The Effects of Biotic Disturbances on Carbon Budgets of North American Forests

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Forest insects and pathogens are major disturbance agents that have affected millions of hectares in North America in recent decades. The extensive area of outbreaks and large number of trees affected suggest significant impacts to the carbon (C) cycle. Here we present a review and synthesis of published studies of biotic forest disturbances in North America and their effects on C budgets. Substantial variability exists among major disturbance agents in terms of area affected, life history and drivers, and effects on trees. Insects and pathogens can greatly affect carbon budgets. Primary productivity can be reduced considerably following insect or pathogen attack. Following repeated growth reductions or a single infestation by some bark beetles, tree mortality can occur, modifying subsequent stand-level productivity and decomposition for decades. Net C sinks may be reduced following attack, with stands often becoming C sources. Recovery of C stocks and fluxes following disturbance is influenced by the number of trees killed and presence of remaining vegetation, including herbs, shrubs, and trees. We identified substantial knowledge gaps regarding the extent, drivers, and dynamics of biotic disturbance agents and their C cycling impacts, including comparisons across disturbance types and times since infestation and behavior at landscape and regional scales. We conclude that although these outbreaks are broad-scale and highly significant forest disturbances, the limited knowledge about biotic agents, few number of C cycle studies, and diversity of outbreak conditions impart large uncertainties for quantifying effects on North America's carbon budget.

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1. Introduction

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Atmospheric CO₂ is a key driver of past and predicted future climate change [IPCC, 2007]. In addition to anthropogenic emissions of carbon (C) from fossil fuel burning and other activities, exchange of C with the biosphere is crucial for governing the amount of CO₂ (the primary anthropogenic greenhouse gas) in the atmosphere [CCSP, 2007]. Biogenic fluxes of C occur mainly through the interplay of CO₂ uptake by plants (photosynthesis) and release of C back into the atmosphere via autotrophic and heterotrophic respiration (the latter also known as decomposition). There is considerable interest in quantifying the North American carbon budget that has, in part, been driven by national and international reporting obligations [Chapin et al., 2008; Kurz et al., 2008b]. Over the past few decades, North American terrestrial ecosystems are thought to have been a carbon sink as a result of forest regrowth following harvest in past centuries, woody encroachment, and sequestration by wetlands and agricultural soils [CCSP, 2007]. The First State of the Carbon Cycle report estimated a North America sink of 0.5 Pg C yr⁻¹ (1 Pg C = 10^{15} g C = 1 Gt C = 1 billion metric tons of C) in 2003, though with large uncertainty [CCSP, 2007]. Reducing this uncertainty to increase confidence in future predictions of C dynamics and support climate change adaptation and mitigation activities will require research to quantify the magnitude of C sources and sinks and their spatial patterns and controls [Denning, 2005]. Forests play major roles in the global carbon budget through their influence on the dynamics of the terrestrial carbon cycle [Bonan, 2008; Canadell and Raupach, 2008]. Forests contain 422 Pg C, 76% of the C in terrestrial vegetation and about the same amount as in the

atmosphere [Houghton and Skole, 1990]. In the Northern Hemisphere, forests are major carbon

sinks, taking up 0.6-0.7 Pg C yr⁻¹ [Goodale et al., 2002], about 12% of the annual emission of CO₂ by humans [Schimel et al., 2001]. Human and natural changes to forests and environmental conditions have resulted in significant effects on the global carbon cycle. For example, large releases of carbon to the atmosphere have been reported from tropical deforestation [Canadell et al., 2007] and fire and insect outbreaks [Kurz et al., 2008b].

Insects and pathogens are agents of significant disturbance in North American forests. Many outbreaks are highly localized, but some range across millions of hectares and may affect up to 45 times the area of wildfire [Dale et al., 2001]. These disturbances in the US have an average annual economic cost of \$1,500,000,000, five times that of wildfire [Dale et al., 2001]. Wood volume lost to insects in Canada from 1977-1987 was nearly 1/3 of harvest volume, on par with volume lost to disease, and exceeds that from fire [Fleming, 2000]. Insects can have substantial effects on forest productivity [Maclean, 1990b], causing, for example, growth loss and mortality that exceeds 50×10^6 m³/year in Canadian forests [Fleming, 2000]. The large areas of tree mortality or reduced tree growth resulting from major epidemics suggest substantial impacts to the North American carbon cycle. Additionally, biosphere-atmosphere exchanges of carbon influence climate, and climate is an important driver of insect and disease outbreaks [Ayres and Lombardero, 2000; CCSP, 2008]. Thus, important feedbacks exist between climate change and biotic disturbances through the carbon cycle.

Here we review the effects of forest insect and disease disturbances on the North American carbon cycle. We focus on major biotic disturbance events, which we define as large (>100 km²) forest areas affected by insects or disease over time scales less than a decade. We do not consider smaller disturbance events or background herbivory or disease, which we recognize

as important, but instead concentrate on those outbreaks with significant impacts to the North American carbon cycle.

Our review is written for carbon cycle scientists interested in learning more about biotic disturbances and for scientists and land managers familiar with insects and pathogens who want to learn more about how these agents affect carbon budgets in forests. We describe the major insect and pathogen disturbance agents in North American forests and provide an overview of the extent of biotic disturbances in the region. We discuss drivers of outbreaks and note interactions with other disturbance agents. We then review the impacts to the carbon cycle, including expected responses as well as studies that document the influence of insects and pathogens on carbon cycling. We end by identifying gaps in knowledge and describing tools available for closing these gaps.

2. Major insect and pathogen species contributing to forest disturbance

2.1 Insects

Insects contribute highly to the species diversity of terrestrial ecosystems. Most insects exert relatively chronic background herbivory and do not undergo large-scale outbreaks.

However, a few have acute landscape-scale impacts, existing at barely detectable levels for extended periods and then rapidly irrupting to outbreak levels with substantial impacts on forests. It is these landscape-level insect outbreaks that likely have the largest implications for forest carbon cycling. The major tree-level impacts caused by insects include growth reduction, predisposition to subsequent biotic and abiotic stress agents, vectoring or creating infection

courts for phytopathogens, and tree mortality. Each of these tree-level impacts may amplify to stand- and landscape-scale impacts such as canopy thinning, gap formation, or alterations in species composition, stand architecture, and successional trajectories [*Raffa et al.*, 2008].

Here we discuss the major insects of deciduous and coniferous forests common to the United States and Canada (Table 1). All of these insects may cause significant growth reductions, tree mortality, or both [*Maclean*, 1990a]. Areas affected by several representative species were compiled to illustrate variability among species and variability over a 9-year time series from 1999-2007 [*Natural Resources Canada*, 2009; *USDA Forest Service*, 2000; 2002; 2003a; b; 2004; 2005a; 2006; 2007; 2009c]. These time series illustrate that outbreaks of some insect species are more extensive than others, the timing of outbreaks varies among species and across regions, and major outbreaks occur throughout North America (Figure 1).

2.1.1 Bark beetles

Bark beetles (Coleoptera: Curculionidae: Scolytinae) feed within phloem and exert the highest forest impacts of any insect group. Most important species are native. This large group of insects can be classified according to the physiological condition of trees they typically colonize. Most species colonize dead trees, and most of the remainder colonize stressed trees. A few bark beetle species kill healthy trees, generate their own positive feedback with respect to population dynamics, and exert landscape-scale impacts. Tree-killing bark beetles exhibit highly eruptive dynamics, and typically reside in an endemic condition, during which a suite of biotic and abiotic constraints limits their reproductive success. Large-scale factors such as canopy structure, forest homogeneity, and regional climate affect microscale interactions that determine

whether various thresholds are breached that lead to epidemics, and conversely, outcomes of microscale interactions have landscape-scale consequences [*Raffa et al.*, 2008]. All of the tree-killing bark beetles have close associations with fungi, and most are host-specific at the level of tree genus.

Hopkins), which attacks pines across western North America (Table 1). Major outbreaks have occurred in the past, with extensive epidemics ongoing in British Columbia and Colorado, among other locations, and large losses expected in the near future [Krist et al., 2007; Kurz et al., 2008a]. Another major insect, spruce beetle (Dendroctonus rufipennis Kirby), is considered the most important insect of spruce-fir forests in western North America and is one of the major biotic disturbance agents in Alaska [USDA Forest Service, 2009c]. Southern pine beetle (Dendroctonus frontalis Zimmermann) occurs across the southeastern US, attacking multiple species of pines over large areas, and is predicted to cause large losses (54 Mm²) of basal area in the next 15 years [Krist et al., 2007]. Various Ips species in combination with severe drought were responsible for a major mortality event in pinyon pine (Pinus edulis Engelm.) in southwestern US in the early 2000s. Western balsam bark beetle (Dryocoetes confuses Swaine) is currently killing subalpine fir (Abies lasiocarpa (Hook.) Nutt.) across the western US.

2.1.2 Defoliators

Defoliators also exert a high level of impact on tree growth and mortality, but their effects are substantially different from those of bark beetles. Defoliators are the largest group of insects affecting trees and include several orders, of which Lepidoptera is most important. The

most common effects of folivory are reduced growth and reallocation of carbon, although repeated severe defoliation may cause tree mortality [Candau et al., 1998; Elkinton and Liebhold, 1990]. Some defoliator species undergo dramatic cyclic or intermittent population outbreaks [e.g., Zeiraphera diniana Gn., larch budmoth, Esper et al., 2007], but most exert more chronic effects. Folivores do not have the "all or nothing" relationship with their hosts that is characteristic of bark beetles (i.e., killed tree versus failed attack), and tree defenses more often affect insect size, development time, and fecundity [Stevens et al., 2007].

Most leaf-feeders are host-specific at the level of tree genus, but in North America two major species, forest tent caterpillar (*Malacosoma disstria* Hubner) and gypsy moth (*Lymantria dispar* L.; discussed below), have very broad feeding ranges (Table 1). Forest tent caterpillar is a major defoliator of several deciduous trees and has a wide distribution in North America, but has had relatively few reports of widespread mortality [*Volney and Fleming*, 2000]. Severe defoliation in combination with climate-related drought, however, is resulting in increasing areas of aspen (*Populus tremuloides* Michx.) decline [*Allen et al.*, 2010; *Hogg et al.*, 2008].

Budworms feed primarily on the buds of new conifer needles, and defoliation over several years can lead to tree mortality by constraining tree growth or vigor and preventing establishment of needles. Eastern spruce budworm (*Choristoneura fumiferana* [Clem.]) causes more damage than any other insect in North America's boreal forests [*Volney and Fleming*, 2000]. Western spruce budworm (*Choristoneura occidentalis* Freeman) is considered the most widely distributed and destructive conifer defoliator in western North America [*Fellin and Dewey*, 1982].

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Species compositions and relationships are shifting rapidly with accelerated introductions accompanying intercontinental trade [Liebhold and Tobin, 2008]. The underlying causes behind the severe impacts of invasive insects can be classified into two main categories, although each instance is a blend of many forces. First, some insects such as gypsy moth are more eruptive in their introduced region because this region lacks the natural enemy complex present in the native range. Second, a lack of coadaptation between the introduced insect and its new host leads to low resistance and or tolerance [Gandhi and Herms, 2009]. For example, emerald ash borer (Agrilus planipennis Fairmaire) colonizes only highly stressed or dead trees in its native Asian range, as opposed to commonly killing live trees of the genus *Fraxinus* where introduced in North America [Kovacs et al., 2010; Poland and McCullough, 2006]. Likewise, hemlock woolly adelgid (Adelges tsugae Annand) is better tolerated by coadapted trees within its native range, allowing predators time to respond and potentially reduce populations; this lack of coadaptation of trees in the adelgid's introduced range does not permit a similar predator response. Although both mechanisms occur, the latter source of outbreak dynamics (lack of host adaptation) is more threatening to tree species. Some invasive insects are more cryptic than others and so their effects are often unnoticed. For example, a complex of root-feeding weevils is currently affecting fine root processes in northern hardwood ecosystems (Table 1).

Two invasive insects are important forest disturbances whose carbon cycle consequences have been studied. Gypsy moth is an invasive species in North America that was established in the middle of the 19th century and spread throughout the eastern US [*USDA Forest Service*, 2009c] and eastern Canada [*Régnière et al.*, 2009]. This exotic insect is projected to affect 41

Mm² of basal area within the next 15 years [Krist et al., 2007]. A major disturbance agent in forests of the eastern US, the hemlock woolly adelgid attacks hemlocks (Tsuga spp.), feeding on tree fluids. Hemlocks in this region have poor natural defenses against or tolerance of hemlock woolly adelgid, leading to high mortality probability from outbreaks.

2.2 Pathogens and declines

A plant disease is defined as "any malfunctioning of host cells and tissues that results from continuous irritation by a pathogenic agent or environmental factor and causes symptoms" [Agrios, 2005]. Here we focus on pathogens, including infectious diseases (Table 2). Forest pathogens are taxonomically diverse and include fungi, bacteria, viruses, nematodes, and parasitic plants. Fungal pathogens, which are the most common causes of forest diseases, are primarily in the phyla Basidiomycota and Ascomycota [Durall et al., 2005]. Many forest pathogens are specific to a host genus (e.g., Cronartium ribicola J.C. Fisch on Pinus spp.), others have a wide host range including both coniferous and deciduous species (e.g., Armillaria spp.), and a few are host-species specific (e.g., most dwarf mistletoes).

Native forest pathogens tend to act more slowly than other disturbance agents but their effects on carbon cycling can be dramatic, causing significant damage to trees by killing them, slowing their growth, predisposing them to windfall or insect attack, and/or affecting overall wood quality [Hansen, 2000; Tkacz and Hansen, 1982; Woods et al., 2005]. Although some pathogens kill trees that are already stressed or weakened, others (e.g., root pathogens) are able to kill vigorous, dominant trees [Hansen and Goheen, 2000]. Estimates of plant pathogen impacts in North America are not well documented, but some figures are available. Harausz and

Pimentel [2002] reported that pathogens have affected approximately 17.4 million ha and 6% of total annual production in the US. Average Canadian losses in 1982-1986 have been estimated at 45 million m³ per year, which is approximately 16% of production [*Singh*, 1993].

We briefly describe some of the most important functional groups of pathogens (Table 2). Cankers occur as localized wounds or dead areas in the bark of stems or branches. Native cankers generally deform and reduce the growth of trees and predispose them to more destructive agents, although some occasionally girdle trees completely [Agrios, 2005]. Important invasive canker fungi include chestnut blight (Cryphonectria parasitica (Murrill) Barr.), which has extirpated American chestnut (Castanea dentate (Marshall) Borkhausen) through the eastern US [Loo, 2009], and beech bark disease (Neonectria spp.).

Foliar pathogens defoliate trees to varying degrees, reducing photosynthetic capacity and causing concomitant growth loss. Foliar fungi are very sensitive to changes in environmental conditions and can be particularly damaging to host trees planted outside their current range [Kliejunas et al., 2009]. Major outbreaks of Dothistroma needle blight and Swiss needle cast (Phaeocryptopus gäumannii (Rohde) Petrak) have occurred in western North America.

The hyphae of vascular wilt and some blue-stain or sapstain fungi kill trees by plugging their hosts' water- and nutrient-conducting tissues. These fungi are often vectored by insects, such as in the case of Dutch elm disease, a pathosystem that includes species of *Ophiostoma* and two species of bark beetles. Bark beetle-vectored fungi such as *Grosmannia clavigera* (Robinson-Jeffrey and Davidson) Zipfel, de Beer and Wingfield may assist beetles in overcoming tree defenses [*Lieutier et al.*, 2009] and may also be moderately pathogenic [*Plattner et al.*, 2008], although their specific roles remain in dispute [*Six*, 2003].

Root disease fungi usually infect trees through the root system. Many pathogenic root fungi readily infect and kill healthy, otherwise unstressed trees and have long-term impacts on tree growth and survival and on overall forest productivity [Cruickshank, 2000; Cruickshank et al., 2009; Klepzig et al., 1991]. Armillaria and Annosus root diseases have the most widespread occurrence in the forests of North America [Goheen and Otrosina, 1998]. In Canada, stem and root rots have destroyed 25 million and 24 million m³, respectively, of timber per year [Singh, 1993].

Rust fungi require live hosts and usually two botanically unrelated hosts to complete their life cycle. The most important rusts in North American forests are the stem and cone rusts of pines [Sinclair et al., 2005]. The native fusiform rust (Cronartium fusiforme Hedgcock & Hunt ex Cummins) infects loblolly pine (Pinus taeda L.) and slash pine (Pinus elliottii var. elliottii Englem.) in the US [Phelps and Czabator, 1978]. The invasive white pine blister rust fungus (Cronartium ribicola J.C. Fisch Ex. Raben) has severely diminished five-needle pine populations across much of their range in Canada and the US [Geils et al., 2010; Kinloch, 2003; Maloy, 1997].

Phytophthora species infect a wide variety of hosts and have been responsible for severe forest epidemics worldwide, such as the recent sudden oak death epidemic in California and Oregon caused by Phytophthora ramorum sp. nov. The success of Phytophthoras as pathogens is due in part to their rapid reproduction under favorable environmental conditions, which allows inoculum levels to increase to high levels within a few days or weeks [Kliejunas et al., 2009].

Dwarf mistletoes (*Arceuthobium* spp.) are parasitic vascular plants causing significant impacts to a range of conifer species in North America [*Geils et al.*, 2002; *Hawksworth and*

Wiens, 1970; Loomis et al., 1985; Singh, 1993]. Dwarf mistletoes reduce host growth and increase tree mortality and susceptibility to other damage agents.

The term "forest decline" describes stands and/or landscapes where trees, typically of a single species, progressively lose vigor and often die, frequently due to a complex of abiotic and biotic factors [Klepzig et al., 1991; Manion and Lachance, 1992]. Although declines are not solely caused by biotic agents, insects and pathogens may amplify the effects of the predisposing stress factors [Raffa et al., 2008]. A current example of decline is sudden aspen decline in western North America, the causes of which have not yet been delineated [Worrall et al., 2008]. Decline events can result in widespread, cumulative mortality with a concomitant effect on the carbon cycle in forests.

Although they generally do not cause major disturbance events in forested landscapes, wood decay organisms, predominately fungi, play a major role in decomposition and carbon cycling in forests. Dozens of fungal species cause decay or "cull" in millions of trees across North America, many of which remain standing, live or dead, for several years to many decades. In southeast Alaska alone it is estimated that >30% of the volume in living, old-growth Sitka spruce (*Picea sitchensis* (Bong.) Carrière) and western hemlock (*Tsuga heterophylla* (Raf.) Sarg.) trees is cull [*Farr et al.*, 1976]. Wood decay fungi may also have different roles in carbon cycling: brown rot fungi, which leave behind decay high in lignin and recalcitrant to further breakdown, may contribute to more storage of carbon in forests than do white rot fungi, which progressively digest all wood components, including lignin.

Invasive pathogens tend to have more pronounced acute impacts than native species, and, when considered with invasive insects, pose a greater threat to forest ecosystems than do all other non-invasive species combined [Lovett et al., 2006]. Indeed, invasive pathogens are

responsible for some of the most significant long-term changes to North American forests that have been observed, including the functional extinction of a number of keystone taxa across all or most of their range.

3. Drivers of biotic disturbances

Several different types of factors act to initiate, maintain, and terminate outbreaks of insects and pathogens, including climate/weather, predators/pathogens, and host plant condition (stressed versus healthy). Below we discuss the major drivers and their effects on the different biotic disturbance agents.

3.1 Drivers that directly affect insects and pathogens

Temperature is a major driver of both insect and pathogen outbreaks, and operates in multiple ways. For insects, life stage development rates increase with temperature, potentially leading to a switch from two- or three-year life cycles ("semi-voltinism") to one-year life cycles ("univoltinism"), resulting in faster population growth [Bentz et al., 1991; Bentz et al., 2001]. Some insects such as mountain pine beetle are under direct temperature control, facilitating the ability of the insect to take advantage of warmer conditions through more rapid development. Such control also leads to population synchrony for mass attack, often required to overcome tree defenses [Raffa and Berryman, 1987]. Other insects undergo an intrinsic resting state called diapause, which is triggered by low temperatures or shortened photoperiod and acts to maintain

beetle populations in life stages most suited for withstanding cold conditions [*Tauber et al.*, 1986].

Winter temperatures also govern population levels of insects through mortality of overwintering life stages in cold conditions. In response, some insects have evolved the capacity to increase cold-tolerance as the winter progresses [*Wygant*, 1940]. This cold-hardening offers protection at low temperatures, though populations are still vulnerable to extremely cold winter conditions as well as low temperatures during late fall or early spring when cold-hardening is not present [*Régnière and Bentz*, 2007].

Pathogens similarly require temperatures above a minimum to grow and survive. However, pathogens are more directly affected by moisture [*Desprez-Loustau et al.*, 2006]. Several pathogens require a minimum continuous time of wet conditions for infection to take place, and for most pathogens, moisture is necessary for reproduction and dispersal [*Agrios*, 2005].

Disease epidemics are more likely to occur when host susceptibility or pathogen virulence increases, as environmental conditions approach optimum for pathogen growth, reproduction and spread, and as the duration of all conditions favorable to the pathogen is prolonged or repeated [Agrios, 2005]. Virulent pathogens are able to infect hosts quickly to ensure higher amounts of inoculum (= high inoculum potential) and thus disease. Pathogens producing inoculum on aerial parts of hosts are often quickly disseminated by wind and can be dispersed several km [Agrios, 2005]. Pathogens reproducing inside hosts and those that are soilborne and rootborne tend to spread more slowly than airborne pathogens.

Biotic interactions also affect insect/pathogen populations. Predators and pathogens of biotic agents reduce population levels. For the most important bark beetle species discussed in

this review (e.g., mountain pine beetle, spruce beetle), these factors are more important at lower, background population levels. When bark beetle populations are at epidemic levels, the effects of predators and pathogens are greatly reduced [*Turchin et al.*, 1999]. In contrast, defoliators are affected by these factors even during outbreaks [*Hajek*, 1999; *Quayle et al.*, 2003; *Régnière and Nealis*, 2007]. In addition to predation, competition can affect populations. For example, wood borers can reduce bark beetle populations through competition for food resources and facultative predation, potentially reducing the likelihood of a transition from background levels to an epidemic [*Dodds et al.*, 2001].

3.2 Drivers that affect insects and pathogens indirectly

Another major driver of outbreaks is the condition of potential host trees. Most insects and pathogens have one or more preferred host tree species, and so forest species composition determines the extent of a potential outbreak. Outbreaks occurring over large areas require large areas of suitable and susceptible hosts. Host age can play a role, with some insects or pathogens preferring younger hosts, others older hosts. Bark beetles prefer larger-diameter trees that provide more food resource [Shore and Safranyik, 1992]. For pathogens and to a lesser-known extent insects, the degree of genetic uniformity of hosts is an important factor: the higher the degree of uniformity, the greater the likelihood of an epidemic. Diseases of long-lived host plants such as trees usually take longer to develop than those of annual crops.

Some insects and pathogens prefer or have easier access to stressed or low vigor trees, which may be less well defended against attack [*Raffa et al.*, 2008]. Hosts predisposed to attack by drought, defoliation, or other stresses often lead to population increase of bark beetles and

latent pathogens. Older trees can be less vigorous and are less able to defend themselves. High-density stands with resultant increased competition for resources, reduced soil fertility, and severe drought can lead to stressed individuals and reduced resistance. Such relationships are not universal, however. For example, folivorous insects generally perform equally well on vigorous and stressed trees. Some insects prefer healthy trees for their superior nutritional content. For example, some sap feeding and shoot feeding insects typically perform better on healthy trees [Koricheva et al., 1998]. Likewise, pathogens show complex and species-specific relationships with host stress. In addition to resistance effects, a tree's ability to tolerate a given amount of feeding or infection often declines with increasing stress.

Host phenological synchrony and nitrogen availability are particularly important for defoliators [*Régnière and Nealis*, 2007]. Early-season foliage tends to be more succulent and lower in defensive compounds than later-season foliage. Many herbivores are nitrogen-limited, so any factor that increases available foliar nitrogen may shorten insect development time, increase survival, and enhance fecundity.

3.3 Outbreak collapse

Outbreaks of insects and pathogens collapse for varying reasons. Within stands or in extensive outbreaks that occur over large areas, outbreaks can collapse from lack of available remaining food resources [Safranyik and Carroll, 2006a]. Natural enemies play important roles in the population dynamics of folivores [Elkinton and Liebhold, 1990; Régnière and Nealis, 2007], which are more exposed than internally feeding insects such as bark beetles. A common trend is for generalist predators such as birds, small mammals, and spiders to constrain low

populations, but once an outbreak commences, these generalists often lack the reproductive capability to respond, and so parasitic insects and entomopathogens become more important.

Weather factors such as extremely cold winters [Safranyik and Linton, 1991] or low year-round temperatures [Logan and Powell, in press] can lead to outbreak collapse.

3.4 Human influences

Anthropogenic emissions of greenhouse gases have resulted in a warming climate that has facilitated outbreaks [Carroll et al., 2004; Logan et al., 2003], and future patterns of insect and pathogen infestations are likely to change in response to projected climate change [Bentz et al., 2010; Hicke et al., 2006; Kliejunas et al., 2009]. Management activities that include selective harvesting, fire suppression, and use of pesticides have led to older, denser stands and/or stands with a higher percentage of host tree species in some forest types, increasing susceptibility to attack by insects and pathogens [Anderson et al., 1987; Blais, 1983; Taylor and Carroll, 2004]. Other forest management practices such as planting even-aged monocultures have led to higher incidences of insect and pathogen outbreaks [Jactel and Brockerhoff, 2007]. The increase in commerce has resulted in increases in invasions by exotic insects and pathogens.

The ability of humans to control epidemics once started varies among insect and pathogen species. Some groups such as bark beetles and most invasive pathogens are effectively immune to human control during epidemics except at a very limited scale such as by treatment of high-value trees with pesticides. Populations of other insect and pathogen species causing defoliation can be controlled by aerial spraying of microbial or synthetic pesticides [*Carisey et al.*, 2004; *Johnson et al.*, 2003; *Van Frankenhuyzen*, 2000]. Silvicultural practices can modify

forest stand densities, species composition, size classes, and risk of fungal infection (e.g., sterilizing cut stump surfaces) that affect tree vigor and resistance to attack by biotic agents [Durall et al., 2005; Fettig et al., 2007].

4. Disturbance interactions

Insect or pathogen impacts are best understood within the context of interactions with other biotic and abiotic agents. For example, root pathogens, root herbivores, and defoliators can predispose trees to attack by bark beetles and wood borers, and defoliators can release latent pathogens such as some species of *Armillaria*. Insects are frequently the vectors for pathogen dispersal, e.g., symbioses between fungi (*Ophiostoma montium* (Rumbold) von Arx, *Grosmannia clavigera*) and *Dendroctonus* bark beetles [*Bleiker and Six*, 2007], while conversely trees whose defenses are compromised by pathogens or fire also become more susceptible to insects [*Klepzig et al.*, 1995; *Lombardero et al.*, 2006]. Complex biotic interactions can strongly affect the resilience of forest ecosystems to herbivory. For example, whitebark pine (*Pinus albicaulis* Engelm.) faces the combined and interacting threats of mountain pine beetle and white pine blister rust, which affects all age categories [*Six and Adams*, 2007]. Outbreaks of forest tent caterpillar or gypsy moth in oak stands are susceptible to subsequent lethal colonization by two-lined chestnut borer (*Agrilus bilineatus* (Weber)).

Insect and pathogens interact with drought and heat to affect forest carbon cycling. A recent global assessment showed that over the past few decades there has been a significant, increasing trend in reporting of drought- and heat-related forest die-off events, including numerous events where biotic agents played either a primary or secondary role [Allen et al.,

2010]. Biotic disturbances and interaction have also be suggested as potential causes of increased background tree mortality rates [van Mantgem et al., 2009]. Trees suffering from drought or other abiotic stress generally have a higher likelihood of infection by pathogens, even though drought generally has a negative effect on most pathogens [Desprez-Loustau et al., 2006].

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Insects and pathogens interact with other abiotic disturbances as well. Infestations interact with wildfire in two ways [Jenkins et al., 2008; Parker et al., 2006; Romme et al., 2006]. First, outbreaks kill trees, which modifies fuel and fire hazard in complex patterns that vary with time since outbreak [e.g., Jenkins et al., 2008]. However, changes to fire probability or behavior is less well known [Bentz et al., 2009]. Second, fires may affect the likelihood and severity of insect and pathogen outbreaks by altering species composition or stand structure in the long term and by injuring surviving trees in the short term. Large, stand-replacing fires in some areas of North America historically produced landscapes dominated by forests of similar age, structure, and host susceptibility that favor higher outbreak severity and larger extent. Trees that are damaged by fire but still alive become stressed, resulting in a predisposition to secondary host infection that can contribute to physiological weakening, reduced growth rates, and mortality. Empirical studies suggest that insect attack rates are often higher on fire-injured trees, though the strength of these effects varies by insect species, even within a feeding group [Simard et al., 2008]. In addition to fire, interactions occur between biotic disturbances and wind. Nonhost tree species may be more exposed to wind during large-scale defoliator outbreaks. Pathogens can also increase the risk of wind damage by weakening stems or roots [*Papaik et al.*, 2005].

5. Impacts on carbon cycling

Various terms are used to describe carbon cycling. *Carbon stocks* (units of g C or g C m⁻²) are reservoirs of carbon in various tree and soil pools, such as stem, foliage, roots, and organic matter. *Carbon fluxes* (g C m⁻² year⁻¹) are transfers between stocks; here, we focus principally on exchanges between the atmosphere and either vegetation or soil. *Gross primary production* (GPP) is the total amount of atmospheric C fixed by plants. *Autotrophic respiration* (R_a) releases C to the atmosphere by the plants themselves; *heterotrophic respiration* (R_h) is C released by microbes through the decomposition process. *Net primary production* (NPP) is GPP – R_a, and represents the net amount of carbon fixed by plants and available for subsequent use by heterotrophs. *Net ecosystem production* (NEP) is NPP – R_h and defined as positive when ecosystems are sinks of C. *Net ecosystem exchange* (NEE) is the exchange of carbon between the ecosystem and the atmosphere, is defined as positive when there is a net transfer of C to the atmosphere, and is often the same as (but opposite in sign to) NEP. *Net biome production* (NBP) includes NEP as well as losses of C through such processes as combustion or leaching. Chapin *et al.* [2006] describe the differences between these fluxes in greater detail.

5.1 Expected responses of terrestrial carbon cycle to biotic disturbances

Effects of insect and pathogen outbreaks on forest carbon cycling vary with biotic disturbance and host species, forest ecosystem type, and severity of outbreak, but general patterns regarding these interactions exist, based in part on understanding of forest processes following disturbances [Fleming et al., 2002; Hicke et al., 2003; Kashian et al., 2006; Litvak et

al., 2003; Odum, 1969] (Figure 2). First, biotic disturbances alter the productivity (GPP) of stands through defoliation or other growth reductions or tree mortality. Impacts on productivity may be positive or negative depending on the severity of the outbreak and time since the disturbance, though for large-scale outbreaks, short-term effects are typically negative.

Second, trees killed by biotic disturbances alter carbon cycling through increases in heterotrophic respiration (decomposition). Carbon in more labile structures such as leaves is released to the atmosphere first, although in killed conifers, needles remain on the dead trees for 1-3 years following bark beetle attack, delaying this decomposition. During outbreaks, defoliators consume C in leaves and release large fluxes of nitrogen, carbon, and other nutrients into forest ecosystems through leaf fragments, frass, and exuviae [*Frost and Hunter*, 2004]. Snags may remain standing for decades following tree death [*Mitchell and Preisler*, 1998], further delaying decomposition. As a result of these processes as well as the long decay time of wood, the release of carbon through R_h following disturbance occurs over a period of decades to centuries.

Reductions in plant productivity (NPP) and increases in decomposition of dead organic matter (R_h) result in large reductions of NEP. The amount of change of these individual fluxes and therefore of the net carbon flux is determined by the magnitude of an outbreak's impact on stands. For severe outbreaks, the magnitude of the effect is large enough to cause a stand to switch from a carbon sink to a carbon source in a manner similar to the effects of wildfire [Fleming et al., 2002].

Responses of the carbon cycle are highly dependent on type of biotic disturbance.

Several examples illustrate this point. A short-term (e.g., one month) outbreak of a defoliator may have a large but short-lived impact on carbon fluxes. A bark beetle epidemic can kill many

rees within a stand within one year, having large and long-lasting effects on carbon budgets. Pathogen and defoliator epidemics that occur over several years before killing a tree can have a large, long-lived effect that is more muted than the bark beetle example. Variability in C cycle responses occurs not only at the tree level but also at the stand level, where the number of trees affected and length of outbreak are important factors.

Insect and pathogen outbreaks exert a number of indirect effects on carbon cycling as well. Perhaps most importantly, moderate to severe outbreaks may alter functional attributes of the ecosystem (e.g., light regime, water availability and loss, nutrient cycling) in a way that affects the trajectory of succession and recovery. Outbreaks severe enough to alter soil moisture and nutrients, for example, cause changes in tree density or species composition such that long-term carbon storage or the rate of carbon cycling is also altered. At longer and broader scales, carbon loss to the atmosphere from severe outbreak areas may drive climate feedbacks that could alter the frequency and severity of future insect or pathogen outbreaks. Finally, killed trees resulting from insect and pathogen outbreaks may interact with other disturbances to affect carbon cycling, particularly with fire and wind damage (see discussion above).

There is increasing evidence that invasive insect and pathogen outbreaks may have a large impact on forest carbon cycling because of their effects on disturbance regimes, nutrient cycling, and other processes in ecosystems with which they have not co-evolved [*Peltzer et al.*, 2010].

Biotic disturbances that cause extensive tree mortality may be comparable to fires in terms of impacts to carbon budgets [Fleming et al., 2002; Kashian et al., 2006]. Many biotic disturbances have less immediate impacts than fires, such as defoliation events that kill trees only after several consecutive years of attack [Fleming et al., 2002]. Other outbreaks, such as

those of some bark beetles, can kill many trees within a stand within one year, acting more similar to fire. Differences exist, however: fires result in an immediate release of some carbon to the atmosphere via combustion; severe fires kill nonhost species, understory trees, shrubs, and herbs in addition to host trees; and although outbreaks modify some soil properties such as biogeochemical cycling [e.g., *le Mellec and Michalzik*, 2008; *Morehouse et al.*, 2008], fires can also cause microbial mortality through heating, increase weathering rates, reduce surface albedo, and affect infiltration and runoff rates [*Neary et al.*, 2005].

5.2 Documented impacts of insects and disease on forest carbon cycling

Relatively few studies have quantified the impacts of insect and disease outbreaks on forest carbon budgets. To facilitate comparison, these studies were summarized by insects (Table 3) and pathogens (Table 4). For summarizing and comparing studies, key factors included the type of insect or pathogen (especially whether or not the damage agent killed trees); the severity of disturbance in terms of the number of trees attacked within a stand and the amount of tree mortality; the time since disturbance; the methods of the study and the carbon variable(s) studied; and the existence and type of study controls that illustrate effects relative to unaffected stands or preoutbreak conditions.

We identified 16 studies that assess impacts to North American carbon cycling by insect outbreaks (Table 3) and four studies that documented impacts by pathogens (Table 4). The insect studies were roughly divided among bark beetles, defoliators, and hemlock woolly adelgid, a fluid feeder. Multiple studies of mountain pine beetle, spruce budworm, forest tent caterpillar, gypsy moth, and hemlock woolly adelgid have been published.

Most studies were field observations of carbon cycling [Forrester et al., 2003; Jenkins et al., 1999; Kizlinski et al., 2002; Morehouse et al., 2008; Nuckolls et al., 2009; Pfeifer et al., 2010; Romme et al., 1986], including eddy flux tower measurements [Brown et al., 2010; Clark et al., 2009; Cook et al., 2008; Hadley et al., 2008]. As such, there was an emphasis among studies of tree- to stand-scales. Analyses of eddy flux tower observations were opportunistic (i.e., unplanned), although in one study sites were selected specifically to investigate mountain pine beetle impacts [Brown et al., 2010]. The pathogen studies addressed needle to plot scales. Several studies used carbon cycle models informed by or parameterized with field observations and concentrated on recent times since outbreak [Cook et al., 2008; Schäfer et al., 2009]. A few studies estimated regional impacts using upscaling of finer scale results, simulation modeling, or remote sensing [Albani et al., 2010; Clark et al., 2009; Dymond et al., 2010; Hicke et al., 2002; Kurz and Apps, 1999; Kurz et al., 2008a], and several assessed effects on time scales of decades to centuries [Dymond et al., 2010; Hogg, 1999; Kurz and Apps, 1999; Kurz et al., 2008a; Pfeifer et al., 2010]. Only one study considered the effects of multiple species [Kurz and Apps, 1999]. Insect and pathogens typically caused reductions in tree- and stand-level NPP immediately following attack [Clark et al., 2009; Cook et al., 2008; Dymond et al., 2010; Hogg, 1999; Manter et al., 2003; Meinzer et al., 2004; Pfeifer et al., 2010; Romme et al., 1986; Schäfer et al., 2009], although one study of beech bark disease reported no change in aboveground NPP across a gradient of impact [Hancock et al., 2008]. Following a disturbance, NEP decreased from large positive values (strong C sinks) to weaker positive values or switched to negative NEP values (C sources) [Albani et al., 2010; Brown et al., 2010; Clark et al., 2009; Cook et al., 2008; Dymond et al., 2010; Kurz and Apps, 1999; Kurz et al., 2008a]. Variability in NEP

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reduction was related to the number of trees affected by defoliation or mortality, amount of leaf area reduced by defoliation, and contribution of the understory to carbon uptake. These patterns of NEP change occurred at local scales as well as regional scales during extensive tree mortality. Studies were inconsistent in their conclusions about soil and forest floor carbon impacts, with some noting modified carbon stocks, C:N values, and soil CO₂ efflux in infested stands compared with uninfested stands [*Hancock et al.*, 2008; *Kizlinski et al.*, 2002; *Nuckolls et al.*, 2009], and some reporting no change in one or more variables [*Jenkins et al.*, 1999; *Kizlinski et al.*, 2002; *Morehouse et al.*, 2008].

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Fewer studies assessed recovery and long-term impacts. Transient (one year) defoliation resulted in recovery of NEP in the year after the outbreak [Cook et al., 2008]. In contrast, the effects on NPP and NEP from tree mortality following bark beetle outbreaks or repeated defoliation persist longer [Kurz and Apps, 1999; Kurz et al., 2008a; Pfeifer et al., 2010]. Regionally, simulated NEP in a region of an extended outbreak of mountain pine beetle remained a net carbon source for the study period of two decades [Kurz et al., 2008a], and a simulated spruce budworm epidemic affected NEP for over a decade [Dymond et al., 2010]. Localized studies indicated rapid recovery of aboveground carbon stocks or NPP to preoutbreak values or uninfested stands within one to several decades [Forrester et al., 2003; Pfeifer et al., 2010; Romme et al., 1986]. Several decades following an outbreak, stand productivity may be higher than preoutbreak values because the growth of surviving trees accelerates after release from competition [Romme et al., 1986], although this pattern did not occur in simulated NPP recovery following an outbreak in a different area [Pfeifer et al., 2010]. The speed and magnitude of the recovery of productivity depend on the severity of the outbreak (number of trees affected), composition of the surviving trees within the stand, and seedling establishment

rate [Pfeifer et al., 2010].

Several studies noted some of the regional forest management implications of bark beetle outbreaks. *Kurz et al.* [2008b] described the significant implications of the 1990 baseline used by the UN Framework Convention on Climate Change for forest carbon stocks, a time when Canadian insect outbreaks were minimal, compared with inventory and models suggesting that the trajectory of Canadian insect outbreaks under climate change and land management may approach 30% of fossil fuel emissions. Using simulation modeling, *Seidl et al.* [2008] show that the interaction of European spruce bark beetle (*Ips typographus* L.) outbreak severity and climate change strongly modifies future carbon stocks of central European Norway spruce (*Picea abies* (L.) Karst.) forests.

Although the focus of this paper is on North America, we include a brief discussion on the few relevant studies outside of this region. Studies of short-term carbon cycling impacts based on eddy flux tower observations reported reductions in photosynthesis and NEP for sapsucking [Kirschbaum et al., 2007] and defoliating [Allard et al., 2008] insects. Carbon fluxes to the soil and C:N ratios in frass were greatly increased in areas attacked by pine lappet moth (Dendrolimus pini L.) [le Mellec and Michalzik, 2008; le Mellec et al., 2009], and experiments of the effects of a fungal pathogen found reduced plant productivity following infection [Luque et al., 1999]. These few findings from studies in Europe and Australia are consistent with those in North America.

6. Knowledge gaps

Our review of published studies indicates substantial gaps in knowledge. A clear need is additional scientific understanding of a broader range of biotic disturbance agent types. The natural history, ecology, and population drivers of only a few insects and pathogens (i.e., those of historical importance) have been studied. Moreover, even for the best-studied insects (e.g., mountain pine beetle), we lack sufficient knowledge about drivers to build robust predictive models of population dynamics important for carbon cycle studies, and some of the better-understood drivers are rapidly changing with a variety of anthropogenic conditions. Less is known about most pathogens, and biotic disturbance agents in some North American regions (especially Mexico) are little understood though may be quite important [Salinas-Moreno et al., 2010].

In addition to natural history, we *lack quantitative understanding of the spatial and temporal characteristics of insect and pathogen outbreaks*. Aerial surveys are undertaken annually in the US and Canada to provide information about general trends and are rich in spatial, temporal, and attribute information. However, these data sets have limitations as noted below in Section 7.2. Even if impact areas were well known, *uncertainties exist on how to convert area or number of affected trees to metrics important for carbon cycle studies* (e.g., leaf area or carbon pools). For insects and pathogens that do not kill trees, identification of impacts to carbon cycling based on affected area is very difficult because these disturbances often progress slowly, making detection difficult, and we lack much understanding of effects of these agents on trees.

For most species, we lack knowledge of life history traits and drivers that can be used as a basis for predictive modeling. For those insects and pathogens for which we have such knowledge, additional research is needed to integrate insect/pathogen responses with host responses as well as to evaluate predictions. In addition, our predictive capability of invasions by exotic insects and pathogens is limited. Invasions may take the form of range expansion into formerly unoccupied habitat or successful introductions of exotic species. Each has the potential to exert large impacts to carbon stocks and fluxes, yet our ability to estimate when, where, and in which host species invasions will occur is very limited.

General conclusions about C cycle processes are subject to *large uncertainty as a result* of the lack of replication of studies. Few studies of impacts on carbon cycling exist, limiting comparisons among studies. Furthermore, replicating impacts of similar insect/pathogen and host type under similar conditions, particularly amount of defoliation and/or mortality and time since disturbance, is challenging. For example, whether a bark beetle-infested stand is a net C source or sink depends on the extent of tree mortality, number and size of surviving trees, spatial pattern of mortality, and time since outbreak, among other processes. Attempts to match conditions among published studies resulted in very few meaningful comparisons (Tables 3 and 4).

The lack of existing C cycle studies implies several gaps in knowledge. The majority of existing studies concern insect outbreaks; *very few studies of impacts of pathogen outbreaks on C cycle* have been reported. Even so, the number of insect species studied is limited to mountain pine beetle, hemlock woolly adelgid, gypsy moth, forest tent caterpillar, and a few other defoliators. *Additional studies are needed to quantify the changes to carbon pools and fluxes associated with different types and severities of outbreaks*. These changes include those from

direct effects, such as loss of GPP, loss of leaf area, changes in plant allocation patterns, and changes in rates of leaf, root, and coarse woody debris inputs to the soil. In addition, studies of indirect effects and ecosystem feedbacks such as changes in microclimate, albedo, heterotrophic respiration, and species composition are needed. The production and decomposition of coarse woody debris is not well characterized. Snagfall rates are important for controlling rates of decomposition, yet little is known about rates and drivers. Seedling establishment is another important yet poorly understood process often following outbreaks. The size and number of surviving trees is key to determining how rapidly postoutbreak NPP increases, yet we have little information about how these factors vary within and among outbreaks. More information is needed about the dynamics of C stocks and fluxes following these disturbances.

Interactions with other disturbances are not well quantified. The interactions between drought and heat stress and biotic disturbances as drivers of tree mortality events are poorly understood. There is currently only limited mechanistic knowledge of the physiological processes and linkages involved [Adams et al., 2009; Breshears et al., 2009; McDowell et al., 2008]. Another feedback loop that has received little attention to date is how large-scale outbreaks of biotic agents might affect such biophysical processes such as leaf area, forest transpiration/stomatal conductance, and albedo [Adams et al., 2010; Bonan, 2002; Hogg et al., 2000]. Modifications to these processes drive changes in surface energy and water budgets, affecting local and regional weather and climate patterns (including amplification of drought and heat stress).

Limited information exists on landscape- to continental-scale impacts on carbon budgets.

Effects of major outbreaks of most large-scale insect and pathogen outbreaks have not been studied. In addition to North America, forest dieoff on other continents has been reported, due in

part to biotic disturbance [Allen et al., 2010], suggesting major modifications to carbon budgets in these other regions.

Biotic disturbances are missing from many carbon cycle models. Possible model modifications to include these disturbances may range from simple (fractional removal of carbon pools as a function of biotic outbreak drivers) to statistical (probability based stochastic outbreaks) to predictive, dynamic (coupling biotic population models to carbon cycle models) changes. Key challenges remain in conceptualizing the level of complexity needed to accurately constrain carbon budget impacts from insect and pathogen outbreaks.

7. Available tools

7.1 Field observations

Methods and tools available for understanding processes associated with carbon cycle impacts using field observations are well established. Techniques to measure carbon stocks and fluxes with field equipment have been widely applied. Similarly, carbon budgets using measurements from eddy flux towers are well understood. One issue that arises in some regions is that forests and their biotic disturbances occur in complex terrain. Such terrain induces advective air flows within and above the canopy that may pose significant challenges for interpreting results from eddy flux towers [*Sun et al.*, 2010]. In addition, flux tower data are spatially integrative, leading to challenges of differentiating effects of variations in outbreak severity that may occur within their spatial footprint.

Ground surveys and pheromone trapping are critical for providing precise information regarding field conditions. Surveys allow for the identification of specific insects or diseases, collection of samples for further analysis (e.g., genomic/molecular characterization, tree defense chemistry), assessment of wood damage/growth reduction, the persistence of reservoir populations at levels not detectable by other means, and early detection of the incidence or population increase of most pathogens and insects. The limitations of ground surveys are their cost, personnel, and difficulty of capturing fine-scale, patch patterns across large spatial scales through point sampling. However, some networks such as the USDA Forest Service Forest Inventory and Analysis (FIA) program or the Canadian Forest Inventory (CanFI) may provide the ability to assess changes to carbon budgets following outbreaks [*Thompson*, 2009a; *Thompson*, 2009b]. Additional relevant information such as disease is collected on a subset of the FIA plots. The Mexican CONAFOR (Comisión Nacional Forestal) is currently in the planning stages for a forest health survey (B. Tkacz, pers. comm.).

Several notable challenges exist when using field observations. Biotic disturbances are relatively sparse in time and space, and we currently lack the capability to generate accurate short-term predictions of these disturbances at local spatial scales. This inability to forecast outbreaks suggests difficulty in setting up studies in advance of a disturbance. Establishing a control for comparison of effects is also a challenge, whether in time (forecasting an outbreak to obtain pre-outbreak measurements) or space (finding similar stands and environmental conditions). Developing chronosequences of time since disturbance that substitute space for time requires identification of similar stands with similar levels of impact across decades, yet identifying old stands becomes more difficult as the biotic disturbance evidence fades with time.

7.2 Aerial survey databases

The detection, mapping, and monitoring of the spatial location, areal extent, and severity of forest disturbances have been long-standing activities in North America [*Tkacz et al.*, 2008]. Aerial surveys are important tools and consist of trained observers delineating damaged areas from aircraft and estimating disturbance severity, host type, and causal agent [*Ciesla*, 2000; *McConnell et al.*, 2000]. In the United States, the annual Aerial Detection Survey program is undertaken in collaboration with various state and federal agencies, with regional data available from 1997 to the present and subsets available further back in time [e.g., *USDA Forest Service*, 2009c]. In Canada, the Forest Insect and Disease Survey of the Canadian Forest Service conducted annual surveys [*Hall et al.*, 1998] until 1996, when surveys largely became the responsibility of provincial and territorial governments.

Historical and current survey maps can contain limitations that make comparisons problematic, including its subjective nature and range of spatial precision [de Beurs and Townsend, 2008; Harris and Dawson, 1979; McConnell et al., 2000]. Furthermore, spatial and temporal extents are limited. In Canada, aerial surveys are largely confined to managed forests, resulting in an omission of damage information in remote areas, and are limited to the extent that they can be used alone to relate defoliated areas to impact [MacLean, 1990]. In the US, surveys were historically limited to timber-producing tree species and did not include national parks or wilderness areas. Aerial surveys include live trees within damage polygons, and this "affected area" is an overestimate of the area of mortality. Approaches to address this problem include deflation factors to convert the affected area to area of tree mortality [Kurz and Apps, 1999] as

well as combining aerial survey information with remotely sensed imagery to identify the specific area in which the disturbance has taken place [Rogan et al., 2006].

Aerial surveys have two additional important limitations. First, they are generally ineffective for identifying outbreaks of most pathogens, except in cases where a single agent damages large patches of one dominant host species that responds with a distinct foliar pathology. Unfortunately, the patchy, discontinuous nature of disease occurrence associated with many native pathogens makes their distribution and severity very difficult to accurately assess from the air. For example, *Morrison et al.* [2000] found incidence of *Armillaria* root disease to be significantly higher than indicated by aboveground symptoms only. Secondly, aerial surveys typically cannot detect signals of insect activity until key thresholds separating different population phases have already been breached [*Safranyik and Carroll*, 2006b], and so have limited utility for studying sub-outbreak dynamics or underlying mechanisms of transition.

7.3 Satellite imagery

There is a long history in using remotely sensed satellite imagery for assessing insect outbreaks in forest canopies. Medium-resolution (30-80 m) satellite images have been used to assess defoliation caused by a range of species, including forest tent caterpillar [Hall et al., 2003; Hall et al., 2006], gypsy moth [Nelson, 1983; Williams and Nelson, 1986], spruce budworm [Franklin and Raske, 1994; Leckie and Ostaff, 1988], mountain pine beetle [Franklin et al., 2003; Wulder et al., 2006], jack pine budworm (Choristoneura pinus) [Hopkins et al., 1988; Radeloff et al., 1999], and hemlock wooly adelgid [Royle and Lathrop, 1997]. Reasonably accurate satellite image classification results have been obtained by classifying single images

[e.g., *Ahern*, 1988; *Radeloff et al.*, 1999] as well as by assessing interannual spectral changes using multi-temporal data sets [e.g., *Goodwin et al.*, 2008; *Vogelmann et al.*, 2009].

Several key characteristics determine the usefulness of satellite imagery for quantifying outbreaks. Spatial resolution is important: smaller outbreaks can be detected using data from high spatial resolution sensors (2-4 m), such as IKONOS or QuickBird [Coops et al., 2006; Hicke and Logan, 2009]. Many studies have used medium-resolution imagery (~30 m, such as Landsat) [Hall et al., 2006; Vogelmann et al., 2009; Wulder et al., 2006], and more recent studies of large outbreaks included coarse-resolution imagery such as Systeme Pour l'Observation de la Terre (SPOT) VEGETATION [Fraser and Latifovic, 2005] or Moderate Resolution Imaging Spectroradiometer (MODIS) [Coops et al., 2009; de Beurs and Townsend, 2008]. Addressing the significant knowledge gap of quantifying the extent of biotic disturbances will require medium- and coarse-resolution imagery that sense much larger spatial extents and are much less expensive than high-resolution alternatives.

Temporal resolution is coupled with spatial resolution: higher spatial resolution typically means longer repeat times, whereas coarse-resolution MODIS imagery is available twice daily. Repeat intervals are less important for tree mortality events, but are more important for capturing potentially short-lived defoliation events at the specific time when the physical discoloration or loss of foliage is most visible for detection.

The location of spectral bands is important, although most sensors provide imagery in visible and near-infrared wavelengths useful for mapping tree mortality. Hyperspectral remote sensing appears to be a promising technology for the detection of pathogens because it is able to detect plant stress far in advance of actual defoliation [Govender et al., 2008], quantify fine-scale

patchy damage patterns [*Hatala et al.*, 2010; *Pu et al.*, 2008], and even detect root disease [*Leckie et al.*, 2004].

Aside from sensor characteristics, a multitude of algorithms and approaches ranging from classification to modeling have been used to assess damage from insect defoliators and bark beetles [Hall et al., 2006; Wulder et al., 2006]. The combination of sensor and methods of analysis have tended to be species- and site-specific as a result of the wide variation in physiological responses associated with the physical manifestation of damage that ranges from foliage color change to physical loss of foliage.

7.4 Models of insect and pathogen outbreaks

Models of forest pathogens and insects have been developed from a number of different perspectives and with different goals. "Hazard rating" models estimate the susceptibility of a stand to attack based on its structure (age, stem density, etc.) [Krist et al., 2007], and may include simple representations of climate and populations [Shore and Safranyik, 1992]. Climate suitability models assess whether conditions are conducive to outbreaks based on temperature and/or drought. Such models can be based on empirical analysis [e.g., Carroll et al., 2004], climate envelop models [e.g., Magarey et al., 2007], or fundamental processes measured in the laboratory [e.g., Bentz et al., 1991]. Neither hazard rating nor climate suitability models predict individual outbreaks, but rather are used to assess current conditions that could potentially lead to outbreaks.

Models of population dynamics, which are often based on spatial extensions of traditional epidemiological models, simulate the course of an epidemic. These models may include such

processes as outbreak initiation, dispersal, and collapse, although some processes may be prescribed by the user instead of simulated [e.g., *Beukema et al.*, 1997; *Cairns et al.*, 2008; *Logan et al.*, 1998]. Biotic interactions between host and insect/pathogen are not present in hazard rating or climate suitability models but are included in population dynamics models. For example, grid-based models have been used to scale single plant studies to populations [*Kleczkowski et al.*, 1997] and to assess the potential role of pathogens in facilitating plant migration through enemy release [*Moorcroft et al.*, 2006]. Newer Bayesian population models show promise for data-driven assessments of disease population dynamics that can account for multiple data sources and the uncertainty associated with the often indirect evidence for pathogen abundance [*Hatala et al.*, in press].

A number of detailed modules for specific insects, pathogens, or functional groups have been developed for the US Forest Service's Forest Vegetation Simulator (FVS), a forestry-oriented stand simulator [Beukema et al., 1997; Frankel, 1998; Hagle et al., 1995; Hansen and Goheen, 2000; Robinson and Geils, 2006]. Similar modules have been developed for the British Columbia Ministry of Forests' Tree and Stand Simulator (TASS), such as the Canadian Forest Service's Root Rot Simulator (ROTSIM), originally developed for laminated root rot (Phellinus weirii (Murrill) R.L. Gibertson) in Douglas-fir (Pseudotsuga menziesii (Mirb.) Franco) [Bloomberg, 1988] and recently adapted for Armillaria root disease [Morrison, 2005]. Detailed "stand-alone" forest pathogen simulators also exist, such as the "Root Rot Tracker" model of forest root diseases [Peet and Hunt, 2005]. For large regional assessments, mechanistic models of pathogen dispersal have been developed that account for both wind characteristics on a continental scale and environmental determinants of spore viability (relative humidity, temperature, UV-B radiation) [Aylor, 2003].

Variability in drivers across species or groups of species implies that modeling outbreaks of biotic disturbance agents requires understanding the natural history of each individual species, or at a minimum, functional groups of similar species. For a limited number of economically important insect species such as mountain pine beetle, spruce budworm, or gypsy moth, predictive models have been developed either empirically or through laboratory studies [Hansen et al., 2001; Logan et al., 2003]. These models usually predict one specific driver (e.g., probability that a given winter's temperatures will result in insect mortality). In one case, mountain pine beetle, models exist for most important drivers, including winter mortality, year-round temperature control of life stage development rates, and host susceptibility [Carroll et al., 2004; Logan and Bentz, 1999; Logan and Powell, 2001; Régnière and Bentz, 2007; Shore and Safranyik, 1992], but limited predictive capability exists for the vast majority of insects and pathogens. Full integration of stand conditions, climate suitability, population dynamics, and host/biotic disturbance agent interactions that allow for more complete simulations have yet to be accomplished.

7.5 Models of the carbon cycle

Models of the carbon cycle often include wildfire and harvest disturbances, but few include biotic disturbance agents. Exceptions include studies of the role of defoliation [Cook et al., 2008], aspen decline [Hogg, 1999; 2001], and tree mortality [Albani et al., 2010; Kurz and Apps, 1999; Kurz et al., 2008a; Pfeifer et al., 2010]. In general, C cycle models are positioned to include biotic disturbances, though challenges remain. In addition to prescribing or predicting the number of trees infested (though an outbreak submodel, for instance), correct simulation

requires understanding how to model response of the carbon cycle to insect or pathogen outbreak in terms tree growth reduction and mortality. Subsequent ecosystem impacts important to carbon cycling such as decomposition, snagfall rate, regeneration, and surviving stand composition also suggest their importance for inclusion in simulations of impacts.

8. Summary and conclusions

Numerous insect and pathogen species impact forests of North America. A few particularly important species affect large areas across many regions, suggesting major impacts to carbon budgets. Life histories of some well-studied species are relatively well understood, but little is known about the majority of important biotic disturbance agents.

Relatively few studies have documented effects of insect or pathogen impacts on stand-level carbon cycling. Tree productivity is reduced in outbreaks, and tree mortality caused by insects or pathogens creates a large amount of dead wood that subsequently decomposes. As a result, NEP is often reduced substantially, sometime switching from a typical sink of carbon to a source. NPP following tree mortality events can recover within a decade or so, though effects on NEP associated with decomposition last longer.

Fewer studies have quantified effects at larger spatial scales. Some of these studies illustrate large impacts on carbon cycling, others more minor impacts, depending on the number of trees affected regionally.

Difficulties arise in obtaining consistency among studies due to several factors. The number of affected trees varies within and among outbreaks of the same species, and impacts (defoliation, growth reduction, mortality) vary among types of disturbance agents. In addition,

on different periods. Future studies of biotic disturbance effects on carbon budgets should include key information that permits comparisons across studies, including spatial and temporal extent of damage, number of trees infested within study region, and time since disturbance. Studies should provide controls for comparison, either preoutbreak and/or in uninfested stands, to quantify the relative magnitude of impact.

Biotic disturbances are often strongly influenced by abiotic factors. Climate change effects on host trees through heat or drought stress are often associated with insect or pathogen outbreaks. Insects and pathogens often interact to affect forest ecosystems. Such interactions make identifying the individual role of biotic agents relative to other potential causal factors difficult, particularly climatic and forest stand condition stressors of forest tree condition.

Invasive insects or pathogens have great potential to affect tree species and therefore carbon budgets. We have, however, little capability for predicting invasions, however.

It is clear that biotic disturbances are critical elements of the North American carbon budget, and considerable uncertainties and knowledge gaps exist in our ability to predict or estimate their effects on the carbon cycle. Additional research across a range of disturbance types and spatial and temporal scales is needed to reduce uncertainties of the effects of insect and pathogen outbreaks on carbon cycling.

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884	References
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886	Adams, H. D., M. Guardiola-Claramonte, G. A. Barron-Gafford, J. C. Villegas, D. D. Breshears,
887	C. B. Zou, P. A. Troch, and T. E. Huxman (2009), Temperature sensitivity of drought-
888	induced tree mortality portends increased regional die-off under global-change-type
889	drought, Proceedings of the National Academy of Sciences of the United States of
890	America, 106(17), 7063-7066.
891	Adams, H. D., A. K. Macalady, D. D. Breshears, C. D. Allen, N. L. Stephenson, S. R. Saleska, T.
892	E. Huxman, and N. G. McDowell (2010), Climate-induced tree mortality: Earth system
893	consequences, EOS, Transactions of the American Geophysical Union, 91(17), 153.
894	Agrios, G. N. (2005), Plant Pathology, Fifth Edition ed., Elsevier Academic Press, USA.
895	Ahern, F. J. (1988), The effects of bark beetle stress on the foliar spectral reflectance of
896	lodgepole pine, Int. J. Remote Sens., 9(9), 1451-1468.
897	Albani, M., P. R. Moorcroft, A. M. Ellison, D. A. Orwig, and D. R. Foster (2010), Predicting the
898	impact of hemlock woolly adelgid on carbon dynamics of eastern United States forests,
899	Canadian Journal of Forest Research, 40(1), 119-133.
900	Allard, V., J. M. Ourcival, S. Rambal, R. Joffre, and A. Rocheteau (2008), Seasonal and annual
901	variation of carbon exchange in an evergreen Mediterranean forest in southern France,
902	Global Change Biol., 14(4), 714-725.
903	Allen, C. D., et al. (2010), A global overview of drought and heat-induced tree mortality reveals
904	emerging climate change risks for forests, Forest Ecology and Management, 259(4), 660-
905	684.

906 Anagnostakis, S. L. (1987), Chestnut blight: The classical problem of an introduced pathogen, 907 *Mycologia*, 79(1), 23-37. 908 Anderson, L., C. E. Carlson, and R. H. Wakimoto (1987), Forest fire frequency and western 909 spruce budworm outbreaks in western Montana, Forest Ecology and Management, 22(3-910 4), 251-260. 911 Anderson, R. L., J. P. McClure, N. D. Cost, and R. J. Uhler (1986), Estimating fusiform rust 912 losses in five southeast states, Southern Journal of Applied Forestry, 10, 237-240. 913 Aylor, D. E. (2003), Spread of plant disease on a continental scale: Role of aerial dispersal of 914 pathogens, *Ecology*, 84(8), 1989-1997. 915 Ayres, M. P., and M. J. Lombardero (2000), Assessing the consequences of global change for 916 forest disturbance from herbivores and pathogens, The Science of the Total Environment, 917 *262*, 263-286. 918 Bentz, B., et al. (2009), Bark Beetle Outbreaks in Western North America: Causes and 919 Consequences, University of Utah Press, Salt Lake City, UT. 920 Bentz, B. J., J. A. Logan, and G. D. Amman (1991), Temperature-dependent development of 921 mountain pine beetle and simulation of its phenology, Canadian Entomologist, 123, 922 1083-1094. 923 Bentz, B. J., J. A. Logan, and J. C. Vandygriff (2001), Latitudinal variation in *Dendroctonus* 924 ponderosae (Coleoptera: Scolytidae) development time and adult size, Canadian 925 Entomologist, 133(3), 375-387. 926 Bentz, B. J., J. Régnière, C. J. Fettig, E. M. Hansen, J. L. Hayes, J. A. Hicke, R. G. Kelsey, J. F. 927 Negrón, and S. J. Seybold (2010), Climate change and bark beetles of the western United

States and Canada: Direct and indirect effects, *BioScience*, 60(8), 602-613.

928

929 Beukema, S. J., J. A. Greenough, D. C. E. Robinson, W. A. Kurz, E. L. Smith, and B. B. Eav 930 (1997), The Westwide Pine Beetle Model: a spatially-explicit contagion model, paper 931 presented at Proceedings of the Forest Vegetation Simulator Conference, Gen. Tech. Rep. 932 INT-GTR-373., U.S. Department of Agriculture, Forest Service, Intermountain Research 933 Station. 934 Blais, J. R. (1983), Trends in the frequency, extent, and severity of spruce budworm outbreaks in 935 eastern Canada, Canadian Journal of Forest Research, 13(4), 539-547. 936 Bleiker, K., and D. L. Six (2007), Dietary benefits of fungal associates to an eruptive herbivore: 937 potential implications of multiple associates on host population dynamics, *Environ*. 938 Entomol., 36, 1384-1396. 939 Bloomberg, W. J. (1988), Modeling control strategies for laminated root rot in managed 940 Douglas-fir stands: Model development, *Phytopathology*, 78(4), 403-409. 941 Bonan, G. (2002), Ecological climatology, 678 pp., Cambridge University Press, Cambridge UK. 942 Bonan, G. B. (2008), Forests and climate change: Forcings, feedbacks, and the climate benefits 943 of forests, Science, 320(5882), 1444-1449. 944 Breshears, D. D., et al. (2005), Regional vegetation die-off in response to global-change-type 945 drought, Proceedings of the National Academy of Sciences of the United States of 946 America, 102(42), 15144-15148. 947 Breshears, D. D., O. B. Myers, C. W. Meyer, F. J. Barnes, C. B. Zou, C. D. Allen, N. G. 948 McDowell, and W. T. Pockman (2009), Tree die-off in response to global change-type 949 drought: mortality insights from a decade of plant water potential measurements, 950 *Frontiers in Ecology and the Environment*, 7(4), 185-189.

951 Brown, M., T. A. Black, Z. Nesic, V. N. Foord, D. L. Spittlehouse, A. L. Fredeen, N. J. Grant, P. 952 J. Burton, and J. A. Trofymow (2010), Impact of mountain pine beetle on the net 953 ecosystem production of lodgepole pine stands in British Columbia, Agricultural and 954 Forest Meteorology, 150, 254-264. 955 Cairns, D. M., C. W. Lafon, J. D. Waldron, M. Tchakerian, R. N. Coulson, K. D. Klepzig, A. G. 956 Birt, and W. Xi (2008), Simulating the reciprocal interaction of forest landscape structure 957 and southern pine beetle herbivory using LANDIS, Landsc. Ecol., 23(4), 403-415. 958 Canadell, J. G., et al. (2007), Contributions to accelerating atmospheric CO2 growth from 959 economic activity, carbon intensity, and efficiency of natural sinks, *Proceedings of the* 960 National Academy of Sciences of the United States of America, 104(47), 18866-18870. 961 Canadell, J. G., and M. R. Raupach (2008), Managing forests for climate change mitigation, 962 Science, 320(5882), 1456-1457. 963 Candau, J. N., R. A. Fleming, and A. Hopkin (1998), Spatiotemporal patterns of large-scale 964 defoliation caused by the spruce budworm in Ontario since 1941, Canadian Journal of 965 Forest Research, 28(11), 1733-1741. 966 Carisey, N., T. Bauce, A. Dupont, and S. Miron (2004), Effects of bud phenology and foliage 967 chemistry of balsam fir and white spruce trees on the efficacy of Bacillus thuringiensis 968 against the spruce budworm, Choristoneura fumiferana, Agricultural and Forest 969 *Entomology*, *6*(1), 55-69. 970 Carroll, A. L., S. W. Taylor, J. Régnière, and L. Safranyik (2004), Effects of climate change on 971 range expansion by the mountain pine beetle in British Columbia, paper presented at 972 Mountain Pine Beetle Symposium: Challenges and Solutions, Natural Resources Canada, 973 Canadian Forest Service, Pacific Forestry Centre, Kelowna, BC, October 30-31, 2003.

974 CCSP (2007), The First State of the Carbon Cycle Report (SOCCR): The North American 975 Carbon Budget and Implications for the Global Carbon Cycle. A Report by the U.S. 976 Climate Change Science Program and the Subcommittee on Global Change Research, 977 242 pp, National Oceanic and Atmospheric Administration, National Climatic Data 978 Center, Asheville, NC, USA. 979 CCSP (2008), The effects of climate change on agriculture, land resources, water resources, and 980 biodiversity. A Report by the U.S. Climate Change Science Program and the 981 Subcommittee on Global Change Research, 362 pp., U.S. Environmental Protection 982 Agency, Washington, DC, USA. 983 Chapin, F. S., et al. (2006), Reconciling carbon-cycle concepts, terminology, and methods, 984 Ecosystems, 9(7), 1041-1050. 985 Chapin, F. S., J. T. Randerson, A. D. McGuire, J. A. Foley, and C. B. Field (2008), Changing 986 feedbacks in the climate-biosphere system, Frontiers in Ecology and the Environment, 987 *6*(6), 313-320. 988 Ciesla, W. M. (2000), Remote sensing in forest health protection, USDA Forest Service, Salt 989 Lake City, Utah. 990 Clark, K. L., N. Skowronski, and J. Hom (2009), Invasive insects impact forest carbon dynamics, 991 Global Change Biol. 992 Cook, B. D., P. V. Bolstad, J. G. Martin, F. A. Heinsch, K. J. Davis, W. G. Wang, A. R. Desai, 993 and R. M. Teclaw (2008), Using light-use and production efficiency models to predict 994 photosynthesis and net carbon exchange during forest canopy disturbance, *Ecosystems*, 995 *11*(1), 26-44.

996 Coops, N. C., M. Johnson, M. A. Wulder, and J. C. White (2006), Assessment of QuickBird high 997 spatial resolution imagery to detect red attack damage due to mountain pine beetle 998 infestation, Remote Sens. Environ., 103(1), 67-80. 999 Coops, N. C., M. A. Wulder, and D. Iwanicka (2009), Large area monitoring with a MODIS-1000 based Disturbance Index (DI) sensitive to annual and seasonal variations, *Remote Sens*. 1001 Environ., 113(6), 1250-1261. 1002 Coyle, D., W. Mattson, and K. Raffa (2008), Invasive root feeding insects in natural forest 1003 ecosystems of North America, in *Root Feeders: an ecosystem perspective*, edited by S. 1004 Johnson and P. Murray, pp. 146-162, CABI. 1005 Cruickshank, M. (2000), Volume loss of Douglas-fir infected with Armillaria ostovae, paper 1006 presented at From science to management and back: A science forum for southern 1007 interior ecosystems of British Columbia, Southern Interior Forest Extension and Research 1008 Partnership, Kamloops, BC. 1009 Cruickshank, M., D. J. Morrison, and A. Lalumière (2009), The interaction between competition 1010 in interior Douglas-fir plantations and disease caused by Armillaria ostoyae in British 1011 Columbia, Forest Ecology and Management, 257, 443-452. 1012 Dale, V. H., et al. (2001), Climate change and forest disturbances, *Bioscience*, 51(9), 723-734. 1013 de Beurs, K. M., and P. A. Townsend (2008), Estimating the effect of gypsy moth defoliation 1014 using MODIS, Remote Sens. Environ., 112(10), 3983-3990. 1015 Denning, A. S., et al., (2005), Science Implementation Strategy for the North American Carbon 1016 Program. Report of the NACP Implementation Strategy Group of the U.S. Carbon Cycle 1017 Interagency Working Group, 68 pp., U.S. Carbon Cycle Science Program, Washington, 1018 DC.

1019 Desprez-Loustau, M. L., B. Marcais, L. M. Nageleisen, D. Piou, and A. Vannini (2006), 1020 Interactive effects of drought and pathogens in forest trees, *Annals of Forest Science*, 1021 *63*(6), 597-612. 1022 Dodds, K. J., C. Graber, and F. M. Stephen (2001), Facultative intraguild predation by larval 1023 Cerambycidae (Coleoptera) on bark beetle larvae (Coleoptera: Scolytidae), Environ. 1024 Entomol., 30(1), 17-22. 1025 Drummond, D. B. (1982), Timber loss estimates for the coniferous forests in the U.S. due to 1026 dwarf mistletoes, USDA Forest Service Methods Application Group. 1027 Durall, D. M., M. D. Jones, and K. J. Lewis (2005), Effects of forest management on fungal 1028 communities, in *The fungal community, its organization and role in ecosystems*, edited by 1029 J. Dighton, et al., pp. 833-856, CRC Press, London. 1030 Dymond, C. C., E. T. Neilson, G. Stinson, K. Porter, D. A. MacLean, D. R. Gray, M. Campagna, 1031 and W. A. Kurz (2010), Future spruce budworm outbreak may create a carbon source in 1032 eastern Canadian forests, *Ecosystems*, 13, 917-931. 1033 Elkinton, J. S., and A. M. Liebhold (1990), Population dynamics of gypsy moth in North 1034 America, Annu. Rev. Entomol., 35, 571-596. 1035 Esper, J., U. Buntgen, D. C. Frank, D. Nievergelt, and A. Liebhold (2007), 1200 years of regular 1036 outbreaks in alpine insects, Proceedings of the Royal Society of London Series B-Biological Sciences, 274, 671-679. 1037 1038 Evans, H. J., A. A. Hopkin, and T. A. Scarr (2006), Status of important forest pests in Ontario in 1039 2006, www.glfc.forestry.ca/VLF/foresthealth/foresthealth2006e.pdf.

1040 Farr, W. A., V. J. LaBau, and T. H. Laurent (1976), Estimation of decay in old-growth western 1041 hemlock and sitka spruce in southeast Alaska, 24 pp, U. S. Department of Agriculture, 1042 Forest Service, Pacific Northwest Forest and Range Experiment Station, Portland, OR. 1043 Farrar, J. L., Ottawa (1995), Trees in Canada, Fitzhenry & Whiteside Ltd. and Canadian Forest 1044 Service, Ottowa. 1045 Fellin, D. G., and J. E. Dewey (1982), Western spruce budworm, United States Department of 1046 Agriculture, Forest Service. 1047 Fettig, C. J., K. D. Klepzig, R. F. Billings, A. S. Munson, T. E. Nebeker, J. F. Negron, and J. T. 1048 Nowak (2007), The effectiveness of vegetation management practices for prevention and 1049 control of bark beetle infestations in coniferous forests of the western and southern 1050 United States, Forest Ecology and Management, 238(1-3), 24-53. 1051 Fleming, R. A. (2000), Climate change and insect disturbance regimes in Canada's boreal forests, 1052 World Resources Review, 12(3), 520-555. 1053 Fleming, R. A., J. N. Candau, and R. S. McAlpine (2002), Landscape-scale analysis of 1054 interactions between insect defoliation and forest fire in Central Canada, Clim. Change, 1055 55(1-2), 251-272. 1056 Forrester, J. A., G. G. McGee, and M. J. Mitchell (2003), Effects of beech bark disease on 1057 aboveground biomass and species composition in a mature northern hardwood forest, 1058 1985 to 2000, J. Torrey Bot. Soc., 130(2), 70-78. 1059 Frankel, S. J. (1998), User's Guide to the Western Root Disease Model, Version 3.0, USFS. 1060 Franklin, S. E., and A. G. Raske (1994), Satellite remote sensing of spruce budworm forest 1061 defoliation in western Newfoundland, Can. J. Rem. Sens., 20, 37-48.

1062 Franklin, S. E., M. A. Wulder, R. S. Skakun, and A. L. Carroll (2003), Mountain pine beetle red-1063 attack forest damage classification using stratified Landsat TM data in British Columbia, 1064 Canada, Photogramm. Eng. Remote Sens., 69(3), 283-288. 1065 Fraser, R. H., and R. Latifovic (2005), Mapping insect-induced tree defoliation and mortality 1066 using coarse spatial resolution satellite imagery, Int. J. Remote Sens., 26(1), 193-200. 1067 Frost, C. J., and M. D. Hunter (2004), Insect canopy herbivory and frass deposition affect soil 1068 nutrient dynamics and export in oak mesocosms, *Ecology*, 85(12), 3335-3347. 1069 Gandhi, K. J., and D. A. Herms (2009), Direct and indirect effects of invasive exotic insect 1070 herbivores on ecological processes and interactions in forests of eastern North America, 1071 Biological Invasions, 12, 389-405. 1072 Geils, B. W., J. Cibrian-Tovar, and B. Moody (Eds.) (2002), Mistletoes of North American 1073 conifers, Gen. Tech. Rep. RMRS-GTR-98, 123 pp., U.S. Department of Agriculture, 1074 Forest Service, Rocky Mountain Research Station, Ogden, UT. 1075 Geils, B. W., K. E. Hummer, and R. S. Hunt (2010), White pines, Ribes, and blister rust: a 1076 review and synthesis, Forest Pathology, 40, 147-185. 1077 Gill, L. S. (1954), Dwarf mistletoe of ponderosa pine in the Southwest, 9 pp, Rocky Mountain 1078 Forest and Range Experiment Station, U.S. Department of Agriculture, Forest Service, 1079 Ft. Collins, CO. 1080 Goheen, D. J., and W. J. Otrosina (1998), Characteristics and consequences of root diseases in 1081 forests of western North America, in User's guide to the western rood disease model, 1082 version 3.0, Gen. Tech. Rep. PSW-GTR-165, edited by S. J. Frankel, pp. 3-8, U.S. 1083 Department of Agriculture, Forest Service, Pacific Southwest Research Station, Berkeley, 1084 CA.

1085 Goheen, E. M., E. Hansen, A. Kanaskie, N. Osterbauer, J. Parke, J. Pscheidt, and G. Chastagner 1086 (2006), Sudden oak death and *Phytophthora ramorum*, a guide for forest managers, 1087 Christmas tree growers, and forest-tree nursery operators in Oregon and Washington, 16 1088 pp, OSU Extension Service. 1089 Goodale, C. L., et al. (2002), Forest carbon sinks in the Northern Hemisphere, *Ecol. Appl.*, 12(3), 1090 891-899. 1091 Goodwin, N. R., N. C. Coops, M. A. Wulder, S. Gillanders, T. A. Schroeder, and T. Nelson 1092 (2008), Estimation of insect infestation dynamics using a temporal sequence of Landsat 1093 data, Remote Sens. Environ., 112(9), 3680-3689. 1094 Govender, M., K. Chetty, V. Naiken, and H. Bulcock (2008), A comparison of satellite 1095 hyperspectral and multispectral remote sensing imagery for improved classification and 1096 mapping of vegetation, Water SA, 34(2), 147-154. 1097 Griffin, J. M., G. M. Lovett, M. A. Arthur, and K. C. Weathers (2003), The distribution and 1098 severity of beech bark disease in the Catskill Mountains, NY, Canadian Journal of 1099 Forest Research, 33, 1754-1760. 1100 Hadley, J. L., P. S. Kuzeja, M. J. Daley, N. G. Phillips, T. Mulcahy, and S. Singh (2008), Water 1101 use and carbon exchange of red oak- and eastern hemlock-dominated forests in the 1102 northeastern USA: implications for ecosystem-level effects of hemlock woolly adelgid. 1103 Hagle, S. K., S. J. Kegley, and S. B. Williams (1995), Assessing pathogen and insect succession 1104 functions in forest ecosystems, in Forest health through silviculture: Proceedings of the 1105 1995 National Silviculture Workshop, Gen. Tech. Rep. RM-GTR-267, edited by L. G. 1106 Eskew, pp. 117-127, U.S. Department of Agriculture, Forest Service, Rocky Mountain 1107 Forest and Range Experiment Station, Fort Collins, CO.

1108 Hajek, A. E. (1999), Pathology and epizootiology of *Entomophaga maimaiga* infections in forest 1109 lepidoptera, Microbiology and Molecular Biology Reviews, 63(4), 814-+. 1110 Hall, P. J., W. W. Bowers, and H. Hirvonen (1998), Forest insect and disease conditions in 1111 Canada, Natural Resources Canada, Canadian Forest Service, Ottawa, Ontario. 1112 Hall, R. J., R. A. Fernandes, E. H. Hogg, J. P. Brandt, C. Butson, B. S. Case, and S. G. Leblanc 1113 (2003), Relating aspen defoliation to changes in leaf area derived from field and satellite 1114 remote sensing data, Can. J. Rem. Sens., 28, 1317-1327. 1115 Hall, R. J., R. S. Skakun, and E. J. Arsenault (2006), Remotely sensed data in the mapping of 1116 insect defoliation, in *Understanding forest disturbance and spatial pattern: Remote* 1117 sensing and GIS approaches, edited by M. A. Wulder and S. E. Franklin, pp. 85-111, 1118 Taylor and Francis, CRC Press, Boca Raton. 1119 Hancock, J. E., M. A. Arthur, K. C. Weathers, and G. M. Lovett (2008), Carbon cycling along a 1120 gradient of beech bark disease impact in the Catskill Mountains, New York, Canadian 1121 *Journal of Forest Research*, *38*(5), 1267-1274. 1122 Hansen, E. M., and E. M. Goheen (2000), *Phellinus weirii* and other native root pathogens as 1123 determinants of forest structure and process in western North America, Annual Review of 1124 *Phytopathology*, 38, 515-539. 1125 Hansen, E. M., J. K. Stone, B. R. Capitano, P. Rosso, W. Sutton, L. Winton, A. Kanaskie, and 1126 M. McWilliams (2000), Incidence and impact of Swiss needle cast in forest plantations of 1127 Douglas-fir in coastal Oregon, *Plant Disease*, 84, 773-778. 1128 Hansen, M. E., B. J. Bentz, and D. L. Turner (2001), Temperature-based model for predicting 1129 univoltine brood proportions in spruce beetle (Coleoptera: Scolytidae), Canadian 1130 Entomologist, 133, 827-841.

1131 Harausz, E., and D. Pimentel (2002), North American forest losses due to insects and plant 1132 pathogens, in *Encyclopedia of Pest Management*, edited by D. Pimentel, Marcel Dekker, 1133 Inc. 1134 Harris, J. W. E., and A. F. Dawson (1979), Evaluation of aerial forest pest damage survey 1135 techniques in British Columbia, 1-22 pp, Canadian Forest Service, Pacific Forestry 1136 Centre, Victoria, British Columbia. 1137 Hatala, J. A., R. L. Crabtree, K. Q. Halligan, and P. R. Moorcroft (2010), Landscape-scale 1138 patterns of forest pest and pathogen damage in the Greater Yellowstone Ecosystem, 1139 Remote Sens. Environ., 114, 375-384. 1140 Hatala, J. A., M. C. Dietze, R. L. Crabtree, K. Kendall, D. Six, and P. R. Moorcroft (in press), 1141 An ecosystem-scale model for the spread of a host-specific forest pathogen in the Greater 1142 Yellowstone Ecosystem, *Ecol. Appl.* 1143 Hawksworth, F. G., and D. Wiens (1970), Biology and taxonomy of the dwarf mistletoes, 1144 Annual Review of Phytopathology, 8, 187-&. 1145 Hicke, J. A., G. P. Asner, J. T. Randerson, C. Tucker, S. Los, R. Birdsey, J. C. Jenkins, and C. 1146 Field (2002), Trends in North American net primary productivity derived from satellite 1147 observations, 1982-1998, Global Biogeochem. Cycles, 16, 10.1029/2001GB001550. 1148 Hicke, J. A., et al. (2003), Postfire response of North American boreal forest net primary 1149 productivity analyzed with satellite observations, Global Change Biol., 9(8), 1145-1157. 1150 Hicke, J. A., J. A. Logan, J. Powell, and D. S. Ojima (2006), Changing temperatures influence 1151 suitability for modeled mountain pine beetle (*Dendroctonus ponderosae*) outbreaks in the 1152 western United States, Journal of Geophysical Research-Biogeosciences, 111, G02019, 1153 doi:02010.01029/02005JG000101.

1154 Hicke, J. A., and J. A. Logan (2009), Mapping whitebark pine mortality caused by a mountain 1155 pine beetle outbreak with high spatial resolution satellite imagery, Int. J. Remote Sens., 1156 *30*, 4427-4441. 1157 Hogg, E. H. (1999), Simulation of inter-annual responses of trembling aspen stands to climatic 1158 variation and insect defoliation in western Canada, Ecol. Model., 114(2-3), 175-193. 1159 Hogg, E. H., D. T. Price, and T. A. Black (2000), Postulated feedbacks of deciduous forest 1160 phenology on seasonal climate patterns in the western Canadian interior, J. Clim., 13(24), 1161 4229-4243. 1162 Hogg, E. H. (2001), Modeling Aspen Responses to Climatic Warming and Insect Defoliation in 1163 Western Canada, in *Proceedings RMRS-P-18*, edited, pp. 325-338, USDA Forest Service. 1164 Hogg, E. H., J. P. Brandt, and M. Michaelian (2008), Impacts of a regional drought on the 1165 productivity, dieback and biomass of western Canadian aspen forests, Canadian Journal 1166 of Forest Research, 38(6), 1373-1384. Hopkins, P. F., A. L. Maclean, and T. M. Lillesand (1988), Assessment of Thematic Mapper 1167 1168 imagery for forestry applications under Lake States conditions, *Photogramm. Eng.* 1169 *Remote Sens.*, 54(1), 61-68. 1170 Houghton, R. A., and D. L. Skole (1990), Carbon, in The Earth as Transformed by Human 1171 Action, edited by B. L. Turner, et al., pp. 393-408, Cambridge University Press, 1172 Cambridge. 1173 Houston, D. R. (1994), Major new tree disease epidemics: Beech bark disease, *Annual Review of* 1174 Phytopathology, 32, 75-87. 1175 IPCC (2007), Summary for Policymakers, in Climate Change 2007: The Physical Science Basis. 1176 Contribution of Working Group I to the Fourth Assessment Report of the

1177	Intergovernmental Panel on Climate Change, edited by S. Solomon, et al., Cambridge
1178	University Press, Cambridge, United Kingdom and New York, NY, USA.
1179	Jactel, H., and E. G. Brockerhoff (2007), Tree diversity reduces herbivory by forest insects,
1180	Ecology Letters, 10(9), 835-848.
1181	Jenkins, J. C., J. D. Aber, and C. D. Canham (1999), Hemlock woolly adelgid impacts on
1182	community structure and N cycling rates in eastern hemlock forests, Canadian Journal of
1183	Forest Research, 29(5), 630-645.
1184	Jenkins, M. J., E. Hebertson, W. Page, and C. A. Jorgensen (2008), Bark beetles, fuels, fires and
1185	implications for forest management in the Intermountain West, Forest Ecology and
1186	Management, 254(1), 16-34.
1187	Johnson, G. R., B. L. Gartner, D. Maguire, and A. Kanaskie (2003), Influence of Bravo fungicide
1188	applications on wood density and moisture content of Swiss needle cast affected
1189	Douglas-fir trees, Forest Ecology and Management, 186, 339-348.
1190	Kashian, D. M., W. H. Romme, D. B. Tinker, M. G. Turner, and M. G. Ryan (2006), Carbon
1191	storage on landscapes with stand-replacing fires, Bioscience, 56(7), 598-606.
1192	Kinloch, B. B. (2003), White pine blister rust in North America: Past and prognosis,
1193	Phytopathology, 93, 1044-1047.
1194	Kirschbaum, M. U. F., H. Keith, R. Leuning, H. A. Cleugh, K. L. Jacobsen, E. van Gorsel, and
1195	R. J. Raison (2007), Modelling net ecosystem carbon and water exchange of a temperate
1196	Eucalyptus delegatensis forest using multiple constraints, Agricultural and Forest
1197	Meteorology 145(1-2) 48-68

1198 Kizlinski, M. L., D. A. Orwig, R. C. Cobb, and D. R. Foster (2002), Direct and indirect 1199 ecosystem consequences of an invasive pest on forests dominated by eastern hemlock, 1200 Journal of Biogeography, 29(10-11), 1489-1503. 1201 Kleczkowski, A., C. A. Gilligan, and D. J. Bailey (1997), Scaling and spatial dynamics in plant-1202 pathogen systems: From individuals to populations, *Proceedings of the Royal Society of* 1203 London Series B-Biological Sciences, 264(1384), 979-984. 1204 Klepzig, K. D., K. F. Raffa, and E. B. Smalley (1991), Association of insect-fungal complexes 1205 with Red Pine Decline in Wisconsin, For. Sci., 37, 1119-1139. 1206 Klepzig, K. D., E. L. Kruger, E. B. Smalley, and K. F. Raffa (1995), Effects of biotic and abiotic 1207 stress on the induced accumulation of terpenes and phenolics in red pines inoculated with 1208 a bark beetle vectored fungus, Journal of Chemical Ecology, 21(5), 601-626. 1209 Kliejunas, J. T., et al. (2009), Review of Literature on Climate Change and Forest Diseases of 1210 Western North America, General Technical Report, PSW-GTR-225, 54 pp., U.S. 1211 Department of Agriculture, Forest Service, Pacific Southwest Research Station, Albany, 1212 CA. 1213 Koricheva, J., S. Larsson, and E. Haukioja (1998), Insect performance on experimentally 1214 stressed woody plants: A meta-analysis, *Annu. Rev. Entomol.*, 43, 195-216. 1215 Kovacs, K. F., R. G. Haight, D. G. McCullough, R. J. Mercader, N. W. Siegert, and A. M. 1216 Liebhold (2010), Cost of potential emerald ash borer damage in U.S. communities, 2009-1217 2019, *Ecological Economics*, 69, 569-578. 1218 Krist, F. J., F. J. Sapio, and B. M. Tkacz (2007), Mapping Risk from Forest Insects and Diseases, 1219 USDA Forest Service.

1220 Kurz, W. A., and M. J. Apps (1999), A 70-year retrospective analysis of carbon fluxes in the 1221 Canadian forest sector, Ecol. Appl., 9(2), 526-547. 1222 Kurz, W. A., C. C. Dymond, G. Stinson, G. J. Rampley, E. T. Neilson, A. L. Carroll, T. Ebata, 1223 and L. Safranyik (2008a), Mountain pine beetle and forest carbon feedback to climate 1224 change, *Nature*, 452, 987-990. 1225 Kurz, W. A., G. Stinson, G. J. Rampley, C. C. Dymond, and E. T. Neilson (2008b), Risk of 1226 natural disturbances makes future contribution of Canada's forests to the global carbon 1227 cycle highly uncerain, Proceedings of the National Academy of Sciences of the United 1228 *States of America*, 105(5), 1551-1555. 1229 le Mellec, A., and B. Michalzik (2008), Impact of a pine lappet (*Dendrolimus pini*) mass 1230 outbreak on C and N fluxes to the forest floor and soil microbial properties in a Scots 1231 pine forest in Germany, Canadian Journal of Forest Research, 38(7), 1829-1841. 1232 le Mellec, A., M. Habermann, and B. Michalzik (2009), Canopy herbivory altering C to N ratios 1233 and soil input patterns of different organic matter fractions in a Scots pine forest, *Plant* 1234 and Soil, 325(1-2), 255-262. 1235 Leckie, D. G., and D. P. Ostaff (1988), Classification of airborne multispectral scanner data for 1236 mapping current defoliation caused by the spruce budworm, For. Sci., 34(2), 259-275. 1237 Leckie, D. G., C. Jay, F. A. Gougeon, R. N. Sturrock, and D. Paradine (2004), Detection and 1238 assessment of trees with Phellinus weirii (laminated root rot) using high resolution multi-1239 spectral imagery, Int. J. Remote Sens., 25(4), 793-818. 1240 Liebhold, A. M., and P. C. Tobin (2008), Population ecology of insect invasions and their 1241 management, Annu. Rev. Entomol., 53, 387-408.

1242 Lieutier, F., A. Yart, and A. Salle (2009), Stimulation of tree defenses by Ophiostomatoid fungi 1243 can explain attack success of bark beetles on conifers, Annals of Forest Science, 66(8), 1244 Article Number: 801 1245 Littke, W. R., and J. E. Browning (1989), Heterobasidion (Fomes) Annosum Incidence in Pre-1246 Commercially Thinned Coastal Washington Western Hemlock Stands, paper presented at 1247 Proceedings of the symposium on research and management of annosus root disease 1248 (Heterobasidion annosum) in western North. America, Pacific Southwest Forest and 1249 Range Experiment Station, Forest Service, U.S. Department of Agriculture, Monterey, 1250 CA, April 18-21, 1989. 1251 Litvak, M., S. Miller, S. C. Wofsy, and M. Goulden (2003), Effect of stand age on whole 1252 ecosystem CO2 exchange in the Canadian boreal forest, Journal of Geophysical 1253 Research, 108(D3), doi:10.1029/2001JD000854. 1254 Logan, J., J. Régnière, and J. A. Powell (2003), Assessing the impacts of global warming on 1255 forest pest dynamics, Frontiers in Ecology and the Environment, 1(3), 130-137. 1256 Logan, J. A., P. White, B. J. Bentz, and J. A. Powell (1998), Model analysis of spatial patterns in 1257 mountain pine beetle outbreaks, *Theoretical Population Biology*, 53(3), 236-255. 1258 Logan, J. A., and B. J. Bentz (1999), Model analysis of mountain pine beetle (Coleoptera: 1259 Scolytidae) seasonality, Environ. Entomol., 28(6), 924-934. 1260 Logan, J. A., and J. A. Powell (2001), Ghost forests, global warming and the mountain pine 1261 beetle (Coleoptera: Scolytidae), American Entomologist, 47(3), 160-173. 1262 Logan, J. A., and J. A. Powell (in press), Ecological consequences of climate change altered 1263 forest insect disturbance regimes, in Climate change in western North America: evidence 1264 and environmental effects, edited by F. H. Wagner, University of Utah Press.

1265 Lombardero, M. J., M. P. Ayres, and B. D. Ayres (2006), Effects of fire and mechanical 1266 wounding on Pinus resinosa resin defenses, beetle attacks, and pathogens, Forest Ecology 1267 and Management, 225(1-3), 349-358. 1268 Loo, J. (2009), Ecological impacts of non-indigenous invasive fungi as forest pathogens, 1269 Biological Invasions, 11(1), 81-96. 1270 Loomis, R. C., S. Tucker, and T. H. Hofacker (1985), *Insect and Disease Conditions in the* 1271 United States 1979–1983, U. S. Department of Agriculture, Washington, DC. 1272 Lovett, G. M., C. D. Canham, M. A. Arthur, K. C. Weathers, and R. D. Fitzhugh (2006), Forest 1273 ecosystem responses to exotic pests and pathogens in eastern North America, BioScience, 1274 *56*(5), 395-405. 1275 Luque, J., M. Cohen, R. Save, C. Biel, and I. F. Alvarez (1999), Effects of three fungal 1276 pathogens on water relations, chlorophyll fluorescence and growth of *Quercus suber* L, 1277 Annals of Forest Science, 56(1), 19-26. 1278 MacLean, D. A. (1990), Impact of forest pests and fire on stand growth and timber yield: 1279 implications for forest management planning, Canadian Journal of Forest Research, 20, 1280 391-404. 1281 Maclean, D. A. (1990a), Impact of forest pests and fire on stand growth and timber yield -1282 Implications for forest management planning, Canadian Journal of Forest Research, 1283 *20*(4), 391-404. 1284 Maclean, D. A. (1990b), Impact of forest pests and fire on stand growth and timber yield: 1285 Implications for forest management planning, Canadian Journal of Forest Research, 1286 *20*(4), 391-404.

1287 Magarey, R. D., G. A. Fowler, D. M. Borchert, T. B. Sutton, M. Colunga-Garcia, and J. A. 1288 Simpson (2007), NAPPFAST: An internet system for the weather-based mapping of plant 1289 pathogens, Plant Disease, 91, 336-345. 1290 Mainwaring, D. B., D. A. Maguire, J. Kanaskie, and J. Brandt (2005), Growth responses to 1291 commercial thinning in Douglas-fir stands with varying intensity of Swiss needle cast, 1292 Canadian Journal of Forest Research, 35, 2394–2402. 1293 Maloy, O. C. (1997), White pine blister rust control in North America: A case history, Annual 1294 Review of Phytopathology, 35, 87-109. 1295 Manion, P. D., and D. Lachance (1992), Forest Decline Concepts, 249 pp., APS Press, St. Paul, 1296 MN. 1297 Manter, D. K., B. J. Bond, K. L. Kavanagh, J. K. Stone, and G. M. Filip (2003), Modelling the 1298 impacts of the foliar pathogen, *Phaeocryptopus gaeumannii*, on Douglas-fir physiology: 1299 net canopy carbon assimilation, needle abscission and growth, Ecol. Model., 164(2-3), 1300 211-226. 1301 Marosy, M., and J. R. Parmeter (1989), The Incidence and Impact of *Heterobasidion annosum* 1302 on Pine and Incense-Cedar in California Forests, paper presented at Proceedings of the 1303 symposium on research and management of annosus root disease (Heterobasidion 1304 annosum) in western North. America, Pacific Southwest Forest and Range Experiment 1305 Station, Forest Service, U.S. Department of Agriculture, Monterey, CA, April 18-21, 1306 1989. 1307 McConnell, T. J., E. W. Johnson, and B. Burns (2000), A guide to conducting aerial 1308 sketchmapping surveys, USDA Forest Service, Forest Health Technology Enterprise 1309 Team, Fort Collins, Colorado.

1310	McDowell, N., et al. (2008), Mechanisms of plant survival and mortality during drought: why do
1311	some plants survive while others succumb to drought?, New Phytologist, 178(4), 719-
1312	739.
1313	Meinzer, F. C., D. R. Woodruff, and D. C. Shaw (2004), Integrated responses of hydraulic
1314	architecture, water and carbon relations of western hemlock to dwarf mistletoe infection,
1315	Plant Cell and Environment, 27(7), 937-946.
1316	Mitchell, R. G., and H. K. Preisler (1998), Fall rate of lodgepole pine killed by mountain pine
1317	beetle in central Oregon, Western Journal of Applied Forestry, 13(1), 23-26.
1318	Moorcroft, P. R., S. W. Pacala, and M. A. Lewis (2006), Potential role of natural enemies during
1319	tree range expansions following climate change, Journal of Theoretical Biology, 241(3),
1320	601-616.
1321	Morehouse, K., T. Johns, J. Kaye, and A. Kaye (2008), Carbon and nitrogen cycling immediately
1322	following bark beetle outbreaks in southwestern ponderosa pine forests, Forest Ecology
1323	and Management, 255(7), 2698-2708.
1324	Morrison, D. J., and K. I. Mallet (1996), Silvicultural management of Armillaria root disease in
1325	western Canadian forests, Canadian Journal of Forest Research, 18, 194-199.
1326	Morrison, D. J., K. W. Pellow, D. J. Norris, and A. F. L. Nemec (2000), Visible versus actual
1327	incidence of Armillaria root disease in juvenile coniferous stands in the southern interior
1328	of British Columbia, Canadian Journal of Forest Research, 30(3), 405-414.
1329	Morrison, D. J. (2005), Past and current research on the biology of Armillaria ostoyae in the
1330	southern interior of British Columbia, paper presented at Proceedings of the 53rd
1331	Western International Forest Disease Work Conference, Jackson, Wyoming, August 26-
1332	29, 2005.

1333 Natural Resources Canada (2009), Statistical data - Canada's forests. 1334 Neary, D. G., K. C. Ryan, and L. F. DeBano (2005), Wildland fire in ecosystems: effects of fire 1335 on soils and water, 250 pp, U.S. Department of Agriculture, Forest Service, Rocky 1336 Mountain Research Station, Ogden, UT. 1337 Nelson, R. F. (1983), Detecting forest canopy change due to insect activity using Landsat MSS, 1338 Photogramm. Eng. Remote Sens., 49(9), 1303-1314. 1339 Neuenschwander, L. F., J. W. Byler, A. E. Harvey, G. I. McDonald, D. S. Ortiz, H. L. Osborne, 1340 G. C. Snyder, and A. Zack (1999), White pine in the American West: a vanishing 1341 species—Can we save it?, 20 pp, U.S. Department of Agriculture, Forest Service, Rocky 1342 Mountain Research Station, Moscow, ID. 1343 Nuckolls, A. E., N. Wurzburger, C. R. Ford, R. L. Hendrick, J. M. Vose, and B. D. Kloeppel 1344 (2009), Hemlock declines rapidly with hemlock woolly adelgid infestation: Impacts on 1345 the carbon cycle of southern Appalachian forests, *Ecosystems*, 12(2), 179-190. 1346 Odum, E. P. (1969), Strategy of ecosystem development, Science, 164(3877), 262-&. 1347 Papaik, M. J., C. D. Canham, E. F. Latty, and K. D. Woods (2005), Effects of an introduced 1348 pathogen on resistance to natural disturbance: beech bark disease and windthrow, 1349 Canadian Journal of Forest Research, 35(8), 1832-1843. 1350 Parker, T. J., K. M. Clancy, and R. L. Mathiasen (2006), Interactions among fire, insects and 1351 pathogens in coniferous forests of the interior western United States and Canada, 1352 *Agricultural and Forest Entomology*, 8(3), 167-189. 1353 Peet, F. G., and R. S. Hunt (2005), A new simulator for the spread of forest root diseases by 1354 individual root contacts, For. Sci., 51(5), 425-437.

1355 Peltzer, D. A., R. B. Allen, G. M. Lovett, D. Whitehead, and D. A. Wardle (2010), Effects of 1356 biological invasions on forest carbon sequestration, Global Change Biol., 16(2), 732-746. 1357 Pfeifer, E. M., J. A. Hicke, and A. J. H. Meddens (2010), Observations and modeling of 1358 aboveground tree carbon stocks and fluxes following a bark beetle outbreak in the 1359 western United States Global Change Biol. 1360 Phelps, W. R., and F. L. Czabator (1978), Fusiform rust of southern pines, USDA Forest Service. 1361 Plattner, A., J. J. Kim, S. DiGuistini, and C. Breuil (2008), Variation in pathogenicity of a 1362 mountain pine beetle-associated blue-stain fungus, Grosmannia clavigera, on young 1363 lodgepole pine in British Columbia, Canadian Journal of Plant Pathology-Revue 1364 Canadienne De Phytopathologie, 30(3), 457-466. 1365 Poland, T. M., and D. G. McCullough (2006), Emerald ash borer: Invasion of the urban forest 1366 and the threat to North America's ash resource, Journal of Forestry, 104, 118-124. 1367 Powers, H. R., Jr., R. A. Schmidt, and G. A. Snow (1981), Current status and management of 1368 fusiform rust on southern pines, Annual Review of Phytopathology, 19, 353-371. 1369 Pu, R., M. Kelly, G. L. Anderson, and P. Gong (2008), Using CASI hyperspectral imagery to 1370 detect mortality and vegetation stress associated with a new hardwood forest disease, 1371 Photogramm. Eng. Remote Sens., 74(65-75). 1372 Quayle, D., J. Régnière, N. Cappuccino, and A. Dupont (2003), Forest composition, host-1373 population density, and parasitism of spruce budworm *Choristoneura fumiferana* eggs by 1374 Trichogramma minutum, Entomologia Experimentalis Et Applicata, 107(3), 215-227. 1375 Radeloff, V. C., D. J. Mladenoff, and M. S. Boyce (1999), Detecting jack pine budworm 1376 defoliation using spectral mixture analysis: Separating effects from determinants, Remote 1377 Sens. Environ., 69(2), 156-169.

1378 Raffa, K. F., and A. A. Berryman (1987), Interacting selective pressures in conifer-bark beetle 1379 systems: A basis for reciprocal adaptations?, Am. Nat., 129(2), 234-262. 1380 Raffa, K. F., B. H. Aukema, B. J. Bentz, A. L. Carroll, J. A. Hicke, M. G. Turner, and W. H. 1381 Romme (2008), Cross-scale drivers of natural disturbances prone to anthropogenic 1382 amplification: The dynamics of bark beetle eruptions, *BioScience*, 58, 501-517. 1383 Régnière, J., and B. Bentz (2007), Modeling cold tolerance in the mountain pine beetle, 1384 *Dendroctonus ponderosae, Journal of Insect Physiology*, 53(6), 559-572. 1385 Régnière, J., and V. G. Nealis (2007), Ecological mechanisms of population change during 1386 outbreaks of the spruce budworm, *Ecological Entomology*, 32(5), 461-477. 1387 Régnière, J., V. Nealis, and K. Porter (2009), Climate suitability and management of the gypsy 1388 moth invasion into Canada, *Biological Invasions*, 11, 135-148. 1389 Robinson, D. C. E., and B. W. Geils (2006), Modelling dwarf mistletoe at three scales: life 1390 history, ballistics and contagion, Ecol. Model., 199(1), 23-38. 1391 Rogan, J., J. Miller, M.A. Wulder, and S.E. Franklin (2006), Integrating GIS and remotely sensed 1392 data for mapping forest disturbance and change, in *Understanding Forest Disturbance* 1393 and Spatial Pattern: Remote Sensing and GIS Approaches, edited, pp. 133-171, Taylor & 1394 Francis (CRC Press), Boca Raton. 1395 Romme, W. H., D. H. Knight, and J. B. Yavitt (1986), Mountain pine beetle outbreaks in the 1396 Rocky Mountains: Regulators of primary productivity, Am. Nat., 127(4), 484-494. 1397 Romme, W. H., J. Clement, J. Hicke, D. Kulakowski, L. H. MacDonald, T. L. Schoennagel, and 1398 T. T. Veblen (2006), Recent Forest Insect Outbreaks and Fire Risk in Colorado Forests: 1399 A Brief Synthesis of Relevant Research, edited, Colorado Forest Restoration Institute, 1400 Colorado State University.

1401	Royle, D. D., and R. G. Lathrop (1997), Monitoring hemlock forest health in New Jersey using
1402	Landsat TM data and change detection techniques, For. Sci., 42(3), 327-335.
1403	Safranyik, L., and D. A. Linton (1991), Unseasonably low fall and winter temperatures affecting
1404	mountain pine beetle and pine engraver beetle populations and damage in the British
1405	Columbia Chilcotin Region, Journal of the Entomological Society of British Columbia,
1406	88, 17-21.
1407	Safranyik, L., and A. Carroll (2006a), The biology and epidemiology of the mountain pine beetle
1408	in lodgepole pine forests, in The mountain pine beetle: a synthesis of biology,
1409	management, and impacts on lodgepole pine, edited by L. Safranyik and W. R. Wilson,
1410	pp. 3-66, Natural Resources Canada, Canadian Forest Service, Pacific Forestry Centre,
1411	Victoria, British Columbia.
1412	Safranyik, L., and A. L. Carroll (2006b), The biology and epidemiology of the mountain pine
1413	beetle in lodgepole pine forests, in The mountain pine beetle: a synthesis of biology,
1414	management, and impacts on lodgepole pine, edited by L. Safranyik and W. R. Wilson,
1415	pp. 3-66, Natural Resources Canada, Canadian Forest Service, Pacific Forestry Centre,
1416	Victoria, British Columbia.
1417	Salinas-Moreno, Y., A. Ager, C. F. Vargas, J. L. Hayes, and G. Zuniga (2010), Determining the
1418	vulnerability of Mexican pine forests to bark beetles of the genus Dendroctonus Erichson
1419	(Coleoptera: Curculionidae: Scolytinae), Forest Ecology and Management, 260(1), 52-
1420	61.
1421	Schäfer, K. V. R., K. L. Clark, N. Skowronski, and E. P. Hamerlynck (2009), Impact of insect
1422	defoliation on forest carbon balance as assessed with a canopy assimilation model,
1423	Global Change Biol.

1424 Schimel, D. S., et al. (2001), Recent patterns and mechanisms of carbon exchange by terrestrial 1425 ecosystems, Nature, 414(6860), 169-172. 1426 Schmidt, R. A., R. C. Holley, M. C. Klapproth, and T. Miller (1986), Temporal and spatial 1427 patterns of fusiform rust epidemics in young plantations of susceptible and resistant slash 1428 and loblolly pines, *Plant Disease*, 70, 661-666. 1429 Seidl, R., W. Rammer, D. Jager, and M. J. Lexer (2008), Impact of bark beetle (*Ips typographus* 1430 L.) disturbance on timber production and carbon sequestration in different management 1431 strategies under climate change, Forest Ecology and Management, 256(3), 209-220. 1432 Shore, T. L., and L. Safranyik (1992), Susceptibility and risk rating systems for the mountain 1433 pine beetle in lodgepole pine stands, Forestry Canada. 1434 Simard, M., E. N. Powell, J. M. Griffin, K. F. Raffa, and M. G. Turner (2008), Annotated 1435 Bibliography for Forest Managers on Fire-Bark Beetle Interactions, University of 1436 Wisconsin, http://www.fs.fed.us/wwetac/publications/WWETAC Fire-1437 BB InterX 25Feb2008.pdf. 1438 Sinclair, W. A., H. H. Lyon, and W. T. Johnson (2005), Diseases of Trees and Shrubs, 2nd ed., 1439 Cornell University Press. 1440 Singh, P. (1993), Research and management strategies for major tree diseases in Canada: 1441 Synthesis .1, For. Chron., 69(2), 151-162. 1442 Six, D. L. (2003), Bark beetle-fungus symbiosis, in *Insect symbiosis*, edited by T. Miller and K. 1443 Kourtzis, pp. 97–114, CRC Press. 1444 Six, D. L., and J. Adams (2007), White pine blister rust severity and selection of individual 1445 whitebark pine by the mountain pine beetle (Coleoptera: Curculionidae, Scolytinae), J. 1446 Entomol. Sci., 42(3), 345-353.

1447	Stevens, M. T., D. M. Waller, and R. L. Lindroth (2007), Resistance and tolerance in Populus
1448	tremuloides: genetic variation, costs, and environmental dependency, Evolutionary
1449	Ecology, 21(6), 829-847.
1450	Sun, J., et al. (2010), A multiscale and multidisciplinary investigation of ecosystem-atmosphere
1451	CO ₂ exchange over the Rocky Mountains of Colorado, Bull. Amer. Meteorol. Soc., 91(2)
1452	209-230.
1453	Tauber, M. J., C. A. Tauber, and S. Masaki (1986), Seasonal Adaptations of Insects, Oxford
1454	University Press.
1455	Taylor, S. W., and A. L. Carroll (2004), Disturbance, forest age dynamics and mountain pine
1456	beetle outbreaks in BC: A historical perspective, in Mountain Pine Beetle Symposium:
1457	Challenges and Solutions, edited by T. L. Shore, et al., pp. 41-51, Natural Resources
1458	Canada, Canadian Forest Service, Pacific Forestry Centre, Victoria, BC, Canada.
1459	Thompson, M. T. (2009a), Analysis of Conifer Mortality in Colorado Using Forest Inventory
1460	and Analysis's Annual Forest Inventory, Western Journal of Applied Forestry, 24(4),
1461	193-197.
1462	Thompson, M. T. (2009b), Mountain pine beetle infestations and sudden aspen decline in
1463	Colorado: Can the Forest Inventory and Analysis annual inventory system address the
1464	issues?, paper presented at Forest Inventory and Analysis (FIA) Symposium 2008, U.S.
1465	Department of Agriculture, Forest Service, Rocky Mountain Research Station, Park City,
1466	UT, October 21-23, 2008.
1467	Tkacz, B., B. Moody, J. Villa Castillo, and M. E. Fenn (2008), Forest health conditions in North
1468	America, Environmental Pollution, 155, 409-425.

1469 Tkacz, B. M., and E. M. Hansen (1982), Damage by laminated root rot in two succeeding stands 1470 of Douglas-fir, Journal of Forestry, 80(12), 788-791. 1471 Turchin, P., A. D. Taylor, and J. D. Reeve (1999), Dynamical role of predators in population 1472 cycles of a forest insect: an experimental test, *Science*, 285, 1068-1071. 1473 USDA Forest Service (2000), Forest insect and disease conditions in the United States 1999, 1474 United States Department of Agriculture, Forest Service, Washington, D.C. 1475 USDA Forest Service (2002), Major forest insect and disease conditions in the United States 1476 2000, 1-110 pp, United States Department of Agriculture, Forest Service, Washington, 1477 D.C. 1478 USDA Forest Service (2003a), Forest Insect and Disease Conditions in the Rocky Mountain 1479 Region, 2002, 28 pp, Washington, D.C. 1480 USDA Forest Service (2003b), Forest Insect and Disease Conditions in the United States 2001, 1481 1-124 pp, United States Department of Agriculture, Forest Service, Washington, D.C. 1482 USDA Forest Service (2004), Forest Insect and Disease Conditions in the United States, 2003, 1483 156 pp, Washington, D.C. 1484 USDA Forest Service (2005a), Forest Insect and Disease Conditions in the United States 2004, 1485 United States Department of Agriculture, Forest Service, Washington, D.C. 1486 USDA Forest Service (2005b), Forest Insect and Disease Conditions in the United States, 2004, 1487 154 pp, Washington, D.C. 1488 USDA Forest Service (2006), Forest Insect and Disease Conditions in the United States, 2005, 1489 159 pp, Washington, D.C. 1490 USDA Forest Service (2007), Forest Insect and Disease Conditions in the United States 2006, 1491 176 pp. pp, Forest Health Protection, USDA Forest Service.

1492 USDA Forest Service (2009a), Sudden Aspen Decline in Colorado Update, Forest Heath 1493 Management, Rocky Mountain Region. 1494 USDA Forest Service (2009b), Major Forest Insect and Disease Conditions in the United States 1495 2008 Update, 37 pp, Washington, D.C. 1496 USDA Forest Service (2009c), Major Forest Insect and Disease Conditions in the United States 1497 2007, 78 pp, Washington, D.C. 1498 Van Frankenhuyzen, K. (2000), Application of *Bacillus thuringiensis* in forestry, in 1499 Entomopathogenic Bacteria: From Laboratory to Field Application, edited by J.-F. 1500 Charles, et al., pp. 371-382, Kluwer Academic Publishers, The Netherlands. 1501 van Mantgem, P. J., et al. (2009), Widespread increase of tree mortality rates in the western 1502 United States, Science, 323(5913), 521-524. 1503 Vogelmann, J. E., B. Tolk, and Z. L. Zhu (2009), Monitoring forest changes in the southwestern 1504 United States using multitemporal Landsat data, Remote Sens. Environ., 113(8), 1739-1505 1748. 1506 Volney, W. J. A., and R. A. Fleming (2000), Climate change and impacts of boreal forest insects, 1507 Agriculture Ecosystems & Environment, 82(1-3), 283-294. 1508 Watt, M. S., D. J. Kriticos, S. Alcaraz, A. V. Brown, and A. Leriche (2008), The hosts and 1509 potential geographic range of Dothistroma needle blight, Forest Ecology and 1510 Management, 257, 1505-1519. 1511 Williams, D. L., and R. F. Nelson (1986), Use of remotely sensed data for assessing forest stand 1512 conditions in the eastern United States, IEEE Trans. Geosci. Remote Sensing, 24(1), 130-1513 138.

1514	Woods, A., K. D. Coates, and H. A (2005), Is an unprecedented Dothistroma needle blight
1515	epidemic related to climate change?, BioScience, 55, 761-769.
1516	Worrall, J. J., L. Egeland, T. Eager, R. A. Mask, E. W. Johnson, P. A. Kemp, and W. D.
1517	Shepperd (2008), Rapid mortality of Populus tremuloides in southwestern Colorado,
1518	USA, Forest Ecology and Management, 255(3-4), 686-696.
1519	Wulder, M. A., C. C. Dymond, J. C. White, D. G. Leckie, and A. L. Carroll (2006), Surveying
1520	mountain pine beetle damage of forests: A review of remote sensing opportunities, Forest
1521	Ecology and Management, 221(1-3), 27-41.
1522	Wygant, N. D. (1940), Effects of low temperature on the Black Hills beetle (Dendroctonus
1523	ponderosae Hopkins), Ph.D. dissertation thesis, 57 pp, State College of New York,
1524	Syracuse, NY.
1525	
1526	
1527	

Figure Captions 1527 1528 1529 Figure 1. Area affected by major insect species from US and Canadian aerial survey databases. 1530 Example species chosen to illustrate variability among species, through time, and between 1531 regions. "Affected area" includes live and dead trees. Note that impact on carbon cycle varies 1532 across species depending on whether insect is a defoliator (a, b, c, d), which may or may not kill 1533 trees, or tree-killing bark beetles (e, f). 1534 1535 Figure 2. Conceptual diagram showing impacts of insect and pathogen outbreaks on carbon 1536 budgets. Numbers outside of boxes indicate studies in Tables 3 and 4 that provide evidence. 1537 Studies that provide evidence counter to that indicated in diagram noted in bold underlined text 1538 (two cases). 1539

Table 1. Insect species that cause major impacts to the North American carbon budget.

Insect type	Insect species	Host tree species	Origin	Major outbreak examples (see also Figure 1)	US basal area risk ^a (million m ²)
bark beetles	mountain pine beetle (<i>Dendroc-tonus ponderosae</i> Hopkins)	pines of western North America (Pinus)	native	 1930s: 200,000 ha, Idaho^b 1970s-1980s: 2 Mha, western US^c 2000s: 12 Mha, western Canada^d 	69
	spruce beetle (Dendroctonus rufipennis Kirby)	spruces of western and northeastern North America (<i>Picea</i>)	native	• 1990s: 1.5 Mha, Alaska ^e • current: 112,000 ha, Alaska ^e	18
	western balsam bark beetle (<i>Dryocoetes</i> confuses Swaine)	subalpine fir (Abies lasiocarpa (Hook.) Nutt.)	native	• ongoing: 216,000 ha, western US ^c	5
	southern pine beetle (<i>Dendroc-</i> tonus frontalis Zimmermann)	pines of the southern US (Pinus)	native	 mid-1980s: 10.5 Mha^c (different system for area; see Footnote f) early 2000s: 5 Mha^{c,e} 	54
	conifer engraver species (<i>Ips</i> spp.)	pines and spruces throughout North America (<i>Pinus</i> and <i>Picea</i>)	native	• 2000s: 1.2 Mha, southwestern US ^g	33
	fir engraver beetle (Scolytus ventralis LeConte)	true firs (Abies)	native	• ongoing: 200,00 ha, western US ^e	25
defoliators	eastern spruce budworm (<i>Cho-</i> ristoneura fumif- erana [Clem.])	firs (Abies), spruces (Picea)	native	 1940s: 16 Mha, eastern Canada^h 1970s-1980s: 20 Mha eastern Canada^h 	8
	western spruce budworm (<i>Cho-</i> ristoneura occi- dentalis Freeman)	firs (Abies), spruces (Picea), Douglas-fir (Pseudotsuga menzi- esii (Mirb.) Franco)	native	• 1980s: 5 Mha, western US ^e • ongoing: 769,000 ha, western US ^e	0.5

	gypsy moth (<i>Ly</i> -	multiple trees species throughout	invasive	• early 1980s: 5.3 Mha, northeast-	
	mantria dispar	northeastern US; primarily		ern US ^e	
	L.)	angiosperms but also <i>Larix</i> and		• ongoing: 600,00 ha, northeastern	
		Pinus strobus L.		US ^e	41
	forest tent cater-	broadleaf species in North	native	• early 2000s	
	pillar (<i>Mala</i> -	America		• central US: 2.8 Mha ⁱ	
	cosoma disstria			• Canada: 14 Mha ^d	
	Hubner)				1
fluid feeders	hemlock woolly	hemlock (<i>Tsuga</i> spp.)	invasive	• ongoing and expanding; in 17	
	adelgid (Adelges			states in eastern US ^e	
	tsugae Annand)				2
wood borers	emerald ash borer	ash spp. (Fraxinus spp.)	invasive	• introduced in 2002; ongoing	
	(Agrilus			outbreak threatening species	
	planipennis			extirpations ^j	
	Fairmaire)			1	5
root feeders	weevils	maple (Acer), birch (Betula)	invasive	• 15% of fine root biomass of	
	(Curculionidae			northern hardwood forests in up-	
	spp.)			per Midwestern US consumed	
an		C. d. HG FT 11 2 . W 1 2000	73 : '1	annually	N/A

^aEstimates of basal area loss in the next 15 years for the US [Table 3 in *Krist et al.*, 2007]; similar standardized estimates not available for Canada or Mexico ^bhttp://www.usu.edu/beetle/databases_bark_beetle.htm

^cUSDA Forest Service [2009b]

^dNatural Resources Canada [2009]

^eUSDA Forest Service [2009c]

^fFor southern pine beetle, all acres within a county were counted if that county contained a single infested tree.

^gBreshears *et al.* [2005]

^hCandau *et al.* [1998]

ⁱUSDA Forest Service [2005b]

^jPoland and McCullough [2006]

^kCoyle *et al.* [2008]

Table 2. Pathogens and declines that cause major impacts to the North American carbon budget.

Pathogen type	Disease name (pathogen species)	Host tree species	Origin	Examples of major epidemics and/or long-term perturbations and impacts	US basal area risk ^a (million m ²)
canker	chestnut blight (Cryphonectria parasitica (Murrill) Barr.)	American chestnut (Castanea dentate (Marshall) Borkhausen)	invasive	 1920s: 3.5 billion trees infected^b 1950s: 3.6 Mha dead or dying^c C. dentata extirpated from its original range^d 	host extir- pated
	beech bark disease (Neonectria spp.)	American beech (Fagus grandifolia Ehrh.)	invasive	 1970: most of the NE USA and SE Canada affected 1977: Vermont lost 708,000 cubic m^f 	7
foliar	Red band needle blight (Dothistroma septosporum)	pines (<i>Pinus</i>) and other conifers	native	• 2002-2004: 38,000 ha impacted, mortality of 2,700 ha in NW BC ^g • Extensive mortality in plantations in BC ^h	N/A
	Swiss needle cast (<i>Phaeocryptopus gäu-mannii</i> (Rohde) Petrak)	Douglas-fir (Pseudotsuga menziesii (Mirb.) Franco)	native	 2010: 160,000 ha affected in Oregonⁱ 2000: stands with the most severe SNC had volume growth loss of approximately 52%^j 	N/A
wilt	Dutch elm disease (Ophiostoma ulmi (Buisman) Nannf and O. novo-ulmi)	elms (<i>Ulmus</i> spp.)	invasive	• 1930-1973: widespread loss of large trees across NA ^{d,k}	1
root	Armillaria root disease (Armillaria solidipes = A. ostoyae)	mainly conifers	native	 responsible for volume losses of 2-3 Mm³ year¹ in Canada's Pacific NW¹ in some forests of western NA, 35% of annual tree mortality caused by Armillaria^m 	48 (all root diseases)

Pathogen type	Disease name (pathogen species)	Host tree species	Origin	Examples of major epidemics and/or long-term perturbations and impacts	US basal area risk ^a (million m ²)
	Annosus root disease (Heterobasidion annosum = H. occidentale & H. parviporum)	many conifer and hardwood species	native	 infections range from 10-50%; in severe cases 30% of a stand may be killedⁿ 12.5% of CA commercial forest land is infested, annual losses of 262,000 cubic m^o 	48 (all root diseases)
stem rust	fusiform rust (Cronartium fusiforme Hedgcock & Hunt ex Cummins)	loblolly (<i>P. taeda</i>), slash (<i>P.elliottii</i> var. elliottii) pines	native	 1930s: recognized as problem in MS & LA, regionally up to 30% of seedlings infected^p 1960-70s disease incidence: 5.5 Mha affected >10%, 1 Mha >50%^{p,q,r} 	2
	white pine blister rust (<i>Cronartium ribicola</i> J.C. Fisch Ex. Raben)	Five-needle pines (<i>Pinus</i> spp.)	invasive	 in northern Idaho, trees decreased 90% where previously dominant^s 0.6 Mm³ estimated mortality loss in Canada, 1976^t 	2
dwarf mistletoe	Arceuthobium spp.	Pinaceae and Cupressaceae	native	 in Canada, 3.5 Mm³ loss to growth reduction and mortality in 1981 and 1987^t in US, 3.8 Mha, 11 Mm³ (10-50% of annual commercial harvest) lost/year in growth, mortality^{u,v} 	6
Phy- tophthora	sudden oak death (<i>Phytophthora</i> ramorum sp. nov)	Numerous spp. identified as potential hosts	invasive	• 1990s: in CA, caused mortality of >3 million oaks (<i>Quercus agrifolia</i> Née) and tanoaks (<i>Lithocarpus densiflorus</i> (Hook. & Arn.) Rehd.) ^w	5
decline	aspen decline (multiple causes including drought, damage by insects and pathogens)	Trembling aspen (<i>Populus tremuloides</i> Michx.)	began recently	 2008 mortality in CO forests was > 222,500 ha^x in Ontario, Canada, area affected >500,000 ha in 2000-2004; plot-level mortality of 74% in 2005^y 2000s: mortality across 10 Mha in Saskatchewan and Alberta^z 	6

^aEstimates of basal area loss in the next 15 years for the US [Table 3 in Krist et al., 2007]; similar standardized estimates not available for Canada or Mexico ^yEvans *et al.* [2006]

^bAgrios [2005]

^cAnagnostakis [1987]

^dLoo [2009]

^eGriffin et al. [2003]

fHouston [1994]

^gWatt *et al*. [2008]

^hWoods *et al.* [2005]

iwww.cof.orst.edu/coops/snc c/2010Results.htm

^kFarrar [1995]

^jMainwaring *et al.* [2005]

¹Morrison and Mallet [1996]

^mSinclair *et al.* [2005] ⁿLittke and Browning [1989]

^oMarosy and Parmeter [1989] ^pPowers *et al.* [1981]

^qSchmidt *et al.* [1986]

^rAnderson et al. [1986]

^sNeuenschwander *et al.* [1999]

^zHogg *et al.* [2008]

^tSingh [1993]

^uGill [1954] ^vDrummond [1982]

WGoheen et al. [2006]

*USDA Forest Service [2009a]

Table 3. Summary of studies that report effects of insect outbreaks on carbon budgets.

	Outbreak cha			Study characteristics						Study results	
Insect type	Insect spe- cies	Host type	Location	Methods	Spatial extent	Time since disturbance	Outbreak severity	Control	Carbon variable ^a	Results	Refer- ence
bark beetles	mountain pine beetle	lodge- pole pine	British Colum- bia	two years of field measure- ments (eddy flux tower),	two stands, MPB-03 and MPB-	• MPB-03: 4-5 years	• >95% red- and gray-attack	• none	NEP	• -56 and 4 g C m ⁻² yr ⁻¹ in Years 1 and 2	Brown et al. [2010]
1 ^b				2006-2007	06	• MPB-06: 0-1 years	• Year 1: 50% green-attack • Year 2: 73% dead	• Year 1, preat-tack		• -82 and -33 g C m ⁻² yr ⁻¹ in Years 1 and 2	
2				simulation modeling	regional	21 years during and following outbreak	varies	years prior to outbreak	NBP	NBP change from sink to a source; magnitude of source was reduced but >0 after 21 years	Kurz et al. [2008a]
3			Yellow- stone area	field measure- ments (dendrochro- nological)	4 attacked stands	decades be- fore, 10-20 years fol- lowing infestation	41-67%	preout- break and uninfes- tated con- trol stands	NPP as indicated by bole volume increment	10 years to recovery to preoutbreak values and values from unattacked stands	Romme et al. [1986]
			central Idaho	field measure- ments, simulation modeling	12 stands	100+ years following outbreak	18-52% number of trees, 31-83% C stocks	years prior to outbreak and uninfes- tated con- trol simu- lations	 above- ground C stocks above- ground woody C production 	 recovered to preoutbreak values in 1-25 years and to unattacked simulations in 56-185 years never recovered to preoutbreak values; recovered to unattacked simulations in 30-160 	Pfeifer et al. [2010]
4	Ips and Den- droctonus	ponder- osa pine	south- western US	field measure- ments	10 in- fested	0-2 years following infestation	≥80%	10 unin- fested stands	above- ground C stocks soil respi-	60% in killed trees similar among infested and	More- house et al. [2008]
5 defo- liators	forest tent caterpillar outbreaks (with drought)	aspen	two sites in central Canada	dendro- chronology, simulation modeling	multiple stands	decades be- fore and after out- break	70% of leaf area lost per tree (prescribed); #trees attacked from tree ring analysis (19-97%)	results in years before and years after outbreak	• stem growth	 uninfested stands reduction by 80-90% reduction by 15-25% 	Hogg [1999]

	Outbreak cha	racteristic	es .		5	Study character		Study results			
Insect type	Insect spe- cies	Host type	Location	Methods	Spatial extent	Time since disturbance	Outbreak severity	Control	Carbon variable ^a	Results	Refer- ence
7	forest tent caterpillar outbreaks	north- ern hard- wood	Wisconsin	field measure- ments, eddy flux tower, simulation modeling	one stand	year of, year before, year after outbreak	40% of leaf area	measure- ments in year be- fore and year after outbreak	• GPP • NEP	24% decrease during year of outbreak reduction in sink (77 g C m² year¹ vs. 323-422 g C m² year¹)	Cook et al. [2008]
	gypsy moth	pine/ oak	New Jersey	field measure- ments, eddy flux tower, modeling upscaling	three stands	two years during out- break	partial to complete defoliation in second and third years 20% of landscape	year prior to out- break	• GPP • NEP • annual	 25-58% reduction reduced sink or switch to C source 41% reduction; 55% 	Clark et al. [2009] and Schäfer et al.
8				using aerial surveys		break	had ≥75% defoliation	without outbreak	NEE	reduction in most impacted stands	[2009]
9	eastern spruce bud- worm	eastern forests	Canada, US	remote sensing, modeling	regional	5-40 years following defoliation	unspecified	none	NPP	recovery indicated by increasing NPP over 17-year period	Hicke et al. [2002]
			Quebec, Canada	simulation modeling	regional (106,000 km²)	outbreak and study period 2011-2024	various: 0-30% mortality per year; 0-86% growth loss per year; 95% of study region experienced	simula- tions without outbreak	 C stocks NPP NBP	 reductions of 11-90% in merchantable C and 2-10% in ecosystem C reduction of 3.5% switch from sink (4.6 g C 	Dy- mond et al. [2010]
10							defoliation			m ² yr ⁻¹) to source (-16.8 g C m ² yr ⁻¹) during outbreak	
fluid feeders	hemlock woolly adelgid	eastern hem- lock species	north- eastern US	eddy flux towers	hemlock and oak stands	N/A; effects estimated by comparing pre- and postoutbreak forest types	N/A; outbreak effects estimated by comparing pre- and postoutbreak forest types	N/A	NEE	maximum postoutbreak forest (oak) uptake 60% greater than hemlock annual NEE similar	Hadley et al. [2008]
12				field measure- ments	six sites	5-6 years after initial infestation	ranged from 0- 98% basal area in infested hemlocks	sites across gradient of damage	soil C and C:N in total soil, forest floor, or mineral soil	no differences among sites with different levels of infestation	Jenkins et al. [1999]

	Outbreak cha	racteristic	S		S	tudy character	Study results				
Insect	Insect spe-	Host	Location	Methods	Spatial	Time since	Outbreak	Control	Carbon	Results	Refer-
type	cies	type			extent	disturbance	severity		variable ^a		ence
				field measure- ments	six in- fested sites	unspecified	43% of hemlock trees killed, 35% of basal area	four control sites	• soil C, mineral C:N	no differences compared with uninfested stands	Kizlin- ski et al. [2002]
13									• forest floor mass, forest floor C:N	lower in damaged sites	,
			southern US	field measure- ments	plots	three years of infestation	unspecified	year prior to infesta- tion and	basal area increment	• decreased by 50-90%	Nuck- olls et al.
								hardwood plots without infestation	very fine root bio- mass	decreased by 20-40% in two years	[2009]
14									• soil CO ₂ efflux	• decreased by 20% in one year	
15			eastern US	spread and ecosystem modeling	hemlock locations in eastern US	continuous	linear increase 0- 50% for 0-20 years following initial infestation	simulation without insect outbreak	regional NEP	for 2000-2040 period, reduction by 8%; for 2040- 2100 period, increase by 12%	Albani et al. [2010]
multiple	eastern spruce bud- worm, mountain pine beetle, jack pine budworm, hemlock	various	Canadian forests	simulation modeling	regional	1920-1989	various	years prior to infestation	NEP	increased disturbance (insect outbreaks, fire) in latter decades contributed to switch from sink to source	Kurz and Apps [1999]
16	looper										

^aSee text for explanation of abbreviations. ^bStudy identifier used in Figure 2.

Table 4. Summary of studies that report effects of pathogen outbreaks on carbon budgets.

	Outbreak cha	racteristic	·s		S	Study character	Results				
Patho- gen type	Pathogen species	Host type	Location	Methods	Spatial extent	Time since disturbance	Outbreak severity	Control	Carbon variable ^a	Results	Refer- ence
canker	beech bark disease	American beech	northeast ern US	field observa- tions	eight plots	unspecified	5-95% live beech by basal area; no information pro- vided about %beech killed	gradient of beech bark disease damage	above-ground NPP growing season soil CO ₂ efflux	 no differences across disease gradient decreased by 40% in infested stands 	Han- cock et al. [2008]
17				field observa- tions during (1985) and after (2000) outbreak	49 plots	15 years	see biomass	preout- break measure- ments	above- ground live bio- mass of beech	1% reduction in live and 99% increase in dead beech	Forrester et al. [2003]
18									• litterfall, % lignin	no change	
foliar	Swiss nee- dle cast, Phaeoc- ryptopus gaeumannii	Doug- las-fir	western Oregon, US	field measure- ments and modeling	trees at three sites	ongoing	density of pathogen: 0-70% of needle stomata	uninfected control trees sprayed	• needle net as- similation	• negative needle net carbon balance when pathogen colonization >25% of stomata	Manter et al. [2003]
19									• whole- canopy net assimila- tion	• reduced by 85%	
parasite 20	dwarf mistletoe (<i>Arceuthobium</i> spp.)	western hem- lock	Wash- ington State, US	field measurements	eight trees at one site	ongoing	five severely infested trees	three uni- fested trees	maximum photosyn- thetic rates	in heavily infested trees, photosynthetic rate was half that of uninfested trees	Meinzer et al. [2004]

^aSee text for explanation of abbreviations. ^bStudy identifier used in Figure 2.

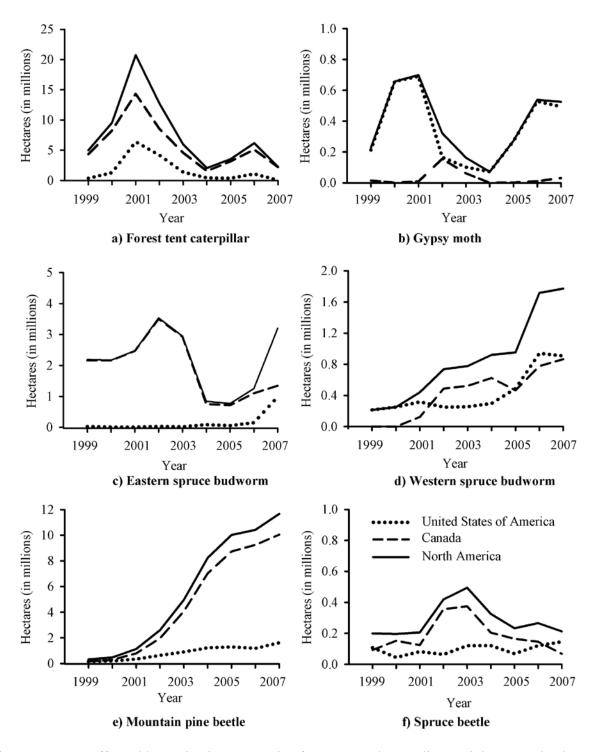


Figure 1. Area affected by major insect species from US and Canadian aerial survey databases. Example species chosen to illustrate variability among species, through time, and between regions. "Affected area" includes live and dead trees. Note that impact on carbon cycle varies across species depending on whether insect is a defoliator (a, b, c, d), which may or may not kill trees, or tree-killing bark beetles (e, f).

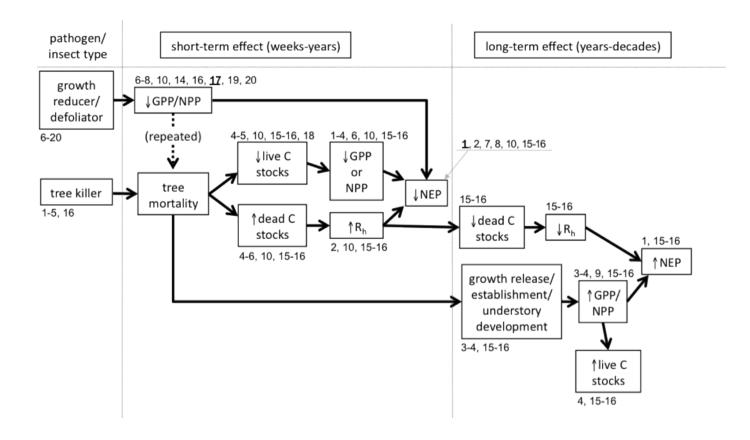


Figure 2. Conceptual diagram showing impacts of insect and pathogen outbreaks on carbon budgets. Numbers outside of boxes indicate studies in Tables 3 and 4 that provide evidence. Studies that provide evidence counter to that indicated in diagram noted in bold underlined text (two cases).