

# Triple Echo Steady-State (TESS) Relaxometry

Rahel Heule, 1\* Carl Ganter, 2 and Oliver Bieri 1

**Purpose:** Rapid imaging techniques have attracted increased interest for relaxometry, but none are perfect: they are prone to static  $(B_0)$  and transmit  $(B_1)$  field heterogeneities, and commonly biased by  $T_2/T_1$ . The purpose of this study is the development of a rapid  $T_1$  and  $T_2$  relaxometry method that is completely  $(T_2)$  or partly  $(T_1)$  bias-free.

**Methods:** A new method is introduced to simultaneously quantify  $T_1$  and  $T_2$  within one single scan based on a triple echo steady-state (TESS) approach in combination with an iterative golden section search. TESS relaxometry is optimized and evaluated from simulations, in vitro studies, and in vivo experiments.

**Results:** It is found that relaxometry with TESS is not biased by  $T_2/T_1$ , insensitive to  $B_0$  heterogeneities, and, surprisingly, that TESS- $T_2$  is not affected by  $B_1$  field errors. Consequently, excellent correspondence between TESS and reference spin echo data is observed for  $T_2$  in vitro at 1.5 T and in vivo at 3 T. **Conclusion:** TESS offers rapid  $T_1$  and  $T_2$  quantification within one single scan, and in particular  $B_1$ -insensitive  $T_2$  estimation. As a result, the new proposed method is of high interest for fast and reliable high-resolution  $T_2$  mapping, especially of the musculoskeletal system at high to ultra-high fields. **Magn Reson Med 71:230–237, 2014.** © **2013 Wiley Periodicals, Inc.** 

**Key words:** triple echo steady-state; relaxometry; fast imaging;  $T_1$ ;  $T_2$ ; quantification

Since its introduction more than half a century ago, the use of steady-state free precession (SSFP) (1) has become increasingly popular, and a large number of SSFP imaging techniques has been described so far (e.g., see Handbook of MRI Pulse Sequences (2). Besides morphological imaging, SSFP has also attracted considerable interest for fast quantitative MRI. Quantitative imaging is thought to represent an important future step toward a significant improvement of the diagnostic potential of MRI, for the early detection of subtle or diffuse pathological changes with high specificity and sensitivity, for an unbiased assessment of treatment or drug effects, as well as for clinical trials in drug research across different sites. When compared with morphological imaging, however,

quantification requires longer scan time and the overall success and applicability of quantitative MRI methods in the clinical setting will depend greatly on the overall acquisition speed. As a result, SSFP-based imaging techniques have come into the research focus, e.g., for relaxation time mapping (3–10), for measuring molecular proton diffusion (11–15), for the assessment of magnetization transfer effects (16–18), or for the characterization of flow or motion (19–21).

Relaxation is one of the most fundamental fingerprints

of NMR. It not only defines contrast in conventional MRI but also reflects the local interaction of water on a molecular and thus very fundamental level. Longitudinal relaxation  $(T_1)$  is typically assessed from inversion recovery (IR) spin echo (SE) techniques, whereas transverse relaxation  $(T_2)$  is commonly based on sampling the decay of the transverse magnetization using single-echo or multiecho SE methods. Acquiring the complete  $T_1$  recovery or T2 decay curve is time consuming and frequently requires segmented imaging strategies. In contrast, quantification techniques that make use of the functional dependence of the steady state on its intrinsic and extrinsic parameters are considerably faster. A common feature of rapid SSFP sequences is their mixed  $T_2$ /  $T_1$  imaging contrast; a natural consequence of a pulse repetition time (TR) being much shorter than  $T_2$ . As a result, accurate quantification of relaxation times using SSFP-based imaging techniques is hampered by a varying marginal  $T_2$ -related bias in  $T_1$  estimates, as seen in radiofrequency spoiled SSFP (22), or by a more or less pronounced  $T_1$ -related bias in  $T_2$ , as seen in balanced SSFP (23), partially spoiled SSFP (10), and double echo SSFP (9). Moreover, all SSFP imaging techniques proposed so far, as well as multiecho SE techniques commonly used for  $T_2$  mapping, are highly sensitive to transmit field  $(B_1)$  inhomogeneities that become especially prominent at high (3 T) to ultra-high (7 T and higher) field strength. As a result, without additional corrective data, accurate and thus fast and reliable quantification of relaxation times is not feasible in practice.

In summary, several different steady-state methods have been proposed for relaxometry, but none are perfect: all of them are  $B_1$ -sensitive, require multiple scans (except for the dual echo approach (9)), suffer from a  $T_2/T_1$  related bias, or are prone to static ( $B_0$ ) field inhomogeneities (24). Hence, there is much room for methodological improvement. In this work, we present a completely new approach for rapid quantification of both,  $T_1$  and  $T_2$  relaxation times, within one single scan using a triple echo steady-state (TESS) approach. Here, we make use of the different functional dependence of the two lowest order SSFP-FID modes (free induction decay) and of the lowest order SSFP-Echo mode, acquired within every TR, for rapid relaxometry using an iterative golden section search algorithm. The accuracy of TESS-based

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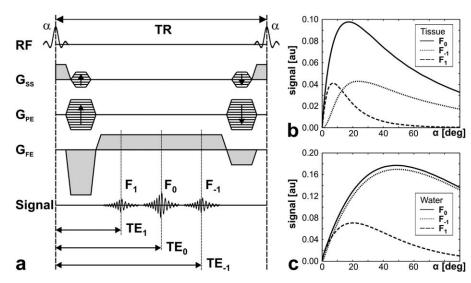
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FIG. 1. (a) Illustration of a triple echo steady-state (TESS) sequence. The center free induction decay (FID) ( $F_0$ ) is flanked by a higher order FID to the left ( $F_1$ ) and by the lowest order Echo ( $F_{-1}$ ) to the right, with echo times TE<sub>0</sub>, TE<sub>1</sub>, and TE<sub>-1</sub>, respectively. (b and c) Simulated TESS signals for tissues (b) and water (c) as a function of the flip angle  $\alpha$ . Simulation parameters: TR=16 ms, TE<sub>1</sub>=TE<sub>0</sub>  $\rightarrow$  0, and TE<sub>-1</sub>  $\rightarrow$  TR, tissues:  $T_1/T_2$ =1000 ms/50 ms, water:  $T_1/T_2$ =3000 ms/1000 ms.



relaxometry is evaluated from simulations and in vitro experiments, and the feasibility of high-resolution three-dimensional (3D)  $T_1$  and  $T_2$  mapping is presented in vivo for human articular cartilage at 3 T.

#### **METHODS**

All numerical simulations, data analysis, and visualization were done using Matlab 7.5 (The MathWorks, Inc., Natick, MA). Measurements and calibrations were performed on a clinical 1.5 T and a 3 T system equipped with actively shielded magnetic field gradient coils.

### Triple Echo Steady State

Multiecho SSFP, as displayed in Figure 1a, was already proposed by Mizumoto and Yoshitome for imaging different contrast dependencies (25). Generally, the ideal SSFP signal (no motion, no diffusion, quasi-instantaneous radiofrequency pulses) can be written in a representation that is closely related to configuration theory (26). Expressions for the two lowest order modes ( $F_0$ , representing the FID, and  $F_{-1}$  representing the Echo) immediately following the excitation pulse can be found, e.g., in (27),

$$F_0 \propto 1 - (E_1 - \cos \alpha) \cdot r \tag{1}$$

$$F_{-1} \propto (1 - (1 - E_1 \cos \alpha) \cdot r) E_2^{-1}$$
 [2]

with definitions

$$E_{1,2} := \exp\left(-TR/T_{1,2}\right)$$
 $p := 1 - E_1 \cos \alpha - E_2{}^2(E_1 - \cos \alpha)$ 
 $q := E_2(1 - E_1)(1 + \cos \alpha)$ 
 $r := \left(1 - E_2^2\right)\left(p^2 - q^2\right)^{-1/2}$ 

All higher order modes can then be derived from  $F_0$  and  $F_{-1}$  as

$$F_{n} = \begin{cases} \left(\frac{u_{1}}{u_{0}}\right)^{n} \cdot F_{0} & for \quad n \geq 0\\ \left(\frac{u_{1}}{u_{0}}\right)^{|n|-1} \cdot F_{-1} & for \quad n < 0 \end{cases}$$
[3]

using

$$u_0 := p(p^2 - q^2)^{-1/2}, u_1 := \frac{p}{q}(u_0 - 1)$$

For the  $F_1$  mode, we thus find

$$F_1 \propto q^{-1} \cdot \left(p - \sqrt{p^2 - q^2}\right) \cdot \left(1 - (E_1 - \cos \alpha) \cdot r\right)$$
 [4]

Note that all modes  $F_n$  in Eq. [3] are positive by definition (in contrast to Eq. 21 given in Ref. 27, where the modes are negative for n < 0). Exemplary  $F_1$ ,  $F_0$ , and  $F_{-1}$  steady-state signal levels for tissues and fluids are shown in Figure 1b,c as a function of the flip angle. Taking now into account that the signals are acquired with t > 0 (rather than t = 0), the signal expressions become weighted by the corresponding echo time (TE<sub>1</sub>, TE<sub>0</sub>, and TE<sub>-1</sub>; for details, see Fig. 1a)

$$F_{1.0.-1} \to F_{1.0.-1} \times \exp\left(-\text{TE}_{1.0.-1}/T_2\right)$$
 [5]

For short enough TEs, a  $T_2$  rather than a  $T_2^*$  weighting is introduced to Eqs. [1], [2], and [4]. As reported in (28), the error arising from such an approximation is small.

#### Relaxometry With TESS

As proposed for rapid quantification of  $T_2$  from double echo steady state (DESS) (9), we follow the initial idea of exploiting the dependencies of the modes on relaxation to quantify  $T_1$  and  $T_2$ . To this end, we investigate the following signal ratios (see Fig. 2):

$$s_{T_2}(T_1) := \frac{F_1}{F_0}, \quad s_{T_1}(T_2) := \frac{F_{-1}}{F_0 - F_1}$$
 [6]

Here, the subscript  $T_2$   $(T_1)$  is used to indicate that for the ratio  $s_{T_2}(T_1)$   $(s_{T_1}(T_2))$  the relaxation time  $T_2$   $(T_1)$  is

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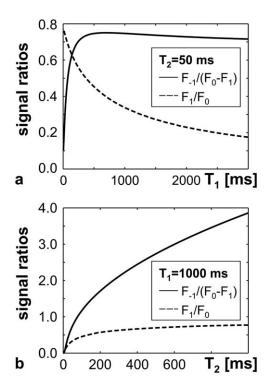


FIG. 2. Simulation of relevant signal ratios  $(s_{T_1}, s_{T_2})$  according to Eq. [6] using a flip angle of  $15^\circ$ , derived from the three base modes, as acquired with TESS (see Fig. 1). (a) Signal ratios as a function of  $T_1$  for a fixed  $T_2$  of 50 ms. Only the  $F_1/F_0$  signal ratio shows a bijective behavior and a good sensitivity for  $T_2$  <<  $T_1$ . (b) Signal ratios as a function of  $T_2$  for a fixed  $T_1$  of 1000 ms. Only the  $F_{-1}/(F_0-F_1)$  signal ratio is sensitive to the limit of  $T_2/T_1 \rightarrow 1$ .

considered to be a bound variable: formally, the signal ratios then depend for any given set of extrinsic parameters (TR,  $\text{TE}_{1,0,-1}$ , and  $\alpha$ ) only on one running variable, namely,  $T_1$  or  $T_2$ .

From this, an easy and fast iterative procedure based on a one-dimensional numerical minimization can be used for the derivation of  $T_1$  and  $T_2$ , as described in the following: A search interval for  $T_1$  and  $T_2$  with a (dummy) upper guess of  $T_{1,\mathrm{u}}=T_{2,\mathrm{u}}=5$  s is defined. The iteration is initialized with a global estimate for the longitudinal relaxation time,  $T_{1,i=0}\in[0,T_{1,\mathrm{u}}]$ , and a golden section search (29) performed to calculate an estimate for the transverse relaxation time,  $T_{2,i+1}$ , based on the measured signal ratio  $s_{T_1}$  (see Eq. [6]),

$$T_{2,i+1} = rg \min \left\{ T_2 \in \left[ 0, T_{2,u} 
ight] : \left| s_{T_1, ext{meas}} - s_{T_{1,i}}(T_2) 
ight| 
ight\} \ \ \ [7]$$

Now, the obtained guess of  $T_{2,i+1}$  is used to yield an improved  $T_{1,i+1}$  estimate based on the signal ratio  $s_{T_2}$  (see Eq. [6]), according to

$$T_{1,i+1} = rg \min \left\{ T_1 \in \left[ 0, T_{1,u} \right] : \left| s_{T_2, \text{meas}} - s_{T_{2,i+1}}(T_1) \right| \right\}$$
[8]

again using a golden section search, but now on  $T_1$ . The improved  $T_{1,i+1}$  estimate can then be used again to cal-

culate an updated  $T_{2,i+2}$  estimate based on Eq. [7], and so on. The iteration stops as soon as the change in both  $T_1$  and  $T_2$  is less than a user defined convergence tolerance, i.e., 0.1 ms. While iterating, the whole search procedure becomes independent of the initial  $T_1$  estimate. In this work, we use a consistent rough global estimate of  $T_1 = 1000$  ms for all simulations, as well as in vitro and in vivo relaxometry. Even with such a rough guess, typically less than 10 iterations are required for convergence of the algorithm.

The specific choice of the signal ratios given in Eq. [6] is motivated as follows: Instead of the basic ratio  $F_{-1}/F_0$  which is used with DESS for  $T_2$  quantification, we investigate the ratio  $F_{-1}/(F_0-F_1)$ , because it shows a stronger sensitivity to  $T_2$  (see Fig. 2b where the good  $T_2$ -sensitivity of this ratio is apparent). For  $T_1$  estimation using TESS, another independent ratio is required and we choose  $F_1/F_0$  because of its strong sensitivity to  $T_1$  (see Fig. 2a). The investigated signal ratios necessarily have to be bijective functions of either  $T_1$  or  $T_2$ . More precisely,  $S_{T_2}(T_1)$  ( $S_{T_1}(T_2)$ ) has to be bijective with respect to  $T_1$  ( $T_2$ ) which holds for our choice. Otherwise, the minimization might run into local rather than global minima.

#### Simulations

Optimal signal-to-noise ratio (SNR) is not only achieved at different flip angles for different modes but also depends on relaxation times (see Fig. 1b,c). We aim at finding optimal flip angle settings for TESS-based relaxometry and analyze the impact of noise on the  $T_1$  and  $T_2$ calculation. To this end, a Monte-Carlo simulation is performed for a range of idealized homogeneous  $T_2/T_1$ probes with signal amplitudes according to Eqs. [1-5]. Optimal flip angle settings are explored based on 100,000  $F_{1,0,-1}$  drawings with mean zero and  $0.001M_0$ standard deviation. The value of the standard deviation directly relates to the amount of noise in the acquired MR images. From the sample of noisy signal amplitudes, distributions of  $T_{1,2}$  with mean  $\langle T_{1,2} \rangle$  and standard deviation  $\Delta T_{1,2}$  are calculated based on Eqs. [6–8]. The SNR for relaxation time mapping using TESS can then be estimated from  $SNR_{T_1,T_2} := \langle T_{1,2} \rangle / \Delta T_{1,2}$ . Relative SNR is evaluated according to  ${\rm rSNR}_{\,T_1,T_2}\,:=\,{\rm SNR}_{\,T_1,T_2}/$ max{SNR<sub>T1</sub>,SNR<sub>T2</sub>} for flip angles ranging from 5° to 40° and for  $T_2/T_1$  ratios ranging from 0.01 to 1.0, using logarithmically spaced  $T_2$  values between 10 ms and 1000 ms, but a fixed  $T_1$  of 1000 ms.

#### Measurements

A contemporary DESS sequence was adapted for acquisition of  $F_1$ ,  $F_0$ , and  $F_{-1}$  within every TR, as displayed in Figure 1a. Generally, although not limited to, the same bandwidth was used for the acquisition of all echoes. To provide enough SNR in the base data for  $T_1$  and  $T_2$  calculation, some averages of the TESS scan were taken as specified below.

In vitro relaxation time mapping with TESS was performed at 1.5 T using manganese-doped spherical phantoms (0 mM, 0.05 mM, 0.125 mM, 0.25 mM, 0.5 mM MnCl<sub>2</sub> in  $\rm H_2O$ ) of about 14 cm in diameter. Longitudinal relaxation was assessed from single-slice IR turbo spin

echo (TSE) experiments with inversion times ranging between 25 ms and 9.6 s. For a single IR-TSE scan with  $2.5 \times 2.5 \text{ mm}^2$  in-plane resolution (128  $\times$  64 matrix) and 5 mm slice thickness, using a turbo factor of 7, a TR of 10 s, a TE of 12 ms, and a bandwidth of 130 Hz/pixel, image acquisition was completed within 1 min 22 s. Transverse relaxation was measured from a contemporary single-echo SE method with a TE ranging from 5 ms up to 3.2 s. One single scan was completed within 3 min 18 s, yielding 2.5  $\times$  5.0 mm<sup>2</sup> in-plane resolution (128  $\times$ 32 matrix) and 5 mm slice thickness, using a turbo factor of 1, a TR of 6 s, and a bandwidth of 592 Hz/pixel. A single-echo rather than a multiecho approach was used to avoid any possible bias in  $T_2$  from stimulated echo contributions (30). TESS imaging was performed in 3D with 4 mm isotropic resolution (64  $\times$  64  $\times$  44 matrix). Scans were performed with a nominal flip angle of 40° and a constant bandwidth of 240 Hz/pixel for all three echoes, yielding an overall TR of 15.7 ms with corresponding  $TE_1 = 3.6$  ms,  $TE_0 = 8.0$  ms, and  $TE_{-1} = 12.4$ ms. Four averages were taken and the 3D TESS scan was completed within 2 min and 58 s.

In vivo human knee scans at 3 T of two healthy volunteers were performed with 3D TESS in axial and sagittal orientation (the slab consisted of 12 slices with 3 mm resolution) using a dedicated 15-channel transmit and receive knee coil (QED) yielding  $0.6 \times 0.6 \text{ mm}^2$  in-plane resolution (256  $\times$  232  $\times$  18 image encoding matrix). Imaging was performed with water-selective excitation pulses  $(1\bar{2}1\text{-binomials})$  of nominal  $15^{\circ}$  flip angle. The bandwidth was set to 230 Hz/pixel, yielding a TR of 20.6 ms, and corresponding echo times  $TE_1 = 6.6$  ms,  $TE_0 = 11.0$  ms, and  $TE_{-1} = 15.4$  ms. Seven averages were taken. The 3D TESS scan was completed within 4 min and 24 s. Reference  $T_1$ values were calculated from six consecutive IR-TSE scans (12 slices, 0.3 mm gap, 2.7 mm slice thickness,  $0.6 \times 0.6$ mm<sup>2</sup> in-plane resolution using a 256  $\times$  232 matrix, turbo factor 8, a TR of 6 s, a TE of 10 ms, and a bandwidth of 227 Hz/pixel) with inversion times ranging between 25 ms and 2.35 s. A single scan took 1 min 44 s to complete. Reference  $T_2$  values were derived based on nine consecutive single-echo SE scans (12 slices, no gap, 3 mm slice thickness, 0.6 × 0.6 mm<sup>2</sup> in-plane resolution using a 256 × 232 matrix, turbo factor 1, a TR of 1210 ms, and a bandwidth of 227 Hz/pixel) with corresponding TEs of 10, 20, 30,..., 90 ms. One single-echo SE scan was completed within 2 min 39 s. For comparison, a multiecho SE approach (nine echoes: starting from 10 ms, and having an echo spacing of 10 ms) was also used with identical settings as the single-echo scan.

#### **RESULTS**

Relaxometry based on TESS is exemplarily illustrated in Figure 3 at 1.5 T for one of the manganese-doped spherical probes (0.25 mM MnCl $_2$  in H $_2$ O) with a nominal  $T_1$  of 456 ms and a nominal  $T_2$  of 48.5 ms, derived by SE techniques (see "Methods" section for details) and regarded in the following as gold standard (31). From a single 3D TESS scan, three different sets of images are acquired (see also Fig. 1a) with formal descriptions as stated in Eqs. [1–5]. From the  $s_{T_1}$  signal ratio (see Eq.

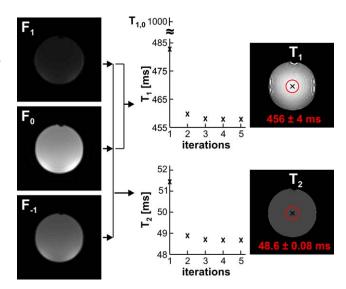


FIG. 3. Illustration of iterative relaxometry calculations from  $F_1$ ,  $F_0$ , and  $F_{-1}$  base images, as presented in Eqs. [7] and [8]. The iteration is exemplarily shown for a pixel in the center of the phantom (black cross). Based on the  $s_{T_1}$  signal ratio (with an initial guess of  $T_{1,0}=1$  s), a guess of  $T_{2,1}\sim51.5$  ms is derived from Eq. [7] using a golden section search. This first guess of  $T_2$  is then used to find an updated estimate of  $T_{1,1}\sim483$  ms based on Eq. [8], again using a golden section search. This procedure is repeated until the change in both  $T_1$  and  $T_2$  falls below a certain threshold (here, 0.1 ms); typically, requiring less than 10 iterations. Relaxation parameters were assessed for a region of interest, as indicated in the relaxation maps by the circle. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

[6]), an estimated  $T_2$  value is found (using Eq. [7], and an initial estimate of  $T_{1,0} = 1$  s). The obtained value is then used to calculate an updated estimate of  $T_1$ , based on  $s_{T_a}$  (see Eqs. [6] and [8]). This procedure is repeated until the iterative change in  $T_1$  and  $T_2$  falls below a certain threshold (here, 0.1 ms). Typically, the iterative procedure converges very quickly (see Fig. 3). The resulting  $T_1$  and  $T_2$  maps are also shown in Figure 3, with the relaxometry results summarized in Table 1 for all probes (evaluated for a circular region of interest, placed in the center of the phantoms as indicated in Fig. 3). Generally, excellent agreement is found between reference singleecho SE and TESS transverse relaxometry data. However, some variation is observed for  $T_1$  using TESS. Interestingly, no spatial variation can be seen for  $T_2$ , whereas  $T_1$ decreases by about 20% toward the rim of the phantom.

The spatial variation of  $T_1$ , as observed in vitro (see Fig. 3), is likely to be due to  $B_1$  field heterogeneities

Table 1 In Vitro Comparison of Single-Echo Spin Echo and TESS Relaxometry Data ( $T_1$  and  $T_2$ ) on Manganese-Doped Aqueous Probes at 1.5 T (for Protocol Details, see "Methods" Section)

	TESS	IR-SE	TESS	SE
[MnCl <sub>2</sub> ]	$T_1$ (ms)	$T_1$ (ms)	$T_2$ (ms)	$T_2$ (ms)
0.000 mM	$3208 \pm 126$	$2995 \pm 2$	$2082 \pm 79$	$1903 \pm 7$
0.050 mM	$1481 \pm 12$	$1485\pm2$	$240 \pm 1.0$	$241 \pm 1.1$
0.125 mM	$871 \pm 10$	$858 \pm 1$	$104.0\pm0.27$	$104.7\pm0.28$
0.250 mM	$456 \pm 4$	$456 \pm 1$	$48.6\pm0.08$	$48.5\pm0.17$
0.500 mM	$263\pm2$	$272\pm1$	$27.0 \pm 0.02$	$27.0 \pm 0.11$

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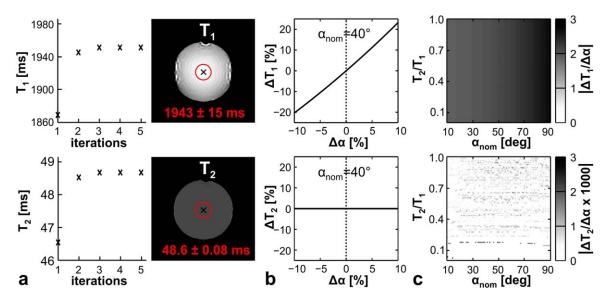


FIG. 4. (a) Recalculation of  $T_1$  and  $T_2$ , as presented in Figure 3, but using only half of the nominal flip angle for the calculation, i.e.,  $20^{\circ}$ , instead of  $40^{\circ}$ . As a result of the dramatic underestimation of the actual flip angle by 50%,  $T_1$  is considerably overestimated (1943 ms, instead of 456 ms), but exactly the same value is derived for  $T_2$ . (b)  $B_1$ -sensitivity of  $T_1$  (top) and  $T_2$  (bottom) simulated for the flip angle and relaxation times of the phantom shown in (a). (c) Simulated  $B_1$ -sensitivity, as shown in (b), over a complete range of flip angles and  $T_2/T_1$  times. Note the different scales in the sensitivity maps for  $T_1$  and  $T_2$ . [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

and thus related to a miscalibration between the nominal  $(\alpha_{\rm nom})$  and the actual flip angle  $(\alpha_{\rm act})$  in the probes. This is confirmed by rerunning the calculation, but this time assuming a halved excitation angle, that is with  $\alpha_{\rm nom} = 20^{\circ}$ , instead of  $\alpha_{\rm nom} = 40^{\circ}$ . As a result, a strongly falsified  $T_1$  is observed (see Fig. 4a); however,  $T_2$  proves to be completely inert. The sensitivity to transmit field errors,  $\Delta \alpha := (\alpha_{act} - \alpha_{nom})/\alpha_{nom}$ , is now systematically evaluated, as exemplarily demonstrated in Figure 4b for  $\alpha_{\mathrm{nom}} = 40^{\circ}$  and  $T_1/T_2/\mathrm{TR} = 456$  ms/48.5 ms/16 ms. The  $B_1$ -sensitivity of  $T_1$ ,  $\Delta T_1/\Delta \alpha$ , is about 2, but  $\Delta T_2/\Delta \alpha$  is barely noticeable, where  $\Delta T_{1,2}$ : =  $(T_{1,2}(\alpha_{\rm act})$  $T_{1,2}(\alpha_{\text{nom}}))/T_{1,2}(\alpha_{\text{nom}})$ . This surprising result is not a remarkable coincidence related to a lucky choice of relaxation and sequence parameters, but can be confirmed over a large range of relaxation parameters and flip angles (Fig. 4c). Thus, TESS offers a unique opportunity for fast, accurate, and unbiased  $T_2$  quantification, whereas  $T_1$  estimates show the expected typical prominent sensitivity of contemporary steady-state methods to  $B_1$  field errors, requiring corrective data. The atypical behavior of TESS-T2 is of special advantage and benefit in vivo.

For high-resolution in vivo TESS relaxometry, SNR is expected to become critical and propagation of noise must be evaluated. Corresponding simulation results are shown in Figure 5. The expected relative SNR for  $T_1$  (Fig. 5a) and  $T_2$  (Fig. 5b) mapping using TESS is strongly affected by the choice of the flip angle (as might already be expected from the corresponding signal curves, see Fig. 1b,c). Generally, a pronounced relative SNR maximum can be observed in both  $T_1$  and  $T_2$  near  $\alpha \sim 15^\circ$  with respect to a  $T_2/T_1 \sim 0.05-0.1$ , that is, for tissues. As a result, optimal SNR for TESS-based relaxometry can be achieved in the low flip angle and low  $T_2/T_1$  limit.

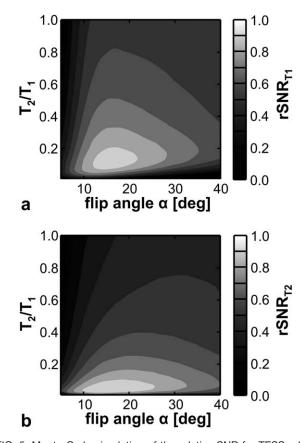


FIG. 5. Monte-Carlo simulation of the relative SNR for TESS relaxometry as a function of the flip angle and relaxation times ( $T_2 = 10$ –1000 ms,  $T_1 = 1000$  ms) for a TR of 16 ms (for simulation details, see "Methods" section). (a) Relative SNR for  $T_1$  mapping (rSNR<sub>T<sub>1</sub></sub>). (b) Relative SNR for  $T_2$  mapping (rSNR<sub>T<sub>2</sub></sub>).

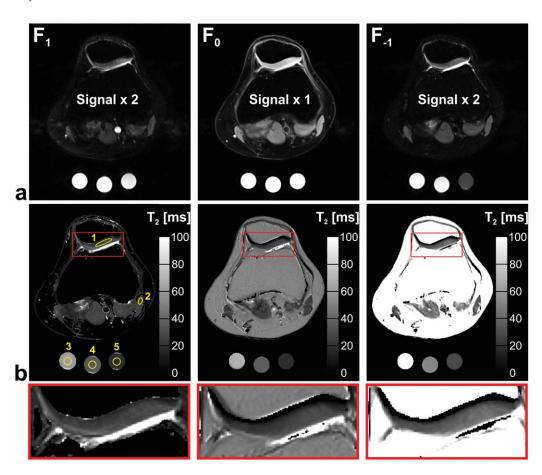


FIG. 6. (a) Exemplary axial TESS base images ( $F_1$ ,  $F_0$ , and  $F_{-1}$ ) of the knee joint at 3 T obtained with water-selective excitation pulses. For better visibility, the signals corresponding to the  $F_1$  and  $F_{-1}$  mode were multiplied by a factor of 2. (b)  $T_2$  values derived either by TESS (left), or by using a single-echo SE approach (middle), or by using a multiecho SE method (right). The manganese-doped test tubes (0.125 mM, 0.25 mM, and 0.5 mM) serve as internal controls. For selected regions of interest (indicated by the yellow numbers),  $T_2$  values are summarized in Table 2. Again, excellent correspondence is observed in  $T_2$  between TESS and single-echo SE data (see also Table 1), whereas the multiecho SE approach shows a clear overestimation of  $T_2$  from stimulated echo contributions.

High-resolution in vivo 3D TESS relaxometry is demonstrated in the knee joint at 3 T in axial (Fig. 6) and sagittal (Fig. 7) slice orientation using optimal flip angles for tissues. As an internal control, three small test tubes containing 0.125 mM, 0.25 mM, and 0.5 mM MnCl<sub>2</sub> were placed adjacent to the knee for the axial scan.  $B_1$ -insensitive TESS- $T_2$  quantification is illustrated in Figure 6 and compared with reference  $T_2$  maps calculated based on single-echo and multiecho SE experiments. Transverse relaxation time values are assessed for selected regions of interest (for definition of regions of interest, see Fig. 6) with corresponding results summarized in Table 2. Overall, excellent correspondence between TESS and single-echo SE transverse relaxometry data is found, whereas  $T_2$  values, derived from multiecho SE data, show a pronounced overestimation of about 30-40% for cartilage, muscle, and for the internal controls due to stimulated echo contributions (from imperfect refocusing pulses and thus due to  $B_1$  errors).

The distinct  $B_1$ -insensitivity of TESS- $T_2$  becomes especially apparent in the sagittal  $T_2$  scan (Fig. 7b) where no variation in muscle- $T_2$  can be observed over the complete field of view. This is quite contrary to the corre-

sponding TESS- $T_1$  map (Fig. 7c, right). It appears more inhomogeneous and shows clearly visible variations in muscle- $T_1$  over the field of view originating from  $B_1$  heterogeneities. Comparison of TESS- $T_1$  with a reference measurement based on six consecutive IR-TSE scans (Fig. 7c, left) reveals that the  $T_1$  map calculated with TESS is flawed as expected. However, good correspondence can be observed in regions where  $B_1$  field errors are not that prominent, for instance in the patella: There, TESS relaxometry yields a  $T_1$  of  $869 \pm 45$  ms for the region of interest indicated by the arrow in Figure 7c compared with a  $T_1$  of  $839 \pm 40$  ms obtained with IR-TSE.

#### DISCUSSION

A variety of SSFP methods have been proposed thus far and are known in the literature for fast relaxometry, but all of them are sensitive to  $B_1$  and show some more or less pronounced mixed  $T_2/T_1$ -sensitivity that can be mitigated by companion scans. As a result, without the use of additional corrective data, current SSFP techniques fail

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to deliver accurate, reliable, and stable quantification results.

In this work, we report on a similar approach, as proposed for rapid  $T_2$  quantification of cartilage using DESS imaging (28,32). In the limit of  $\alpha \sim 90^{\circ}$ , the ratio between the  $F_{-1}$  and  $F_0$  signal, as acquired with DESS, becomes independent of  $T_1$ , allowing accurate quantification of  $T_2$ (9). Signal-to-noise, however, is especially poor in this limit, requiring considerably lower flip angles in practice and, hence, leading to a systematic  $T_1$ -related bias in the estimated  $T_2$  values using DESS. In principle, we solve this issue by acquiring a third independent mode, namely,  $F_1$ . Based on two independent signal ratios, the interacting  $T_1$ - and  $T_2$ -sensitivity can be tackled by a simple and fast ping-pong approach, using a golden section search, until the calculated signal ratios converge to the actual measured ones. As a result, TESS offers the possibility of acquiring both  $T_1$  and  $T_2$  within one single scan and without the confounding influence of either  $T_1$ on  $T_2$  or  $T_2$  on  $T_1$  (as observed with any other steadystate method). Moreover, from the application of crusher gradients within every TR, TESS is not affected by static field inhomogeneities. However, some motion sensitivity must be taken into account. Therefore, proper fixation seemed mandatory and was carefully conducted before every measurement. Diffusion effects are expected to be negligible for tissues, but reduce the signal of fluids for high-resolution scans (33).

As with any other fast quantitative SSFP imaging technique, TESS-based relaxometry is expected to be affected by  $B_1$  field heterogeneities, and the impact of such transmit field calibration errors on  $T_1$  and  $T_2$  quantification

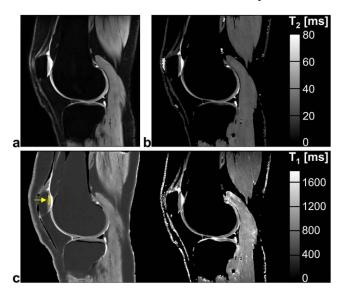


FIG. 7. (a) Morphological TESS image, calculated from a weighted combination of the  $F_0$  and  $F_{-1}$  signal ( $F_0+3\times F_{-1}$ ) to accentuate the contrast between synovial fluid and cartilage, similar to DESS. (b)  $B_1$ -insensitive sagittal  $T_2$  map calculated from a single TESS scan. (c) Sagittal TESS- $T_1$  map (right) is compared with a reference  $T_1$  measurement derived from 2D multislice IR-TSE scans. TESS- $T_1$  is clearly affected by  $B_1$ ; however, good correspondence to the IR-TSE technique can be observed for instance in the patella (region of interest indicated by the arrow). [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Table 2 In Vivo Comparison of Spin Echo and TESS  $T_2$  Relaxometry Data in the Knee Joint at 3 T (for Protocol Details, see "Methods" Section)

	•				
Ī		TESS	SE <sup>a</sup>	SE <sup>b</sup>	SE <sup>c</sup>
			Single-echo	Multiecho	Multiecho
	Tissue/[MnCl <sub>2</sub> ]	$T_2$ (ms)	$T_2$ (ms)	$T_2$ (ms)	$T_2$ (ms)
	Cartilage <sup>d</sup>	$27.3 \pm 3.2$	$26.5 \pm 3.2$	$32.9 \pm 4.5$	$40.4 \pm 5.2$
	Muscle <sup>e</sup>	$26.3 \pm 0.6$	$24.6 \pm 1.1$	$31.1\pm4.5$	$37.6 \pm 4.9$
	0.125 mM <sup>f</sup>	$64.2 \pm 0.9$	$69.1 \pm 0.6$	$84.0 \pm 0.6$	$102.6\pm0.7$
	0.250 mM <sup>g</sup>	$34.9 \pm 0.3$	$36.6 \pm 0.1$	$44.4\pm0.2$	$53.0 \pm 0.3$
	0.500 mM <sup>h</sup>	$18.0 \pm 0.2$	$18.7 \pm 0.1$	$23.0 \pm 0.1$	$28.9 \pm 0.1$

 $<sup>^{\</sup>mathrm{a}}T_{2}$  value derived based on nine single-echo SE scans using a nonlinear least-squares fit.

was analyzed in Figure 4. Surprisingly,  $T_2$  relaxometry with TESS revealed to be independent of  $B_1$ , whereas  $T_1$ quantification showed the expected pronounced  $B_1$ related estimation errors. This extraordinary feature is not only of special interest for high to ultra-high field  $T_2$ relaxometry where prominent  $B_1$  variations can be expected and applicability of SE techniques might be limited due to SAR constraints but also provides accurate quantification results in combination with spectralspatial excitation pulses that typically entail flip angle calibration errors in the presence of  $B_0$  heterogeneities. Spectral-spatial excitation, as demonstrated in Figures 6 and 7 for TESS, is especially beneficial for musculoskeletal imaging where fat is often found adjacent or interspersed within the tissues of interest. Fat suppression may thus not only enhance diagnostic information (34) but also eliminates the chemical shift artefact and thereby allows readout of the SSFP signal modes with a reduced receiver bandwidth which in turn results in an overall increased SNR.

## CONCLUSIONS

In contrast to all other existing SSFP quantification techniques, TESS offers rapid  $T_1$  and  $T_2$  estimation within one single scan. Moreover, quantification of  $T_2$  with TESS is markedly insensitive to  $B_1$ . As a result, the new proposed method is of high interest for fast and reliable relaxometry in the clinical routine, especially for rapid and bias-free  $T_2$  imaging of the musculoskeletal system at high to ultra-high field strength.

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 $<sup>^{</sup>b}T_{2}$  value derived based on a multiecho SE scan (nine echoes), using a nonlinear least-squares fit. For the fit, the first echo was discarded to mitigate stimulated echo contributions.

<sup>&</sup>lt;sup>c</sup>T<sub>2</sub> value derived based on a multiecho SE scan (nine echoes), using a nonlinear least-squares fit.

<sup>&</sup>lt;sup>d</sup>For definition, see Figure 6: region of interest 1.

<sup>&</sup>lt;sup>e</sup>For definition, see Figure 6: region of interest 2.

<sup>&</sup>lt;sup>f</sup>For definition, see Figure 6: region of interest 3.

<sup>&</sup>lt;sup>g</sup>For definition, see Figure 6: region of interest 4.

<sup>&</sup>lt;sup>h</sup>For definition, see Figure 6: region of interest 5.

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