i+4	C=O of residue <i>i</i> bonds to N-H of <i>i</i> +4	
<b>B-SHEET SPACING (BETWEEN STRANDS)</b>	~4.5 – 5.0 Å	Between aligned residues across strands	

When validating interactions or modeling:

- **H-bonds:** Check donor–acceptor distance and angle
- **Salt bridges:** Charged groups <4.0 Å apart
- **π–π:** Parallel rings within ~3.5–4 Å
- **Cation–π:** Aromatic ring within 6 Å of positive group
- **Metal ions:** Coordinating atoms within ~2 Å (depends on metal)
- **Hydrophobic contacts:** Any nonpolar groups within 4 Å
- **Angles:** Especially for hydrogen bonds and halogen bonds